DATA NOTE

The old and grumpy biogas metagenome

Andreas Bremges^{1,2*}, Irena Maus¹, Alfred Pühler¹, Andreas Schlüter^{1†} and Alexander Sczyrba^{1,2†}

abremges@cebitec.uni-bielefeld.de

¹Center for Biotechnology,
Bielefeld University, Germany
Full list of author information is
available at the end of the article

† Equal contributor

Abstract

Background: a presentation of the interest or relevance of these data for the broader community

Findings: a very brief preview of the data type(s) produced, the methods used, and information relevant to data validation

Conclusions: a short summary of the potential uses of these data and implications for the field

Keywords: Biogas; Metagenome; Sequencing; Assembly; Annotation

Data description

Background

Biogas important energy source. Clean and awesome. Number of biogas plants in Germany/Europe/worldwide. Key is process optimization, not building more and more plants. Little is known about the microbial community responsible for everything.

section not final!

Here, we report the first deeply sequenced metagenome (and metatranscriptome?) of an agricultural production-scale biogas plant on the Illumina platform [1]. We sequenced $27.3 \times$ and $19.3 \times$ deeper than previous studies relying on 454 [2] or SOLiD [3] sequencing. These data will enable a deeper exploration of the biogas-producting microbial community, a key step towards process optimization.

phrasing

Digester management

The biogas plant, located in North Rhine Westphalia, Germany, features a mesophilic continuous wet fermentation technology and was designed for a capacity of $537\,kW_{el}$ combined heat and power (CHP) generation. The process comprises two digesters: a primary digester, where biogas and methane is produced, and a secondary digester, insignificant for production. The primary digester is fed hourly with a mixture of maize silage and liquid pig manure. The biogas and methane yields at the time of sampling were at 810.5 and 417.8 liters per kg organic dry matter $(l/kg\,oDM)$, respectively. After a theoretical retention time of 55 days, the digestate is stored in the closed, non-heated final storage tank. Further metadata are summarized in Table 1.

Sampling and DNA isolation

Samples from the primary digester of the aforementioned biogas plant were taken in November 2010. Prior to sampling process, approximately $15\,L$ of fermenter substrate were discarded before aliquots of $1\,L$ were transferred into clean gastight sampling vessels and transported directly to the laboratory.

^{*}Correspondence:

Bremges et al. Page 2 of 7

> A 20 g aliquot of the fermentation sample was used for total community DNA preparation as described previously [4].

Metagenomic sequencing

we need actual numbers somewhere In total, we sequenced four different metagenome shotgun libraries with different insert sizes, resulting in over 23 gigabases scattered across 144 million reads. On Illumina's Genome Analyzer IIx system, we sequenced two libraries with an average insert size of 250 nt and 450 nt, respectively, applying the Paired-End DNA Sample Preparation Kit (Illumina Inc.) as described by the manufacturer. On Illumina's MiSeq system, we sequenced two further libraries with an average insert size of 190 nt and 690 nt, respectively, applying the Nextera DNA Sample Preparation Kit (Illumina Inc.) as described by the manufacturer. In all cases, we sequenced for approximately 300 cycles, applying the $2 \times 150 \, nt$ paired-end protocol.

Eher kein 2x250: MiSeq v2 öffentlich 10/2012, Alex und ich hatten die MiSeq Daten in 07/2012!

Quality control with FastQC, version 0.11.2. Adapter contamination in Lane 7.

We used Trimmomatic [5], version 0.32, for adapter removal and moderate quality trimming. After adapter clipping, using Trimmomatic's Truseq2-PE and Nextera-

PE templates, we removed leading and trailing low quality bases (below quality 3).

Then, we performed an adaptive quality trimming, balancing read length against error rate (target read length of 100 bases, strictness of 0.2, thus favouring read length over error sensitivity). Finally, reads shorter than 36 bases were discarded.

Sequence quality control

cite if needed

mention at

all?

section not final!

Metagenome assembly and quality assessment

Either another table or brief summary post-QC.

2.3.1

We assembled the metagenome with Ray Meta [6], version 2.3.0, using a k-mer size of 31 and a minimum contig length of 1,000 bp. This resulted in a total assembly size of approximately 217 megabases in 55,563 contigs, with an N50 value of 8,137 bp. Table 2 summarizes our results.

2.2.4

Run LAP or ALE

We aligned the post-QC sequencing reads to the assembled contigs with bowtie2 [7], version 2.2.1, and used samtools [8], version 1.1, to convert SAM to BAM and thereafter sort the alignment file.

Gene prediction and annotation

We then used MetaProdigal [9], version 2.6.1, to predict 239,412 protein-coding genes on the assembled contigs. Table 3 also includes these results.

We blasted all predicted genes against the KEGG database [10], release 72.0, using Protein-Protein BLAST [11], version 2.2.29+. Of the 239, 412 predicted genes, 230, 354 were found in the KEGG database. We determined the KEGG Orthology (KO) for each gene by mapping the top-scoring BLAST hit to its KO.

Coverage analysis

0.6.1p2

We counted aligned reads in predicted genes with HTSeq [12], version 0.6.1p1. Figure 1 relates the metagenome and the metatranscriptome.

work in progress and up for discussion

Bremges et al. Page 3 of 7

Availability

Data accession

The datasets supporting the results of this article are available in the [repository name] repository, [unique persistent identifier and hyperlink to datasets in http://format].

Reproducibility

The complete workflow is organized in a single GNU Makefile and available on GitHub [13]. Starting from the raw read files, available from SRA and/or GigaDB, all data and results can be reproduced by a simple invocation of *make*. Excluding KEGG analysis, which relies on a commercial license of the KEGG database, all steps are performed using free and open-source software.

phrasing

Requirements

The assembly of the metagenome data sets with Ray Meta requires 70 GB of RAM. Using only 40 cores, it completed within 12 hours wall-clock time. Annotation with blast took, of course, forever.

Discussion

Potential use cases. Metagenomic (and metatranscriptomic?) profiling of the biogas-producing microbial community. Identification of metaproteomic data out there (cite Vera, in preparation).

Ultimate goal: process optimization by biological insights.

final run!

log run-

time and

memory in

section not final!

Bremges et al. Page 4 of 7

Competing interests

The authors declare that they have no competing interests.

Author's contributions

AB conveived and performed all bioinformatic analyses and wrote the paper. IM investigated all metadata and drafted part of the data description. AScz conveived of many of the analyses and revised the paper. ASch and AScz jointly directed the project. All authors read and approved the final manuscript.

Acknowledgements AB and IM are supp

Funding

Andreas S.

Stadtwerke

AB and IM are supported by a fellowship from the CLIB Graduate Cluster Industrial Biotechnology.

Author details

 1 Center for Biotechnology, Bielefeld University, Germany. 2 Faculty of Technology, Bielefeld University, Germany.

References

- Bremges, A., Maus, I., Pühler, A., Schlüter, A., Sczyrba, A.: Name of the GigaScience repository (2014). [DOI:foo/bar]
- Jaenicke, S., Ander, C., Bekel, T., Bisdorf, R., Dröge, M., Gartemann, K.H., Jünemann, S., Kaiser, O., Krause, L., Tille, F., Zakrzewski, M., Pühler, A., Schlüter, A., Goesmann, A.: Comparative and joint analysis of two metagenomic datasets from a biogas fermenter obtained by 454-pyrosequencing. PLoS ONE 6(1), 14519 (2011). [PubMed Central:PMC3027613] [DOI:10.1371/journal.pone.0014519] [PubMed:21297863]
- Wirth, R., Kovács, E., Maróti, G., Bagi, Z., Rákhely, G., Kovács, K.L.: Characterization of a biogas-producing microbial community by short-read next generation DNA sequencing. Biotechnol Biofuels 5, 41 (2012). [PubMed Central:PMC3395570] [DOI:10.1186/1754-6834-5-41] [PubMed:22673110]
- Schlüter, A., Bekel, T., Diaz, N.N., Dondrup, M., Eichenlaub, R., Gartemann, K.H., Krahn, I., Krause, L., Krömeke, H., Kruse, O., Mussgnug, J.H., Neuweger, H., Niehaus, K., Pühler, A., Runte, K.J., Szczepanowski, R., Tauch, A., Tilker, A., Viehöver, P., Goesmann, A.: The metagenome of a biogas-producing microbial community of a production-scale biogas plant fermenter analysed by the 454-pyrosequencing technology. J. Biotechnol. 136(1-2), 77–90 (2008). [DOI:10.1016/j.jbiotec.2008.05.008] [PubMed:18597880]
- Bolger, A.M., Lohse, M., Usadel, B.: Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30(15), 2114–2120 (2014). [PubMed Central:PMC4103590] [DOI:10.1093/bioinformatics/btu170] [PubMed:24695404]
- Boisvert, S., Raymond, F., Godzaridis, E., Laviolette, F., Corbeil, J.: Ray Meta: scalable de novo metagenome assembly and profiling. Genome Biol. 13(12), 122 (2012). [PubMed Central:PMC4056372] [DOI:10.1186/gb-2012-13-12-r122] [PubMed:23259615]
- Langmead, B., Salzberg, S.L.: Fast gapped-read alignment with Bowtie 2. Nat. Methods 9(4), 357–359 (2012).
 [PubMed Central:PMC3322381] [DOI:10.1038/nmeth.1923] [PubMed:22388286]
- Li, H., Handsaker, B., Wysoker, A., Fennell, T., Ruan, J., Homer, N., Marth, G., Abecasis, G., Durbin, R.: The Sequence Alignment/Map format and SAMtools. Bioinformatics 25(16), 2078–2079 (2009). [PubMed Central:PMC2723002] [DOI:10.1093/bioinformatics/btp352] [PubMed:19505943]
- Hyatt, D., LoCascio, P.F., Hauser, L.J., Uberbacher, E.C.: Gene and translation initiation site prediction in metagenomic sequences. Bioinformatics 28(17), 2223–2230 (2012). [DOI:10.1093/bioinformatics/bts429] [PubMed:22796954]
- Kanehisa, M., Goto, S., Sato, Y., Kawashima, M., Furumichi, M., Tanabe, M.: Data, information, knowledge and principle: back to metabolism in KEGG. Nucleic Acids Res. 42(Database issue), 199–205 (2014). [PubMed Central:PMC3965122] [DOI:10.1093/nar/gkt1076] [PubMed:24214961]
- Camacho, C., Coulouris, G., Avagyan, V., Ma, N., Papadopoulos, J., Bealer, K., Madden, T.L.: BLAST+: architecture and applications. BMC Bioinformatics 10, 421 (2009). [PubMed Central:PMC2803857] [DOI:10.1186/1471-2105-10-421] [PubMed:20003500]
- 12. Anders, S., Pyl, P.T., Huber, W.: HTSeq-a Python framework to work with high-throughput sequencing data. Bioinformatics (2014). [DOI:10.1093/bioinformatics/btu638] [PubMed:25260700]
- 13. Bremges, A.: GitHub (2014). [https://github.com/abremges/2014-biogas]

Less author names, remove PMC?

Bremges et al. Page 5 of 7

Figures

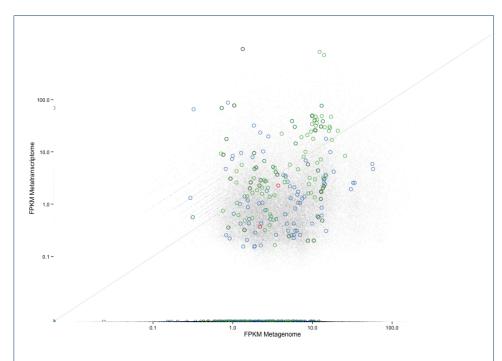


Figure 1 Relating the metagenome and metatranscriptome. Low-quality image as a placeholder. FPKM values, work in progress! Highlighted are genes involved in methanogenesis, color coded by pathway type: CO2 to methane [MD:M00567] green, methanol to methane [MD:M00356] red, and acetate to methane [MD:M00357] blue.

Bremges et al. Page 6 of 7

Tables

 Table 1 Characteristics of the studied biogas plant. Primary digester, sampling date: Nov 15, 2010.

Process parameter	Sample
Net volume	$2041 m^3$
Dimensions	6.4m high, diameter of $21m$
Electrical capacity	$537 kW_{el}$
рН	7.83
Temperature	40 °C
Conductivity	22.10 mS/cm
Volative organic acids (VOA)	5327 mg/l
Total inorganic carbon (TIC)	14397 mg/l
VOA/TIC	0.37
Ammoniacal nitrogen	2.93 g/l
Acetic acid	863mg/l
Propionic acid	76mg/l
Fed substrates	72% maize silage, $28%$ pig manure
Organic load	$4.0 kg oDM m^{-3} d^{-1}$
Biogas yield	810.5l/kgoDM
Methane yield	417.8 l/kg oDM

TIC only calculated, might be a few digits off

Table 2 Metagenome assembly. Some assembly statistics, minimum contig size of $1,000\,bp$.

Assembly metric	Our assembly
Total size	216,554,757 bp
Number of contigs	55,563
N50 value	8,137 bp
Largest contig	319,083 bp

Table 3 Gene prediction and annotation. Whatever.

Annotation metric	Our annotaion				
Predicted genes	239,412				
of these, full-length	160,124 (66.9%)				
KEGG stuff	?				

Bremges et al. Page 7 of 7

_		••	-
10	പറ	lis	٠
	uυ	113	·L

section not final!						
phrasing						
we need actual numbers somewhere						
Eher kein 2x250: MiSeq v2 öffentlich 10/2012, Alex und ich hatten die	e I	Mi	S_{ϵ}	eq	l	
Daten in 07/2012!						
cite if needed						
mention at all?						
section not final!						
2.3.1						
2.2.4						
Run LAP or ALE						
$0.6.1 \mathrm{p}2$						
work in progress and up for discussion						
phrasing						
log runtime and memory in final run!						
section not final!						
Funding Andreas S, Stadtwerke?						
Less author names, remove PMC?						
TIC only calculated, might be a few digits off						