

An R interface to the Ensembl REST API

Tim Yates

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1 Introduction

This package uses the Ensembl REST API¹ (currently in beta) to extract data from Ensembl into R.

As the REST API is in Beta, this package should also be considered to be in flux and functions/-parameters/etc are subject to change as things get finalized.

It could also do with your help. If you find a problem, something you think could be better, or a better way of doing things, please consider visiting the GitHub project at <https://github.com/acbb/EnsemblRest> and posting an issue or a Pull Request. Thanks!

2 Available Methods

To begin with (assuming you have installed this package), you need to load it into your R session:

```
> library( EnsemblRest )
```

The following subsections then list the methods available to you.

¹<http://beta.rest.ensembl.org/>

2.1 Information

2.1.1 isAlive

Firstly, we can check to see that the REST API is accepting calls:

```
> isAlive()  
[1] TRUE
```

2.1.2 infoSpecies

To get a list of available species on the server, you can use the `infoSpecies` call².

```
> infoSpecies()[1:3] # Just the first 3  
[[1]]  
name      : saccharomyces_cerevisiae  
aliases   : 4932, saccer, saccharomyces cerevisiae (baker's yeast), baker's yeast, scer, sacchar  
groups    : core, otherfeatures, variation  
release   : 69  
  
[[2]]  
name      : ciona_savignyi  
aliases   : ciosav, 51511, ciona savignyi, csavignyi, c.savignyi, csav, sea squirt ciona savigny  
groups    : core, otherfeatures  
release   : 69  
  
[[3]]  
name      : myotis_lucifugus  
aliases   : little brown bat, mlucifugus, myoluc, mluc, 59463, myotis lucifugus, myotis_lucifugu  
groups    : core, otherfeatures  
release   : 69
```

2.1.3 infoAssembly

The `infoAssembly` call³ returns information about the currently available assemblies in the given species.

```
> infoAssembly( 'human' )  
assembly_name      : GRCh37.p8  
assembly_date      : 2009-02  
coord_system_versions : , GRCh37, NCBI36, NCBI34, NCBI35  
schema_build       : 69_37  
genebuild_start_date : 2010-07-Ensembl  
genebuild_initial_release_date : 2011-04  
genebuild_last_geneset_update : 2012-10  
genebuild_method    : full_genebuild  
top_level_seq_region_names : 1, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 2, 20, 21, 22, 3,
```

²<http://beta.rest.ensembl.org/documentation/info/species>

³http://beta.rest.ensembl.org/documentation/info/assembly_info

2.1.4 assemblyDetails

The `assemblyDetails` call⁴ returns information about one of these assemblies.

```
> assemblyDetails( 'X', 'human' )
is_chromosome      : TRUE
length            : 155270560
assembly_exception_type : REF
coordinate_system  : chromosome
```

2.1.5 infoComparas

The `infoComparas` call⁵ lists the available comparative genomics databases.

```
> infoComparas()
multi
"69"
```

2.1.6 infoData

The `infoData` call⁶ shows the data releases available to the REST service

```
> infoData()
[1] "69"
```

2.1.7 infoRest

`infoRest` shows the current version⁷ of the REST service

```
> infoRest()
[1] "1.1.1"
```

2.1.8 infoSoftware

And finally in the `info` section, `infoSoftware` shows⁸ the current version of the Ensembl API.

```
> infoSoftware()
[1] 69
```

⁴http://beta.rest.ensembl.org/documentation/info/assembly_stats

⁵<http://beta.rest.ensembl.org/documentation/info/comparas>

⁶<http://beta.rest.ensembl.org/documentation/info/data>

⁷<http://beta.rest.ensembl.org/documentation/info/rest>

⁸<http://beta.rest.ensembl.org/documentation/info/software>

2.2 Comparative Genomics

2.2.1 geneTree

This method⁹ fetches the gene tree in New Hampshire format for a given Ensembl gene tree identifier.

```
> geneTree( 'ENSGT00390000003602' )  
[1] "(((((((ENSXMAP00000006983:0.2452,ENSORLP00000004773:0.3910):0.1067,ENSONIP00000006940:0.2
```

it is also possible to specify the NH format you require (ie: for full format):

```
> geneTree( 'ENSGT00390000003602', nh_format='full' )  
[1] "(((((((ENSXMAP00000006983:0.2452,ENSORLP00000004773:0.3910):0.1067,ENSONIP00000006940:0.2
```

2.2.2 homologyById

When given an Ensembl Gene ID, returns the homology¹⁰ information for it.

```
> hResponse = homologyById( 'ENSG00000170037' )  
> hResponse # The response object  
id : ENSG00000170037  
containing 50 homologies  
> hResponse$homologies[1:2] # Just the top 2 homologies to save room  
[[1]]  
dn_ds : 0.29412  
type : ortholog_one2one  
subtype : Homininae  
source :  
id : ENSG00000170037  
species : homo_sapiens  
protein_id : ENSP00000369614  
perc_pos : 99  
perc_id : 99  
cigar_line : 925M  
align_seq : MATSADSPSSPLGAEDLLSDSSEPPGLNQVSSEVTSQLYASRLSRQAEATARAQLYLPSTSPPEGLDGFAQELSRSLSV  
  
target :  
id : ENSPTRG00000008719  
species : pan_troglodytes  
protein_id : ENSPTRP00000014861  
perc_pos : 100  
perc_id : 100  
cigar_line : 850MD74M  
align_seq : MATSADSPSSPLGAEDLLSDSSEPPGLNQVSSEVTSQLYASRLSRQAEATARAQLYLPSTSPPEGLDGLAQELSRSLSV  
  
[[2]]  
dn_ds :  
type : ortholog_one2one  
subtype : Homininae  
source :  
id : ENSG00000170037
```

⁹<http://beta.rest.ensembl.org/documentation/info/rest>

¹⁰http://beta.rest.ensembl.org/documentation/info/homology_ensemblgene

```

species      : homo_sapiens
protein_id   : ENSP00000369614
perc_pos     : 99
perc_id      : 98
cigar_line   : 925M
align_seq    : MATSADSPSSPLGAEDLLSDSSEPPGLNQVSSEVTSQLYASRLSRQAEATARAQLYLPSTSPPEGLDGFAGELSRSLSV

target :
id       : ENSGGOG00000006451
species  : gorilla_gorilla
protein_id : ENSGGOP00000006314
perc_pos  : 99
perc_id   : 98
cigar_line : 925M
align_seq : MATSADSPSSPLGAEDLLSDSSEPPGLNQVSSEVTSQLYASRLSRQAEATARAQLYLPSTSPPEGLDGLAQELSRSLSV

```

2.2.3 homologyBySymbol

You can also retrieve homology information¹¹ given a symbol and a species;

```

> hResponse = homologyBySymbol( 'BRCA2', 'human' )
> hResponse                                     # The response object

id : ENSG00000139618
containing 55 homologies

> hResponse$homologies[1:2] # Again, just the top 2 homologies to save room

[[1]]
dn_ds      : 0.29371
type       : ortholog_one2one
subtype    : Homininae
source     :
id          : ENSG00000139618
species     : homo_sapiens
protein_id  : ENSP00000369497
perc_pos    : 99
perc_id     : 99
cigar_line  : 3418M
align_seq   : MPIGSKERPTFFEIFKTRCNKADLGPISLWFEELSSEAPPYNSEPAEESSEHKNNNYEPNLFKTPQRKPSYNQLASTPIIF

target :
id       : ENSPTRG00000005766
species  : pan_troglodytes
protein_id : ENSPTRP00000009812
perc_pos  : 99
perc_id   : 99
cigar_line : 3418M
align_seq : MPIGSKERPTFFEIFKTRCNKADLGPISLWFEELSSEAPPYNSEPAEESSEHKNNNYEPNLFKTPQRKPSYNQLASTPIIF

[[2]]
dn_ds      : 0.61742
type       : ortholog_one2one
subtype    : Hominidae

```

¹¹http://beta.rest.ensembl.org/documentation/info/homology_symbol

```
source :
id      : ENSG00000139618
species : homo_sapiens
protein_id : ENSP00000369497
perc_pos : 98
perc_id  : 96
cigar_line : 3418M
align_seq : MPIGSKERPTFFEIFKTRCNKADLGPISLWFEELSSEAPPYNSEPAEESSEHKNNNYEPNLFKTPQRKPSYNQLASTPIIF

target :
id      : ENSPPYG00000005264
species : pongo_abelii
protein_id : ENSPPYP00000005997
perc_pos : 98
perc_id  : 96
cigar_line : 975MD302MD442MD1696M
align_seq : MPVGSKERPTFFEIFKTRCNKADLGPISLHWFEELSSEAPPYNSEPAEESSEHKNNNYEPNLFKTPQRKPSYNQLASTPIIF
```

2.3 Cross References

Cross references are links to other data about the object of interest. It should be noted that these other data hold different fields and datatypes, so the results are returned in a data.frame containing a superset of column names, with non-applicable columns for a given result filled with <NA>.

As a single object may have multiple synonyms, this will cause the object to exist in multiple rows, one for each synonym.

2.3.1 xrefsById

Firstly we can get all external references¹² for a given Ensembl ID:

```
> xrefsById( 'ENSG00000170037' )
```

	display_id	primary_id	version
1	OTTHUMG00000172932	OTTHUMG00000172932	2
2	ENSG00000170037	ENSG00000170037	0
3	CNTROB	116840	0
4	CNTROB	29616	0
5	CNTROB	29616	0
6	CENTROSOMAL BRCA2-INTERACTING PROT [*611425]	611425	0
7	Hs.348012	Hs.348012	0
8	Hs.732863	Hs.732863	0
9	CNTROB	CNTROB	0
10	CNTROB	116840	0

	dbname	info_type	info_text	db_display_name
1	OTTG	NONE		Havana gene
2	ArrayExpress	DIRECT		ArrayExpress
3	EntrezGene	DEPENDENT		EntrezGene
4	HGNC	DIRECT	Generated via ccds	HGNC Symbol
5	HGNC	DIRECT	Generated via ccds	HGNC Symbol
6	MIM_GENE	DEPENDENT		MIM gene
7	UniGene	SEQUENCE_MATCH		UniGene
8	UniGene	SEQUENCE_MATCH		UniGene
9	Uniprot_genename	DEPENDENT		UniProtKB Gene Name
10	WikiGene	DEPENDENT		WikiGene

	description
1	<NA>
2	
3	centrobin, centrosomal BRCA2 interacting protein
4	centrobin, centrosomal BRCA2 interacting protein
5	centrobin, centrosomal BRCA2 interacting protein
6	CENTROSOMAL BRCA2-INTERACTING PROTEIN; CNTROB
7	Centrobin, centrosomal BRCA2 interacting protein
8	Transcribed locus, moderately similar to NP_444279.2 centrobin isoform alpha [Homo sapiens]
9	
10	centrobin, centrosomal BRCA2 interacting protein

	synonyms	ensembl_identity	ensembl_start	xref_start	xref_end	ensembl_end
1	<NA>	NA	NA	NA	NA	NA
2	<NA>	NA	NA	NA	NA	NA
3	LIP8	NA	NA	NA	NA	NA
4	LIP8	NA	NA	NA	NA	NA
5	PP1221	NA	NA	NA	NA	NA
6	<NA>	NA	NA	NA	NA	NA

¹²http://beta.rest.ensembl.org/documentation/info/xref_id

7	<NA>	99	1	32	3794	3769
8	<NA>	99	1	6	752	752
9	LIP8	NA	NA	NA	NA	NA
10	<NA>	NA	NA	NA	NA	NA

	score	cigar_line	xref_identity
1	NA	<NA>	NA
2	NA	<NA>	NA
3	NA	<NA>	NA
4	NA	<NA>	NA
5	NA	<NA>	NA
6	NA	<NA>	NA
7	18783	376M6D3387M	90
8	3677	289M6D406M1I51M	99
9	NA	<NA>	NA
10	NA	<NA>	NA

2.3.2 xrefsByName

Or, we can look for an external reference primary accession¹³ (given a species):

```
> xrefsByName( 'NM_004333', 'human' )
      display_id primary_id version
1 NM_004333.4   NM_004333      4
                                     description
1 Homo sapiens v-raf murine sarcoma viral oncogene homolog B1 (BRAF), mRNA.
      dbname info_type      info_text db_display_name
1 RefSeq_mRNA   DIRECT Generated via ccds   RefSeq mRNA
```

2.3.3 xrefsBySymbol

And we can finally look up all Ensembl objects referenced by an external symbol for a given species¹⁴:

```
> xrefsBySymbol( 'BRAF', 'human' )
      type      id
1      gene ENSG00000157764
2 transcript ENST00000288602
```

¹³http://beta.rest.ensembl.org/documentation/info/xref_name

¹⁴http://beta.rest.ensembl.org/documentation/info/xref_external

2.4 Features

2.4.1 featuresByRegion

We can also look for features along a given range¹⁵ (by default this will just look for genes):

```
> featuresByRegion( '7:140424943-140624564', 'human' )
GRanges with 2 ranges and 6 elementMetadata cols:
      seqnames      ranges strand |      ID
      <Rle>        <IRanges> <Rle> |      <factor>
[1]          7 [140424943, 140624564] - | ENSG00000157764
[2]          7 [140583872, 140583978] + | ENSG00000207040
      logic_name feature_type external_name
      <factor>      <factor>      <factor>
[1] ensembl_havana_gene      gene      BRAF
[2]              ncrna      gene      U6
      description
      <factor>
[1] v-raf murine sarcoma viral oncogene homolog B1 [Source:HGNC Symbol;Acc:1097]
[2]              U6 spliceosomal RNA [Source:RFAM;Acc:RF00026]
      biotype
      <factor>
[1] protein_coding
[2]          snRNA
---
seqlengths:
      7
      NA
```

And by using the feature parameter, we can specify what we're looking for

```
> featuresByRegion( '7:140424943-140624564', 'human', feature='transcript' )
GRanges with 6 ranges and 5 elementMetadata cols:
      seqnames      ranges strand |      ID
      <Rle>        <IRanges> <Rle> |      <factor>
[1]          7 [140424943, 140482957] - | ENST00000496384
[2]          7 [140434279, 140624564] - | ENST00000288602
[3]          7 [140434321, 140454011] - | ENST00000479537
[4]          7 [140434397, 140624458] - | ENST00000497784
[5]          7 [140533861, 140624509] - | ENST00000469930
[6]          7 [140583872, 140583978] + | ENST00000384313
      logic_name feature_type      Parent
      <factor>      <factor>      <factor>
[1]          havana      transcript ENSG00000157764
[2] ensembl_havana_transcript      transcript ENSG00000157764
[3]          havana      transcript ENSG00000157764
[4]          havana      transcript ENSG00000157764
[5]          havana      transcript ENSG00000157764
[6]          ncrna      transcript ENSG00000207040
      biotype
      <factor>
[1]      protein_coding
[2]      protein_coding
```

¹⁵http://beta.rest.ensembl.org/documentation/info/feature_region

```

[3] nonsense_mediated_decay
[4] nonsense_mediated_decay
[5]      retained_intron
[6]                snRNA
---
seqlengths:
  7
NA

```

You can specify multiple features (columns which don't exist for a given type of result will be filled with NA)

```
> featuresByRegion( '7:140424943-140624564', 'human', feature=c('gene','transcript') )
```

GRanges with 8 ranges and 7 elementMetadata cols:

	seqnames	ranges	strand	ID
	<Rle>	<IRanges>	<Rle>	<factor>
[1]	7	[140424943, 140624564]	-	ENSG00000157764
[2]	7	[140583872, 140583978]	+	ENSG00000207040
[3]	7	[140424943, 140482957]	-	ENST00000496384
[4]	7	[140434279, 140624564]	-	ENST00000288602
[5]	7	[140434321, 140454011]	-	ENST00000479537
[6]	7	[140434397, 140624458]	-	ENST00000497784
[7]	7	[140533861, 140624509]	-	ENST00000469930
[8]	7	[140583872, 140583978]	+	ENST00000384313

	logic_name	feature_type	external_name
	<factor>	<factor>	<factor>
[1]	ensembl_havana_gene	gene	BRAF
[2]	ncrna	gene	U6
[3]	havana	transcript	<NA>
[4]	ensembl_havana_transcript	transcript	<NA>
[5]	havana	transcript	<NA>
[6]	havana	transcript	<NA>
[7]	havana	transcript	<NA>
[8]	ncrna	transcript	<NA>

	description
	<factor>
[1]	v-raf murine sarcoma viral oncogene homolog B1 [Source:HGNC Symbol;Acc:1097]
[2]	U6 spliceosomal RNA [Source:RFAM;Acc:RF00026]
[3]	<NA>
[4]	<NA>
[5]	<NA>
[6]	<NA>
[7]	<NA>
[8]	<NA>

	biotype	Parent
	<factor>	<factor>
[1]	protein_coding	<NA>
[2]	snRNA	<NA>
[3]	protein_coding	ENSG00000157764
[4]	protein_coding	ENSG00000157764
[5]	nonsense_mediated_decay	ENSG00000157764
[6]	nonsense_mediated_decay	ENSG00000157764
[7]	retained_intron	ENSG00000157764
[8]	snRNA	ENSG00000207040

```
---  
seqlengths:  
  7  
NA
```

2.5 Lookup

2.5.1 lookupId

To find the database and species containing a known Ensembl id, you can use the lookup function¹⁶ like so:

```
> lookupId( 'ENSG00000170037' )  
      id      species object_type db_type  
1 ENSG00000170037 homo_sapiens      Gene    core
```

¹⁶<http://beta.rest.ensembl.org/documentation/info/lookup>

2.6 Mapping

The mapping functions are used to convert co-ordinates between systems or databases.

There is currently an issue with `mappingCdna`, `mappingCds` and `mappingTranslation` in that the `seq_region_name` is not returned from the REST interface. This has been reported to Ensembl and should be fixed in the next release.

2.6.1 mapping

The `mapping` function¹⁷ converts the co-ordinates in one assembly into another, ie:

```
> mapping( 'NCBI36', 'X:1..10000:1', 'GRCh37', 'human' )
[[1]]
GRanges with 2 ranges and 3 elementMetadata cols:
      seqnames      ranges strand |   assembly coordinate_system
      <Rle>        <IRanges> <Rle> | <character>      <character>
[1]          X [ 1, 10000]      + |   NCBI36        chromosome
[2]          X [60001, 70000]    + |   GRCh37        chromosome
      type
      <character>
[1]   original
[2]    mapped
---
seqlengths:
      X
      NA
```

As you can see, it returns one `GRanges` object per result, with an original row and a mapped row.

2.6.2 mappingCdna

This function¹⁸ converts CDNA co-ordinates for a given Ensembl Transcript to genomic co-ordinates.

```
> mappingCdna( 'ENST00000288602', '100..300' )
GRanges with 2 ranges and 2 elementMetadata cols:
      seqnames      ranges strand |   gap   rank
      <Rle>        <IRanges> <Rle> | <numeric> <numeric>
[1]          7 [140624366, 140624465] - |       0       0
[2]          7 [140549912, 140550012] - |       0       0
---
seqlengths:
      7
      NA
```

2.6.3 mappingCds

Or you can convert CDS co-ordinates¹⁹ instead of CDNA ones:

```
> mappingCds( 'ENST00000288602', '100..300' )
```

¹⁷http://beta.rest.ensembl.org/documentation/info/assembly_map

¹⁸http://beta.rest.ensembl.org/documentation/info/assembly_cdna

¹⁹http://beta.rest.ensembl.org/documentation/info/assembly_cds

GRanges with 3 ranges and 2 elementMetadata cols:

	seqnames	ranges	strand	gap	rank
	<Rle>	<IRanges>	<Rle>	<numeric>	<numeric>
[1]	7	[140624366, 140624404]	-	0	0
[2]	7	[140549911, 140550012]	-	0	0
[3]	7	[140534613, 140534672]	-	0	0

seqlengths:

7

NA

2.6.4 mappingTranslation

And finally, it is possible to convert from protein co-ordinates to genomic ones using the mappingTranslation method²⁰:

```
> mappingTranslation( 'ENSP00000288602', '100..300' )
```

GRanges with 5 ranges and 2 elementMetadata cols:

	seqnames	ranges	strand	gap	rank
	<Rle>	<IRanges>	<Rle>	<numeric>	<numeric>
[1]	7	[140534409, 140534615]	-	0	0
[2]	7	[140508692, 140508795]	-	0	0
[3]	7	[140507760, 140507862]	-	0	0
[4]	7	[140501212, 140501360]	-	0	0
[5]	7	[140500242, 140500281]	-	0	0

seqlengths:

7

NA

²⁰http://beta.rest.ensembl.org/documentation/info/assembly_translation

2.7 Sequences

2.7.1 sequenceById

Fetch a sequence based on the stable id of an Ensembl feature²¹ (I'm using `str` here to avoid overflowing the pdf too much):

```
> str( sequenceById( 'ENSG00000157764' ), give.head=F, strict.width='cut' )
List of 4
 $ desc      : "chromosome:GRCh37:7:140424943:140624564:-1"
 $ id        : "ENSG00000157764"
 $ seq       : "CGCCTCCCTTCCCCCTCCCCGCCGACAGCGGCCGCTCGGGCCCCGGCTCTCGGTTATAAGATGG..
 $ molecule  : "dna"
```

You can also get different types of sequence, here is an example for the spliced CDNA sequence of a transcript:

```
> str( sequenceById( 'ENST00000408384', type='cdna' ), give.head=F, strict.width='cut' )
List of 4
 $ desc      : NULL
 $ id        : "ENST00000408384"
 $ seq       : "GGATGCCCGAGCTAGTTTGAATTTTAGATAAAACAACGAATAATTCGTAGCATAAATATGTCCCAA..
 $ molecule  : "dna"
```

And again, for the protein coding

```
> str( sequenceById( 'ENSP00000334393', type='protein' ), give.head=F, strict.width='cut' )
List of 4
 $ desc      : NULL
 $ id        : "ENSP00000334393"
 $ seq       : "MVTEFIFLGLSDSQELQTFLEMLFFVFYGGIVFGNLLIVITVVSDSLHSPMYFLLANLSLIDLS..
 $ molecule  : "protein"
```

2.7.2 sequenceByRegion

You can also just query for a region of a given species²²:

```
> str( sequenceByRegion( 'X:1_000_000..1_000_100:1', 'human' ), give.head=F, strict.width='cut' )
List of 3
 $ id        : "chromosome:GRCh37:X:1000000:1000100:1"
 $ seq       : "GAAACAGCTACTTGGAAGGCTGAAGCAGGAGGATTGTTGAGTCTAGGAGTTTGAGGCTGCAGTG..
 $ molecule  : "dna"
```

If you pass `format='fasta'` to the above method, it will just return you a character vector containing a FastA formatted sequence.

²¹http://beta.rest.ensembl.org/documentation/info/sequence_id

²²http://beta.rest.ensembl.org/documentation/info/sequence_region

2.8 Variation

2.8.1 variationAllele

```
> var = variationAllele( 'C', '9:22125503-22125502:1', 'human' )
> var                                     # The response object

[[1]]
hgvs      :
  C = 9:g.22125502_22125503insC
transcripts : 10 in total

> var[[1]]$transcripts[1:2] # Just the top 2 transcripts of the first response to save room

[[1]]
data      :
      name      gene_id  transcript_id  biotype  cdna_allele_string
1 CDKN2B-AS1 ENSG00000240498 ENST00000585267 antisense      -/C
  is_canonical
1      FALSE

alleles      :
      consequence_terms
1 downstream_gene_variant

[[2]]
data      :
      name      gene_id  transcript_id  biotype  cdna_allele_string
1 CDKN2B-AS1 ENSG00000240498 ENST00000580576 antisense      -/C
  is_canonical
1      FALSE

alleles      :
      consequence_terms
1 downstream_gene_variant
```

2.8.2 variationId

```
> var = variationId( 'COSM476', 'human' )
> var                                     # The response object

[[1]]
name      : COSM476
is_somatic : TRUE
hgvs      :
  T = 7:g.140453136A>T
transcripts : 4 in total

> var[[1]]$transcripts[1:2] # Again, just the top 2 transcripts to save room

[[1]]
data      :
      name      gene_id  transcript_id  biotype  ccds
1 BRAF ENSG00000157764 ENST00000288602 protein_coding CCDS5863.1
  cdna_allele_string codon_position translation_stable_id translation_start
1      T/A      2      ENSP00000288602      600
  translation_end exon_number cdna_start cdna_end cds_start cds_end
```

```

1           600           15/18           1860           1860           1799           1799
  is_canonical
1           TRUE

alleles      :
  display_codon_allele_string pep_allele_string codon_allele_string
1           gTg/gAg           V/E           GTG/GAG
           hgvs_transcript           hgvs_protein polyphen_score
1 ENST00000288602.6:c.1799T>A ENSP00000288602.6:p.Val600Glu           0.999
  polyphen_prediction sift_score sift_prediction consequence_terms
1  probably damaging           0.02      deleterious  missense_variant

protein_features :
      name      db
1 PF07714      Pfam domain
2 PF00069      Pfam domain
3 SSF56112 Superfamily domains
4 SM00220      SMART domains
5 SM00219      SMART domains
6 PS50011      PROSITE profiles

[[2]]
data      :
  name      gene_id      transcript_id      biotype
1 BRAF ENSG00000157764 ENST00000479537 nonsense_mediated_decay
  cdna_allele_string codon_position translation_stable_id translation_start
1           T/A           2           ENSP00000418033           28
  translation_end exon_number cdna_start cdna_end cds_start cds_end
1           28           2/6           83           83           83           83
  is_canonical
1           FALSE

alleles      :
  display_codon_allele_string pep_allele_string codon_allele_string
1           gTg/gAg           V/E           GTG/GAG
           hgvs_transcript           hgvs_protein polyphen_score
1 ENST00000479537.1:c.83T>A ENSP00000418033.1:p.Val28Glu           0.946
  polyphen_prediction sift_score sift_prediction           V10
1  probably damaging           0.12      tolerated  missense_variant
  consequence_terms
1 NMD_transcript_variant

protein_features :
      name      db
1 PF00069      Pfam domain
2 PF07714      Pfam domain
3 SSF56112 Superfamily domains
4 PS50011      PROSITE profiles

```