

An R interface to the Ensembl REST API

Tim Yates

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1 Introduction

This package uses the Ensembl REST API¹ (currently in beta) to extract data from Ensembl into R.

As the REST API is in Beta, this package should also be considered to be in flux and functions/-parameters/etc are subject to change as things get finalized.

It could also do with your help. If you find a problem, something you think could be better, or a better way of doing things, please consider visiting the GitHub project at <https://github.com/acbb/EnsemblRest> and posting an issue or a Pull Request. Thanks!

2 Available Methods

To begin with (assuming you have installed this package), you need to load it into your R session:

```
> library( EnsemblRest )
```

The following subsections then list the methods available to you.

¹<http://beta.rest.ensembl.org/>

2.1 Information

2.1.1 isAlive

Firstly, we can check to see that the REST API is accepting calls:

```
> isAlive()  
[1] TRUE
```

2.1.2 infoSpecies

To get a list of available species on the server, you can use the `infoSpecies` call².

```
> infoSpecies()[1:3] # Just the first 3  
[[1]]  
name      : saccharomyces_cerevisiae  
aliases   : 4932, saccar, saccharomyces cerevisiae (baker's yeast), baker's yeast, scer, sacchar  
groups    : core, otherfeatures, variation, funcgen  
release   : 75  
  
[[2]]  
name      : ciona_savignyi  
aliases   : ciosav, 51511, ciona savignyi, csavignyi, c.savignyi, csav, sea squirt ciona savigny  
groups    : core, otherfeatures  
release   : 75  
  
[[3]]  
name      : myotis_lucifugus  
aliases   : little brown bat, mlucifugus, myoluc, mluc, 59463, myotis lucifugus, little_brown_ba  
groups    : core, otherfeatures  
release   : 75
```

2.1.3 infoAssembly

The `infoAssembly` call³ returns information about the currently available assemblies in the given species.

```
> infoAssembly( 'human' )  
assembly_name      : GRCh37.p13  
assembly_date      : 2009-02  
coord_system_versions : GRCh37, NCBI36, NCBI35, NCBI34  
genebuild_start_date : 2010-07-Ensembl  
genebuild_initial_release_date : 2011-04  
genebuild_last_geneset_update : 2013-09  
genebuild_method    : full_genebuild  
default_coord_system_version : GRCh37  
top_level_region count : 84
```

²<http://beta.rest.ensembl.org/documentation/info/species>

³http://beta.rest.ensembl.org/documentation/info/assembly_info

2.1.4 assemblyDetails

The `assemblyDetails` call⁴ returns information about one of these assemblies.

```
> assemblyDetails( 'X', 'human' )
is_chromosome      : TRUE
length            : 155270560
assembly_exception_type : REF
coordinate_system  : chromosome
```

2.1.5 infoComparas

The `infoComparas` call⁵ lists the available comparative genomics databases.

```
> infoComparas()
multi
75
```

2.1.6 infoData

The `infoData` call⁶ shows the data releases available to the REST service

```
> infoData()
[1] 75
```

2.1.7 infoRest

`infoRest` shows the current version⁷ of the REST service

```
> infoRest()
[1] "1.6.0"
```

2.1.8 infoSoftware

And finally in the `info` section, `infoSoftware` shows⁸ the current version of the Ensembl API.

```
> infoSoftware()
[1] 75
```

⁴http://beta.rest.ensembl.org/documentation/info/assembly_stats

⁵<http://beta.rest.ensembl.org/documentation/info/comparas>

⁶<http://beta.rest.ensembl.org/documentation/info/data>

⁷<http://beta.rest.ensembl.org/documentation/info/rest>

⁸<http://beta.rest.ensembl.org/documentation/info/software>

2.2 Comparative Genomics

2.2.1 geneTree

This method⁹ fetches the gene tree in New Hampshire format for a given Ensembl gene tree identifier.

```
> geneTree( 'ENSGT00390000003602' )
[1] "(((((((ENSXMAP00000006983:0.250602,ENSORLP00000004773:0.577127):0.196061,ENSGACP0000000151
```

it is also possible to specify the NH format you require (ie: for full format):

```
> geneTree( 'ENSGT00390000003602', nh_format='full' )
[1] "(((((((ENSXMAP00000006983:0.250602,ENSORLP00000004773:0.577127):0.196061,ENSGACP0000000151
```

2.2.2 homologyById

When given an Ensembl Gene ID, returns the homology¹⁰ information for it.

```
> hResponse = homologyById( 'ENSG00000170037' )
> hResponse # The response object
$ENSG00000170037
$id: ENSG00000170037 $homologies: 18 homologies
> hResponse[[1]]$homologies[1:2,] # Just the top 2 homologies to save room
  source.perc_pos
1             99
2             99

1 MATSADSPSSPLGAEDLLSDSSEPPGLNQVSSEVTSQLYASRLRSRQAEATARAQLYLPSTSPPEGLDGFAGELSRSLSVGLEKNLKKKDG
2 MATSADSPSSPLGAEDLLSDSSEPPGLNQVSSEVTSQLYASRLRSRQAEATARAQLYLPSTSPPEGLDGFAGELSRSLSVGLEKNLKKKDG
  source.protein_id source.perc_id source.cigar_line source.species
1 ENSP00000369614             99             925M homo_sapiens
2 ENSP00000369614             98             925M homo_sapiens
  source.id dn_ds target.perc_pos
1 ENSG00000170037 0.95             99
2 ENSG00000170037 NA              99

1 MATSADSPSSPLGAEDLLSDSSEPPGLNQVSSEVTSQLYASRLRSRQAEATARAQLYLPSTSPPEGLDGLAQELSRSLSVGLENNLKKKDG
2 MATSADSPSSPLGAEDLLSDSSEPPGLNQVSSEVTSQLYASRLRSRQAEATARAQLYLPSTSPPEGLDGLAQELSRSLSVGLENNLKKKDG
  target.protein_id target.perc_id target.cigar_line target.species
1 ENSPTRP00000014861             99             856MD68M pan_troglodytes
2 ENSGGOP00000006314             98             925M gorilla_gorilla
  target.id taxonomy_level type method_link_type
1 ENSPTRG00000008719 Homininae ortholog_one2one ENSEMBL_ORTHOLOGUES
2 ENSGGOG00000006451 Homininae ortholog_one2one ENSEMBL_ORTHOLOGUES
```

2.2.3 homologyBySymbol

You can also retrieve homology information¹¹ given a symbol and a species;

⁹<http://beta.rest.ensembl.org/documentation/info/rest>

¹⁰http://beta.rest.ensembl.org/documentation/info/homology_ensemblgene

¹¹http://beta.rest.ensembl.org/documentation/info/homology_symbol


```

> hResponse = homologyBySymbol( 'BRCA2', 'human' )
> hResponse                                     # The response object

$BRCA2
$id: ENSG00000139618 $homologies: 18 homologies
> hResponse[[1]]$homologies[1:2,] # Again, just the top 2 homologies to save room
  source.perc_pos
1              99
2              95

1  MPIGSKERPTFFEIFKTRCNKADLGPISLNWFEELSSEAPPYNSEPAEESSEHKNNNYEPNLFKTPQRKPSYNQLASTPIIFKEQGLTLPLY
2  MPIGSKERPTFFEIFKTRCNKA-DLGPISLNWFEELSSEAPPYNSEPAEESSEHKNNNYEPNLFKTPQRKPSYNQLASTPIIFKEQGLTLPLY
  source.protein_id source.perc_id source.cigar_line source.species
1  ENSP00000439902          99          3418M  homo_sapiens
2  ENSP00000439902          94          22MD3396M  homo_sapiens
  source.id  dn_ds target.perc_pos
1 ENSG00000139618 0.29371          99
2 ENSG00000139618    NA          98

1  MPIGSKERPTFFEIFKTRCNKADLGPISLNWFEELSSEAPPYNSEPAEESSEHKNNNYEPNLFKTPQRKPSYNQLASTPIIFKEQGLTLPLY
2  MPIGSKERPTFFEIFKTRCNKAVDLGPISLNWFEELSSEAPPYNSEPAEESSEHKNNNYEPNLFKTPQRKPSYNQLASTPIIFKEQGLTLPLY
  target.protein_id target.perc_id          target.cigar_line
1 ENSPTRP00000009812          99          3418M
2 ENSGGOP00000015446          97 98M7D563M4D615MD560M7D606M19D891M48D
  target.species          target.id taxonomy_level          type
1 pan_troglodytes ENSPTRG00000005766          Homininae ortholog_one2one
2 gorilla_gorilla ENSGGOG00000015808          Homininae ortholog_one2one
  method_link_type
1 ENSEMBL_ORTHOLOGUES
2 ENSEMBL_ORTHOLOGUES

```

And using format='condensed', you can get a more condensed result:

```

> hResponse = homologyBySymbol( 'BRCA2', 'human', format='condensed' )
> hResponse                                     # The response object

$BRCA2
$id: ENSG00000139618 $homologies: 6 homologies
> hResponse[[1]]$homologies[1:10,]
  taxonomy_level          protein_id          species          id
1          Homininae ENSPTRP00000009812 pan_troglodytes ENSPTRG00000005766
2          Homininae ENSGGOP00000015446 gorilla_gorilla ENSGGOG00000015808
3          Hominidae ENSPPYP00000005997 pongo_abelii ENSPPYG00000005264
4          Hominoidea ENSNLEP00000001277 nomascus_leucogenys ENSNLEG00000001048
5          Catarrhini ENSMMUP00000009432 macaca_mulatta ENSMMUG00000007197
6          Simiiformes ENSCJAP00000034250 callithrix_jacchus ENSCJAG00000018462
7          Haplorrhini ENSTSY00000000441 tarsius_syrichta ENSTSYG00000000478
8          Primates ENSMICP00000010933 microcebus_murinus ENSMICG00000011994
9          Primates ENSOGAP00000009477 otolemur_garnettii ENSOGAG00000010588
10 Euarchontoglires ENSTBEP00000013856 tupaia_belangeri ENSTBEG00000015907
  type          method_link_type
1 ortholog_one2one ENSEMBL_ORTHOLOGUES
2 ortholog_one2one ENSEMBL_ORTHOLOGUES
3 ortholog_one2one ENSEMBL_ORTHOLOGUES

```


4 ortholog_one2one ENSEMBL_ORTHOLOGUES
5 ortholog_one2one ENSEMBL_ORTHOLOGUES
6 ortholog_one2one ENSEMBL_ORTHOLOGUES
7 ortholog_one2one ENSEMBL_ORTHOLOGUES
8 ortholog_one2one ENSEMBL_ORTHOLOGUES
9 ortholog_one2one ENSEMBL_ORTHOLOGUES
10 ortholog_one2one ENSEMBL_ORTHOLOGUES

2.3 Cross References

Cross references are links to other data about the object of interest. It should be noted that these other data hold different fields and datatypes, so the results are returned in a `data.frame` containing a superset of column names, with non-applicable columns for a given result filled with `<NA>`.

As a single object may have multiple synonyms, this will cause the object to exist in multiple rows, one for each synonym.

2.3.1 xrefsById

Firstly we can get all external references¹² for a given Ensembl ID:

```
> xrefsById( 'ENSG00000170037' )
```

	display_id	primary_id	version
1	OTTHUMG00000172932	OTTHUMG00000172932	2
2	Hs.732863	Hs.732863	0
3	Hs.348012	Hs.348012	0
4	CENTROSOMAL BRCA2-INTERACTING PROT [*611425]	611425	0
5	CNTROB	29616	0
6	CNTROB	29616	0
7	CNTROB	116840	0
8	ENSG00000170037	ENSG00000170037	0
9	CNTROB	116840	0
10	CNTROB	CNTROB	0

	dbname	info_type	info_text	db_display_name
1	OTTG	NONE		Havana gene
2	UniGene	SEQUENCE_MATCH		UniGene
3	UniGene	SEQUENCE_MATCH		UniGene
4	MIM_GENE	DEPENDENT		MIM gene
5	HGNC	DIRECT	Generated via ccds	HGNC Symbol
6	HGNC	DIRECT	Generated via ccds	HGNC Symbol
7	EntrezGene	DEPENDENT		EntrezGene
8	ArrayExpress	DIRECT		ArrayExpress
9	WikiGene	DEPENDENT		WikiGene
10	Uniprot_gn	DEPENDENT		UniProtKB Gene Name

	ensembl_identity	ensembl_start	xref_start	xref_end	ensembl_end	score
1	NA	NA	NA	NA	NA	NA
2	99	1	6	752	752	3677
3	99	1	32	3794	3769	18783
4	NA	NA	NA	NA	NA	NA
5	NA	NA	NA	NA	NA	NA
6	NA	NA	NA	NA	NA	NA
7	NA	NA	NA	NA	NA	NA
8	NA	NA	NA	NA	NA	NA
9	NA	NA	NA	NA	NA	NA
10	NA	NA	NA	NA	NA	NA

	cigar_line	description
1	<NA>	<NA>
2	289M6D406M1I51M	Transcribed locus
3	376M6D3387M	Centrobin, centrosomal BRCA2 interacting protein
4	<NA>	CENTROSOMAL BRCA2-INTERACTING PROTEIN; CNTROB
5	<NA>	centrobin, centrosomal BRCA2 interacting protein
6	<NA>	centrobin, centrosomal BRCA2 interacting protein

¹²http://beta.rest.ensembl.org/documentation/info/xref_id


```

7          <NA> centrobins, centrosomal BRCA2 interacting protein
8          <NA>
9          <NA> centrobins, centrosomal BRCA2 interacting protein
10         <NA>
      xref_identity synonyms
1      NA      <NA>
2      99      <NA>
3      90      <NA>
4      NA      <NA>
5      NA      LIP8
6      NA      PP1221
7      NA      LIP8
8      NA      <NA>
9      NA      <NA>
10     NA      LIP8

```

2.3.2 xrefsByName

Or, we can look for an external reference primary accession¹³ (given a species):

```

> xrefsByName( 'NM_004333', 'human' )
      display_id primary_id version description      dbname info_type
1 NM_004333.4   NM_004333      4          RefSeq_mRNA    DIRECT
      info_text db_display_name
1 Generated via otherfeatures    RefSeq mRNA

```

2.3.3 xrefsBySymbol

And we can finally look up all Ensembl objects referenced by an external symbol for a given species¹⁴:

```

> xrefsBySymbol( 'BRAF', 'human' )
      type      id
1      gene ENSG00000157764
2 transcript ENST00000288602

```

¹³http://beta.rest.ensembl.org/documentation/info/xref_name

¹⁴http://beta.rest.ensembl.org/documentation/info/xref_external

2.4 Features

2.4.1 featuresByRegion

We can also look for features along a given range¹⁵ (by default this will just look for genes):

```
> featuresByRegion( '7:140424943-140624564', 'human' )
GRanges with 2 ranges and 7 metadata columns:
      seqnames      ranges strand |      source      ID
      <Rle>        <IRanges> <Rle> |      <factor>      <factor>
[1]          7 [140419127, 140624564] - | ensembl_havana ENSG00000157764
[2]          7 [140583872, 140583978] + |      ensembl ENSG00000271932
      logic_name feature_type external_name
      <factor>      <factor>      <factor>
[1] ensembl_havana_gene      gene      BRAF
[2]              ncrna      gene      U6
      description
      <factor>
[1] v-raf murine sarcoma viral oncogene homolog B [Source:HGNC Symbol;Acc:1097]
[2]              U6 spliceosomal RNA [Source:RFAM;Acc:RF00026]
      biotype
      <factor>
[1] protein_coding
[2]          snRNA
---
seqlengths:
  7
NA
```

And by using the feature parameter, we can specify what we're looking for

```
> featuresByRegion( '7:140424943-140624564', 'human', feature='transcript' )
GRanges with 6 ranges and 7 metadata columns:
      seqnames      ranges strand |      source      ID
      <Rle>        <IRanges> <Rle> |      <factor>      <factor>
[1]          7 [140419127, 140482957] - | ensembl_havana ENST00000496384
[2]          7 [140434279, 140624564] - | ensembl_havana ENST00000288602
[3]          7 [140434321, 140454011] - | ensembl_havana ENST00000479537
[4]          7 [140434397, 140624458] - | ensembl_havana ENST00000497784
[5]          7 [140533861, 140624509] - | ensembl_havana ENST00000469930
[6]          7 [140583872, 140583978] + |      ensembl ENST00000605989
      logic_name feature_type external_name      Parent
      <factor>      <factor>      <factor>      <factor>
[1]          havana      transcript      BRAF-003 ENSG00000157764
[2] ensembl_havana_transcript      transcript      BRAF-001 ENSG00000157764
[3]          havana      transcript      BRAF-005 ENSG00000157764
[4]          havana      transcript      BRAF-002 ENSG00000157764
[5]          havana      transcript      BRAF-004 ENSG00000157764
[6]              ncrna      transcript      U6.53-201 ENSG00000271932
      biotype
      <factor>
[1]      protein_coding
[2]      protein_coding
```

¹⁵http://beta.rest.ensembl.org/documentation/info/feature_region


```

[3] nonsense_mediated_decay
[4] nonsense_mediated_decay
[5]      retained_intron
[6]                snRNA
---
seqlengths:
  7
NA

```

You can specify multiple features (columns which don't exist for a given type of result will be filled with NA)

```
> featuresByRegion( '7:140424943-140624564', 'human', feature=c('gene','transcript') )
```

GRanges with 8 ranges and 8 metadata columns:

	seqnames	ranges	strand	source	ID
	<Rle>	<IRanges>	<Rle>	<factor>	<factor>
[1]	7	[140419127, 140624564]	-	ensembl_havana	ENSG00000157764
[2]	7	[140583872, 140583978]	+	ensembl	ENSG00000271932
[3]	7	[140419127, 140482957]	-	ensembl_havana	ENST00000496384
[4]	7	[140434279, 140624564]	-	ensembl_havana	ENST00000288602
[5]	7	[140434321, 140454011]	-	ensembl_havana	ENST00000479537
[6]	7	[140434397, 140624458]	-	ensembl_havana	ENST00000497784
[7]	7	[140533861, 140624509]	-	ensembl_havana	ENST00000469930
[8]	7	[140583872, 140583978]	+	ensembl	ENST00000605989

	logic_name	feature_type	external_name
	<factor>	<factor>	<factor>
[1]	ensembl_havana_gene	gene	BRAF
[2]	ncrna	gene	U6
[3]	havana	transcript	BRAF-003
[4]	ensembl_havana_transcript	transcript	BRAF-001
[5]	havana	transcript	BRAF-005
[6]	havana	transcript	BRAF-002
[7]	havana	transcript	BRAF-004
[8]	ncrna	transcript	U6.53-201

	description
	<factor>
[1]	v-raf murine sarcoma viral oncogene homolog B [Source:HGNC Symbol;Acc:1097]
[2]	U6 spliceosomal RNA [Source:RFAM;Acc:RF00026]
[3]	<NA>
[4]	<NA>
[5]	<NA>
[6]	<NA>
[7]	<NA>
[8]	<NA>

	biotype	Parent
	<factor>	<factor>
[1]	protein_coding	<NA>
[2]	snRNA	<NA>
[3]	protein_coding	ENSG00000157764
[4]	protein_coding	ENSG00000157764
[5]	nonsense_mediated_decay	ENSG00000157764
[6]	nonsense_mediated_decay	ENSG00000157764
[7]	retained_intron	ENSG00000157764
[8]	snRNA	ENSG00000271932


```
---  
seqlengths:  
  7  
NA
```


2.5 Lookup

2.5.1 lookupId

To find the database and species containing a known Ensembl id, you can use the lookup function¹⁶ like so:

```
> lookupId( 'ENSG00000170037' )  
      id      species object_type db_type  
1 ENSG00000170037 homo_sapiens      Gene    core
```

¹⁶<http://beta.rest.ensembl.org/documentation/info/lookup>

2.6 Mapping

The mapping functions are used to convert co-ordinates between systems or databases.

There is currently an issue with `mappingCdna`, `mappingCds` and `mappingTranslation` in that the `seq_region_name` is not returned from the REST interface. This has been reported to Ensembl and should be fixed in the next release.

2.6.1 mapping

The `mapping` function¹⁷ converts the co-ordinates in one assembly into another, ie:

```
> mapping( 'NCBI36', 'X:1..10000:1', 'GRCh37', 'human' )
[[1]]
GRanges with 2 ranges and 3 metadata columns:
      seqnames      ranges strand |   assembly coordinate_system
      <Rle>        <IRanges> <Rle> | <character>      <character>
[1]          X [ 1, 10000]      + |   NCBI36         chromosome
[2]          X [60001, 70000]    + |   GRCh37         chromosome
      type
      <character>
[1]   original
[2]    mapped
---
seqlengths:
      X
      NA
```

As you can see, it returns one `GRanges` object per result, with an original row and a mapped row.

2.6.2 mappingCdna

This function¹⁸ converts CDNA co-ordinates for a given Ensembl Transcript to genomic co-ordinates.

```
> mappingCdna( 'ENST00000288602', '100..300' )
GRanges with 2 ranges and 2 metadata columns:
      seqnames      ranges strand |   gap   rank
      <Rle>        <IRanges> <Rle> | <numeric> <numeric>
[1]          7 [140624366, 140624465] - |         0         0
[2]          7 [140549912, 140550012] - |         0         0
---
seqlengths:
      7
      NA
```

2.6.3 mappingCds

Or you can convert CDS co-ordinates¹⁹ instead of CDNA ones:

```
> mappingCds( 'ENST00000288602', '100..300' )
```

¹⁷http://beta.rest.ensembl.org/documentation/info/assembly_map

¹⁸http://beta.rest.ensembl.org/documentation/info/assembly_cdna

¹⁹http://beta.rest.ensembl.org/documentation/info/assembly_cds

GRanges with 3 ranges and 2 metadata columns:

	seqnames	ranges	strand	gap	rank
	<Rle>	<IRanges>	<Rle>	<numeric>	<numeric>
[1]	7	[140624366, 140624404]	-	0	0
[2]	7	[140549911, 140550012]	-	0	0
[3]	7	[140534613, 140534672]	-	0	0

seqlengths:

7

NA

2.6.4 mappingTranslation

And finally, it is possible to convert from protein co-ordinates to genomic ones using the mappingTranslation method²⁰:

```
> mappingTranslation( 'ENSP00000288602', '100..300' )
```

GRanges with 5 ranges and 2 metadata columns:

	seqnames	ranges	strand	gap	rank
	<Rle>	<IRanges>	<Rle>	<numeric>	<numeric>
[1]	7	[140534409, 140534615]	-	0	0
[2]	7	[140508692, 140508795]	-	0	0
[3]	7	[140507760, 140507862]	-	0	0
[4]	7	[140501212, 140501360]	-	0	0
[5]	7	[140500242, 140500281]	-	0	0

seqlengths:

7

NA

²⁰http://beta.rest.ensembl.org/documentation/info/assembly_translation

2.7 Sequences

2.7.1 sequenceById

Fetch a sequence based on the stable id of an Ensembl feature²¹ (I'm using `str` here to avoid overflowing the pdf too much):

```
> str( sequenceById( 'ENSG00000157764' ), give.head=F, strict.width='cut' )
List of 4
 $ desc      : "chromosome:GRCh37:7:140419127:140624564:-1"
 $ id        : "ENSG00000157764"
 $ seq       : "CGCCTCCCTTCCCCCTCCCCGCCCCGACAGCGGCCGCTCGGGCCCCGGCTCTCGGTTATAAGATG"..
 $ molecule  : "dna"
```

You can also get different types of sequence, here is an example for the spliced CDNA sequence of a transcript:

```
> str( sequenceById( 'ENST00000473358', type='cdna' ), give.head=F, strict.width='cut' )
List of 4
 $ desc      : NULL
 $ id        : "ENST00000473358"
 $ seq       : "GTGCACACGGCTCCCATGCGTTGTCTTCGAGCGTCAGGCCGCCCTACCCGTGCTTTCTGCTC"..
 $ molecule  : "dna"
```

And again, for the protein coding

```
> str( sequenceById( 'ENSP00000334393', type='protein' ), give.head=F, strict.width='cut' )
List of 4
 $ desc      : NULL
 $ id        : "ENSP00000334393"
 $ seq       : "MVTEFIFLGLSDSQELQTFLEMLFFVFYGGIVFGNLLIVITVVSDSLHSPMYFLLANLSLIDL"..
 $ molecule  : "protein"
```

2.7.2 sequenceByRegion

You can also just query for a region of a given species²²:

```
> str( sequenceByRegion( 'X:1000000..1000100:1', 'human' ), give.head=F, strict.width='cut' )
List of 3
 $ id        : "chromosome:GRCh37:X:1000000:1000100:1"
 $ seq       : "GAAACAGCTACTTGGAAGGCTGAAGCAGGAGGATTGTTTGAAGTCTAGGAGTTTGAGGCTGCAGT"..
 $ molecule  : "dna"
```

If you pass `format='fasta'` to the above method, it will just return you a character vector containing a FastA formatted sequence.

²¹http://beta.rest.ensembl.org/documentation/info/sequence_id

²²http://beta.rest.ensembl.org/documentation/info/sequence_region

2.8 Variation

2.8.1 variationAllele

```
> var = variationAllele( 'C', '9:22125503-22125502:1', 'human' )
> var                                     # The response object

[[1]]
hgvs      :
  C = 9:g.22125502_22125503insC
transcripts : 10 in total
> var[[1]]$transcripts[1:2] # Just the top 2 transcripts of the first response to save room

[[1]]
data      :
      name      gene_id  transcript_id  biotype cdna_allele_string
1 CDKN2B-AS1 ENSG00000240498 ENST00000585267 antisense          -/C
  is_canonical
1      FALSE

alleles      :
  consequence_terms
1 downstream_gene_variant

[[2]]
data      :
      name      gene_id  transcript_id  biotype cdna_allele_string
1 CDKN2B-AS1 ENSG00000240498 ENST00000580576 antisense          -/C
  is_canonical
1      FALSE

alleles      :
  consequence_terms
1 downstream_gene_variant
```

2.8.2 variationId

```
> var = variationId( 'COSM476', 'human' )
> var                                     # The response object

[[1]]
name      : COSM476
is_somatic : TRUE
hgvs      :
  T = 7:g.140453136A>T
transcripts : 4 in total
> var[[1]]$transcripts[1:2] # Again, just the top 2 transcripts to save room

[[1]]
data      :
      name      gene_id  transcript_id  biotype
1 BRAF ENSG00000157764 ENST00000479537 nonsense_mediated_decay
  cdna_allele_string codon_position translation_stable_id translation_start
1      T/A          2      ENSP00000418033          28
  translation_end exon_number cdna_start cdna_end cds_start cds_end
```



```

1           28           2/6           83           83           83           83
  is_canonical
1          FALSE

alleles      :
  display_codon_allele_string pep_allele_string codon_allele_string
1           gTg/gAg           V/E           GTG/GAG
           hgvs_transcript           hgvs_protein polyphen_score
1 ENST00000479537.1:c.83T>A ENSP00000418033.1:p.Val28Glu           0.676
  polyphen_prediction sift_score sift_prediction           V10
1  possibly damaging           0.12           tolerated missense_variant
           consequence_terms
1 NMD_transcript_variant

protein_features :
           name           db
1 PF00069           Pfam domain
2 PF07714           Pfam domain
3 SSF56112 Superfamily domains
4 PS50011           PROSITE profiles

[[2]]
data      :
  name      gene_id      transcript_id      biotype      ccds
1 BRAF ENSG00000157764 ENST00000288602 protein_coding CCDS5863.1
  cdna_allele_string codon_position translation_stable_id translation_start
1           T/A           2           ENSP00000288602           600
  translation_end exon_number cdna_start cdna_end cds_start cds_end
1           600           15/18           1860           1860           1799           1799
  is_canonical
1          TRUE

alleles      :
  display_codon_allele_string pep_allele_string codon_allele_string
1           gTg/gAg           V/E           GTG/GAG
           hgvs_transcript           hgvs_protein polyphen_score
1 ENST00000288602.6:c.1799T>A ENSP00000288602.6:p.Val600Glu           0.967
  polyphen_prediction sift_score sift_prediction consequence_terms
1  probably damaging           0           deleterious missense_variant

protein_features :
           name           db
1 PF07714           Pfam domain
2 PF00069           Pfam domain
3 SSF56112 Superfamily domains
4 SM00220           SMART domains
5 SM00219           SMART domains
6 PS50011           PROSITE profiles

```