

### ANOVA (CRD)

- General format of ANOVA in R
- Testing the assumption of homogeneity of variances using Levene's Test
- One-way ANOVA of nested design
- Obtaining and interpreting Components of Variance

### Linear Models in R

The primary R function for the analysis of variance of a fixed-effects model is the linear model (**lm()**) function. The **lm()** function has the following basic syntax:

**lm(model, dataset)**

The **model** is an explanatory linear model for the response variable. It is a minimalist representation of a general linear effects model, where a response (i.e. dependent variable) is explained by a host of known additive deviations from a base mean:

$$Y_i = \mu + \kappa_1 + \kappa_2 + \dots + \kappa_n + \varepsilon_i$$

For a completely randomized design (CRD) with one treatment variable, the independent effect is the treatment and the dependent variable is the response. The general syntax is:

*dependent variable ~ independent effects*

For a single-factor CRD, the R script would be:

**lm(Response ~ Treatment, data = dataname)**

There can be more complex models, like those below; and the **lm()** function can handle them:

$y \sim a \ b$	-> main effects of a factorial experiment
$y \sim a \ b \ a*b$	-> including their interaction

### ANOVAs in R

Now, to view the results of the above linear model analysis in the familiar form of an ANOVA table, one must apply a second R function (**anova()**) to the object produced by the **lm()** function. The general workflow involves the following two lines of code:

```
data.mod <- lm(Response ~ Treatment, data = dataname)
anova(data.mod)
```

## Example 1

[Lab3ex1.R]

In this experiment, the nitrogen fixation capacities of six different strains of rhizobia on clover are compared. The experiment is arranged as a CRD with 6 treatments (i.e. 6 different strains) and five independent replications (e.g. plots) per treatment.

```
#This script performs a CRD analysis (one-way ANOVA) and tests for
homogeneity of variance
```

```
#Load the data, then use the following function to examine the structure of
your data
```

```
str(clover_dat, give.attr = F)
```

```
#Convert clover_dat to a dataframe and tell R that "Culture" is a
classification variable; re-examine
```

```
clover_dat <- as.data.frame(clover_dat)
```

```
clover_dat$Culture <- as.factor(clover_dat$Culture)
```

```
str(clover_dat, give.attr = F)
```

```
#Apply the linear model and look at the results (ANOVA table)
```

```
clover_mod<-lm(NLevel ~ Culture, data = clover_dat)
```

```
anova(clover_mod)
```

```
#Some more results
```

```
summary(clover_mod)
```

```
#Create box-and-whisker plot of the data
```

```
plot(clover_dat)
```

### Results of the `lm()` and `anova()` functions:

Analysis of Variance Table

Response: NLevel

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Culture	5	847.05	169.409	14.37	1.485e-06 ***
Residuals	24	282.93	11.789		

---

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**Interpretation** Recall that the null hypothesis of an ANOVA is that all means are equal ( $H_0: \mu_1 = \mu_2 = \dots = \mu_n$ ) while the alternate hypothesis is that at least one mean is not equal ( $H_1: \mu_i \neq \mu_j$ ). With a p-value of less than 0.00001, we soundly reject  $H_0$ . A significant component of variation exists in the dataset due to treatments.

### Results of the `summary()` function:

```
Residual standard error: 3.433 on 24 degrees of freedom
Multiple R-squared:  0.7496, Adjusted R-squared:  0.6975
F-statistic: 14.37 on 5 and 24 DF,  p-value: 1.485e-06
```

**Interpretation** The "residual standard error" is the "root MSE" (i.e. the square root of the mean square error  $\sqrt{11.789} = 3.433$ ). The R-Square value is a measure of the amount of variation explained by the model:

$$\text{R-Square} = 847.046667 / 1129.974667 = 0.749616$$

### Testing the Assumption of Homogeneity of Variances (HOV)

When you perform an ANOVA, you assert that the data under consideration meet the assumptions for which an ANOVA is valid. We briefly discussed normality in Lab 1. Now we will cover a second assumption, that the variances of all compared treatments are homogeneous.

#### ...by an ANOVA of residuals

*#extract residual values*

```
clover_res<-residuals(clover_mod)
```

*#add these residual values to the original dataframe*

```
clover_dat$res<-clover_res
```

```
clover_dat$absres<-abs(clover_res) #Levene's original test
```

```
clover_dat$res2<-clover_res^2 #more robust test, using squared residuals
```

```
head(clover_dat)
```

*#Test for homogeneity of variances (Levene's Test)*

```
leveneABS_mod<-lm(absres ~ Culture, data = clover_dat)
```

```
anova(leveneABS_mod)
```

```
leveneRES2_mod<-lm(res2 ~ Culture, data = clover_dat)
```

```
anova(leveneRES2_mod)
```

#### Results:

Response: absres

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Culture	5	48.957	9.7915	3.1451	0.02531 *
Residuals	24	74.718	3.1133		

Response: res2

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Culture	5	2563.0	512.59	1.8433	0.1423
Residuals	24	6674.1	278.09		

**Interpretation** The original Levene test indicates a violation of HOV, while the more robust test based on squared residuals does not. In 1974, Brown and Forsythe investigated this sort of disparity. They extended the Levene test to use residuals calculated on group medians and trimmed means, in addition to just means, and found *the method based on group medians to be the most generally robust technique*. This is the default technique used by R and the one we will be using:

```
#The Levene Test is available through the "car" package
#install.packages("car")
#library(car)
```

```
leveneTest(clover.mod, center = mean) #same as abs(res)
leveneTest(clover.mod, center = median) #the default
```

### Results:

#### Levene's Test for Homogeneity of Variance (center = mean)

	Df	F value	Pr(>F)
group	5	3.1451	0.02531 *
	24		

#### Levene's Test for Homogeneity of Variance (center = median)

	Df	F value	Pr(>F)
group	5	0.9295	0.4794
	24		

**Interpretation**  $H_0$  for Levene's Test is that the variances of the treatments are homogeneous, and  $H_a$  is that they are not homogeneous. With a p-value of 0.4794, we fail to reject  $H_0$ ; thus we have no evidence, at the 95% confidence level, that the variances are not homogeneous. *If you would like to see how to perform an ANOVA on the absolute values of median-based residuals, refer to the code on the next page.*

Finally, there are other HOV tests, also available through the "car" package:

#### #alternative HOV tests

```
bartlett.test(NLevel ~ Culture, data = clover.dat) #parametric test
fligner.test(NLevel ~ Culture, data = clover.dat) #non-parametric test
```

#### Bartlett test of homogeneity of variances

```
data: NLevel by Culture
Bartlett's K-squared = 14.2207, df = 5, p-value = 0.01427
```

#### Fligner-Killeen test of homogeneity of variances

```
data: NLevel by Culture
Fligner-Killeen:med chi-squared = 3.7264, df = 5, p-value = 0.5894
```

**For the interested:**

*Performing an ANOVA on the absolute values of the median-based residuals (Brown and Forsythe's improved Levene's Test)*

```
#Testing for homogeneity of variances using Brown and Forsythe's improved
  Levene's Test,
#based on an ANOVA of the absolute values of median-based residuals

#To find residuals based on MEDIANs, first calculate the medians for the 6
  treatment classes
aggregate(clover_dat$NLevel, list(clover_dat$Culture), median)
#Create a medians column in the original dataset
clover_dat$medians<-
  c(rep(32.1,5),rep(24.8,5),rep(15.8,5),rep(20.5,5),rep(14.2,5),rep(19.1,5))
#Calculate residuals, based on medians
clover_dat$median_resids<-clover_dat$NLevel - clover_dat$medians
#Find the absolute values of those residuals
clover_dat$median_resids_abs<-abs(clover_dat$median_resids)
clover_dat

#Perform the ANOVA
leveneMEDIAN_ABS_mod<-lm(median_resids_abs ~ Culture, data = clover_dat)
anova(leveneMEDIAN_ABS_mod)
```

**Results:**

Analysis of Variance Table

```
Response: median.resids.abs
      Df Sum Sq Mean Sq F value Pr(>F)
Culture   5  37.062   7.4123   0.9295  0.4794
Residuals 24 191.392   7.9747
```

The results match those of the default Levene's Test procedure in the "car" package.

## One-Way ANOVA of Nested Design

### Example 2

[Lab3ex2.R]

In this experiment, the effects of different treatments on the growth of mint plants are being measured. It is a purely nested CRD with six levels of treatment, three replications (i.e. pots) per level, and four subsamples (i.e. plants) per replication.

#This script performs a one-way ANOVA for a perfectly nested CRD and finds variance components

#read in, re-classify, and inspect the data

```
mint_dat <- as.data.frame(mint_dat)
mint_dat$Trtmt <- as.factor(mint_dat$Trtmt)
mint_dat$Pot <- as.factor(mint_dat$Pot)
mint_dat$Plant <- as.factor(mint_dat$Plant)
str(mint_dat, give.attr = F)
head(mint_dat)
tail(mint_dat)
```

'data.frame': 72 obs. of 4 variables:

```
$ Trtmt : Factor w/ 6 levels "Cond1","Cond2",...: 1 1 1 1 1 1 1 1 1 1 ...
$ Pot   : Factor w/ 3 levels "Pot1","Pot2",...: 1 1 1 1 2 2 2 2 3 3 ...
$ Plant : Factor w/ 4 levels "Plant1","Plant2",...: 1 2 3 4 1 2 3 4 1 2 ...
$ Growth: num 3.5 4 3 4.5 2.5 4.5 5.5 5 3 3 ...
```

	Trtmt	Pot	Plant	Growth
1	Cond1	Pot1	Plant1	3.5
2	Cond1	Pot1	Plant2	4.0
3	Cond1	Pot1	Plant3	3.0
4	Cond1	Pot1	Plant4	4.5
5	Cond1	Pot2	Plant1	2.5
...				
68	Cond6	Pot2	Plant4	7
69	Cond6	Pot3	Plant1	11
70	Cond6	Pot3	Plant2	7
71	Cond6	Pot3	Plant3	9
72	Cond6	Pot3	Plant4	8

```
#Calculating components of variance
#install.packages("lme4")
#library(lme4)
mintCV_mod<-lmer(Growth ~ Trtmt + (1|Trtmt:Pot), data = mint_dat)
summary(mintCV_mod)
```

## Output

```
Random effects:
  Groups      Name      Variance Std.Dev.
Pot:Trtmt (Intercept) 0.3047    0.5520
Residual      0.9340    0.9665
Number of obs: 72, groups: Pot:Trtmt, 18
```

## #The ANOVA

```
mint_mod<-lm(Growth ~ Trtmt/Pot, data = mint_dat)
anova(mint_mod)
```

## #The above model is short-hand for this:

```
mint2_mod<-lm(Growth ~ Trtmt + Trtmt:Pot, data = mint_dat)
anova(mint2_mod)
```

With the above scripts, R will generate the following ANOVA table, which presents an *incorrect* F test for the effect of treatment:

```
Response: Growth
      Df  Sum Sq Mean Sq F value Pr(>F)
Trtmt    5  179.642   35.928  38.4662 < 2e-16 ***
Trtmt:Pot 12   25.833    2.153   2.3048 0.01858 *
Residuals 54   50.438    0.934
---
***
```

**WRONG !**

We know from the expected mean squares that the correct error term to use for testing the effect of treatment is **MS<sub>Pot</sub>** (the MSEE), not **MS<sub>Plant</sub>** (MSSE). To conduct the correct test, we type:

## #Custom F-test for Trtmt

```
F_Trmtmt<-35.928/2.153
F_Trmtmt
pf(F_Trmtmt, 5, 12, lower.tail = F)
```

This returns the correct F-value (**16.69**) and p-value (**4.88e-05**) for the effect of treatment.

If this seems unnecessarily complicated, it is. Note that the same F value will result if you simply consider the averages of the subsamples, though the MS's will be different. **If you are not concerned with the components of the variance, there is no need to carry out a nested analysis.** Just average the subsamples and treat it as an un-nested experiment, as shown in the following example.

### Example 3

[Lab3ex3.R]

This is the same experiment as above, but here the measurements of the four plants (subsamples) in each pot have been averaged.

#This script performs a one-way ANOVA, where subsamples have been averaged

#read in, re-classify, and inspect the data

```
mint_dat <- as.data.frame(mint_dat)
mint_dat$Trtmt <- as.factor(mint_dat$Trtmt)
mint_dat$Pot <- as.factor(mint_dat$Pot)
str(mint_dat, give.attr = F)
```

#The ANOVA

```
mint_mod<-lm(Growth ~ Trtmt, data = mint_dat)
anova(mint_mod)
```

```
'data.frame': 18 obs. of 3 variables:
 $ Trtmt : Factor w/ 6 levels "Cond1","Cond2",...: 1 1 1 2 2 2 3 3 3 4 ...
 $ Pot : Factor w/ 3 levels "Pot1","Pot2",...: 1 2 3 1 2 3 1 2 3 1 ...
 $ Growth: num 3.75 4.38 2.88 4.5 3.5 ...
```

	Trtmt	Pot	Growth
1	Cond1	Pot1	3.750
2	Cond1	Pot2	4.375
3	Cond1	Pot3	2.875
4	Cond2	Pot1	4.500
5	Cond2	Pot2	3.500
6	Cond2	Pot3	4.375
...			
13	Cond5	Pot1	5.500
14	Cond5	Pot2	6.625
15	Cond5	Pot3	7.250
16	Cond6	Pot1	8.250
17	Cond6	Pot2	6.750
18	Cond6	Pot3	8.750



```
#The ANOVA
mint_mod<-lm(Growth ~ Trtmt, data = mint_dat)
anova(mint_mod)
```

## Output

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Trtmt	5	44.911	8.9821	<b>16.689</b>	<b>4.881e-05</b> ***
Residuals	12	6.458	0.5382		

We arrive at the same result as before, but with much less complication.

## For the interested – some sweet code

Wouldn't it be great if you could run an ANOVA on the Pot means without having to first calculate those means in Excel and then re-import a reduced dataset? The following code, which relies on the `ddply()` function within the "plyr" library, is what you're looking for:

```
pot_means_dat <- ddply(mint_dat, c("Trtmt", "Pot"), summarise,
  Growth = mean(Growth, na.rm=TRUE))
```

This one-liner creates a reduced dataset for you (I call it "pot\_means\_dat"), in which the Growth measurements of all the Plants in a given Pot have been averaged. The elements in green are the terms you would need to change, based on your specific dataset. Give it a try! And thanks to Natalie, a previous BIOL933 student, for figuring this one out.