Preparing spatial data for species distribution modelling in R

Simon Kapitza* Adriano Fernandez Kaulich[†]
November 28, 2014



Abstract

This report serves as documentation for an Rscript produced in the masters course "Best practice for data analysis and R: writing and coding geographic and statistical analyses for theses and reports". The Rscript loads and processes a user-specified species distribution shapefile and a multitude of user-specified environmentally relevant parameters in raster format, combining both file formats into one data frame representing equal area raster cells. The resulting data frames can be used for statistical analysis, such as species distribution modelling. The aim is to automatise a relatively easy GIS task so that users, who are more interested in statistics and less in GIS, are able to prepare their data set with minimal user input and without relying on conventional GIS software. Excerpts from the Rscript are interlaced in this report. The entire Rscript is appended.

*MtrklNr.: 3710110 †MtrklNr: 2920170

Contents

1	Introduction	2
2	Rscript documentation	2
3	Short-comings	6

1 Introduction

The aim of this project is to facilitate the use of R as a GIS-Software. GIS-Software is often costly. Hence, the free statistical software R becomes a worthwhile alternative. In order to help students and researchers explore R as an alternative to GIS, many common GIS functions need to be implemented in a comfortable way in R.

In this document we explain how our Rscript can help researchers prepare their data for species distribution modeling (SDM). Data used in SDM is often presented as a spatial grid (raster-data) with every grid cell containing data about the presence of the species and environmental parameters like altitude and land cover or precipitation and temperature records. Our Rscript brings data of this type into one grid of user-specified properties.

2 Rscript documentation

The first step is to acquire all the necessary data for the analysis. This requires the user to input some basic parameters like the species name (in Latin), the desired cell size of the grid (in meters) and the EPSG-code for the coordinate reference system best suited to the extent of the study site (equal area projection).

Spatial data for almost every non-domesticated mammalian, amphibian, reptilian and marine fish species is accessible at www.iucnredlist.org. A function was written that redirects the user to the download page of the requested species at the website of the IUCN. Unfortunately a user login is required, otherwise it would have been possible to directly download the spatial data and load it into R. The user is required to move the downloaded species distribution folder into the set working directory. It is possible to manually download all species distribution shapefiles in bulk (over 2 GB of data in shapefiles) from the website of the IUCN, which could be added as an additional feature to the present workflow. Raster data for climate records is available at www.worldclim.org without a user login, allowing us to directly download and load the data into a temporary directory for use in the workflow. The same applies for land-cover data from www.fao.org.

. Both data sets will be deleted eventually. (Start the workflow: Load all libraries in section 2. of the Rscript and specify your species, WorldClim-parameters, your target resolution and the equal area coordinate reference system of your intended study site in section 3. In order to download all required files and be prompted with a download link for your species' distribution shapefile, execute sections 4. - 5. in the Rscript)

```
#-----4. LOAD WORKFLOW FUNCTIONS-----
# 4. a) Function for the download of WorldClim parameters
```

```
getWorldClim <- function(par, res) {</pre>
 parlist <- list()</pre>
 for (i in 1:length(par)) {
    getData('worldclim', path=path.temp_data, download = T, var=par[i], res=res)
}
# 4. b) Function that creates link to IUCN profile of intended species, where the species
IUCNdata <- function(name) {</pre>
 browseURL(paste("http://maps.iucnredlist.org/map.html?id=", ID, sep=""))
  message("Make sure to move the species folder into your working directory")
# 4. c) Function to round numbers to nearest multiple of a specific number (needed in creations)
mround <- function(number, multiple) {</pre>
  # if number and multiple have different sign, returning an error.
 if (sign(number) != sign(multiple)) {
    stop("number and multiple have different sign")
 n = number / multiple
  if (abs(n - trunc(n)) == 0.5) {
    if (sign(n) > 0) {
      n = n + 0.1
    } else {
      n = n - 0.1
  round(n) * multiple
```

Once all the data is available in R, the species is plotted onto a 2D plane (WGS84 Lat/Lon; EPSG:4326) together with the borders of countries in the distribution area to facilitate the selection of the area of interest. The world borders shapefile was downloaded from www.thematicmapping.org. The user is then asked to select the study area (AoI) on the plotted maps. The extent of a rectangular box sourrounding the AoI is then extracted using the function gEnvelope(). The box is then converted to a Formal Class SpatialPolygon and reprojected into the user-specified CRS. Since the user-defined CRS is metric, the extent()-function allows us to extract the dimensions of the AoI and calculate the desired raster grid (dividing the aoi's extent by the desired cell size, e.g. 50 km). The grid size has to be a multiple of the desired cell size. If this condition is not met, the original extent will be rounded to the next biggest multiple of the specified cell size (e.g. if the original extent between xmin and xmax from gEnvelope() is 887 km, the final grid will be extended to 900km, in order to allow 18 cells of 50 km on the x-axis). In order to round to multiples, the mround function by Alberto Santini: https://gist.github.com/albertosantini/3638434 was applied.

```
#-----6. RASTERISATION------
```

6. a) Select Area of Interest (needs to be within equal area projection specified above

[#] Read species distribution and worldborders shapefiles and plot them into 0-device worldborders <- readOGR(dsn = paste(path.temp_data, "WorldBorders", sep = "/"), layer = "species.shp <- readOGR(dsn = paste(paste(getwd(), "species_", sep = "/"), ID, sep = ""), ID, sep = ""),

```
# plot for AOI selection
plot(species.shp, col = "brown")
plot(worldborders, add = T)
# clip a polygon from distribution map, convert to spatial polygons object
aoi <- getpoly(quiet = F)</pre>
aoi_sp <- Polygon(aoi)</pre>
aoi_sps <- Polygons(list(aoi_sp),ID = ID)</pre>
aoi_spls <- SpatialPolygons(list(aoi_sps), proj4string = CRS(species.shp@proj4string@proj8</pre>
# 6. b) Build template raster covering the Area of Interest
# get the extent of the clipped polygon and convert to target projection
aoi_ext <- gEnvelope(aoi_spls)</pre>
aoi_ext_utm <- spTransform(aoi_ext, crs(crs))</pre>
ext <- extent(aoi_ext_utm)</pre>
# force target grid to be a multiple of desired grid size in order to get integer values :
dimx <- (mround(length(ext@xmin:ext@xmax), target_resolution))/target_resolution</pre>
dimy <- (mround(length(ext@ymin:ext@ymax), target_resolution))/target_resolution</pre>
# build a template raster for the spatial_sync_raster function
aoi_rr_utm <- raster(ext, ncols = dimx, nrows = dimy, crs = crs)</pre>
```

The species distribution shapefile needs to be cropped to the extent of the *AoI* before it can be processed into a raster. The unprojected polygon boxing the selected area of interest is used for this. The resulting cropped species shapefile is then projected into the target CRS. Rasterising this cropped and reprojected spatial shapefile takes into account the area the species distribution polygon covers in each cell. Cells located completely within the extent of the polygon will be assigned the value 1 for full coverage by the polygon. Along the edges of the polygon, the rasterised cells's values range from from 0 to 1, depending on how much of these cells is actually covered by the species shapefile. This is important to adequately weight the influence of these cells in further analysis (e.g. in a linear regression). *To crop your area of interest and calculate a template raster that is then filled cell-by-cell with your specified parameters, execute the entire section 6. of the Rscript. Be aware that processing land cover data may take a while.*

```
# 6. c) fitting species distribution data into template raster

# crop to AOI extent
species_aoi_shp <- crop(species.shp, aoi_ext)

# transfrom to target crs
species_aoi_shp_utm <- spTransform(species_aoi_shp, crs(crs))

# rasterise species shapefile, extracting proportion of each raster cell covered by species_aoi_raster <- rasterize(species_aoi_shp_utm, aoi_rr_utm, getCover = T)
species_aoi_raster@data@values <- species_aoi_raster@data@values/100
species_aoi_raster@data@names <- species</pre>
```

Processing the rasterised environmental data starts with indexing the location of every rasterfile inside the folders they were downloaded to, using the function list.raster.files(). This list of file paths is used by a for-loop that loads every downloaded WorldClim raster and crops, reprojects and resamples them in accordance with the properties of the template raster. The files from www.worldclim.org are detected by list.raster.files() and loaded in alphabetical order. This would be fine except that the WorldClim monthly data are numbered for each month (e.g. tmin1, tmin2, tmin3 ... tmin10, tmin11, tmin12). The rasters for October, November and December are thus loaded before the rasters for February in the list

of file-paths returned by the list.raster.files-function. As a workaround, an empty list with only the file names was created, using the list.files-function. The rasters are called into the loop in the wrong order, however, since the raster names are called into the function in the same order, they can be assigned to the rasters by using their indices. This way, the rasters get their original names while being on an index position that does not correspond to the ID of their month.

Processing itself includes the cropping, projecting and resampling of every rasterfile in rasterlist using the same template raster created for rasterizing the species shapefile. The resampling-function offers two ways of estimating the new cell values: nearest neighbour and bilinear resampling. We found that bilinear resampling yields better results but takes notably longer than the nearest neighbour option. The processed rasters were stored as raster stack.

6. d) fitting WorldClim data into template raster

```
# find WorldClim rasters in their download folder and save their target path and names in
rasterlist <- list.raster.files(paste(paste(path.temp_data,"wc", sep = "/"),res, sep = """)
names <- list.files(paste(paste(path.temp_data, "wc",sep = "/"), res, sep = """), pattern = """)
# load, crop, reproject and resample WorldClim raster to fit template raster
wc_processed <- list()
for (i in 1:length(rasterlist$raster_files)) {
    r <- raster(rasterlist$raster_files[[i]], sep = "") # loading longlat rasters
    rc <- crop(r, aoi_ext)
    rc_utm <- projectRaster(rc, crs = crs)
    wc_processed[[i]] <- resample(rc_utm, aoi_rr_utm, method = "bilinear")
    name <- paste("processed_", names[[i]], sep = "")
}
wc_processed <- stack(wc_processed)</pre>
```

Global land-cover data are loaded using the readGDAL() function. This function did not seem to create a valid spatial object although its projection is in LongLat. It was thus necessary assign the LongLat to the global raster and then load it as Formal Class Raster using the raster() function. It was then cropped to AoI-size and reprojected in the specified target CRS. To extract the land-cover class values for the calculation of the share each land-cover class has in each target grid cell, the template raster was converted to a Spatial Polygon, using the rasterToPolygons function. Values were extracted from the cropped land-cover raster using the extract function and then saved as list. NA values were set to 0. The class proportions were calculated and saved in a list. The values were then written into copies of the template raster, in order to create a raster stack with one layer for each land-cover class.

6. e) fitting land cover data into template raster

```
# 10. e) Titting land tover data into template laster

# load Tiff, assign CRS, transform it to formal raster class and crop it
message("The following 5 steps can take up to 10 minutes to complete. Time for a cuppa!")
glc <- readGDAL(paste(path.temp_data, "GLCdom/glc_shv10_DOM.tif", sep = "/"))
crs(glc) <- crs(aoi_ext)
glc <- raster(glc)
glc_c <- crop(glc, aoi_ext)
glc_c_utm <- projectRaster(glc_c, crs = crs, method = "ngb")

#delete the massive glc raster
remove(glc)
gc()

# use utm template raster to create spatial polygons object as mask for value extraction is
rastercells <- rasterToPolygons(aoi_rr_utm)
lcraster <- extract(glc_c_utm, rastercells)

# set NA to 0 (to equalise length of land cover pixels per target cell)</pre>
```

```
# calculation of proportion of each class in each target cell
lcsharelist <- list()</pre>
for (i in 1:length(lcraster)) {
  lcshares <- numeric()</pre>
  for (j in 1:11) {
    lcshares[j] <- cbind(length(lcraster[[i]][lcraster[[i]]==j])/length(lcraster[[i]]))</pre>
  lcsharelist[[i]] <- lcshares</pre>
}
# creation of rasterstack from land cover shares
landcovernames <- c("ARTIFICIAL", "CROP", "GRASS", "TREE", "SHRUB", "HERBS", "MANGROVES",
lclayers <- stack()</pre>
for (i in 1: length(landcovernames)) {
  layer <- setValues(aoi_rr_utm, do.call(rbind, lcsharelist)[,i])</pre>
  names(layer) <- landcovernames[i]</pre>
  lclayers <- addLayer(lclayers, layer)</pre>
  As a final step, a csv file containing all data processed for the AoI as well as a correspond-
ing multiband GeoTiff file for control of the correct execution of the Rscript are written into
the working directory.
#----7. SET UP FINAL DATASHEET-----
# combine species distribution raster, WorldCLim stack and land cover stack into one stack
alldata <- addLayer(species_aoi_raster, lclayers, wc_processed)</pre>
# get values and save as dataframe
alldata_df <- as.data.frame(rasterToPoints(alldata))</pre>
# assign remaining column names, species-ID-column, add unique identifier column (CELLCOD)
colnames(alldata_df)[1:2] <- c("EOFORIGIN", "NOFORIGIN")</pre>
final <- cbind(ID = ID, CELLCODE = paste0(target_resolution/1000, "KM", "E", round(alldata_
# export sheet as csv with conditional name into working directory
write.csv(final, paste0("species_", ID,"_", target_resolution/1000, "KM_data.csv"))
# create geotiff of all rasterized data
```

writeRaster(alldata, paste0("species_", ID,"_", target_resolution/1000, "KM_data"), forma-

3 Short-comings

for (i in 1:length(lcraster)) {

lcraster[[i]][is.na(lcraster[[i]])] <- 0</pre>

Unfortunately it was not possible to automise the download of species shapefiles from the IUCN website. The function could be extended to get the questionaire and user sign-up at IUCN into the R console and facilitate the download of according files straight into the working directory. An additional useful functionality of this workflow could be to automatically select the best CRS for any specified study area. However, this would require a lot of programming and it may be more appropriate to leave the CRS-selection to the user, as they would be able to decide on the basis of the conducted study. Areas of interest with extraordinary large extends along the x-axis will make it dificult to define an adequat coordinate reference system, thus making our script unable to run. The user must be aware that equal area projections are limited in their ability to represent truly equal areas when moving away from their center of projection. In case the species ID list gets modified by the IUCN, the

github-hosted .txt-file that relates the latin name to the ID necessary for redirecting the user to the download page would also needed to be manually updated.

We did not provide nor did we research what environmental parameters are relevant or available to analyse marine life. But considering that the IUCN provides shapefiles for marine fish species, species distribution modeling data could be prepared for analysis by our Rscript as well.