# AnaCoDa: Analyzing Codon Data with Bayesian mixture models

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#### **ABSTRACT**

AnaCoDa is an R package for estimating biological parameters, such as selection against translation inefficiency, nonsense error rate, and ribosome pausing time, for sets of genes based on codon data. AnaCoDa provides an adaptive Bayesian MCMC algorithm, fully implemented in C++ for high performance with an ergonomic R interface to improve usability. AnaCoDa employs a generic objectoriented design to allow users to extend the framework and implement their own models for analyzing biological data. Current models implemented in AnaCoDa can accurately estimate biologically relevant parameters given either coding sequences or ribosome foot-printing data. Optionally, AnaCoDa can utilize additional data sources, such as gene expression measurements, to improve aid model fitting and parameter estimation. By utilizing a hierarchical object structure, some parameters can vary between sets of genes while others can be shared. Gene membership in a set can either be pre-assigned or estimated by AnaCoDa. This flexibility allows users to estimate the same model parameter under different biological conditions and categorize genes into different sets based on shared model properties embedded within the data. AnaCoDa also allows users to generate simulated data which can be used to aid model development and model analysis as well as evaluate model adequacy. Finally, Ana-CoDa also comes with contains a set of visualization routines and the ability to revisit or reinitiate previous model fittings, providing researchers with a well rounded easy to use framework to analyze genome scale data.

**Availability:** AnaCoDa is freely available under the Mozilla Public License 2.0 on CRAN (https://github.com/clandere/AnaCoDa).

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### INTRODUCTION

The exponential increase in publicly available genomes over the past decade and the addition of novel technologies produced a vast amount of data for researchers. This influx of raw data necessitates the development of computational tools for extracting biological information. Such tools and models have to be developed and provided in easy to use software to allow researchers to analyze classical sequence data as well as novel data like ribosome footprinting counts. Here, we describe AnaCoDa is an open-source software implemented in R (R Core Team, 2015) that allows researchers to analyze genome-scale data like coding sequences and ribosome foot-printing data using a Bayesian framework. Models described by Gilchrist et al. (2015), Wallace et al. (2013), and Shah and Gilchrist (2011) can be effectively fitted using Ana-CoDa. In addition three currently unpublished models are included as well. The FONSE (first order approximation on nonsense error) model allows for the estimation of selection against of nonsense error rates. The PA (pausing time) and PANSE (pausing time + nonsense error) model allows to estimate ribosome pausing time from ribosome footprinting data, including or ignoring the posibility for nonsense errors. AnaCoDa implements an adaptive Gibbs sampler within a Metropolis-Hastings Monte Carlo Markov Chain (MCMC) approach. This allows for the incorporation of prior knowledge and easy sampling from the posterior distribution to estimate parameter values and quantify the degree of uncertainty in these estimates. Currently, AnaCoDa provides four models to analyze codon counts obtained from coding sequences or ribosome foot-printing experiments. However, AnaCoDa provides a modular infrastructure such that additional genome scale or even phylogenetic models can be integrated. AnaCoDa provides a generic, mixture distribution option to all implemented models, allowing for the estimation of condition gene-set specific parameters or the automatic categorization of data into different sets based on differences in their posterior probabilities of set membership.

The AnaCoDa framework works with AnaCoDa requires gene specific data such as codon frequencies obtained from coding sequences or position specific footprint counts. Conceptually, AnaCoDa uses three different types of parameters. The first type of parameters are gene specific parameters such as gene expression protein synthesis or functionality. Gene-specific parameters are estimated separately for each gene and can vary between potential gene categories or sets. The second type of parameters are gene-set specific

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parameters, such as mutation bias terms or translation error rates. These parameters are shared across genes within a set and can be exclusive to a single set or shared with other sets. While the number of gene sets must be pre-defined by the user, set assignment of genes can be pre-defined or estimated as part of the model fitting. Estimation of the set assignment provides the probability of a gene being assigned to a set allowing the user to asses the uncertainty in each assignment. The third type of parameters are hyperparameters, such as parameters controlling the prior distribution for mutation bias or error rate protein synthesis rate. Hyperparameters can be set specific or shared across multiple sets and allow for the construction and analysis of hierarchical models, by controlling prior distributions for gene or gene-set specific parameters. In order to reduce the effect of the 'curse of dimensionality' on the sampling efficiency of the MCMC chain and allow for flexibility in parallelization, AnaCoDa uses an adaptive Gibbs sampling approach where the MCMC sampling of one parameter type is conditioned on the other two types.

### **FEATURES**

**AnaCoDa** provides an interface written in R, a freely available programming language noted for its ease of use for even inexperienced programmers. As a result, **AnaCoDa** is accessible to researchers with minimal computational experience.

The AnaCoDa interface is designed for quick and efficient data analysis. The interface of AnaCoDa is designed for quick and efficient data analysis. Generally, the only input needed for fitting a model to the data are protein-coding nucleotide codon sequences in the form of a FASTA file or a flat-file containing codon counts obtained from ribosome foot-printing experiments. If available, users may also provide additional types of data such as estimates of gene expression. AnaCoDa can simultaneously utilize the information embedded in these additional data types and/or estimate the error associated with it. AnaCoDa also provides visualization functionality, including plotting functions to compare parameter estimates for different mixture distributions and display codon usage patterns. In addition, diagnostic functions such as those for calculating and visualizing the degree of autocorrelation in MCMC samples the parameter traces are provided.

Robust and efficient model fitting AnaCoDa has built-in features designed to improve the robustness and performance of the implemented MCMC approach. For example, the implemented MCMC approach automatically adapts the proposal width for sampled parameters so such that an user defined acceptance rate intervall is met, improving sampling efficiency of the MCMC and computational performance. Even though AnaCoDa is written in C++, analysis of large datasets and/or complex models can be very computationally intensive. In order to protect users from computer failures or aid in the collection of additional MCMC samples, AnaCoDa can periodically produce output files which can be used to restart an MCMC chain from a previous time point. In addition, AnaCoDa is capable of thinning the MCMC chain, automatically thinnes all parameter traces - meaning only every  $k^{th}$  sample is kept - increasing the effective number of samples and reducing the memory foorprint of AnaCoDa. Thinning increases the effective number of samples by reducing the auto-correlation between samples and reduces the

amount of memory required by the underlying data structures. **Ana-CoDa** is also able to create file R compatible representations of its parameter and MCMC objects. These objects can be loaded into the R environment for model analysis and visualization.

Although AnaCoDa is provided as an R package, the main computational work is implemented in C++. Because R does not provide native C++ support, we used the R package Rcpp which allows for the exposure of Rcpp was employed to expose whole C++ classes as modules to R (Edelbuettel and Francois, 2011). Using Rcpp eliminates time consuming data transfera between the R environment and the C++ core during runs model fittings, resulting in improved computational performance and allowed for a fully object-oriented code design (Booch, 1993). As expected, the runtime of **AnaCoDa** scales linearly with genome size and number of iterations, and polynomial with the number of mixture distributions in the data set. The polynomial increase in the number of mixture distributions is explained by the necessity to estimate the protein production synthesis rate for each gene in each mixture distribution, as it is a gene specific parameter and the probability of a gene being assigned to a mixture has to be conditioned on it.

Data Simulation In addition to model fitting to to fitting the models to actual datasets, AnaCoDa can be used to generate simulated data sets as well. On their own, simulated datasets are useful for model development and analysis. Simulating data under different conditions allows the user to explore model behavior and explore theoretical situation. Different conditions can include the addition or elimination of parameters, or simply allowing a set of parameter values to vary. Fitting models to simulated data can provide users insight into potential pitfalls or shortcomings when fitting observational data and can serve as the basis for evaluating model adequacy of a model fit to observational data (Mi et al., 2015). Significant differences between simulated and observational data suggests the current set of parameters or the model as a whole fail to include or adequately represent biological mechanisms underlying the observational observed data.

Available models AnaCoDa currently provides codon models for analyzing genome scale data. The ROC model implements and extends the codon usage bias (CUB) models developed by Gilchrist et al. (2015); Wallace et al. (2013); Shah and Gilchrist (2011), which can reliably estimate the strength of selection on ribosome overhead cost, mutation bias and allows for the inference of protein synthesis rates. This model allows for the separation of effects of mutation and selection based on gene ordering by protein synthesis rate, and the added addition of a mixture distribution allows for gene clustering based on these effects mutation bias and selection for translation efficiency. In addition to identifying the most efficient codons, ROC provides information on the direction of mutation bias allowing the approximation of mutation rates between codons Gilchrist et al. (2015); Wallace et al. (2013). The ability to estimate protein synthesis rates in the absence of empirical data is useful for investigating CUB of non-model organisms for which such data is lacking and enables the usage of protein synthesis rate in comparative frameworks or other analyses which require information about protein synthesis rates Dunn et al. (2018). Use of the mixture model allows for the investigation of CUB heterogeneity at the genome or gene level. Following the same framework, additional models included in AnaCoDa provide estimates of codon-specific nonsense errors rates (FONSE) and ribosome pausing times (PA and PANSE). Furthermore, **AnaCoDa** implements a ribosome <u>Pausing</u> (PA) model to estimate codon specific ribosome pausing times from ribosome foot-printing data. Parameters estimated with the evolutionary models ROC and FONSE represent evolutionary averages and do not depend on experimental conditions. In contrast, the statistical models PA and PANSE describes the distribution of ribosome pausing times along a gene from ribosome footprinting data. The distribution can be dependent (PANSE) or independend of evidence for nonsense errors in the data.

## **REFERENCES**

- Booch, G. (1993). Object-oriented analysis and design with applications. Benjamin-Cummings Publishing Co, Redwood City.
- Dunn, C., Zapata, F., Munro, C., Siebert, S., and Hejnol, A. (2018). Pairwise comparisons across species are problematic when analyzing functional genomic data. *Proc Natl Acad Sci USA*.

- Edelbuettel, D. and Francois, R. (2011). Rcpp: Seamless r and c++ integration. *Journal of Statistical Software*, **40**, 1–18.
- Gilchrist, M., Chen, W., Shah, P., Landerer, C., and Zaretzki, R. (2015). Estimating gene expression and codon-specific translational efficiencies, mutation biases, and selection coefficients from genomic data alone. *Genome Biology and Evolution*, 7, 1559–1579.
- Mi, G., Di, Y., and Schafer, D. (2015). Goodness-of-fit tests and model diagnostics for negative binomial regression of rna sequenceing data. *PLOS ONE*, **10**, e0119254.
- R Core Team (2015). *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Vienna, Austria.
- Shah, P. and Gilchrist, M. (2011). Explaining complex codon usage patterns with selection for translational efficiency, mutation bias, and genetic drift. *Proc Natl Acad Sci USA*, **108**, 10231–6.
- Wallace, E., Airoldi, E., and Drummond, D. (2013). Estimating selection on synonymous codon usage from noisy experimental data. *Molecular Biology and Evolution*, 30, 1438–1453.