Running head: PLANT-POLLINATOR NETWORK ASSEMBLY

The temporal assembly of plant-pollinator networks following restoration

Lauren C. Ponisio^{1,2}, Marilia P. Gaiarsa³, Claire Kremen¹

- Department of Environmental Science, Policy, and Management University of California, Berkeley 130 Mulford Hall Berkeley, California, USA 94720
- Department of Entomology
 University of California, Riverside
 417 Entomology Bldg.
 Riverside, California, USA
 92521
- Departamento de Ecologia Universidade de Sao Paulo São Paulo, SP, Brazil 05508-900

1 Abstract

TO BE RE-WRITTEN The structure of networks is related to ability of communities to
maintain function in the face of species extinction. Understanding network structure and how
it relates to network disassembly, therefore, is a priority for system-level conservation biology.
We explore the assembly of plant-pollinator communities on native plant restorations in the
Central Valley of California.

- 7 Keywords: changing points, temporal networks, hedgerows, species interactions, preferential at-
- 8 tachment, mutualisms

9 Introduction

Global change has created a severe biodiversity crisis, and as species are lost, so are their interactions (Dunn et al., 2009; Barnosky et al., 2011). Because mutualistic interactions are essential 11 for maintaining the diversity their component guilds of species, these systems are particularly at 12 risk from coextinction cascades. The nature of these cascades will depend on the interaction pat-13 terns within a community (Memmott et al., 2004; Rezende et al., 2007; Bascompte & Stouffer, 2009). Recovering the lost biodiversity and interactions through ecological restoration has become increasingly imperative, and a key restoration aim is to facilitate assembly of robust interaction networks (Menz et al., 2010). We know little, however, about how to re-assemble interacting communities through restoration, or the process of ecological network assembly more generally. The most widely explored mechanism of network assembly, preferential attachment (Barabási & Albert, 1999), predicts that a new species is more likely to interact with species that are already well-connected ("the rich-get-richer" principle, Barabási & Albert, 1999). In pollination systems 21 — a particularly ubiquitous mutualism (Ollerton et al., 2011; Klein et al., 2007) — some studies have found support for this mechanism of assembly. Investigating the day-to-day, temporal assembly of a plant-pollinator network within a season, Olesen et al. (2008) found that new species tended to interact with already well-connected species, potentially because these species are either more abundant or more temporally persistent (Olesen et al., 2008). In addition, using a space-26 for-time substitution to study primary succession along a glacier foreland, Albrecht et al. (2010) 27 found some indication assembly occurred through preferential attachment. Network nestedness, a 28 pattern of interactions where a generalist core interacts with both specialist and generalist species, 29 increased as the community aged (Albrecht et al., 2010). Increasing nestedness could result from 30 preferential attachment process where specialist species attach to the well-connected, generalist 31 core.

In contrast to the ordered network build-up described by preferential attachment, significant reorganizations of interactions can punctuate assembly (Peel & Clauset, 2014). Such significant
reorganizations of interactions, or changing points, are observed in social networks responding to
abrupt shifts in the behavior of interactors (Peel & Clauset, 2014). In ecological communities,
such shifts may occur if, as new species colonize, resident species change their interaction partners to optimize their foraging effort. In plant-pollinator communities, theory predicts pollinators
optimize their use of floral resources to reduce interspecific competition and improve resource-use
efficiency (Pyke, 1984; Valdovinos *et al.*, 2010, 2013; Albrecht *et al.*, 2010; Blüthgen *et al.*, 2007).
No studies, however, have examined whether changing points occur during ecological network
assembly, and how these changes relate to the behavior of the interactors.

Understanding network assembly is particularly relevant to ecological restoration, which is essentially 'applied succession' (Parker, 1997, e.g.,). In pollination systems, the time since an area was restored has been shown to affect the structure of networks (Forup *et al.*, 2008a,b; Devoto *et al.*, 2012), suggesting interactions are evolving as the community develops. Understanding the mechanisms of network assembly will help to guide community restoration.

Facilitating effective restoration of networks is especially imperative in areas where species in-48 teractions provide essential ecosystem services, such as crop pollination. In intensively managed 49 agricultural landscapes, the demand for pollination services is the greatest (Kremen, 2008). How-50 ever, honey bees, managed extensively around the world to provide crop pollination, are in global 51 decline (Neumann & Carreck, 2010; van Engelsdorp et al., 2009). In addition, native pollina-52 tors, which are capable of providing sufficient crop pollination (Kremen et al., 2002; Winfree 53 et al., 2007; Kremen et al., 2004), are in short supply because these landscapes make poor habitats 54 for pollinator populations (Kremen et al., 2002). To ensure provision the continued provision of ecosystem services and curb biodiversity loss, effective restoration of pollinators and their interactions in agricultural landscapes is critical.

To promote pollinator services in agriculture, farmers are increasingly turning to the habitat restoration technique of planting strips of native plants along farm edges (hedgerows) to help provide habitat for pollinators without removing arable land from production. Hedgerows augment the richness, abundance and trait diversity of pollinators in agricultural landscapes(Morandin & Kremen, 2013; M'Gonigle *et al.*, 2015; Kremen & M'Gonigle, 2015; Ponisio *et al.*, 2015). In addition, hedgerows promote the persistence and colonization of floral resource specialists (M'Gonigle *et al.*, 2015). Little is known, however, about the assembly of the network following hedgerow restoration.

Using a long-term dataset of plant-pollinator communities assembling following hedgerow restora-66 tion in the highly simplified and intensively managed agricultural landscape of California's Central 67 Valley, we explore the process of network development. We first determine whether the mechanism underlying network assembly is a smooth build up of interactions as would be predicted by 69 preferential attachment, or punctuated by significant reorganizations of interactions (i.e., network 70 changing points). Even with changing points in interaction organization, networks could still be assembling via preferential attachment if the network reorganizations were primarily driven the by peripheral, temporally variable species while a stable, well-connected core of species still persists. We test whether the species are most variable in their network position — and thus important contributors network reorganizations — are less persistent and connected species. To further explore the mechanisms underlying the temporal dynamics in the networks, we examine patterns in the species and interaction temporal turnover. Lastly, we investigate whether networks are assembling toward predictable interaction patterns, and the ramifications for the robustness of the networks to species extinction and cascading perturbations.

Materials & Methods

81 Study sites and collection methods

We surveyed plant-pollinator interaction networks of assembling hedgerows (N=5), as well as in 82 two types of non-assembling communities to serve as controls: unrestored, weedy field margins 83 (N=19) and established hedgerows (greater than 10 years since planting, N=29). The sites were 84 located in the Central Valley of California in Yolo, Colusa and Solano Counties. This area is comprosed of intensively managed agriculture – primarily monocultures of conventional row crops, vineyards and orchards. Hedgerows we planted along field margins where they do not remove valuable land from production. Plantings are between are ca. 3–6 m wide and approximately 350 m long, bordering large (ca. 30-hectare) crop fields. Hedgerows consist of native, perennial, shrub and tree plantings including Rosa californica, Cercis occidentalis, Ceanothus spp., Heteromeles arbutifolia, Sambucus mexicana, Eriogonum spp., Baccharis spp., Salvia spp. and others (Kremen & M'Gonigle, 2015; M'Gonigle et al., 2015). The mean distance between monitoring sites was 15 km, and the minimum distance between sites of the same type sampled in the same year was 2 km. The entire area surveyed spanned almost 300 km². The crop fields adjacent to all sites were similarly managed as intensive, high-input monoculture.

Monitoring began in 2006 and continued through 2015. Surveys of the assembling hedgerows began the year before the area was restored. For logistical reasons, no sampling of assembling hedgerows was conducted in 2010. Sites were sampled between two and five times per year (Tables S1-S3). In each round of sampling, the order in which sites were sampled was randomized.

Surveys were conducted under sunny conditions when the temperature was above 21°C and wind speed was below 2.5 meters/second.

Flower-visitors to plants in hedgerows and unrestored controls were netted for one hour of active search time (the timer was paused when handling specimens). Honeybees (*Apis mellifera*) were

not collected because their abundance is determined largely by hive placement by bee-keepers.

All other insect flower visitors that touched the reproductive parts of the flower were collected;

however, here we focus only on wild bees and syrphids — the most abundant and effective polli
nators in the system (representing 49 and 19 percent of records, respectively, C. Kremen, A. Klein

and L. Morandin, unpublished data). Bee and syrphid specimens were identified to species (or

morpho-species for some bee specimens in the genera *Nomada* and *Sphecodes*) by expert tax
onomists.

Quantitative networks were generated for each site through time. To account for the unequal number of surveys between years (Tables S1-S3), we use the mean of the interactions between a pair of plants and pollinators across surveys within a year to represent interaction frequency.

114 Change point analysis

115 Identifying change points

We employed a change point detection method (Peel & Clauset, 2014) to identify fundamental 116 reorganizations in large-scale interaction patterns. A change point is caused by a merge, split, 117 fragmentation or formation of communities (also called modules or compartments). Change point 118 detection methods have yet to be generalized to quantitative networks, so for this analysis we 119 focused on qualitative (binary) networks. Following Peel & Clauset (2014), we first defined a 120 probability distribution over the networks using the generalized hierarchical random graph model 121 (GHRG). The GHRG model is able to capture both assortative and disassortative community struc-122 ture patterns at all scales in the network (Peel & Clauset, 2014). A network G is composed of vertices V and edges $E \subseteq VV$. The GHRG model decomposes the N vertices into a series of nested groups, the relationships among which are represented by the dendrogram T. The tips of T are the 125 vertices of G, and the probability that two vertices u and v connect is given by the parameter p_r . The probability distribution of the network G thus defined as:

$$P(G|T, pr) = p_r^{E_r} (1 - p_r)^{N_r - E_r}$$
(1)

Where E_r is the observed number of edges between vertices with the common ancestor r, and N_r is the total possible edges.

Using Bayesian posterior inference and techniques from phylogenetic tree reconstruction, we fit the GHRG model to the networks (Peel & Clauset, 2014). This is accomplished by using a Markov chain Monte Carlo (MCMC) procedure to first sample the posterior distribution of bipartitions, from which a consensus tree is derived (Peel & Clauset, 2014). We use β distributions with the hyperparameters $\alpha = \beta = 1$ to define priors for p_r .

Once the GHRG model has been fit to the networks, we determine whether a change point occurred 135 between two time slices. To detect a change point, we compare the fit of two models — one where a change point occurred between two networks, and one where no change occurred using posterior Bayes factors. We chose a sliding window of length, w, of four, within which to 138 find change points. Larger windows allow for more gradual changes, and four was the maximum 139 possible with our maximum of nine years of data. Lastly, to calculate a p-value for the Bayes 140 factors, we use parametric bootstrapping to numerically estimate the null distribution (Peel & 141 Clauset, 2014). We emploied code published online by L. Peel for the change point analysis. 142 Analyses we conducted in Python 3.4. 143

We next test whether the change points occurring in maturing hedgerows were a component of
the assembly process or a product of environmental shifts that lead to network reorganizations
in all types of communities. We model the number of change points as successes and the total
number of years each site was sampled as trails, and use a generalized linear model with Binomial
error to test whether the probability of a change point occurring varied by site type. We used

standard techniques to determine whether the assumptions of the models were met for this and all subsequent models. For the non-assembling hedgerows and weedy field margins, only sites with five or greater survey years were included in this analysis. All statistical analysis were conducted in R 3.2.3 (R Core Team, 2015).

153 Characteristics of species that contribute to change points

To further elucidate the nature of the change points, we examine the characteristics of the species that contributed interaction reorganization. Some species remain in relatively similar network positions through time, whereas others are more variable in their role and thus contribute more strongly to network reorganization. We test the that more persistent species with the highest degree are the most stable in their network position, as would be expected is the networks were assembling via preferential attachment.

We calculate species persistence as the proportion of surveys a plant or pollinator is observed. 160 Species observed consistently within and between years are thus maximally persistent. Weighted 161 species degree is calculated from interaction observations from an extensive dataset from Yolo 162 County (approx. 18000 interaction records) that included both the data included in this study and 163 additional data from sites where we collected flower visitors using the same methods (M'Gonigle 164 et al., 2015; Ponisio et al., 2015). To represent network position variability, we computed the 165 coefficient of variation of weighted closeness at each site through time. Closeness describes the centrality of a species in the network by calculating path lengths to other vertices (species) in the graph. We use linear mixed models to test whether the species closeness variability (log) is related to the persistence or degree of that species (Bates et al., 2014; Kuznetsova et al., 2014). 169 We included random effects for species, as well as site. Because the degree and persistence of pollinators were strongly correlated, ($\rho = 0.84$, p-value $< 2 * 10^{-16}$), we include each explanatory 171 variable in the model separately. Plant degree and persistent were not significantly correlated, but

we use the same models as we did for the pollinators for consistency. A linear increase in closeness, as might be expected with assembly by preferential attachment, would also lead to a high variability in closeness scores, so we test whether closeness increases through time.

Species and interaction turnover

Reorganizations of network structure can be the result of species turnover or species changing their interaction partners (i.e., re-wiring). To better understand the mechanisms underlying the temporal dynamics of the assembling networks, we examined patterns of pollinator species and interaction turnover. Assembling networks may have higher rates of species turnover than non-assembling communities because new species are colonizing and establishing themselves (M'Gonigle *et al.*, 2015). Similarly, if pollinators try to maximize their foraging efficiency based on the species present, interactions may turnover more quickly than in established communities.

To estimate the temporal species and interaction turnover, we use an approach similar to calculat-184 ing spatial β -diversity. Instead of calculating the variation in community composition across sites 185 within a year, however, we estimated turnover across years at a site. We first calculated the pair-186 wise dissimilarity of species and interactions between years within each site using a dissimilarity 187 estimator that incorporates abundances, while also accounting for unobserved records (Chao et al., 2005). Dissimilarity estimates can be affected by the total number of species and individuals sampled at a site (e.g., Ponisio et al., 2015). For example, the probability that two sites do not share any species is higher when there are few individuals at those sites. Following Ponisio et al. (2015), 191 we use null models that constrained species richness to estimate the deviation of the observed dis-192 similarity from that which would be expected under a random community assembly process. With 193 the corrected dissimilarity values, we then calculated the multivariate dispersion of community 194 composition across years (Anderson et al., 2011). In order to test whether assembling hedgerows 195 had more turnover of species and interactions than non-assembling communities, the species and 196

interaction temporal turnover estimates were response variables in a linear mixed models with site type as an explanatory variable along with random effects for both year and site (Bates *et al.*, 2014; Kuznetsova *et al.*, 2014).

Though species may turnover across years, some groups of species may essentially replace each 200 other if they fill similar roles in the network. At non-assembling communities, species turnover 20 may overestimate the temporal changes in the networks if the interactions occurring in one year are similar to those in the next year when they are weighted by the similarity of their constituent 203 species. We develope a method examine the temporal turnover of interactions weighted based 204 on their similarity. We followed Ahn et al. (2010) algorithm to hierarchically cluster all the in-205 teractions (edges) across sites and years based on their similarity, and build a dendrogram. The 206 interaction similarity is based how may plants and pollinators (vertices) two edges share (Ahn 207 et al., 2010; Kalinka & Tomancak, 2011). The more species edges shared in common, the shorter 208 the branch length between them on the dendrogram. We next calculated the temporal turnover 209 of interactions weighted by their similarity, as approximated by "phylogenetic" distance (Graham 210 & Fine, 2008; Kembel et al., 2010). When then use linear models to test whether the weighted 211 turnover of interactions varied between assembling and non-assembling networks (Bates et al., 212 2014; Kuznetsova et al., 2014).

Temporal changes in interaction patterns

Network structure

Any changing points in network structure may contribute to the reorganization of the assembling networks into predictable interaction patterns. Pollination networks exhibit two main structural patterns — modularity (e.g., Olesen *et al.*, 2007) and nestedness (e.g., Bascompte *et al.*, 2006,

2003). In modular networks, interactions are insular, occurring within separate groups or "modules" more often than between modules. Communities in the network may fragment as the network assembles, enhancing modularity. Conversely, nested networks are like a pyramid of interactions, 221 where there are some species that interact with many species, other species that interact with a 222 subset of those species, and so on. If species entering the network tend to interact with the gen-223 eralist base of the network pyramid (i.e., via preferential attachment), nestedness would increase 224 through time. The connectance — the proportion of observed out of possible interactions — is 225 also expected to increase as species preferentially attach to the core. The network specialization 226 may also change as the community assembles. If specialist species colonize the network or species 227 begin to limit their interaction niche breath as the network assembles, network-level specialization 228 will increase (Blüthgen et al., 2006). 229

To evaluate network nestedness, we used the estimator weighted NODF citepnodf. NODF evalu-230 ates whether species with fewer partners interact with subsets of partners with which more con-231 nected species interact (Almeida-Neto et al., 2008). To estimate modularity, we use a hierarchical 232 clustering algorithm (Newman & Girvan, 2004; Csardi & Nepusz, 2006). We calculated standard-233 ized z-scores so that nestedness and modularity metrics could be compared between communities. 234 The z-scores were calculated by generating an ensemble of 999 randomly assembled communities, 235 subtracting the mean of the statistic calculated across these communities from the observed value, 236 and then dividing by the standard deviation. To assemble random communities, we reshuffled 237 the interactions between species but fixed the total number of interactions, species and interaction 238 frequency distributions (Galeano et al., 2009). We evaluate network specialization with the metric 239 H2, which estimate the deviation of the observed interaction frequency between plants and pollina-240 tors from a null expectation where all partners interact in proportion to their abundances (Blüthgen et al., 2006). It ranges from zero for generalized networks to one for specialized networks.

To test whether network modularity, nestedness, connectance or specialization changed linearly

with assembly, we use linear mixed models with the descriptive network metrics as the response variable, year of assembly as the explanatory variable, and random effects of site and year. The number of species in a network affects affects the patterns of interaction possible, so we also examined the change in plant and pollinator species richness through time. We employ generalized linear mixed models with Poisson error to model richness. We scaled explanatory variables.

Network robustness

Lastly, we test the changes in interaction patterns associated with network assembly affect the robustness of the network to species loss and to cascading perturbations. Following Memmott *et al.* (2004), we simulate plant species extinction and the subsequent extinction cascades of pollinator species. Because the reproduction of plant species if facilitated by active restoration efforts, it is unlikely the extinction of pollinator species would affect plant populations in the hedgerows. However, plants ceasing to bloom, for example in response to drought, will likely affect the pollinators that depend on them. We eliminate plants species based on their degree or abundance, and the number of pollinators that secondarily went extinct is calculated. The area below the extinction curve is an estimate of network robustness (Dormann *et al.*, 2008).

We also explored how the robustness to cascading perturbations changed as community assembled, using algebraic connectivity — the second smallest eigenvalue of the Laplacian matrix (Fiedler, 1973) — as a proxy for network robustness (Gaiarsa et al., submitted). Algebraic connectivity relates to how difficult it is to turn a network into completely disconnected groups of vertices, or species (Costa *et al.*, 2007) — the larger the algebraic connectivity, the more robust a network is to cascading perturbations (Gaiarsa et al., submitted), and the harder it is to break the community into isolated groups of species.

266 Results

Over eight years and 747 samples, we collected and identified 19,547 wild bees and syrphids comprising 173 species from 50 genera. We observed 1,521 unique interactions between plants and pollinators.

270 Change point analysis

Identifying change points

The majority (76%) of the sites underwent at least one significant interaction reorganization (Fig. 1).

All five of the assembling hedgerows experienced changing points, whereas only 40% and 81% of non-assembling hedgerows and field margins, respectively, underwent significant interaction reorganizations. Assembling hedgerows experienced significantly more changing points than the non-assembling networks (estimate of the difference in the odds ratios between assembling and non-assembling networks, 3.316, 95% CI [1.314, 8.572], *p*-value= 0.0117). Network assembly following restoration is thus punctuated by more interaction reorganizations than would be expected by environmental shifts that would effect assembling and non-assembling networks equally.

280 Characteristics of species that contribute to change points

In contradiction to the predictions of assembly by preferential attachment, both pollinator persistence and degree were positively related to network position variability (estimate of the slope of closeness variability and persistence \pm standard error of the estimate, 0.653 ± 0.225 , p-value=0.009; slope of closeness variability and degree, 0.008 ± 0.002 , p-value='0.002). In addition, plant persistence and degree were not significantly related to network position variability (estimate of the slope of closeness variability and persistence \pm standard error of the estimate, -2.063 ± 3.091 , p-value='0.002 at 3.091, p-value='0.003 at 3.091, p-val

value=0.5; slope of closeness variability and degree, 0.0018 \pm 0.002, p-value=0.3). The variability of species network position was not the result of closeness linearly increasing through time, and, in fact, plant and pollinator closeness decreased slightly through time (Fig. S1, estimate of the slope of closeness through time \pm standard error of the estimate, pollinators: -0.0003 ± 0.00005 , p-value=2.7 * 10^{-12} ; plants -0.007 ± 0.001 , p-value=1.4 * 10^{-6}). Through statistically significant, the slopes are so slight they may not me biologically significant.

Species and interaction turnover

The rates of species and interaction temporal turnover similar across assembling hedgerows, non-294 assembling hedgerows and field margins, though mature hedgerows has marginally significantly 295 less species turnover than field margins (estimate \pm standard error of the estimate of the differ-296 ence in turnover between field margins and mature hedgerows, -0.0498 ± 0.026 , p-value=0.058). 297 When interactions where weighted by their similarity, both assembling and mature hedgerows had higher rates of turnover than field margins (estimate \pm standard error of the estimate of the difference in turnover between field margins and assembling hedgerows, 0.115 ± 0.027 , p-300 value=0.0002; field margins and mature hedgerows, 0.082 ± 0.024 , p-value=0.002). The weighted 301 interaction turnover at assembling hedgerows, however, was not significantly higher than in non-302 assembling, mature hedgerows. 303

Temporal changes in interaction patterns

Network structure

Network nestedness significantly increased with assembly (estimate of the slope of nestedness through time \pm standard error of the estimate, 1.834 \pm 0.6142, *p*-value=0.022, Fig. 4). Modularity

decreased (Fig. 4), though the slope was not significantly different from zero (estimate of the slope of modularity through time \pm standard error of the estimate, -0.524 ± 0.295 , p-value=0.124). Connectance decreased as the community assembled (Fig. 4, estimate of the slope of connectance through time \pm standard error of the estimate, -0.0434 ± 0.0152 , p-value=0.03). In contrast, specialization remained constant (estimate of the slope of specialization through time \pm standard error of the estimate, 0.003 ± 0.015 , p-value=0.827).

Both plant and pollinator species richness increased through time (Fig. 4, estimate of the slope of richness through time \pm standard error of the estimate, pollinators: 0.193 ± 0.0729 , p-value=0.008; plants: 0.212 ± 0.0653 , p-value=0.001). Pollinator species are colonizing and persisting at the assembling hedgerows. Though some new plant species may eshablish themselves in the hedgerows, because the plant species richness is based on the flowers in the network and not the presence of a plant at a site, the increase in plant richness is likely due to plants attracting more visitors as they offer better rewards.

21 Network robustness

Assembly did not effect the robustness of the networks to species extinction when species where removed incrementally by degree (estimate of the slope of robustness through time \pm standard error of the estimate, $6*10^{-5} \pm 4*10^{-3}$, p-value=0.987) or abundance (0.001 \pm 0.003, p-value=0.65, Fig. 5).

In contrast, the robustness of networks to cascading perturbations, as measured by the algebraic connectivity of the network, increased as the network assembled (estimate of the slope of robustness through time \pm standard error of the estimate, 0.6814 \pm 0.272, p-value=0.042, Fig. 5).

Discussion

We show that the temporal assembly of plant-pollinator networks following restoration is a highly 330 dynamic process where interactions often undergo significant reorganizations. These network or-331 ganizations are unlikely to be a product of environmental forces alone because the network chang-332 ing points in non-assembling communities are less frequent, and there are few consistent trends 333 in when change points occurred across all sites. Several sites had changing points between year 334 2009 and 2011 (Fig. 1). In California, 2011 marked the beginning of a multi-year drought. In the 335 assembling hedgerows were not sampled in 2010, so disentangling whether the changing points 336 are due to skipping a year of assembly or the drought is not possible. Interestingly, most of the 337 assembling hedgerows did not undergo a significant reorganization of interactions immediately the hedgerow was planted (i.e., the transition from weedy field margin to hedgerow). This result is consistent with the finding that in our study system, hedgerow restoration takes several years to have a significant impact on the plant-pollinator communities (Kremen and M'Gonigle, in prep). Given that several changing points in network organization occurred during assembly, we next 342 explored the species most likely responsible for the shifts in interaction patterns. Based on a preferential attachment-like mechanism, we expect that the most persistent and high degree species would remain stable in the network core during assembly, and would thus contribute the least to the changing points in network organization. Surprisingly however, we encountered the opposite: 346 the species that were most variable in their network position and thus contributed most to network 347 reorganizations where species with the highest degrees (i.e., most generalized) and persistence. 348 For example, the five most ubiquitous species in our study landscape — Halictus ligatus, Halictus 349 tripartitus, Lasioglossum (Dialictus) incompletum, and Toxomerus marginatus — were the only 350 species that changed what module (i.e., community), and they were also present in across years in 351 all of the assembling hedgerows. Because species degree and persistence were strongly correlated, 352 it is difficult to disentangle the causal mechanism for why species with those characteristics are so variable in their network position. Generalized species may be able to better exploit the limited
floral resources in the intensively managed agriculture landscape, and thus also most persistent.
More persistent species also have longer phenologies, so they have the opportunity to visit many
different flowers, resulting in a higher degree. Either way, our result suggests that adaptable species
can change their network position to utilize the most advantageous floral resources available, which
may depend on the other pollinator species that are present and the state of the floral resource. Thus
given the opportunity and ability to use different resources, species will often change their network
positions.

The frequent changing points in network organization, dynamic nature of the location of species in 362 networks, and turnover of species and interactions all point to an assembly mechanism other than 363 preferential attachment. Nestedness did increase with years post restoration, as would be expected 364 if colonizing, specialist species attached to a central, generalist core Albrecht et al. (2010). With 365 preferential attachment, however, we would also expect connectance and specialization to increase, 366 and we found no such trends. The stable level of network-level specialization through the assembly 367 process may be due to the increased colonization of specialized species (M'Gonigle et al., 2015) 368 accompanied by an increase in the diet breath of resident species. This would be expected if 369 resident species were able to minimize their foraging time by expanding their diet breath as plant 370 diversity increases with hedgerow maturation (Waser et al., 1996; Pyke, 1984; Blüthgen et al., 371 2007; Albrecht et al., 2010). Such a change in pollinator behavior would also explain the increase 372 in network nestedness. 373

The changes in network patterns with assembly also increased the robustness of the networks to cascading perturbations – providing further evidence that hedgerows are valuable tool for promoting species conservation and ecosystem provision in agricultural areas (M'Gonigle *et al.*, 2015; Ponisio *et al.*, 2015; Kremen & M'Gonigle, 2015). Because the vulnerability of the network to cascading perturbations, as measured by algebraic connectivity, is correlated with species richness,

the increase and plant and pollinator richness following restoration is at least partially responsible for enhancing network robustness to cascading effects. The relationship between diversity and 380 stability in networks has been the subject of considerable debate (e.g., May, 1972; Pimm, 1984; 38 Montoya et al., 2006). Our results provides one of the few empirical examples of how restoring 382 species diversity contributes to enhancing network stability. Though these hedgerows were de-383 signed to promote floral resources across the growing season and not specifically to promote any 384 specific network properties (Menz et al., 2010), the pollinators assembled into resilient interaction 385 patterns. With prior knowledge of the floral preferences of pollinators, future restoration efforts 386 may also be able to incorporate promoting network stability into planning efforts (M'Gonigle et al., 387 2016) 388

Interestingly, however, the robustness of hedgerow communities to species loss did not change with assembly. This is particularly surprising given the observed increase in nestedness, which is often associated with an enhanced in robustness to extinction (Memmott *et al.*, 2004). It may be that assembling hedgerows have not yet reached a sufficient level of nestedness to realize its benefits. Nestedness of the assembling hedgerows, however, did not asymptote within the eight years following restoration that the sites were surveyed, so hedgerow networks may eventually reach sufficient levels of nestedness to gain the robustness advantage.

Plant-pollinator networks in general are highly dynamic, with high turnover of species and inter-396 actions both within and between seasons (Burkle & Alarcón, 2011). Though our non-assembling 397 communities experience fewer network reorganizations than the assembling hedgerows, 82% of 398 field margins and 40% of mature hedgerows underwent at least one changing point in network 399 structure. Pollinators are also highly opportunistic (Petanidou et al., 2008; Vázquez, 2005; Al-400 brecht et al., 2010), though trait complementarity such as tongue length and corolla depth impose 401 some biophysical limits to the interactions between plants and pollinators (Vázquez et al., 2009; 402 Vázquez et al., 2009; Stang et al., 2009, 2006; Santamaría & Rodríguez-Gironés, 2007). Such 403

opportunism may buffer plant-pollinator communities from global change (e.g., Ramos-Jiliberto et al., 2012; Kaiser-Bunbury et al., 2010), but our limited understanding of the assembly of these 405 communities impedes making such predictions (Vázquez et al., 2009; Burkle & Alarcón, 2011). 406 Unlike in the broader food web literature, we have few mechanistic models of mutualistic net-407 work assembly (Valdovinos et al., 2013). In addition, the few that have been developed often 408 borrow their mechanisms from competitive interactions, leading to inaccurate biological assump-409 tions (Holland et al., 2006). We need further development of mechanistic models of mutualistic 410 system to generate testable predictions, along with empirical exploration of network assembly. 411 Plant-pollinator communities and mutualisms in general are vital for the maintenance of biodiver-412 sity and the provision of essential ecosystem services. We must therefore understand the processes 413 underlying their assembly to facilitate restoration and conservation.

15 Acknowledgments

We would like to thank Paulo Guimarães and Aaron Clauset for their invaluable discussions, and
Leto Peel for help with the change point analysis. We thank the growers and land owners that allowed us to work on their property. We also greatly appreciate the identification assistance of expert
taxonomists Martin Hauser, Robbin Thorp and Jason Gibbs. This work was supported by funding
from the Army Research Office (W911NF-11-1-0361 to CK), the Natural Resources Conservation
Service (CIG-69-3A75-12-253, CIG-69-3A75-9-142, CIG-68-9104-6-101 and WLF-69-7482-6277 to The Xerces Society), the National Science Foundation (DEB-0919128 to CK), The U.S.
Department of Agriculture (USDA-NIFA 2012-51181-20105 to Michigan State University). Funding for LCP was provided by an NSF Graduate Research Fellowship and the USDA NIFA Graduate
Fellowship. Funding for MPG was provided by São Paulo Research Foundation (FAPESP, grant
2013/13319-5).

27 References

- Ahn, Y.Y., Bagrow, J.P. & Lehmann, S. (2010). Link communities reveal multiscale complexity in networks. *Nature*, 466, 761–764.
- Alarcón, R., Waser, N.M. & Ollerton, J. (2008). Year-to-year variation in the topology of a plant–pollinator interaction network. *Oikos*, 117, 1796–1807.
- Albrecht, M., Riesen, M. & Schmid, B. (2010). Plant–pollinator network assembly along the chronosequence of a glacier foreland. *Oikos*, 119, 1610–1624.
- Almeida-Neto, M., Gumarães, P., Gumarães, P.R., Loyola, R.D. & Ulrich, W. (2008). A consistent
- metric for nestedness analysis in ecological systems: reconciling concept and measurement.
- 436 Oikos, 117, 1227–1239.
- Anderson, M.J., Crist, T.O., Chase, J.M., Vellend, M., Inouye, B.D., Freestone, A.L., Sanders,
- N.J., Cornell, H.V., Comita, L.S., Davies, K.F. et al. (2011). Navigating the multiple meanings
- of β diversity: a roadmap for the practicing ecologist. *Ecol. Lett.*, 14, 19–28.
- Barabási, A.L. & Albert, R. (1999). Emergence of scaling in random networks. *science*, 286, 509–512.
- Barnosky, A.D., Matzke, N., Tomiya, S., Wogan, G.O., Swartz, B., Quental, T.B., Marshall, C.,
- McGuire, J.L., Lindsey, E.L., Maguire, K.C. et al. (2011). Has the earth/'s sixth mass extinction
- already arrived? *Nature*, 471, 51–57.
- Bascompte, J., Jordano, P., Melián, C.J. & Olesen, J.M. (2003). The nested assembly of plantanimal mutualistic networks. *Proc. Natl. Acad. Sci. USA*, 100, 9383–9387.
- Bascompte, J., Jordano, P. & Olesen, J.M. (2006). Asymmetric coevolutionary networks facilitate biodiversity maintenance. *Science*, 312, 431–433.

- Bascompte, J. & Stouffer, D.B. (2009). The assembly and disassembly of ecological networks.
- 450 Phil. Trans. R. Soc. B, 364, 1781.
- Bates, D., Maechler, M., Bolker, B. & Walker, S. (2014). *lme4: Linear mixed-effects models using*
- Eigen and S4. R package version 1.1-7.
- Blüthgen, N., Menzel, F. & Blüthgen, N. (2006). Measuring specialization in species interaction
- networks. BMC Ecol., 6, 9.
- Blüthgen, N., Menzel, F., Hovestadt, T., Fiala, B. & Blüthgen, N. (2007). Specialization, con-
- straints, and conflicting interests in mutualistic networks. *Curr. Biol.*, 17, 341–346.
- Burkle, L.A. & Alarcón, R. (2011). The future of plant-pollinator diversity: Understanding in-
- teraction networks across time, space, and global change. American Journal of Botany, 98,
- 459 528.
- 460 Chao, A., Chazdon, R.L., Colwell, R.K. & Shen, T.J. (2005). A new statistical approach for
- assessing similarity of species composition with incidence and abundance data. *Ecol. Lett.*, 8,
- 462 148–159.
- Costa, L.d.F., Rodrigues, F.A., Travieso, G. & Villas Boas, P.R. (2007). Characterization of com-
- plex networks: A survey of measurements. Advances in physics, 56, 167–242.
- ⁴⁶⁵ Csardi, G. & Nepusz, T. (2006). The igraph software package for complex network research.
- InterJournal, Complex Systems, 1695.
- ⁴⁶⁷ Devoto, M., Bailey, S., Craze, P. & Memmott, J. (2012). Understanding and planning ecological
- restoration of plant–pollinator networks. *Ecology letters*, 15, 319–328.
- Díaz-Castelazo, C., Guimaraes Jr, P.R., Jordano, P., Thompson, J.N., Marquis, R.J. & Rico-Gray,
- V. (2010). Changes of a mutualistic network over time: reanalysis over a 10-year period. *Ecol-*
- *ogy*, 91, 793–801.

- Dormann, C., Gruber, B. & Fründ, J. (2008). Introducing the bipartite package: analysing ecological networks. *R News*, 8, 8.
- Dunn, R.R., Harris, N.C., Colwell, R.K., Koh, L.P. & Sodhi, N.S. (2009). The sixth mass coextinc-
- tion: are most endangered species parasites and mutualists? *Proceedings of the Royal Society*
- of London B: Biological Sciences, 276, 3037–3045.
- van Engelsdorp, D., Evans, J., Saegerman, C., Mullin, C., Haubruge, E., Nguyen, B., Frazier, M.,
- Frazier, J., Cox-Foster, D., Chen, Y., Underwood, R., Tarpy, D. & Pettis, J. (2009). Colony
- collapse disorder: A descriptive study. *Plos One*, 4, 1–17.
- Fang, Q. & Huang, S.Q. (2012). Relative stability of core groups in pollination networks in a biodiversity hotspot over four years. *PloS one*, 7, e32663.
- Fiedler, M. (1973). Algebraic connectivity of graphs. *Czechoslovak mathematical journal*, 23, 298–305.
- Forup, M., Henson, K., Craze, P. & Memmott, J. (2008a). The restoration of ecological interactions: plant-pollinator networks on ancient and restored heathlands. *J. Appl. Ecol.*, 45, 742–752.
- Forup, M.L., Henson, K.S., Craze, P.G. & Memmott, J. (2008b). The restoration of ecological
- interactions: plant-pollinator networks on ancient and restored heathlands. Journal of Applied
- 488 *Ecology*, 45, 742–752.
- Galeano, J., Pastor, J.M. & Iriondo, J.M. (2009). Weighted-interaction nestedness estimator (wine):
- a new estimator to calculate over frequency matrices. Environmental Modeling & Software, 24,
- 491 1342–1346.
- Graham, C.H. & Fine, P.V. (2008). Phylogenetic beta diversity: linking ecological and evolutionary processes across space in time. *Ecology letters*, 11, 1265–1277.

- Holland, J.N., Okuyama, T. & DeAngelis, D.L. (2006). Comment on asymmetric coevolutionary networks facilitate biodiversity maintenance. *Science*, 313, 1887b–1887b.
- Kaiser-Bunbury, C.N., Muff, S., Memmott, J., Müller, C.B. & Caflisch, A. (2010). The robustness
- of pollination networks to the loss of species and interactions: a quantitative approach incorpo-
- rating pollinator behaviour. *Ecology Letters*, 13, 442–452.
- Kalinka, A.T. & Tomancak, P. (2011). linkcomm: an r package for the generation, visualization, and analysis of link communities in networks of arbitrary size and type. *Bioinformatics*, 27.
- Kembel, S., Cowan, P., Helmus, M., Cornwell, W., Morlon, H., Ackerly, D., Blomberg, S. &
- Webb, C. (2010). Picante: R tools for integrating phylogenies and ecology. *Bioinformatics*, 26,
- 1463–1464.
- Klein, A., Vaissière, B., Cane, J., Steffan-Dewenter, I., Cunningham, S., Kremen, C. & Tscharntke,
- T. (2007). Importance of pollinators in changing landscapes for world crops. *Proc. R. Soc. B*,
- 506 274, 303–313.
- Kremen, C. (2008). *Bee Pollinators in Agricultural Ecosystems*, New York: Oxford University

 Press, chap. Crop Pollination Services From Wild Bees, pp. 10–26.
- Kremen, C. & M'Gonigle, L.K. (2015). Small-scale restoration in intensive agricultural landscapes supports more specialized and less mobile pollinator species. *J. Appl. Ecol.*, 52, 602–610.
- Kremen, C., Williams, N., Bugg, R., Fay, J. & Thorp, R. (2004). The area requirements of an ecosystem service: crop pollination by native bee communities in California. *Ecol. Lett.*, 7, 1109–1119.
- Kremen, C., Williams, N. & Thorp, R. (2002). Crop pollination from native bees at risk from agricultural intensification. *Proc. Natl. Acad. Sci. U.S.A.*, 99, 16812–16816.

- Kuznetsova, A., Bruun Brockhoff, P. & Haubo Bojesen Christensen, R. (2014). *ImerTest: Tests*
- for random and fixed effects for linear mixed effect models (lmer objects of lme4 package). R
- package version 2.0-11.
- May, R.M. (1972). Will a large complex system be stable? *Nature*, 238, 413–414.
- Memmott, J., Waser, N.M. & Price, M.V. (2004). Tolerance of pollination networks to species
- extinctions. *Proc. R. Soc. Lond. B*, 271, 2605–2611.
- Menz, M., Phillips, R., Winfree, R., Kremen, C., Aizen, M., Johnson, S. & Dixon, K. (2010).
- Reconnecting plants and pollinators: challenges in the restoration of pollination mutualisms.
- 524 Trends Plant Sci., 16, 4–12.
- M'Gonigle, L., Ponisio, L., Cutler, K. & Kremen, C. (2015). Habitat restoration promotes pollina-
- tor persistence and colonization in intensively-managed agriculture. *Ecol. Appl.*, 25, 1557–1565.
- ⁵²⁷ M'Gonigle, L.K., Williams, N.M., Lonsdorf, E. & Kremen, C. (2016). A tool for selecting plants
- when restoring habitat for pollinators. *Conservation Letters*.
- Montoya, J.M., Pimm, S.L. & Solé, R.V. (2006). Ecological networks and their fragility. *Nature*,
- 530 442, 259–264.
- Morandin, L. & Kremen, C. (2013). Hedgerow restoration promotes pollinator populations and
- exports native bees to adjacent fields. *Ecol. Appl.*, 23, 829–839.
- Neumann, P. & Carreck, N.L. (2010). Honey bee colony losses. J. Api. Res., 49, 1-6.
- Newman, M.E.J. & Girvan, M. (2004). Finding and evaluating community structure in networks.
- 535 *Phys. Rev. E*, 69, 026113.
- Olesen, J.M., Bascompte, J., Dupont, Y. & Jordano, P. (2007). The modularity of pollination
- networks. *Proc. Natl. Acad. Sci. USA*, 104, 19891–19896.

- Olesen, J.M., Bascompte, J., Elberling, H. & Jordano, P. (2008). Temporal dynamics in a pollination network. *Ecology*, 89, 1573.
- Ollerton, J., Winfree, R. & Tarrant, S. (2011). How many flowering plants are pollinated by animals? *Oikos*, 120, 321–326.
- Parker, V.T. (1997). The scale of successional models and restoration objectives. *Restoration ecology*, 5, 301–306.
- Peel, L. & Clauset, A. (2014). Detecting change points in the large-scale structure of evolving networks. *arXiv preprint arXiv:1403.0989*.
- Petanidou, T., Kallimanis, S., Tzanopoulos, J., Sgardelis, S. & Pantis, J. (2008). Long-term observation of a pollination network: fluctuation in species and interactions, relative invariance of network structure and implications for estimates of specialization. *Ecol. Lett.*, 11, 564–575.
- Pimm, S.L. (1984). The complexity and stability of ecosystems. *Nature*, 307, 321–326.
- Ponisio, L.C., M'Gonigle, L.K. & Kremen, C. (2015). On-farm habitat restoration counters biotic homogenization in intensively managed agriculture. *Global change biology*.
- Pyke, G.H. (1984). Optimal foraging theory: a critical review. *Annual review of ecology and*systematics, pp. 523–575.
- R Core Team (2015). *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Vienna, Austria.
- Ramos-Jiliberto, R., Valdovinos, F.S., Moisset de Espanés, P. & Flores, J.D. (2012). Topological plasticity increases robustness of mutualistic networks. *Journal of Animal Ecology*, 81, 896–904.

- Rezende, E.L., Lavabre, J.E., Guimarães, P.R., Jordano, P. & Bascompte, J. (2007). Non-random coextinctions in phylogenetically structured mutualistic networks. *Nature*, 448, 925–928.
- Santamaría, L. & Rodríguez-Gironés, M.A. (2007). Linkage rules for plant–pollinator networks:

 Trait complementarity or exploitation barriers? *PLoS Biol.*, 5, e31.
- Stang, M., Klinkhamer, P., Waser, N.M., Stang, I. & van der, M.E. (2009). Size-specific interaction patterns and size matching in a plant-pollinator interaction web. *Ann. Bot.*, 103.
- Stang, M., Klinkhamer, P.G. & Van Der Meijden, E. (2006). Size constraints and flower abundance determine the number of interactions in a plant–flower visitor web. *Oikos*, 112, 111–121.
- Valdovinos, F.S., Moisset de Espanés, P., Flores, J.D. & Ramos-Jiliberto, R. (2013). Adaptive foraging allows the maintenance of biodiversity of pollination networks. *Oikos*, 122, 907–917.
- Valdovinos, F.S., Ramos-Jiliberto, R., Garay-Narváez, L., Urbani, P. & Dunne, J.A. (2010). Consequences of adaptive behaviour for the structure and dynamics of food webs. *Ecology Letters*,
 13, 1546–1559.
- Vázquez, D.P. (2005). Degree distribution in plant-animal mutualistic networks: forbidden links or random interactions? *Oikos*, 108, 421–426.
- Vázquez, D.P., Blüthgen, N., Cagnolo, L. & Chacoff, N.P. (2009). Uniting pattern and process in plant-animal mutualistic networks: a review. *Annals of Botany*, 103, 1445–1457.
- Vázquez, D.P., Chacoff, N.P. & Cagnolo, L. (2009). Evaluating multiple determinants of the structure of plant-animal mutualistic networks. *Ecology*, 90, 2039–2046.
- Waser, N.M., Chittka, L., Price, M.V., Williams, N.M. & Ollerton, J. (1996). Generalization in
 pollination systems, and why it matters. *Ecology*, 77, 1043.

Winfree, R., Williams, N., Dushoff, J. & Kremen, C. (2007). Native bees provide insurance against
 ongoing honey bee losses. *Ecol. Lett.*, 10, 1105–1113.

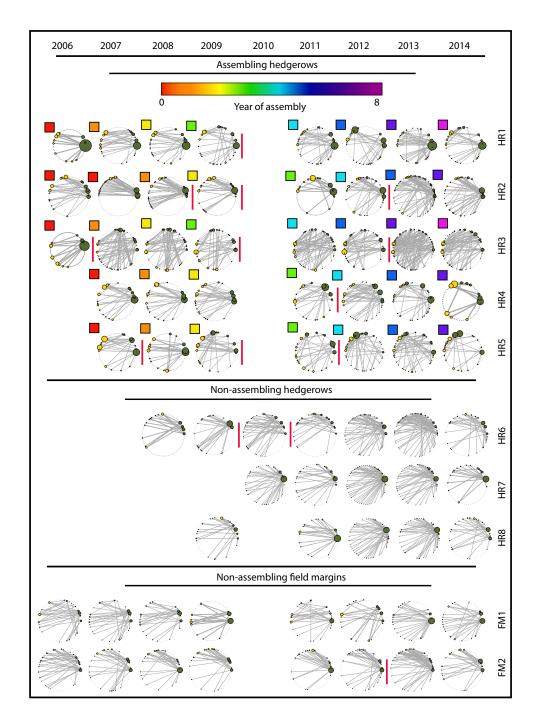


Figure 1: Assembling hedgerow networks had more changing points (vertical red lines) and non-assembling hedgerows and weedy field margins (a represented sample of non-assembling sites are depicted here). In each network, plants and pollinators are represented by green and yellow circles, respectively, weighted by their degree. Each species has has a consistent position in the network across years. In the assembling hedgerows, colored squares in the corner of each network represent the years post restoration.

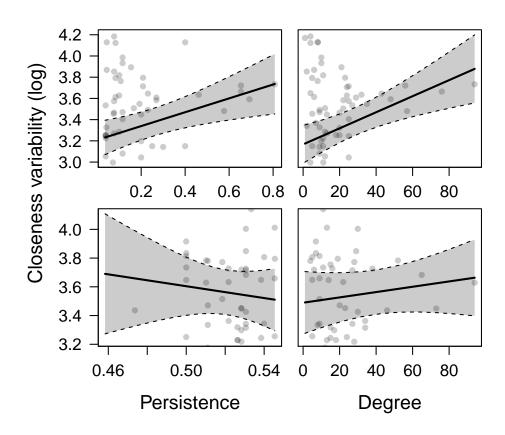


Figure 2: The coefficient of variation of network position, as represented by closeness, plotted against pollinator persistence and degree. Persistence and degree were positively related to network position variability in pollinators, but unrelated in plants. Points represents means for each species across sites. The solid line indicates the mean slope estimate and the dashed lines are the 95% confidence intervals around the estimate.

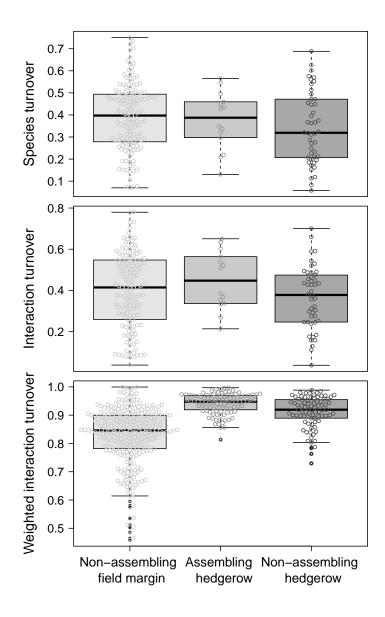


Figure 3: The pollinator species, interaction and weighted interactions turnover of plant-pollinator networks at non-assembling field margins sites, assembling hedgerows, and non-assembling, mature hedgerows. Rates of species and interaction turnover were similar between site types. However, when interactions where weighted by their uniqueness, both hedgerow types had higher turnover that unrestored field margins. Boxplots represent medians (black horizontal line) first and third quartiles (box perimeter) and extremes (whiskers).

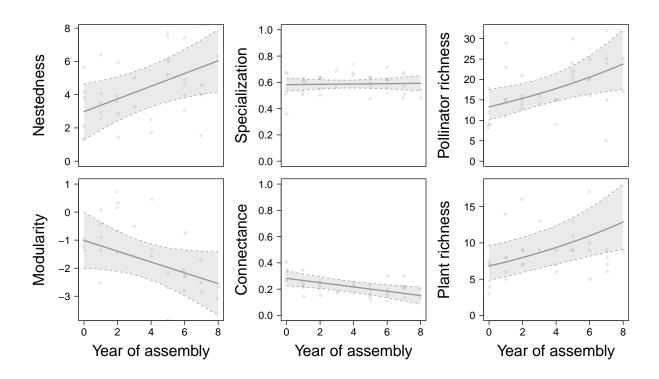


Figure 4: The change in nestedness, modularity, specialization and connectance as the networks assemble. The left panels represent z-scores. Scores greater than ~ 2 or less than ~ -2 are significantly more or less structured than randomly assembled networks. Points are the value for each site at each year of assembly. The solid line indicates the mean slope estimate and the dashed lines are the 95% confidence intervals around the estimate.

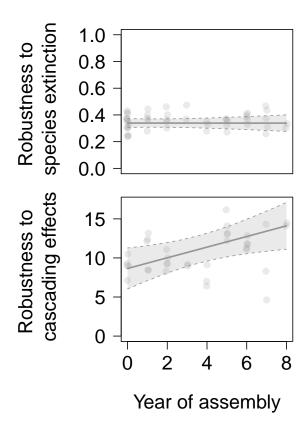


Figure 5: The robustness of networks to species extinction and to cascading perturbations. The robustness to species extinction is measured by incrementally removing species by degree, through removing species by abundance did not yield qualitatively different results. The robustness of networks to cascading perturbations is measured as the algebraic connectivity, the second smallest eigenvalue of the Laplacian matrix. Points are the value for each site at each year of assembly. The solid line indicates the mean slope estimate and the dashed lines are the 95% confidence intervals around the estimate.