

The temporal assembly of plant-pollinator networks following restoration

Lauren C. Ponisio^{1/2}, Marilia P. Gaiarsa³, Claire Kremen¹

1. Department of Environmental Science, Policy, and Management
University of California, Berkeley
130 Mulford Hall
Berkeley, California, USA
94720
2. Department of Entomology
University of California, Riverside
417 Entomology Bldg.
Riverside, California, USA
92521
3. Departamento de Ecologia
Universidade de Sao Paulo
Sao Paulo, SP, Brazil
05508-900

Abstract

TO BE RE-WITTEN The structure of networks is related to ability of communities to maintain function in the face of species extinction. Understanding network structure and how it relates to network disassembly, therefore, is a priority for system-level conservation biology. We explore the assembly of plant-pollinator communities on native plant restorations in the Central Valley of California.

Keywords: changing points, temporal networks, hedgerows, species interactions, preferential attachment, mutualisms

9 Introduction

10 Global change has created a severe biodiversity crisis, and as species are lost, so are their inter-
11 actions (Dunn *et al.*, 2009; Barnosky *et al.*, 2011). Because mutualistic interactions are essential
12 for maintaining the diversity their component guilds of species, these systems are particularly at
13 risk from coextinction cascades. The nature of these coextinction cascades depends on the inter-
14 action patterns within a community (Memmott *et al.*, 2004; Rezende *et al.*, 2007; Bascompte &
15 Stouffer, 2009). Recovering the lost biodiversity and interactions through ecological restoration
16 has become increasingly imperative, and a key restoration aim is to facilitate assembly of robust
17 interaction networks (Menz *et al.*, 2010). We know little, however, about how to re-assemble in-
18 teracting communities through restoration, or the process of ecological network assembly more
19 generally.

20 The mostly widely explored mechanism of network assembly, preferential attachment (Barabási &
21 Albert, 1999), predicts that a new species is more likely to interact with species that are already
22 well-connected (“the rich-get-richer” principle, Barabási & Albert, 1999). In pollination systems
23 — a particularly ubiquitous mutualism (Ollerton *et al.*, 2011; Klein *et al.*, 2007) — some studies
24 have found support for this mechanism of assembly. Investigating the day-to-day, temporal as-
25 sembly of a plant-pollinator network within a season, Olesen *et al.* (2008) found that new species
26 tended to interact with already well-connected species, likely because these species are either more
27 abundant or more temporally persistent (Olesen *et al.*, 2008). In addition, using a space-for-time
28 substitution to study primary succession along a glacier foreland, Albrecht *et al.* (2010) found
29 some indication assembly was occurring through preferential attachment. Network nestedness, a
30 pattern of interactions where a generalist core interacts with both specialist and generalist species,
31 increased as the community aged (Albrecht *et al.*, 2010). Increasing nestedness could result from
32 a process like preferential attachment where specialist species attach to the well-connected, gener-
33 alist core.

In contrast to the ordered network build-up described by preferential attachment, assembly can be punctuated by significant reorganizations of interactions (Peel & Clauset, 2014). For example, as new species are added, resident species change their interaction partners to minimize competition, or become extinct. Such significant reorganizations of interactions, or changing points, have been observed in networks responding to abrupt shifts in the behavior of interactors (Peel & Clauset, 2014). No studies, however, have examined whether changing points occur during ecological network assembly.

Understanding network assembly is particularly relevant to ecological restoration, which is essentially 'applied succession' (Parker, 1997, e.g.,). In pollination systems, the time since an area was restored has been shown to effect the structure of networks (Forup *et al.*, 2008a,b; Devoto *et al.*, 2012), suggesting interactions are evolving as the community develops. Understanding the mechanisms of network assembly will help to guide the restoration of particular communities.

Facilitating effective restoration of networks is particularly imperative in areas where species interactions provide essential ecosystem services, such as crop pollination. In intensively managed agricultural landscapes, the demand for pollination services is the greatest (Kremen, 2008). However, honey bees, managed extensively around the world to provide crop pollination, are in global decline (Neumann & Carreck, 2010; van Engelsdorp *et al.*, 2009). In addition, native pollinators, which have the capacity to provide sufficient crop pollination (Kremen *et al.*, 2002; Winfree *et al.*, 2007; Kremen *et al.*, 2004), are in short supply because these landscapes make poor habitats for pollinator populations (Kremen *et al.*, 2002). To ensure provision the continued provision of ecosystem services and curb biodiversity loss, effective restoration of pollinators and their interactions in agricultural landscapes is critical.

To promote pollinator services in agriculture, farmers are increasingly turning to the habitat restoration technique of planting strips of native plants along farm edges (hedgerows) to help provide habitat for pollinators without removing arable land from production. Hedgerows have been

shown to augment the richness, abundance and trait diversity of pollinators in agricultural landscapes (Morandin & Kremen, 2013; M’Gonigle *et al.*, 2015; Kremen & M’Gonigle, 2015; Ponisio *et al.*, 2015). In addition, hedgerows promote the persistence and colonization of floral resource specialists (M’Gonigle *et al.*, 2015). Little is known however, about the assembly of the network following hedgerow restoration.

Using a long-term data-set of plant-pollinator communities assembling following hedgerow restoration in the highly simplified and intensively managed agricultural landscape of California’s Central Valley, we explore the process of network development. We first determine whether the mechanism underlying network assembly is a smooth build up of interactions as would be predicted by preferential attachment, or punctuated by significant reorganizations of interactions (i.e., changing points). Even with changing points in interaction organization, networks could still be assembling via preferential attachment if the network reorganizations were primarily driven by peripheral, temporally variable species while a stable, well-connected core of species still persists. We thus examine whether the species are most variable in their network position — and thus important contributors to network reorganizations — are less persistent and connected species. Lastly, we examine whether networks are assembling toward predictable interaction patterns, and the ramifications for the robustness of the networks to species extinction and perturbation.

Materials & Methods

Study sites and collection methods

We surveyed plant-pollinator interaction networks of assembling hedgerows (N=5), as well as in two types of non-assembling communities to serve as controls: unrestored, weedy field margins (N=19) and established hedgerows (greater than 10 years since planting, N=29). The sites were

located in the Central Valley of California in Yolo, Colusa and Solano Counties. This area is comprised of intensively managed agriculture – primarily monocultures of conventional row crops, vineyards and orchards. Hedgerows we planted along field margins where they do not remove valuable land from production, and are ca. 3–6m wide and approximately 350m long and border large (ca. 30–hectare) crop fields. Hedgerows consist of native, perennial, shrub and tree plantings including *Rosa californica*, *Cercis occidentalis*, *Ceanothus spp.*, *Heteromeles arbutifolia*, *Sambucus mexicana*, *Eriogonum spp.*, *Baccharis spp.*, *Salvia spp.* and others (Kremen & M’Gonigle, 2015; M’Gonigle *et al.*, 2015). The mean distance between monitoring sites was 15 km, and the minimum distance between sites of the same type sampled in the same year was 2 km. The entire area surveyed spanned almost 300 km². The crop fields adjacent to all sites were similarly managed as intensive, high-input monoculture.

Monitoring began in 2006 and continued through 2015. Sampling of the assembling hedgerows began the year before the area was restored. For logistical reasons, no sampling of assembling hedgerows was conducted in 2010. Sites were sampled between two and five times per year (Tables S1-S3). In each round of sampling, the order in which sites were sampled was randomized. Surveys were conducted under sunny conditions when the temperature was above 21°C and wind speed was below 2.5 meters/second.

Flower-visitors to plants in hedgerows and unrestored controls were netted for one hour of active search time (the timer was paused when handling specimens). Honeybees (*Apis mellifera*) were not collected because their abundance is determined largely by the placement of hives throughout the region by bee-keepers. All other insect flower visitors that touched the reproductive parts of the flower were collected; however, here we focus only on wild bees and syrphids (representing 49 and 19 percent of records, respectively), the most abundant and effective pollinators in the system (C. Kremen, A. Klein and L. Morandin, unpublished data). Bee and syrphid specimens were identified to species (or morpho-species for some bee specimens in the genera *Nomada* and *Sphecodes*) by

106 expert taxonomists.

107 Quantitative networks were generative for each site through time. To account for the unequal num-
108 ber of surveys between years, the mean of the interactions between a pair of plants and pollinators
109 across surveys within a year was used as a representation of the frequency of interactions.

110 **Change point analysis**

111 **Identifying change points**

112 We employed a change point detection method (Peel & Clauset, 2014) to identify fundamental
113 changes in large-scale pattern of interactions of plants and pollinators. A change point is caused by
114 a merge, split, fragmentation or formation of communities (also called modules or compartments).
115 Change point detection methods have yet to be generalized to quantitative networks, so for this
116 analysis we focused on qualitative (binary) networks. Following Peel & Clauset (2014), we first
117 defined a probability distribution over the networks using the generalized hierarchical random
118 graph model (GHRG). The GHRG model is able to capture both assortative and disassortative
119 community structure patterns at all scales in the network (Peel & Clauset, 2014). A network G is
120 composed of vertices V and edges $E \subseteq VV$. The GHRG model decomposes the N vertices into a
121 series of nested groups, the relationships among which are represented by the dendrogram T . The
122 tips of T are the vertices of G , and the probability that two vertices u and v connect is given by the
123 parameter p_r . The probability distribution of the network G thus defined as:

$$P(G|T, pr) = p_r^{E_r} (1 - p_r)^{N_r - E_r} \quad (1)$$

124 Where E_r is the observed number of edges between vertices with the common ancestor r , and N_r
125 is the total possible edges.

Using Bayesian posterior inference and techniques from phylogenetic tree reconstruction, we fit the GHRG model to the networks (Peel & Clauset, 2014). This is accomplished by using a Markov Chain Monte Carlo (MCMC) procedure to first sample the posterior distribution of bipartitions, from which a consensus tree is derived (Peel & Clauset, 2014). We used β distributions with the hyperparameters $\alpha = \beta = 1$ to define priors for p_r .

Once the GHRG model has been fit to the networks, we determine whether a change point occurred between two time slices. To detect a change point, we compare the fit of two models – one where a change point had occurred between two networks, and one where no change occurred – using posterior Bayes factors. We chose a sliding window of length, w , of four, within which to find change points. Larger windows allow for more gradual changes, and four was the maximum possible with our maximum of nine years of data. Lastly, to calculate a p -value for the Bayes factors, we use parametric bootstrapping to numerically estimate the null distribution (Peel & Clauset, 2014). The change point analysis was carried out using code published online by L. Peel. Analyses we conducted in Python 3.4.

We next test whether the change points occurring in maturing hedgerows were a component of the assembly process or a product of environmental shifts that lead to network reorganizations in all types of communities. We model the number of change points as successes and the total number of years each site was sampled as trials, and use a generalized linear model with Binomial error to test whether the probability of a change point occurring varied by site type. For the non-assembling hedgerows and weedy field margins, only sites with five or greater years of sampling was included in this analysis. All statistical analysis were conducted in R 3.2.3 (R Core Team, 2015).

Characteristics of species that contribute to change points

To further elucidate the nature of the change points, we examine the characteristics of the species that contributed the reorganization of interactions. Some species remain in relatively similar net-

work positions through time, whereas others are more variable in their role and thus contribute more strongly to network reorganization. We test the that more persistent species with the highest degree are the most stable in their network position, as would be expected is the networks were assembling via preferential attachment.

We calculate species persistence as the proportion of the surveys a pollinator was observed. Species observed consistently within and between years are thus maximally persistent. Weighted species degree is calculated from interaction observations observed in more extensive data-set from Yolo County (approx. 18000 interaction records) that included both the data included in this study and additional data from sites where we collected flower visitors using the same methods (M’Gonigle *et al.*, 2015; Ponisio *et al.*, 2015). To represent the the variability of species within networks, we computed the coefficient of variation of weighted closeness at each site through time. Closeness describes the centrality of a species in the network by calculating path lengths to other vertices (species) in the graph. We used linear mixed models to test whether the variability of species closeness values was related to the persistence or degree of that species (Bates *et al.*, 2014; Kuznetsova *et al.*, 2014). We included random effects for species, as well as site. We focused on the pollinator species because the hedgerow flowers are planted and thus are not directly assembling. Because degree and persistence were strongly correlated, ($\rho = 0.84$, $p\text{-value} < 2 * 10^{-16}$), each explanatory variable was included in the model separately. Because a linear increase in closeness, as might be expected with assembly by preferential attachment, would lead to a high variability in closeness scores, we also test whether closeness increases through time.

Temporal changes in interaction patterns

Network structure

The changing points in network structure may contribute to the reorganization of the assembling networks into predictable interaction patterns. Pollination networks exhibit two main topologies — modularity (e.g., Olesen *et al.*, 2007) and nestedness (e.g., Bascompte *et al.*, 2006, 2003). Modular community interactions are insular, occurring within separate groups or “modules” more often than between modules. Communities in the network may fragment as the network assembles, enhancing modularity. Conversely, nested networks are like pyramid of interactions, where there are some species that interact with many species, other species that interact with a subset of those species, and so on. If species entering the network tend to interact with the generalist base of the network pyramid (i.e., preferential attachment), nestedness would increase through time. Lastly, if the network is accumulating specialist species or species are beginning to limit their interaction niche breath as the network assembles, this would lead to an increase in the network-level specialization (Blüthgen *et al.*, 2006). To test whether network modularity, nestedness or specialization changed linearly with assembly, we used linear mixed models with the descriptive network metrics as the response variable, year of assembly as the explanatory variable, and random effects of site and year.

We use NODF (weighted or unweighted) to evaluate network nestedness (Almeida-Neto *et al.*, 2008). NODF evaluates whether species with fewer partners interact with subsets of partners with which more connected species interact (Almeida-Neto *et al.*, 2008). To estimate modularity, we use a hierarchical clustering algorithm (Newman & Girvan, 2004; Csardi & Nepusz, 2006). We calculated standardized z-scores so that nestedness and modularity metrics could be compared between communities. The z-scores were calculated by generating an ensemble of 999 randomly assembled communities, subtracting the mean of the statistic calculated across these communities

from the observed value, and then dividing by the standard deviation. To assemble random communities, we reshuffled the interactions between species but fixed the total number of interactions, species and the distribution of the interaction frequencies (Galeano *et al.*, 2009). Lastly, Network specialization was measured using H2, which estimate the deviation of the observed interaction frequency between plants and pollinators from a null expectation where all partners interact in proportion to their abundances (Blüthgen *et al.*, 2006). It ranges from 0 for generalized networks to 1 for specialized networks.

Network robustness

Lastly, we test whether the changes in interaction patterns associated with network assembly affect the robustness. First, following Memmott *et al.* (2004), we simulate the extinction of plant species the subsequent extinction cascades of pollinator species. Because the reproduction of plant species if facilitated by active restoration efforts, it is unlikely the extinction of pollinators would effect plant populations in the hedgerows. However, plants ceasing to bloom, for example in response to drought, will likely effect the pollinators that depend on them. Plants species were eliminated based on their degree or abundance, and the number of pollinators that secondarily went extinct is calculated. The area below the extinction curve is used as a measure of network robustness (Memmott *et al.*, 2004).

Results

Over eight years and 747 samples, we collected and identified 19,547 wild bees and syrphids comprising 173 species from 50 genera. We observed 1,521 unique interactions between plants and pollinators.

Change point analysis

Identifying change points

The majority (76%) of the sites tests underwent at least one significant reorganization of interactions (Fig. 1). There were no consistent trends as to when change points occurred within assembling hedgerows or across all sites, except many site had changing points between year 2009 and 2011 (Fig. 1). In California, 2011 marked the beginning of a multi year drought. In the assembling hedgerows were not sampled in 2010, so disentangling whether the changing points are due to skipping a year of assembly or the drought is not impossible.

All five of the assembling hedgerows experienced changing points, whereas only 40% and 81% of non-assembling hedgerows and field margins, respectively, underwent significant interaction reorganizations. Assembling hedgerows experienced significantly more changing points than the non-assembling networks (difference in the odds ratios between assembling and non-assembling networks, 3.316, 95% CI [1.314, 8.572], p -value= 0.0117). Network assembly following restoration is thus punctuated by more interaction reorganizations than would be expected by environmental shifts that would effect assembling and non-assembling networks equally.

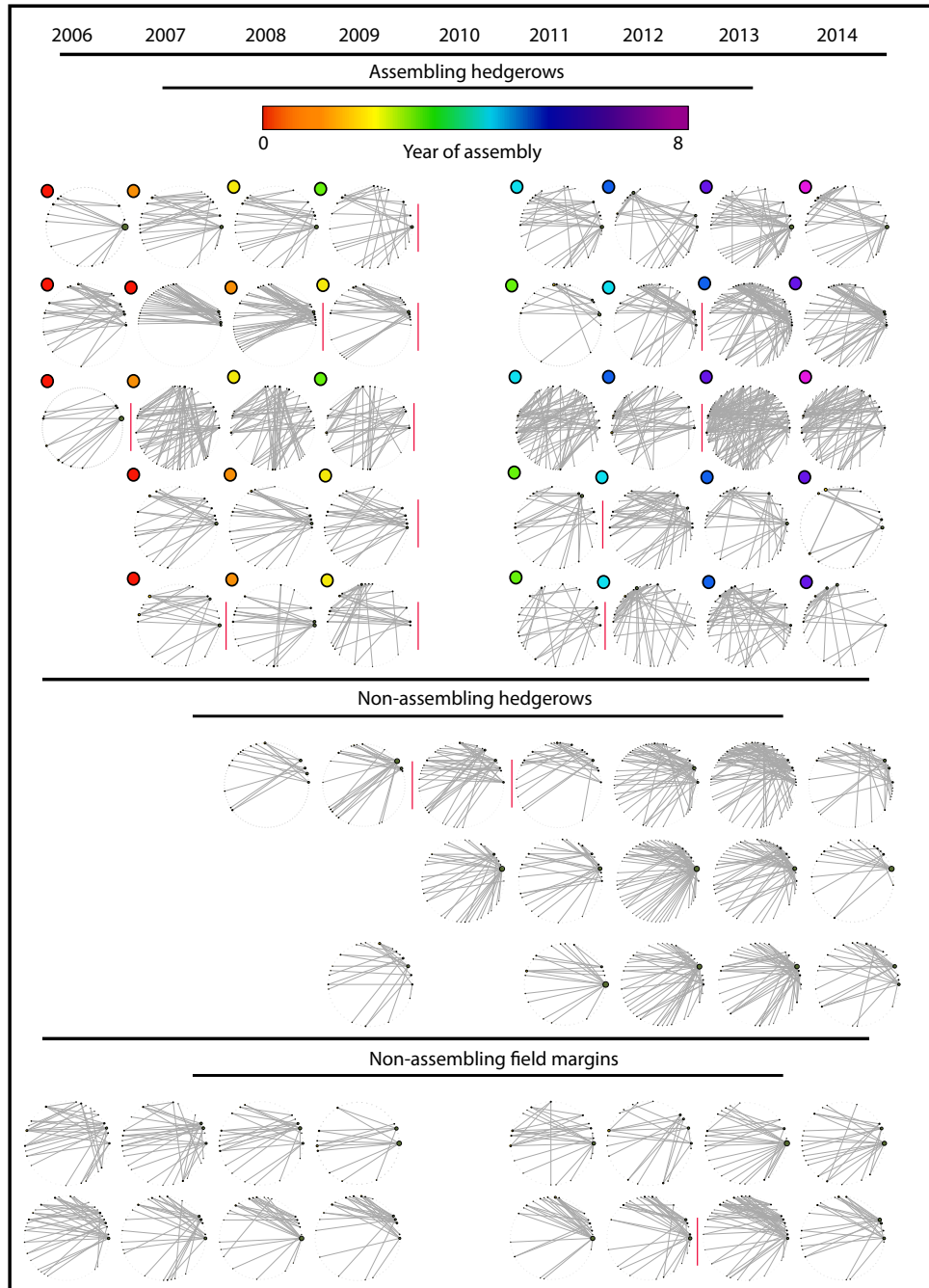


Figure 1: The network structure and changing points (vertical red lines) in assembling hedgerows and a representative sample of non-assembling hedgerows and weedy field margins. In each network, plants and pollinators are represented by green and yellow circles, respectively, weighted by their degree. Each species has a consistent position in the network across years. In the assembling hedgerows, colored circles in the corner of each network represent the years post restoration.

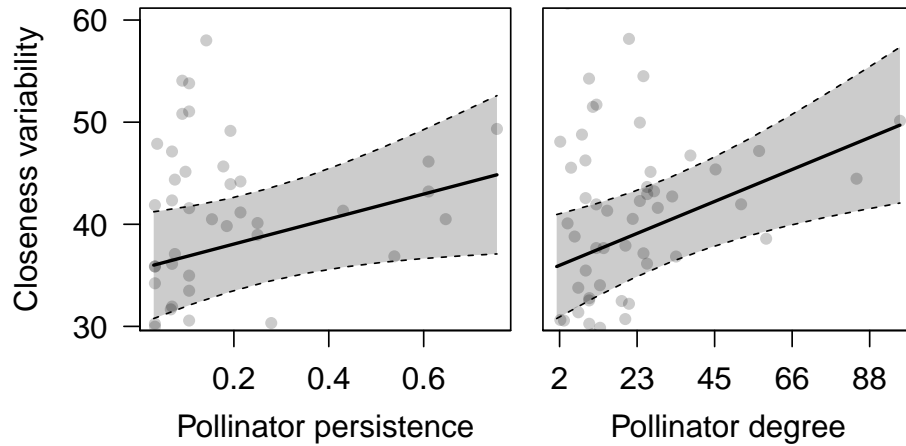


Figure 2: The coefficient of variation of network position, as represented by closeness, plotted against pollinator persistence and degree. Points represents means for each species across sites. The solid line indicates the mean slope estimate and the dashed lines are the 95% confidence intervals around the estimate.

Characteristics of species that contribute to change points

In contradiction to the predictions of assembly by preferential attachment, both pollinator persistence and degree were positively related to network position variability. (ADD STATS IF KEEP-ING RESULT).

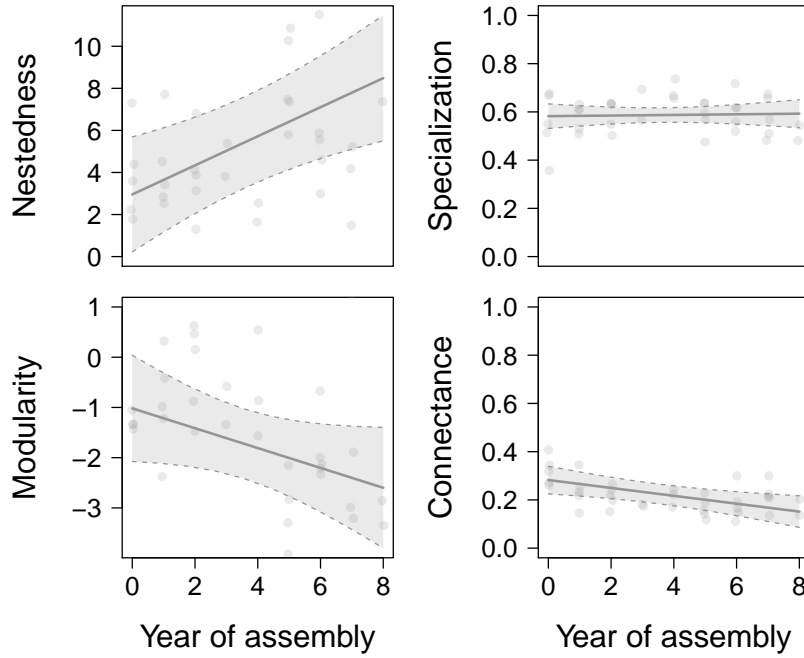


Figure 3: The change in nestedness, modularity, specialization and connectance as the networks assemble. The left panels represent z-scores. Scores greater than ~ 2 or less than ~ -2 are significantly more or less structured than randomly assembled networks. Points are the value for each site at each year of assembly. The solid line indicates the mean slope estimate and the dashed lines are the 95% confidence intervals around the estimate.

Temporal changes in interaction patterns

Network structure

Network nestedness significantly increased with assembly (estimate of the slope of nestedness through time \pm standard error of the estimate, 1.834 ± 0.6142 , p -value=0.022, Fig. 3). Modularity decreased (Fig. 3), though the slope was not significantly different from zero (estimate of the slope of modularity through time \pm standard error of the estimate, -0.524 ± 0.295 , p -value=0.124). Specialization remained relatively constant (estimate of the slope of specialization through time \pm standard error of the estimate, 0.003 ± 0.015 , p -value=0.827).

Network robustness

Assembly did not effect the robustness of the networks to species extinction when species where removed incrementally by degree (estimate of the slope of robustness through time \pm standard error of the estimate, $6 * 10^{-5} \pm 4 * 10^{-3}$, p -value=0.987) or abundance (0.001 ± 0.003 , p -value=0.65, Fig. 4).

In contrast, the robustness of networks to perturbation, as measured by the algebraic connectivity of the network, increased as the network assembled (estimate of the slope of robustness through time \pm standard error of the estimate, 0.6814 ± 0.272 , p -value=0.042, Fig. 4).

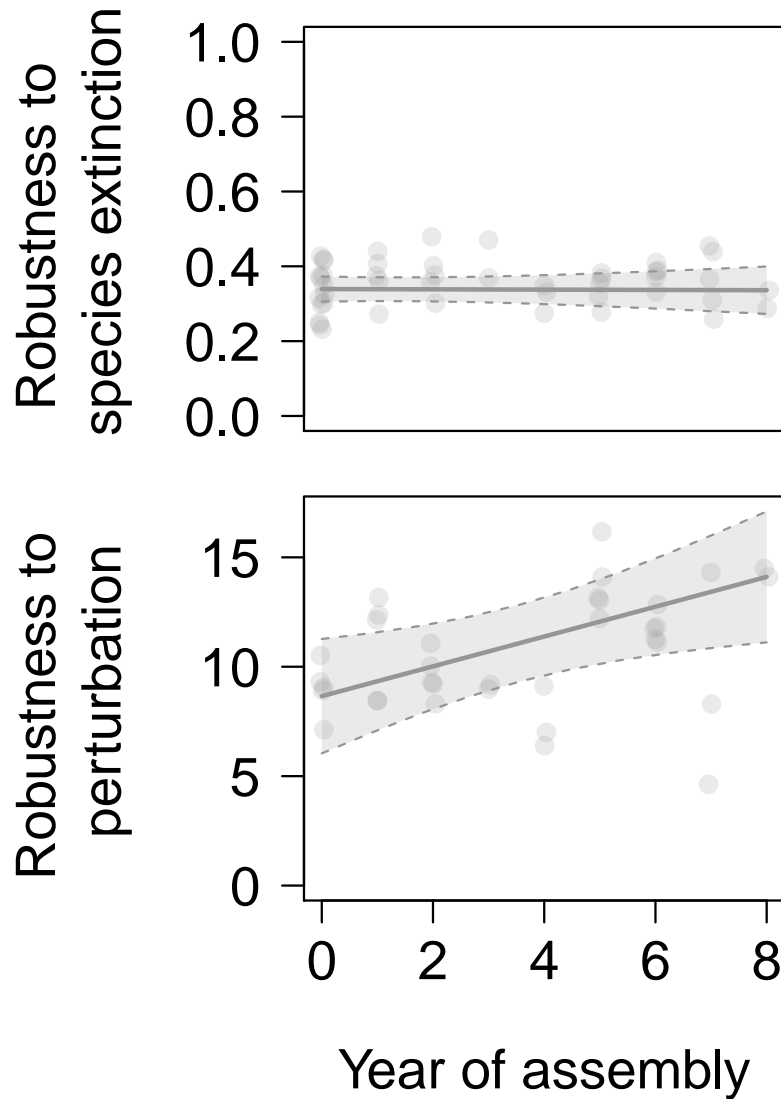


Figure 4: The robustness of networks to species extinction and perturbation. The robustness to species extinction is measured by incrementally removing species by degree, through removing species by abundance did not yield qualitatively different results. Points are the value for each site at each year of assembly. The solid line indicates the mean slope estimate and the dashed lines are the 95% confidence intervals around the estimate.

Discussion

Acknowledgments

We would like to thank Leto Peel and Aaron Clauset for their invaluable discussions and for help with the change point analysis. We thank the growers and land owners that allowed us to work on their property. We also greatly appreciate the identification assistance of expert taxonomists Martin Hauser, Robbin Thorp and Jason Gibbs. This work was supported by funding from the Army Research Office (W911NF-11-1-0361 to CK), the Natural Resources Conservation Service (CIG-69-3A75-12-253, CIG-69-3A75-9-142, CIG-68-9104-6-101 and WLF-69-7482-6-277 to The Xerces Society), the National Science Foundation (DEB-0919128 to CK), The U.S. Department of Agriculture (USDA-NIFA 2012-51181-20105 to Michigan State University). Funding for LCP was provided by an NSF Graduate Research Fellowship and the USDA NIFA Graduate Fellowship.

FUNDING FOR MARILLIA.

References

- Albrecht, M., Riesen, M. & Schmid, B. (2010). Plant–pollinator network assembly along the chronosequence of a glacier foreland. *Oikos*, 119, 1610–1624.
- Almeida-Neto, M., Guimarães, P., Guimarães, P.R., Loyola, R.D. & Ulrich, W. (2008). A consistent metric for nestedness analysis in ecological systems: reconciling concept and measurement. *Oikos*, 117, 1227–1239.
- Barabási, A.L. & Albert, R. (1999). Emergence of scaling in random networks. *science*, 286, 509–512.

270 Barnosky, A.D., Matzke, N., Tomiya, S., Wogan, G.O., Swartz, B., Quental, T.B., Marshall, C.,
271 McGuire, J.L., Lindsey, E.L., Maguire, K.C. *et al.* (2011). Has the earth's sixth mass extinction
272 already arrived? *Nature*, 471, 51–57.

273 Bascompte, J., Jordano, P., Melián, C.J. & Olesen, J.M. (2003). The nested assembly of plant-
274 animal mutualistic networks. *Proc. Natl. Acad. Sci. USA*, 100, 9383–9387.

275 Bascompte, J., Jordano, P. & Olesen, J.M. (2006). Asymmetric coevolutionary networks facilitate
276 biodiversity maintenance. *Science*, 312, 431–433.

277 Bascompte, J. & Stouffer, D.B. (2009). The assembly and disassembly of ecological networks.
278 *Phil. Trans. R. Soc. B*, 364, 1781.

279 Bates, D., Maechler, M., Bolker, B. & Walker, S. (2014). *lme4: Linear mixed-effects models using*
280 *Eigen and S4*. R package version 1.1-7.

281 Blüthgen, N., Menzel, F. & Blüthgen, N. (2006). Measuring specialization in species interaction
282 networks. *BMC Ecol.*, 6, 9.

283 Csardi, G. & Nepusz, T. (2006). The igraph software package for complex network research.
284 *InterJournal*, Complex Systems, 1695.

285 Devoto, M., Bailey, S., Craze, P. & Memmott, J. (2012). Understanding and planning ecological
286 restoration of plant–pollinator networks. *Ecology letters*, 15, 319–328.

287 Dunn, R.R., Harris, N.C., Colwell, R.K., Koh, L.P. & Sodhi, N.S. (2009). The sixth mass coextinc-
288 tion: are most endangered species parasites and mutualists? *Proceedings of the Royal Society*
289 *of London B: Biological Sciences*, 276, 3037–3045.

290 van Engelsdorp, D., Evans, J., Saegerman, C., Mullin, C., Haubruge, E., Nguyen, B., Frazier, M.,
291 Frazier, J., Cox-Foster, D., Chen, Y., Underwood, R., Tarpy, D. & Pettis, J. (2009). Colony
292 collapse disorder: A descriptive study. *Plos One*, 4, 1–17.

- Forup, M., Henson, K., Craze, P. & Memmott, J. (2008a). The restoration of ecological interactions: plant-pollinator networks on ancient and restored heathlands. *J. Appl. Ecol.*, 45, 742–752.
- Forup, M.L., Henson, K.S., Craze, P.G. & Memmott, J. (2008b). The restoration of ecological interactions: plant–pollinator networks on ancient and restored heathlands. *Journal of Applied Ecology*, 45, 742–752.
- Galeano, J., Pastor, J.M. & Iriondo, J.M. (2009). Weighted-interaction nestedness estimator (wine): a new estimator to calculate over frequency matrices. *Environmental Modeling & Software*, 24, 1342–1346.
- Klein, A., Vaissière, B., Cane, J., Steffan-Dewenter, I., Cunningham, S., Kremen, C. & Tscharntke, T. (2007). Importance of pollinators in changing landscapes for world crops. *Proc. R. Soc. B*, 274, 303–313.
- Kremen, C. (2008). *Bee Pollinators in Agricultural Ecosystems*, New York: Oxford University Press, chap. Crop Pollination Services From Wild Bees, pp. 10–26.
- Kremen, C. & M’Gonigle, L.K. (2015). Small-scale restoration in intensive agricultural landscapes supports more specialized and less mobile pollinator species. *J. Appl. Ecol.*, 52, 602–610.
- Kremen, C., Williams, N., Bugg, R., Fay, J. & Thorp, R. (2004). The area requirements of an ecosystem service: crop pollination by native bee communities in California. *Ecol. Lett.*, 7, 1109–1119.
- Kremen, C., Williams, N. & Thorp, R. (2002). Crop pollination from native bees at risk from agricultural intensification. *Proc. Natl. Acad. Sci. U.S.A.*, 99, 16812–16816.
- Kuznetsova, A., Bruun Brockhoff, P. & Haubo Bojesen Christensen, R. (2014). *lmerTest: Tests for random and fixed effects for linear mixed effect models (lmer objects of lme4 package)*. R package version 2.0-11.

316 Memmott, J., Waser, N.M. & Price, M.V. (2004). Tolerance of pollination networks to species
 317 extinctions. *Proc. R. Soc. Lond. B*, 271, 2605–2611.

318 Menz, M., Phillips, R., Winfree, R., Kremen, C., Aizen, M., Johnson, S. & Dixon, K. (2010).
 319 Reconnecting plants and pollinators: challenges in the restoration of pollination mutualisms.
 320 *Trends Plant Sci.*, 16, 4–12.

321 M’Gonigle, L., Ponisio, L., Cutler, K. & Kremen, C. (2015). Habitat restoration promotes pollina-
 322 tor persistence and colonization in intensively-managed agriculture. *Ecol. Appl.*, 25, 1557–1565.

323 Morandin, L. & Kremen, C. (2013). Hedgerow restoration promotes pollinator populations and
 324 exports native bees to adjacent fields. *Ecol. Appl.*, 23, 829–839.

325 Neumann, P. & Carreck, N.L. (2010). Honey bee colony losses. *J. Api. Res.*, 49, 1–6.

326 Newman, M.E.J. & Girvan, M. (2004). Finding and evaluating community structure in networks.
 327 *Phys. Rev. E*, 69, 026113.

328 Olesen, J.M., Bascompte, J., Dupont, Y. & Jordano, P. (2007). The modularity of pollination
 329 networks. *Proc. Natl. Acad. Sci. USA*, 104, 19891–19896.

330 Olesen, J.M., Bascompte, J., Elberling, H. & Jordano, P. (2008). Temporal dynamics in a pollina-
 331 tion network. *Ecology*, 89, 1573.

332 Ollerton, J., Winfree, R. & Tarrant, S. (2011). How many flowering plants are pollinated by
 333 animals? *Oikos*, 120, 321–326.

334 Parker, V.T. (1997). The scale of successional models and restoration objectives. *Restoration*
 335 *ecology*, 5, 301–306.

336 Peel, L. & Clauset, A. (2014). Detecting change points in the large-scale structure of evolving
 337 networks. *arXiv preprint arXiv:1403.0989*.

- 338 Ponisio, L.C., M'Gonigle, L.K. & Kremen, C. (2015). On-farm habitat restoration counters biotic
339 homogenization in intensively managed agriculture. *Global change biology*.
- 340 R Core Team (2015). *R: A Language and Environment for Statistical Computing*. R Foundation
341 for Statistical Computing, Vienna, Austria.
- 342 Rezende, E.L., Lavabre, J.E., Guimarães, P.R., Jordano, P. & Bascompte, J. (2007). Non-random
343 coextinctions in phylogenetically structured mutualistic networks. *Nature*, 448, 925–928.
- 344 Winfree, R., Williams, N., Dushoff, J. & Kremen, C. (2007). Native bees provide insurance against
345 ongoing honey bee losses. *Ecol. Lett.*, 10, 1105–1113.