Running head: PLANT-POLLINATOR NETWORK ASSEMBLY

The temporal assembly of plant-pollinator networks following restoration

Lauren C. Ponisio^{1,2}, Marilia P. Gaiarsa³, Claire Kremen¹

- Department of Environmental Science, Policy, and Management University of California, Berkeley 130 Mulford Hall Berkeley, California, USA 94720
- Department of Entomology
 University of California, Riverside
 417 Entomology Bldg.
 Riverside, California, USA
 92521
- 3. Departamento de Ecologia Universidade de Sao Paulo Sao Paulo, SP, Brazil 05508-900

1 Abstract

TO BE RE-WRITTEN The structure of networks is related to ability of communities to
maintain function in the face of species extinction. Understanding network structure and how
it relates to network disassembly, therefore, is a priority for system-level conservation biology.
We explore the assembly of plant-pollinator communities on native plant restorations in the
Central Valley of California.

- 7 Keywords: changing points, temporal networks, hedgerows, species interactions, preferential at-
- 8 tachment, mutualisms

9 Introduction

Global change has created a severe biodiversity crisis, and as species are lost, so are their interactions (Dunn *et al.*, 2009; Barnosky *et al.*, 2011). Because mutualistic interactions are essential
for maintaining the diversity their component guilds of species, these systems are particularly at
risk from coextinction cascades. The nature of these coextinction cascades depends on the interaction patterns within a community (Memmott *et al.*, 2004; Rezende *et al.*, 2007; Bascompte &
Stouffer, 2009). Recovering the lost biodiversity and interactions through ecological restoration
has become increasingly imperative, and a key restoration aim is to facilitate assembly of robust
interaction networks (Menz *et al.*, 2010). We know little, however, about how to re-assemble interacting communities through restoration, or the process of ecological network assembly more
generally.

The mostly widely explored mechanism of network assembly, preferential attachment (Barabási & 20 Albert, 1999), predicts that a new species is more likely to interact with species that are already 21 well-connected ("the rich-get-richer" principle, Barabási & Albert, 1999). In pollination systems 22 — a particularly ubiquitous mutualism (Ollerton et al., 2011; Klein et al., 2007) — some studies have found support for this mechanism of assembly. Investigating the day-to-day, temporal assembly of a plant-pollinator network within a season, Olesen et al. (2008) found that new species tended to interact with already well-connected species, likely because these species are either more 26 abundant or more temporally persistent (Olesen et al., 2008). In addition, using a space-for-time 27 substitution to study primary succession along a glacier foreland, Albrecht et al. (2010) found 28 some indication assembly was occurring through preferential attachment. Network nestedness, a 29 pattern of interactions where a generalist core interacts with both specialist and generalist species, 30 increased as the community aged (Albrecht et al., 2010). Increasing nestedness could result from a 31 process like preferential attachment where specialist species attach to the well-connected, general-32 ist core. In addition, non-successional temporal dynamics also suggest a stable core of generalists

- persists despite high turnover of peripheral species (Fang & Huang, 2012; Díaz-Castelazo *et al.*, 2010; Alarcón *et al.*, 2008).
- In contrast to the ordered network build-up described by preferential attachment, assembly can be punctuated by significant reorganizations of interactions (Peel & Clauset, 2014). Such significant reorganizations of interactions, or changing points, have been observed in social networks responding to abrupt shifts in the behavior of interactors (Peel & Clauset, 2014). In ecological communities, such shifts my occur if, as new species are added, resident species change their interaction partners to optimize their foraging effort. In plant-pollinator communities, theory predicts pollinators optimize their use of floral resources to reduce interspecific competition and improve resource-use efficiency (Pyke, 1984; Valdovinos *et al.*, 2010, 2013; Albrecht *et al.*, 2010; Blüthgen *et al.*, 2007). No studies, however, have examined whether changing points occur during ecological network assembly and how they relate to the behavior of the interactors.
- Understanding network assembly is particularly relevant to ecological restoration, which is essentially 'applied succession' (Parker, 1997, e.g.,). In pollination systems, the time since an area was restored has been shown to effect the structure of networks (Forup *et al.*, 2008a,b; Devoto *et al.*, 2012), suggesting interactions are evolving as the community develops. Understanding the mechanisms of network assembly will help to guide the restoration of particular communities.
- Facilitating effective restoration of networks is especially imperative in areas where species interactions provide essential ecosystem services, such as crop pollination. In intensively managed
 agricultural landscapes, the demand for pollination services is the greatest (Kremen, 2008). However, honey bees, managed extensively around the world to provide crop pollination, are in global
 decline (Neumann & Carreck, 2010; van Engelsdorp *et al.*, 2009). In addition, native pollinators, which have the capacity to provide sufficient crop pollination (Kremen *et al.*, 2002; Winfree *et al.*, 2007; Kremen *et al.*, 2004), are in short supply because these landscapes make poor habitats
 for pollinator populations (Kremen *et al.*, 2002). To ensure provision the continued provision of

- ecosystem services and curb biodiversity loss, effective restoration of pollinators and their interactions in agricultural landscapes is critical.
- To promote pollinator services in agriculture, farmers are increasingly turning to the habitat restoration technique of planting strips of native plants along farm edges (hedgerows) to help provide habitat for pollinators without removing arable land from production. Hedgerows have been shown to augment the richness, abundance and trait diversity of pollinators in agricultural land-scapes(Morandin & Kremen, 2013; M'Gonigle *et al.*, 2015; Kremen & M'Gonigle, 2015; Ponisio *et al.*, 2015). In addition, hedgerows promote the persistence and colonization of floral resource specialists (M'Gonigle *et al.*, 2015). Little is known however, about the assembly of the network following hedgerow restoration.
- Using a long-term data-set of plant-pollinator communities assembling following hedgerow restoration in the highly simplified and intensively managed agricultural landscape of California's Central 70 Valley, we explore the process of network development. We first determine whether the mechanism underlying network assembly is a smooth build up of interactions as would be predicted by preferential attachment, or punctuated by significant reorganizations of interactions (i.e., changing 73 points). Even with changing points in interaction organization, networks could still be assembling via preferential attachment if the network reorganizations were primarily driven the by peripheral, 75 temporally variable species while a stable, well-connected core of species still persists. We thus 76 examine whether the species are most variable in their network position — and thus important contributors network reorganizations — are less persistent and connected species. Lastly, we examine whether networks are assembling toward predictable interaction patterns, and the ramifications for the robustness of the networks to species extinction and cascading perturbations.

Materials & Methods

102

Study sites and collection methods

We surveyed plant-pollinator interaction networks of assembling hedgerows (N=5), as well as in 83 two types of non-assembling communities to serve as controls: unrestored, weedy field margins 84 (N=19) and established hedgerows (greater than 10 years since planting, N=29). The sites were 85 located in the Central Valley of California in Yolo, Colusa and Solano Counties. This area is comprised of intensively managed agriculture – primarily monocultures of conventional row crops, vineyards and orchards. Hedgerows we planted along field margins where they do not remove valuable land from production, and are ca. 3-6m wide and approximately 350m long and border large (ca. 30-hectare) crop fields. Hedgerows consist of native, perennial, shrub and tree plantings including Rosa californica, Cercis occidentalis, Ceanothus spp., Heteromeles arbutifolia, Sambucus mexicana, Eriogonum spp., Baccharis spp., Salvia spp. and others (Kremen & M'Gonigle, 2015; M'Gonigle et al., 2015). The mean distance between monitoring sites was 15 km, and the minimum distance between sites of the same type sampled in the same year was 2 km. The entire area surveyed spanned almost 300 km². The crop fields adjacent to all sites were similarly managed as intensive, high-input monoculture.

Monitoring began in 2006 and continued through 2015. Sampling of the assembling hedgerows began the year before the area was restored. For logistical reasons, no sampling of assembling hedgerows was conducted in 2010. Sites were sampled between two and five times per year (Tables S1-S3). In each round of sampling, the order in which sites were sampled was randomized. 100 Surveys were conducted under sunny conditions when the temperature was above 21°C and wind 101 speed was below 2.5 meters/second.

Flower-visitors to plants in hedgerows and unrestored controls were netted for one hour of active 103 search time (the timer was paused when handling specimens). Honeybees (Apis mellifera) were 104

not collected because their abundance is determined largely by the placement of hives throughout
the region by bee-keepers. All other insect flower visitors that touched the reproductive parts of the
flower were collected; however, here we focus only on wild bees and syrphids (representing 49 and
19 percent of records, respectively), the most abundant and effective pollinators in the system (C.
Kremen, A. Klein and L. Morandin, unpublished data). Bee and syrphid specimens were identified
to species (or morpho-species for some bee specimens in the genera *Nomada* and *Sphecodes*) by
expert taxonomists.

Quantitative networks were generative for each site through time. To account for the unequal number of surveys between years, the mean of the interactions between a pair of plants and pollinators across surveys within a year was used as a representation of the frequency of interactions.

115 Change point analysis

116 Identifying change points

We employed a change point detection method (Peel & Clauset, 2014) to identify fundamental 117 changes in large-scale pattern of interactions of plants and pollinators. A change point is caused by 118 a merge, split, fragmentation or formation of communities (also called modules or compartments). Change point detection methods have yet to be generalized to quantitative networks, so for this 120 analysis we focused on qualitative (binary) networks. Following Peel & Clauset (2014), we first 121 defined a probability distribution over the networks using the generalized hierarchical random 122 graph model (GHRG). The GHRG model is able to capture both assortative and disassortative 123 community structure patterns at all scales in the network (Peel & Clauset, 2014). A network G is composed of vertices V and edges $E \subseteq VV$. The GHRG model decomposes the N vertices into a 125 series of nested groups, the relationships among which are represented by the dendrogram T. The 126 tips of T are the vertices of G, and the probability that two vertices u and v connect is given by the parameter p_r . The probability distribution of the network G thus defined as:

$$P(G|T, pr) = p_r^{E_r} (1 - p_r)^{N_r - E_r}$$
(1)

Where E_r is the observed number of edges between vertices with the common ancestor r, and N_r is the total possible edges.

Using Bayesian posterior inference and techniques from phylogenetic tree reconstruction, we fit the GHRG model to the networks (Peel & Clauset, 2014). This is accomplished by using a Markov Chain Monte Carlo (MCMC) procedure to first sample the posterior distribution of bipartitions, from which a consensus tree is derived (Peel & Clauset, 2014). We used β distributions with the hyperparameters $\alpha = \beta = 1$ to define priors for p_r .

Once the GHRG model has been fit to the networks, we determine whether a change point occurred 136 between two time slices. To detect a change point, we compare the fit of two models – one where a change point had occurred between two networks, and one where no change occurred – using posterior Bayes factors. We chose a sliding window of length, w, of four, within which to find 139 change points. Larger windows allow for more gradual changes, and four was the maximum 140 possible with our maximum of nine years of data. Lastly, to calculate a p-value for the Bayes 141 factors, we use parametric bootstrapping to numerically estimate the null distribution (Peel & 142 Clauset, 2014). The change point analysis was carried out using code published online by L. Peel. 143 Analyses we conducted in Python 3.4. 144

We next test whether the change points occurring in maturing hedgerows were a component of the assembly process or a product of environmental shifts that lead to network reorganizations in all types of communities. We model the number of change points as successes and the total number of years each site was sampled as trails, and use a generalized linear model with Binomial error to test whether the probability of a change point occurring varied by site type. For the non-assembling

hedgerows and weedy field margins, only sites with five or greater years of sampling was included in this analysis. All statistical analysis were conducted in R 3.2.3 (R Core Team, 2015).

152 Characteristics of species that contribute to change points

To further elucidate the nature of the change points, we examine the characteristics of the species that contributed the reorganization of interactions. Some species remain in relatively similar network positions through time, whereas others are more variable in their role and thus contribute more strongly to network reorganization. We test the that more persistent species with the highest degree are the most stable in their network position, as would be expected is the networks were assembling via preferential attachment.

We calculate species persistence as the proportion of the surveys a pollinator was observed. Species 159 observed consistently within and between years are thus maximally persistent. Weighted species 160 degree is calculated from interaction observations observed in more extensive data-set from Yolo 161 County (approx. 18000 interaction records) that included both the data included in this study and 162 additional data from sites where we collected flower visitors using the same methods (M'Gonigle 163 et al., 2015; Ponisio et al., 2015). To represent the the variability of species within networks, 164 we computed the coefficient of variation of weighted closeness at each site through time. Close-165 ness describes the centrality of a species in the network by calculating path lengths to other vertices 166 (species) in the graph. We used linear mixed models to test whether the variability of species close-167 ness values was related to the persistence or degree of that species (Bates et al., 2014; Kuznetsova et al., 2014). We included random effects for species, as well as site. We focused on the pollinator species because the hedgerow flowers are planted and thus are not directly assembling. Because degree and persistence were strongly correlated, ($\rho = 0.84$, p-value $< 2 * 10^{-16}$), each explana-171 tory variable was included in the model separately. Because a linear increase in closeness, as might 172 be expected with assembly by preferential attachment, would lead to a high variability in closeness scores, we also test whether closeness increases through time.

75 Temporal changes in interaction patterns

6 Network structure

Any changing points in network structure may contribute to the reorganization of the assembling 177 networks into predictable interaction patterns. Pollination networks exhibit two main structural 178 patterns — modularity (e.g., Olesen et al., 2007) and nestedness (e.g., Bascompte et al., 2006, 179 2003). In modular networks, interactions are insular, occurring within separate groups or "mod-180 ules" more often than between modules. Communities in the network may fragment as the network 181 assembles, enhancing modularity. Conversely, nested networks are like pyramid of interactions, 182 where there are some species that interact with many species, other species that interact with a 183 subset of those species, and so on. If species entering the network tend to interact with the gen-184 eralist base of the network pyramid (i.e., via preferential attachment), nestedness would increase 185 through time. Lastly, if the network is accumulating specialist species or species are beginning 186 to limit their interaction niche breath as the network assembles, this would lead to an increase in 187 the network-level specialization (Blüthgen et al., 2006). To test whether network modularity, nest-188 edness or specialization changed linearly with assembly, we used linear mixed models with the descriptive network metrics as the response variable, year of assembly as the explanatory variable, and random effects of site and year.

We use NODF (weighted or unweighted) to evaluate network nestedness (Almeida-Neto *et al.*, 2008). NODF evaluates whether species with fewer partners interact with subsets of partners with which more connected species interact (Almeida-Neto *et al.*, 2008). To estimate modularity, we use a hierarchical clustering algorithm (Newman & Girvan, 2004; Csardi & Nepusz, 2006). We

calculated standardized z-scores so that nestedness and modularity metrics could be compared between communities. The z-scores were calculated by generating an ensemble of 999 randomly 197 assembled communities, subtracting the mean of the statistic calculated across these communities 198 from the observed value, and then dividing by the standard deviation. To assemble random com-199 munities, we reshuffled the interactions between species but fixed the total number of interactions, 200 species and the distribution of the interaction frequencies (Galeano et al., 2009). Lastly, Network 201 specialization was measured using H2, which estimate the deviation of the observed interaction 202 frequency between plants and pollinators from a null expectation where all partners interact in 203 proportion to their abundances (Blüthgen et al., 2006). It ranges from 0 for generalized networks 204 to 1 for specialized networks. 205

06 Network robustness

Lastly, we test the changes in interaction patterns associated with network assembly affect the robustness of the network to species loss and to cascading effects. Following Memmott et al. 208 (2004), we simulate the extinction of plant species the subsequent extinction cascades of pollinator 209 species. Because the reproduction of plant species if facilitated by active restoration efforts, it is unlikely the extinction of pollinators would affect plant populations in the hedgerows. However, 211 plants ceasing to bloom, for example in response to drought, will likely affect the pollinators that 212 depend on them. Plants species were eliminated based on their degree or abundance, and the 213 number of pollinators that secondarily went extinct is calculated. The area below the extinction 214 curve is used as a measure of network robustness (Dormann *et al.*, 2008). 215

We also explored how the robustness to cascading effects changed as community assembled, using
algebraic connectivity — the second smallest eigenvalue of the Laplacian matrix (Fiedler, 1973)
— as a proxy for network robustness (Gaiarsa et al., submitted). Algebraic connectivity is related
to the mean diameter of a graph and to how difficult it is to turn a network into completely dis-

connected groups of nodes, or species (Costa *et al.*, 2007) — the larger the algebraic connectivity, the more robust a network is to cascading effects (Gaiarsa et al., submitted), and the harder it is to break the community into isolated groups of species.

23 Results

Over eight years and 747 samples, we collected and identified 19,547 wild bees and syrphids comprising 173 species from 50 genera. We observed 1,521 unique interactions between plants and pollinators.

227 Change point analysis

228 Identifying change points

The majority (76%) of the sites tests underwent at least one significant reorganization of interactions (Fig. 1). All five of the assembling hedgerows experienced changing points, whereas only 40% and 81% of non-assembling hedgerows and field margins, respectively, underwent significant interaction reorganizations. Assembling hedgerows experienced significantly more changing points than the non-assembling networks (difference in the odds ratios between assembling and non-assembling networks, 3.316, 95% CI [1.314, 8.572], *p*-value= 0.0117). Network assembly following restoration is thus punctuated by more interaction reorganizations than would be expected by environmental shifts that would effect assembling and non-assembling networks equally.

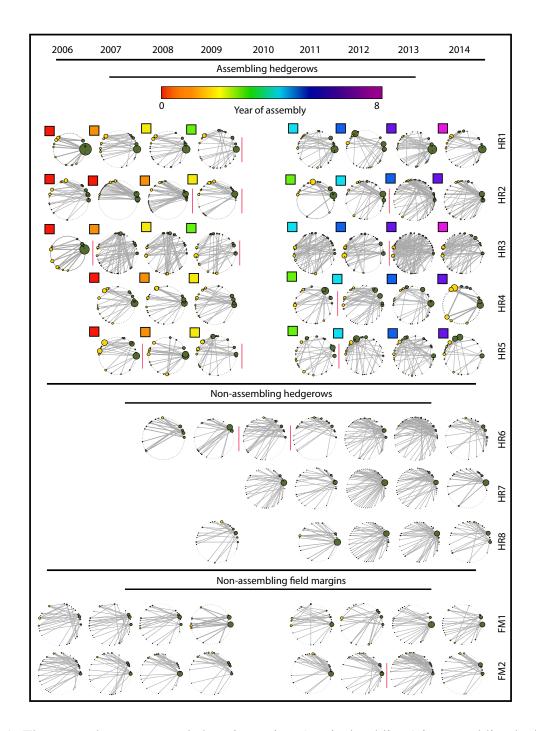


Figure 1: The network structure and changing points (vertical red lines) in assembling hedgerows and a representative sample of non-assembling hedgerows and weedy field margins. In each network, plants and pollinators are represented by green and yellow circles, respectively, weighted by their degree. Each species has has a consistent position in the network across years. In the assembling hedgerows, colored squares in the corner of each network represent the years post restoration.

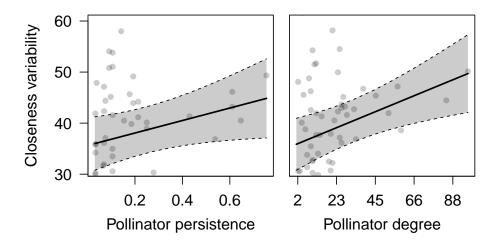


Figure 2: The coefficient of variation of network position, as represented by closeness, plotted against pollinator persistence and degree. Points represents means for each species across sites. The solid line indicates the mean slope estimate and the dashed lines are the 95% confidence intervals around the estimate.

237 Characteristics of species that contribute to change points

- 238 In contradiction to the predictions of assembly by preferential attachment, both pollinator persis-
- 239 tence and degree were positively related to network position variability. (ADD STATS IF KEEP-
- 240 ING RESULT).

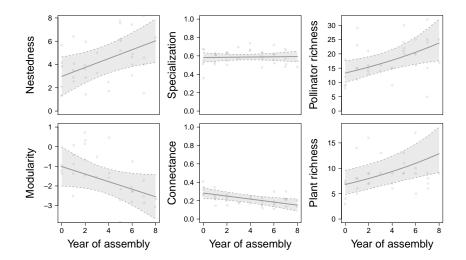


Figure 3: The change in nestedness, modularity, specialization and connectance as the networks assemble. The left panels represent z-scores. Scores greater than ~ 2 or less than ~ -2 are significantly more or less structured than randomly assembled networks. Points are the value for each site at each year of assembly. The solid line indicates the mean slope estimate and the dashed lines are the 95% confidence intervals around the estimate.

Temporal changes in interaction patterns

Network structure

Network nestedness significantly increased with assembly (estimate of the slope of nestedness through time \pm standard error of the estimate, 1.834 ± 0.6142 , p-value=0.022, Fig. 3). Modularity decreased (Fig. 3), though the slope was not significantly different from zero (estimate of the slope of modularity through time \pm standard error of the estimate, -0.524 ± 0.295 , p-value=0.124). Specialization remained relatively constant (estimate of the slope of specialization through time \pm standard error of the estimate, 0.003 ± 0.015 , p-value=0.827).

Network robustness

Fig. 4).

- Assembly did not effect the robustness of the networks to species extinction when species where removed incrementally by degree (estimate of the slope of robustness through time \pm standard error of the estimate, $6*10^{-5} \pm 4*10^{-3}$, p-value=0.987) or abundance (0.001 \pm 0.003, p-value=0.65,
- In contrast, the robustness of networks to cascading perturbations, as measured by the algebraic connectivity of the network, increased as the network assembled (estimate of the slope of robustness through time \pm standard error of the estimate, 0.6814 \pm 0.272, p-value=0.042, Fig. 4).

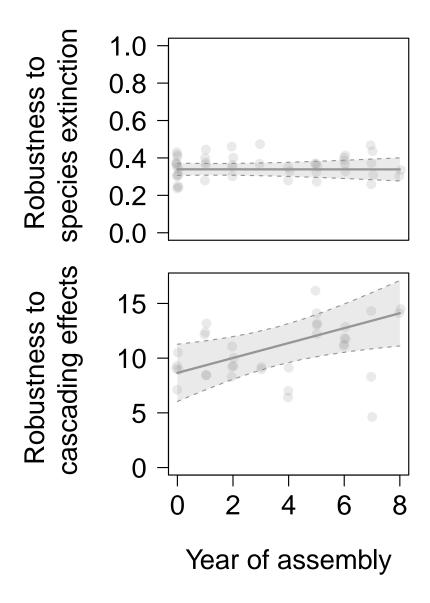


Figure 4: The robustness of networks to species extinction and to cascading effects. The robustness to species extinction is measured by incrementally removing species by degree, through removing species by abundance did not yield qualitatively different results. The robustness of networks to cascading effects is measured as the algebraic connectivity, the second smallest eigenvalue of the Laplacian matrix. Points are the value for each site at each year of assembly. The solid line indicates the mean slope estimate and the dashed lines are the 95% confidence intervals around the estimate.

Discussion

We show that the temporal assembly of plant-pollinator networks following restoration is a highly 258 dynamic process where interactions often undergo significant reorganizations. These network or-259 ganizations are unlikely to be a product of environmental forces alone because the network chang-260 ing points in non-assembling communities are less frequent, and there are few consistent trends 261 in when change points occurred across all sites. Several sites had changing points between year 262 2009 and 2011 (Fig. 1). In California, 2011 marked the beginning of a multi-year drought. In the 263 assembling hedgerows were not sampled in 2010, so disentangling whether the changing points 264 are due to skipping a year of assembly or the drought is not possible. Interestingly, most of the 265 assembling hedgerows did not undergo a significant reorganization of interactions immediately the hedgerow was planted (i.e., the transition from weedy field margin to hedgerow). This result is consistent with the finding that in our study system, hedgerow restoration takes several years to have 268 a significant impact on the plant-pollintor communities (Kremen and M'Gonigle, in prep). 269 Given that several changing points in network organization occured during assembly, we next 270 explored the species most likely responsible for the shifts in interaciton patterns. Based on a preferential attachment-like mechanism, we expect that the most persistent and high degree species would remain stable in the network core during assembly, and would thus contribute the least to the changing points in network organization. Surprisingly however, we encountered the opposite: the species that were most variable in their network position and thus contributed most to network 275 reorganizations where species with the highest degrees (i.e., most generalized) and persistence. 276 For example, the five most ubiquitous species in our study landscape — Halictus ligatus, Halictus 277 tripartitus, Lasioglossum (Dialictus) incompletum, and Toxomerus marginatus — were the only 278 species that changed what module (i.e., community), and they were also present in across years in 279 all of the assembling hedgerows. Because species degree and persistence were strongly correlated, 280 it is difficult to disentangle the causal mechanism for why species with those characteristics are so 28

variable in their network position. Generalized species may be able to better exploit the limited
floral resources the intensively managed agriculture landscape, and thus also most persistent. More
persistent species also have longer phenologies, so they have the opportunity to visit many different
flowers, resulting in a higher degree. Either way, our result suggests that adaptable species can
change their network position to utilize the most advantageous floral resources available, which
my depend the other pollinator species that are present and the state of the floral resource. Thus
given the opportunity and ability to use different resources, species will often change their network
positions.

Given the regional species pool it is possible that specialization might be constant either because 290 species are not changing through time, or because there is a species turnover, but new species have 291 the same role/posotion as the species there are absent in the following time step. We found that 292 node turover was higher in assembling and non-assembling hedgerows when compared to field 293 margin, but there was no difference between them. Thus, maybe looking just at network level 294 metrics, such as nestedness and modularity, when exploring the temporal assembly of ecologi-295 cal networks might be misleading, given that there is a node turnover happening at the species 296 level. 297

The frequent changing points in network organization, dynamic nature of the location of species 298 in networks, and turnover of species and nodes all point to an assembly mechanism other than 299 preferential attachment. Nestedness did increase with years post restoration, as would be expected 300 if colonizing, specialist species attached to a central, generalist core Albrecht et al. (2010). With 301 preferential attachment, however, we would also expect connectance and specialization to increase, 302 and we found no such trends. The stable level of network-level specialization through the assembly 303 process may be due to the increased colonization of specialized species (M'Gonigle et al., 2015) 304 accompanied by an increase in the diet breath of resident species. This would be expected if 305 resident species were able to minimize their foraging time by expanding their diet breath as plant 306

diversity increases with hedgerow maturation (Waser *et al.*, 1996; Pyke, 1984; Blüthgen *et al.*, 2007; Albrecht *et al.*, 2010). Such a change in pollinator behavior would also explain the increase in network nestedness.

The changes in network patterns with assembly also increased the robustness of the networks to 310 cascading perturbations – providing further evidence that hedgerows are valuable tool for promoting species conservation and ecosystem provision in agricultural areas (M'Gonigle et al., 2015; Ponisio et al., 2015; Kremen & M'Gonigle, 2015). Because the vulnerability of the network to cascading effects, as measured by algebraic connectivity, is correlated with species richness, the increase and plant and pollinator richness following restoration is at least partially responsible 315 for enhancing network robustness to cascading effects. The relationship between diversity and 316 stability in networks has been the subject of considerable debate (e.g., May, 1972; Pimm, 1984; 317 Montoya et al., 2006). Our results provides one of the few empirical examples of how restoring 318 species diversity contributes to enhancing network stability. Through these hedgerows were de-319 signed to promote floral resrouces across the growing season and not specifically to promote any 320 specific network properties (Menz et al., 2010), the pollinators assembled into resilent interaction 321 patterns. With prior knowledge of the floral preferences of pollinators, future restoration efforts 322 may also be able to incorporate promoting network stability into planning efforts (M'Gonigle et al., 323 2016) 324

Interestingly, however, the robustness of hedgerow communities to species loss did not change with assembly. This is particularly surprising given the observed increase in nestedness, which is often associated with an enhanced in robustness to extinction (Memmott *et al.*, 2004). It may be that assembling hedgerows have not yet reached a sufficient level of nestedness to realize its benefits. Nestedness of the assembling hedgerows, however, did not asymptote within the eight years following restoration that the sites were surveyed, so hedgerow networks may eventually reach sufficient levels of nestedness to gain the robustness advantage.

Plant-pollinator networks in general are highly dynamic, with high turnover of species and interactions both within and between seasons (Burkle & Alarcón, 2011). Though our non-assembling 333 communities experience fewer network reorganizations than the assembling hedgerows, 82% of 334 field margins and 40% of mature hedgerows underwent at least one changing point in network 335 structure. Pollinators are also highly opportunistic (Petanidou et al., 2008; Vázquez, 2005; Al-336 brecht et al., 2010), though trait complementarity such as tongue length and corolla depth impose 337 some biophysical limits to the interactions between plants and pollinators (Vázquez et al., 2009; 338 Vázquez et al., 2009; Stang et al., 2009, 2006; Santamaría & Rodríguez-Gironés, 2007). Such 339 opportunism may buffer plant-pollinator communities from global change (e.g., Ramos-Jiliberto 340 et al., 2012; Kaiser-Bunbury et al., 2010), but our limited understanding of the assembly of these 341 communities impedes making such predictions (Vázquez et al., 2009; Burkle & Alarcón, 2011). 342 Unlike in the broader food web literature, we have few mechanistic models of mutualistic net-343 work assembly (Valdovinos et al., 2013). In addition, the few that have been developed often borrow their mechanisms from competitive interactions, leading to inaccurate biological assump-345 tions (Holland et al., 2006). We need further development of mechanistic models of mutualistic 346 stystem to generate testable predictions, along with empirical exploration of network assembly. Plant-pollinator communities and mutualisms in general are vital for the maintenance of biodiversity and the provision of essential ecosystem services. We must therefore understand the processes underlying their assembly to facilitate restoration and conservation.

51 Acknowledgments

We would like to thank Paulo Guimarães and Aaron Clauset for their invaluable discussions, and Leto Peel for help with the change point analysis. We thank the growers and land owners that allowed us to work on their property. We also greatly appreciate the identification assistance of expert

- taxonomists Martin Hauser, Robbin Thorp and Jason Gibbs. This work was supported by funding
- from the Army Research Office (W911NF-11-1-0361 to CK), the Natural Resources Conservation
- 357 Service (CIG-69-3A75-12-253, CIG-69-3A75-9-142, CIG-68-9104-6-101 and WLF-69-7482-6-
- 277 to The Xerces Society), the National Science Foundation (DEB-0919128 to CK), The U.S.
- Department of Agriculture (USDA-NIFA 2012-51181-20105 to Michigan State University). Fund-
- ing for LCP was provided by an NSF Graduate Research Fellowship and the USDA NIFA Graduate
- Fellowship. Funding for MPG was provided by São Paulo Research Foundation (FAPESP, grant
- 362 2013/13319-5).

References

- Alarcón, R., Waser, N.M. & Ollerton, J. (2008). Year-to-year variation in the topology of a plant—
- pollinator interaction network. *Oikos*, 117, 1796–1807.
- Albrecht, M., Riesen, M. & Schmid, B. (2010). Plant–pollinator network assembly along the
- chronosequence of a glacier foreland. Oikos, 119, 1610–1624.
- Almeida-Neto, M., Gumarães, P., Gumarães, P.R., Loyola, R.D. & Ulrich, W. (2008). A consistent
- metric for nestedness analysis in ecological systems: reconciling concept and measurement.
- *Oikos*, 117, 1227–1239.
- Barabási, A.L. & Albert, R. (1999). Emergence of scaling in random networks. science, 286,
- ³⁷² 509–512.
- Barnosky, A.D., Matzke, N., Tomiya, S., Wogan, G.O., Swartz, B., Quental, T.B., Marshall, C.,
- McGuire, J.L., Lindsey, E.L., Maguire, K.C. et al. (2011). Has the earth/'s sixth mass extinction
- already arrived? *Nature*, 471, 51–57.

- Bascompte, J., Jordano, P., Melián, C.J. & Olesen, J.M. (2003). The nested assembly of plantanimal mutualistic networks. *Proc. Natl. Acad. Sci. USA*, 100, 9383–9387.
- Bascompte, J., Jordano, P. & Olesen, J.M. (2006). Asymmetric coevolutionary networks facilitate biodiversity maintenance. *Science*, 312, 431–433.
- Bascompte, J. & Stouffer, D.B. (2009). The assembly and disassembly of ecological networks.

 Phil. Trans. R. Soc. B, 364, 1781.
- Bates, D., Maechler, M., Bolker, B. & Walker, S. (2014). *lme4: Linear mixed-effects models using*Eigen and S4. R package version 1.1-7.
- Blüthgen, N., Menzel, F. & Blüthgen, N. (2006). Measuring specialization in species interaction networks. *BMC Ecol.*, 6, 9.
- Blüthgen, N., Menzel, F., Hovestadt, T., Fiala, B. & Blüthgen, N. (2007). Specialization, constraints, and conflicting interests in mutualistic networks. *Curr. Biol.*, 17, 341–346.
- Burkle, L.A. & Alarcón, R. (2011). The future of plant-pollinator diversity: Understanding interaction networks across time, space, and global change. *American Journal of Botany*, 98, 528.
- Costa, L.d.F., Rodrigues, F.A., Travieso, G. & Villas Boas, P.R. (2007). Characterization of complex networks: A survey of measurements. *Advances in physics*, 56, 167–242.
- Csardi, G. & Nepusz, T. (2006). The igraph software package for complex network research. *InterJournal*, Complex Systems, 1695.
- Devoto, M., Bailey, S., Craze, P. & Memmott, J. (2012). Understanding and planning ecological restoration of plant–pollinator networks. *Ecology letters*, 15, 319–328.

- Díaz-Castelazo, C., Guimaraes Jr, P.R., Jordano, P., Thompson, J.N., Marquis, R.J. & Rico-Gray,
- V. (2010). Changes of a mutualistic network over time: reanalysis over a 10-year period. *Ecol-*
- оду, 91, 793–801.
- Dormann, C., Gruber, B. & Fründ, J. (2008). Introducing the bipartite package: analysing ecolog-
- ical networks. *R News*, 8, 8.
- Dunn, R.R., Harris, N.C., Colwell, R.K., Koh, L.P. & Sodhi, N.S. (2009). The sixth mass coextinc-
- tion: are most endangered species parasites and mutualists? Proceedings of the Royal Society
- of London B: Biological Sciences, 276, 3037–3045.
- van Engelsdorp, D., Evans, J., Saegerman, C., Mullin, C., Haubruge, E., Nguyen, B., Frazier, M.,
- Frazier, J., Cox-Foster, D., Chen, Y., Underwood, R., Tarpy, D. & Pettis, J. (2009). Colony
- collapse disorder: A descriptive study. *Plos One*, 4, 1–17.
- Fang, Q. & Huang, S.Q. (2012). Relative stability of core groups in pollination networks in a
- biodiversity hotspot over four years. *PloS one*, 7, e32663.
- Fiedler, M. (1973). Algebraic connectivity of graphs. Czechoslovak mathematical journal, 23,
- 411 298–305.
- Forup, M., Henson, K., Craze, P. & Memmott, J. (2008a). The restoration of ecological interac-
- tions: plant-pollinator networks on ancient and restored heathlands. J. Appl. Ecol., 45, 742–752.
- 414 Forup, M.L., Henson, K.S., Craze, P.G. & Memmott, J. (2008b). The restoration of ecological
- interactions: plant–pollinator networks on ancient and restored heathlands. *Journal of Applied*
- 416 Ecology, 45, 742–752.
- Galeano, J., Pastor, J.M. & Iriondo, J.M. (2009). Weighted-interaction nestedness estimator (wine):
- a new estimator to calculate over frequency matrices. Environmental Modeling & Software, 24,
- 1342–1346.

- Holland, J.N., Okuyama, T. & DeAngelis, D.L. (2006). Comment on asymmetric coevolutionary networks facilitate biodiversity maintenance. *Science*, 313, 1887b–1887b.
- Kaiser-Bunbury, C.N., Muff, S., Memmott, J., Müller, C.B. & Caflisch, A. (2010). The robustness
- of pollination networks to the loss of species and interactions: a quantitative approach incorpo-
- rating pollinator behaviour. *Ecology Letters*, 13, 442–452.
- Klein, A., Vaissière, B., Cane, J., Steffan-Dewenter, I., Cunningham, S., Kremen, C. & Tscharntke,
- T. (2007). Importance of pollinators in changing landscapes for world crops. *Proc. R. Soc. B*,
- 274, 303–313.
- ⁴²⁸ Kremen, C. (2008). Bee Pollinators in Agricultural Ecosystems, New York: Oxford University
- Press, chap. Crop Pollination Services From Wild Bees, pp. 10–26.
- 430 Kremen, C. & M'Gonigle, L.K. (2015). Small-scale restoration in intensive agricultural landscapes
- supports more specialized and less mobile pollinator species. J. Appl. Ecol., 52, 602–610.
- 432 Kremen, C., Williams, N., Bugg, R., Fay, J. & Thorp, R. (2004). The area requirements of an
- ecosystem service: crop pollination by native bee communities in California. Ecol. Lett., 7,
- 1109–1119.
- 435 Kremen, C., Williams, N. & Thorp, R. (2002). Crop pollination from native bees at risk from
- agricultural intensification. *Proc. Natl. Acad. Sci. U.S.A.*, 99, 16812–16816.
- Kuznetsova, A., Bruun Brockhoff, P. & Haubo Bojesen Christensen, R. (2014). *ImerTest: Tests*
- for random and fixed effects for linear mixed effect models (lmer objects of lme4 package). R
- package version 2.0-11.
- May, R.M. (1972). Will a large complex system be stable? *Nature*, 238, 413–414.
- Memmott, J., Waser, N.M. & Price, M.V. (2004). Tolerance of pollination networks to species
- extinctions. *Proc. R. Soc. Lond. B*, 271, 2605–2611.

- 443 Menz, M., Phillips, R., Winfree, R., Kremen, C., Aizen, M., Johnson, S. & Dixon, K. (2010).
- Reconnecting plants and pollinators: challenges in the restoration of pollination mutualisms.
- 445 *Trends Plant Sci.*, 16, 4–12.
- 446 M'Gonigle, L., Ponisio, L., Cutler, K. & Kremen, C. (2015). Habitat restoration promotes pollina-
- tor persistence and colonization in intensively-managed agriculture. *Ecol. Appl.*, 25, 1557–1565.
- 448 M'Gonigle, L.K., Williams, N.M., Lonsdorf, E. & Kremen, C. (2016). A tool for selecting plants
- when restoring habitat for pollinators. *Conservation Letters*.
- 450 Montoya, J.M., Pimm, S.L. & Solé, R.V. (2006). Ecological networks and their fragility. *Nature*,
- 451 442, 259–264.
- 452 Morandin, L. & Kremen, C. (2013). Hedgerow restoration promotes pollinator populations and
- exports native bees to adjacent fields. *Ecol. Appl.*, 23, 829–839.
- Neumann, P. & Carreck, N.L. (2010). Honey bee colony losses. J. Api. Res., 49, 1–6.
- Newman, M.E.J. & Girvan, M. (2004). Finding and evaluating community structure in networks.
- 256 *Phys. Rev. E*, 69, 026113.
- Olesen, J.M., Bascompte, J., Dupont, Y. & Jordano, P. (2007). The modularity of pollination
- networks. *Proc. Natl. Acad. Sci. USA*, 104, 19891–19896.
- Olesen, J.M., Bascompte, J., Elberling, H. & Jordano, P. (2008). Temporal dynamics in a pollina-
- tion network. *Ecology*, 89, 1573.
- Ollerton, J., Winfree, R. & Tarrant, S. (2011). How many flowering plants are pollinated by
- animals? Oikos, 120, 321–326.
- Parker, V.T. (1997). The scale of successional models and restoration objectives. Restoration
- ecology, 5, 301–306.

- Peel, L. & Clauset, A. (2014). Detecting change points in the large-scale structure of evolving networks. *arXiv preprint arXiv:1403.0989*.
- Petanidou, T., Kallimanis, S., Tzanopoulos, J., Sgardelis, S. & Pantis, J. (2008). Long-term ob-
- servation of a pollination network: fluctuation in species and interactions, relative invariance of
- network structure and implications for estimates of specialization. *Ecol. Lett.*, 11, 564–575.
- Pimm, S.L. (1984). The complexity and stability of ecosystems. *Nature*, 307, 321–326.
- Ponisio, L.C., M'Gonigle, L.K. & Kremen, C. (2015). On-farm habitat restoration counters biotic
- homogenization in intensively managed agriculture. *Global change biology*.
- Pyke, G.H. (1984). Optimal foraging theory: a critical review. *Annual review of ecology and*systematics, pp. 523–575.
- R Core Team (2015). *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Vienna, Austria.
- Ramos-Jiliberto, R., Valdovinos, F.S., Moisset de Espanés, P. & Flores, J.D. (2012). Topological
- plasticity increases robustness of mutualistic networks. Journal of Animal Ecology, 81, 896-
- 479 904.
- Rezende, E.L., Lavabre, J.E., Guimarães, P.R., Jordano, P. & Bascompte, J. (2007). Non-random coextinctions in phylogenetically structured mutualistic networks. *Nature*, 448, 925–928.
- Santamaría, L. & Rodríguez-Gironés, M.A. (2007). Linkage rules for plant–pollinator networks:
- Trait complementarity or exploitation barriers? *PLoS Biol.*, 5, e31.
- Stang, M., Klinkhamer, P., Waser, N.M., Stang, I. & van der, M.E. (2009). Size-specific interaction
- patterns and size matching in a plant-pollinator interaction web. *Ann. Bot.*, 103.

- Stang, M., Klinkhamer, P.G. & Van Der Meijden, E. (2006). Size constraints and flower abundance determine the number of interactions in a plant–flower visitor web. *Oikos*, 112, 111–121.
- Valdovinos, F.S., Moisset de Espanés, P., Flores, J.D. & Ramos-Jiliberto, R. (2013). Adaptive foraging allows the maintenance of biodiversity of pollination networks. *Oikos*, 122, 907–917.
- Valdovinos, F.S., Ramos-Jiliberto, R., Garay-Narváez, L., Urbani, P. & Dunne, J.A. (2010). Consequences of adaptive behaviour for the structure and dynamics of food webs. *Ecology Letters*,
 13, 1546–1559.
- Vázquez, D.P. (2005). Degree distribution in plant-animal mutualistic networks: forbidden links or random interactions? *Oikos*, 108, 421–426.
- Vázquez, D.P., Blüthgen, N., Cagnolo, L. & Chacoff, N.P. (2009). Uniting pattern and process in plant-animal mutualistic networks: a review. *Annals of Botany*, 103, 1445–1457.
- Vázquez, D.P., Chacoff, N.P. & Cagnolo, L. (2009). Evaluating multiple determinants of the structure of plant-animal mutualistic networks. *Ecology*, 90, 2039–2046.
- Waser, N.M., Chittka, L., Price, M.V., Williams, N.M. & Ollerton, J. (1996). Generalization in
 pollination systems, and why it matters. *Ecology*, 77, 1043.
- Winfree, R., Williams, N., Dushoff, J. & Kremen, C. (2007). Native bees provide insurance against ongoing honey bee losses. *Ecol. Lett.*, 10, 1105–1113.