Running head: PLANT-POLLINATOR NETWORK ASSEMBLY

# The temporal assembly of plant-pollinator networks following restoration

Lauren C. Ponisio<sup>1,2</sup>, Marilia P. Gaiarsa<sup>3</sup>, Claire Kremen<sup>1</sup>

- Department of Environmental Science, Policy, and Management University of California, Berkeley 130 Mulford Hall Berkeley, California, USA 94720
- Department of Entomology
   University of California, Riverside
   417 Entomology Bldg.
   Riverside, California, USA
   92521
- Departamento de Ecologia Universidade de Sao Paulo São Paulo, SP, Brazil 05508-900

Abstract

1

2

8

10

11

12

13

14

15

16

17

18

19

20

The structure of networks is related to ability of communities to maintain function in the face of species extinction. Understanding network structure and how it relates to network assembly, therefore, is a priority for system-level conservation biology. We explore the assembly of plant-pollinator communities on native plant restorations in the Central Valley of California. Species are highly dynamic in their positions in the networks, such that we find that assembly is punctuated by significant reorganizations of interactions. Non-assembling mature hedgerows and weedy field margins did not undergo as many network reorganizations, suggesting environmental factors alone cannot account for the changes observed in assembling communities. Pollinator species were highly opportunistic, and in fact the most persistent and generalized species were most variable in their network positions — the opposite of what would be expected by preferential attachment. High species and interaction turnover was ubiquitous across assembling and non-assembling communities, though unique interactions turn over at higher rates in assembling hedgerows as the networks continually reorganize. Assembling networks also increased in nestedness through time, though this did not make them more robust to simulated plant extinctions. The resilience of networks to cascading perturbations, however, increased as the communities assembled, at least partially due to accumulating species richness. Our results elucidate some of the mechanisms underlying plant-pollinator network assembly and restoration, while providing further evidence that hedgerows are a valuable tool for promoting species conservation and ecosystem provision in agricultural areas.

- Keywords: changing points, temporal networks, hedgerows, species interactions, preferential at-
- tachment, mutualisms

# 3 Introduction

Global change has created a severe biodiversity crisis, and as species are lost, so are their interactions (Dunn et al., 2009; Barnosky et al., 2011). Because mutualistic interactions are essential 25 for maintaining the diversity their component guilds of species, these systems are particularly at risk from coextinction cascades. The nature of these cascades will depend on the interaction pat-27 terns within a community (Memmott et al., 2004; Rezende et al., 2007; Bascompte & Stouffer, 2009). Recovering the lost biodiversity and interactions through ecological restoration has become increasingly imperative, and a key restoration aim is to facilitate assembly of robust interaction networks (Menz et al., 2010). We know little, however, about how to re-assemble interacting communities through restoration, or the process of ecological network assembly more generally. The most widely explored mechanism of network assembly, preferential attachment (Barabási & Albert, 1999), predicts that species entering a network are more likely to interact with species that are already well-connected ("the rich-get-richer" principle, Barabási & Albert, 1999). In pollination systems — a particularly ubiquitous mutualism (Ollerton et al., 2011; Klein et al., 2007) — some studies have found support for this mechanism of assembly. Investigating the day-today, temporal assembly of a plant-pollinator network within a season, Olesen et al. (2008) found that new species tended to interact with already well-connected species, potentially because these species are either more abundant or more temporally persistent (Olesen et al., 2008). In addition, using a space-for-time substitution to study primary succession along a glacier foreland, Albrecht 41 et al. (2010) found some evidence that assembly occurred through preferential attachment. Network nestedness, a pattern of interactions where a generalist core interacts with both specialist 43 and generalist species, increased as the community aged (Albrecht et al., 2010). Increasing nestedness could result from preferential attachment process where specialist species attach to the 45 well-connected, generalist core.

In contrast to the ordered network build-up described by preferential attachment, significant reorganizations of interactions can punctuate assembly (Peel & Clauset, 2014). Such significant reorganizations of interactions, or network changing points, are observed in social networks responding
to abrupt shifts in the behavior of interactors (Peel & Clauset, 2014). In ecological communities,
such shifts may occur if, as new species colonize, resident species change their interaction partners to optimize their foraging effort. In plant-pollinator communities, theory predicts pollinators
optimize their use of floral resources to reduce interspecific competition and improve resource-use
efficiency (Pyke, 1984; Valdovinos *et al.*, 2010, 2013; Albrecht *et al.*, 2010; Blüthgen *et al.*, 2007).
No studies, however, have examined whether changing points occur during ecological network
assembly, and how these changes relate to the species behavior.

Understanding network assembly is particularly relevant to ecological restoration, which is essentially 'applied succession' (e.g., Parker, 1997). In pollination systems, the time since an area was restored has been shown to affect the structure of networks (Forup *et al.*, 2008a,b; Devoto *et al.*, 2012), suggesting interactions are evolving as the community develops. Understanding the mechanisms of network assembly will help to guide community restoration.

Facilitating network restoration is especially imperative in areas where species interactions provide essential ecosystem services, such as crop pollination. In intensively managed agricultural land-scapes, the demand for pollination services is the greatest (Kremen, 2008). However, honey bees, managed extensively around the world to provide crop pollination, are in global decline (Neumann & Carreck, 2010; van Engelsdorp *et al.*, 2009). In addition, native pollinators, which are capable of providing sufficient crop pollination (Kremen *et al.*, 2002; Winfree *et al.*, 2007; Kremen *et al.*, 2004), are in short supply because these landscapes make poor habitats for pollinator populations (Kremen *et al.*, 2002). To ensure provision the continued provision of ecosystem services and curb biodiversity loss, effective restoration of pollinators and their interactions in agricultural landscapes is critical.

To promote pollinator services in agriculture, farmers may chose to plant strips of native plants along farm edges (hedgerows) to help provide habitat for pollinators without removing arable land from production. Hedgerows augment the richness, abundance and trait diversity of pollinators in agricultural landscapes (Morandin & Kremen, 2013; M'Gonigle *et al.*, 2015; Kremen & M'Gonigle, 2015; Ponisio *et al.*, 2015). In addition, hedgerows promote the persistence and colonization of floral resource specialists (M'Gonigle *et al.*, 2015). Little is known, however, about how new species are being incorporated into the network as the community assembles, or the consequences for interaction patterns and robustness.

Using a long-term dataset of plant-pollinator communities assembling following hedgerow restora-80 tion in the highly simplified and intensively managed agricultural landscape of California's Central 81 Valley, we explore the process of network development. We first determine whether the mecha-82 nism underlying network assembly is a smooth build up of interactions as would be predicted by 83 preferential attachment, or punctuated by significant reorganizations of interactions (i.e., network 84 changing points). Even with changing points in interaction organization, networks could still be 85 assembling via preferential attachment if the network reorganizations were primarily driven the by peripheral, temporally variable species while a stable, well-connected core of species persist. We 87 test whether the species are most variable in their network position — and thus important contributors network reorganizations — are less persistent and connected species. To further explore the mechanisms underlying the temporal dynamics in the networks, we examine patterns in the species and interaction temporal turnover. Lastly, we investigate whether networks are assembling toward 91 predictable interaction patterns, and the ramifications for the robustness of the networks to species extinction and cascading perturbations.

# Materials & Methods

## 95 Study sites and collection methods

We surveyed plant-pollinator interaction networks of assembling hedgerows (N=5), as well as in two types of non-assembling communities to serve as controls: unrestored, weedy field margins 97 (N=19) and established hedgerows (greater than 10 years since planting, N=29). The sites were 98 located in the Central Valley of California in Yolo, Colusa and Solano Counties. This area is composed of intensively managed agriculture — primarily monocultures of conventional row crops, vineyards and orchards. Hedgerows we planted along field margins where they do not remove valuable land from production. Plantings are between are ca. 3–6 m wide and approximately 350 102 m long, bordering large (ca. 30-hectare) crop fields. Hedgerows consist of native, perennial, shrub and tree plantings including Rosa californica, Cercis occidentalis, Ceanothus spp., Heteromeles arbutifolia, Sambucus mexicana, Eriogonum spp., Baccharis spp., Salvia spp. and others (Kremen 105 & M'Gonigle, 2015; M'Gonigle et al., 2015). The mean distance between monitoring sites was 106 15 km, and the minimum distance between sites of the same type sampled in the same year was 2 107 km. The entire area surveyed spanned almost 300 km<sup>2</sup>. The crop fields adjacent to all sites were 108 similarly managed as intensive, high-input monoculture. 109

Monitoring began in 2006 and continued through 2014. Surveys of the assembling hedgerows began the year before the area was restored. For logistical reasons, no sampling of assembling hedgerows was conducted in 2010. Sites were sampled between two and five times per year (Tables S1-S3). In each round of sampling, the order in which sites were sampled was randomized. Surveys were conducted under sunny conditions when the temperature was above 21°C and wind speed was below 2.5 meters/second.

Flower-visitors to plants in hedgerows and unrestored controls were netted for one hour of active search time (the timer was paused when handling specimens). Honeybees (*Apis mellifera*) were

not collected because their abundance is determined largely by hive placement by bee-keepers.

All other insect flower visitors that touched the reproductive parts of the flower were collected;

however, here we focus only on wild bees and syrphids — the most abundant and effective pollinators in the system (representing 49 and 19 percent of records, respectively, C. Kremen, A. Klein and L. Morandin, unpublished data). Bee and syrphid specimens were identified to species (or morpho-species for some bee specimens in the genera *Nomada* and *Sphecodes*) by expert tax-

Quantitative networks were generated for each site through time. To account for the unequal number of surveys between years (Tables S1-S3), we use the mean of the interactions between a pair of plants and pollinators across surveys within a year to represent interaction frequency.

## 128 Change point analysis

## 129 Identifying change points

We employed a change point detection method (Peel & Clauset, 2014) to identify fundamental 130 reorganizations in large-scale interaction patterns. A change point is caused by a merge, split, 131 fragmentation or formation of communities (also called modules or compartments). Change point 132 detection methods have yet to be generalized to quantitative networks, so for this analysis we 133 focused on qualitative (binary) networks. Following Peel & Clauset (2014), we first defined a probability distribution over the networks using the generalized hierarchical random graph model 135 (GHRG). The GHRG model is able to capture both assortative and disassortative community structure patterns at all scales in the network (Peel & Clauset, 2014). A network G is composed of vertices V and edges E. The GHRG model decomposes the N vertices into a series of nested 138 groups, the relationships among which are represented by the dendrogram T. The tips of T are the 139 vertices of G, and the probability that two vertices u and v connect is given by the parameter  $p_r$ .

The probability distribution of the network G thus defined as:

$$P(G|T, pr) = p_r^{E_r} (1 - p_r)^{N_r - E_r}$$
(1)

Where  $E_r$  is the observed number of edges between vertices with the common ancestor r, and  $N_r$  is the total possible edges.

Using Bayesian posterior inference and techniques from phylogenetic tree reconstruction, we fit the GHRG model to the networks (Peel & Clauset, 2014). This is accomplished by using a Markov chain Monte Carlo (MCMC) procedure to first sample the posterior distribution of bipartitions, from which a consensus tree is derived (Peel & Clauset, 2014). We use  $\beta$  distributions with the hyperparameters  $\alpha = \beta = 1$  to define priors for  $p_r$ .

Once the GHRG model has been fit to the networks, we determine whether a change point occurred 149 between two time slices. To detect a change point, we compare the fit of two models — one where a change point occurred between two networks, and one where no change occurred using posterior Bayes factors. We chose a sliding window of length, w, of four, within which to 152 find change points. Larger windows allow for more gradual changes, and four was the maximum 153 possible with our maximum of eight years of data. Lastly, to calculate a p-value for the Bayes 154 factors, we use parametric bootstrapping to numerically estimate the null distribution (Peel & 155 Clauset, 2014). We employed code published online by L. Peel for the change point analysis. 156 Analyses we conducted in Python 3.4. 157

We next test whether the change points occurring in maturing hedgerows were a component of
the assembly process or a product of environmental shifts that lead to network reorganizations
in all types of communities. We model the number of change points as successes and the total
number of years each site was sampled as trails, and use a generalized linear model with Binomial
error to test whether the probability of a change point occurring varied by site type. We used

standard techniques to determine whether the assumptions of the models were met for this and all subsequent models. For the non-assembling hedgerows and weedy field margins, only sites with five or greater survey years were included in this analysis. All statistical analysis were conducted in R 3.2.3 (R Core Team, 2015).

## 167 Characteristics of species that contribute to change points

To further elucidate the nature of the change points, we examine the characteristics of the species that contributed interaction reorganization. Some species remain in relatively similar network positions through time, whereas others are more variable in their role and thus contribute more strongly to network reorganization. We test whether the more persistent species with the highest degree are the most stable in their network positions, as would be expected if the networks were assembling via preferential attachment.

We calculate species persistence as the proportion of surveys a plant or pollinator is observed. 174 Species observed consistently within and between years are thus maximally persistent. Weighted 175 species degree is calculated from interaction observations from an extensive dataset from Yolo 176 County (approx. 18000 interaction records) that included both the data included in this study and 177 additional data from sites where we collected flower visitors using the same methods (M'Gonigle 178 et al., 2015; Ponisio et al., 2015). To represent network position variability, we computed the 179 coefficient of variation of weighted closeness at each site through time. Closeness describes the 180 centrality of a species in the network by calculating path lengths to other vertices (species) in the graph. We use linear mixed models to test whether the species closeness variability (log) is related to the persistence or degree of that species (Bates et al., 2014; Kuznetsova et al., 2014). 183 We included random effects for species, as well as site. Because the degree and persistence of 184 pollinators were strongly correlated, ( $\rho = 0.84$ , p-value  $< 2 * 10^{-16}$ ), we include each explanatory 185 variable in the model separately. Plant degree and persistent were not significantly correlated, but

we use the same models as we did for the pollinators for consistency. A linear increase in closeness, as might be expected with assembly by preferential attachment, would also lead to a high variability in closeness scores, so we test whether closeness increases through time.

## Species and interaction turnover

Reorganizations of network structure can be the result of species turnover or species changing their interaction partners (i.e., re-wiring). To better understand the mechanisms underlying the temporal dynamics of the assembling networks, we examined patterns of species and interaction turnover.

Assembling networks may have higher rates of pollinator turnover than non-assembling communities because new pollinator species are colonizing and establishing themselves (M'Gonigle *et al.*, 2015). Similarly, if pollinators try to maximize their foraging efficiency based on the species present, interactions may turnover more quickly than in established communities. In addition, at assembling hedgerows, plants may "colonize" the networks as they become more attractive resources.

To estimate the temporal species and interaction turnover, we use an approach similar to calcu-200 lating spatial  $\beta$ -diversity. Instead of calculating the variation in community composition across 201 sites within a year, however, we estimated turnover across years at a site. We first calculated the 202 pairwise dissimilarity of plants, pollinators and interactions between years within each site using a dissimilarity estimator that incorporates abundances, while also accounting for unobserved records (Chao et al., 2005). Dissimilarity estimates can be affected by the total number of species and in-205 dividuals sampled at a site (e.g., Ponisio et al., 2015). For example, the probability that two sites 206 do not share any species is higher when there are few individuals at those sites. Following Ponisio 207 et al. (2015), we use null models that constrained species richness to estimate the deviation of the 208 observed dissimilarity from that which would be expected under a random community assembly 209 process. With the corrected dissimilarity values, we then calculated the multivariate dispersion of 210

community composition across years (Anderson *et al.*, 2011). In order to test whether assembling hedgerows had more turnover of species and interactions than non-assembling communities, the species and interaction temporal turnover estimates were included as response variables in a linear mixed models with site type as an explanatory variable along with a random effect of site (Bates *et al.*, 2014; Kuznetsova *et al.*, 2014).

Though species may turnover across years, some groups of species may essentially replace each other if they fill similar roles in the network. At non-assembling communities, species turnover 217 may overestimate the temporal changes in the networks if the interactions occurring in one year 218 are similar to those in the next year when they are weighted by the similarity of their constituent 219 species. We develop a method examine the temporal turnover of interactions weighted based on 220 their similarity. We followed Ahn et al. (2010) algorithm to hierarchically cluster all the interac-221 tions (edges) across sites and years based on their similarity, and build a dendrogram. The interac-222 tion similarity is based how may plants and pollinators (vertices) two edges share (Ahn et al., 2010; 223 Kalinka & Tomancak, 2011). The more species edges shared in common, the shorter the branch 224 length between them on the dendrogram. We next calculated the temporal turnover of interactions 225 weighted by their similarity, as approximated by "phylogenetic" distance (Graham & Fine, 2008; 226 Kembel et al., 2010). When then use linear models to test whether the weighted turnover of inter-227 actions varied between assembling and non-assembling networks (Bates et al., 2014; Kuznetsova 228 et al., 2014). 229

# Temporal changes in interaction patterns

## Network structure

Any changing points in network structure may contribute to the reorganization of the assembling 232 networks into predictable interaction patterns. Pollination networks exhibit two main structural 233 patterns — modularity (e.g., Olesen et al., 2007) and nestedness (e.g., Bascompte et al., 2006, 234 2003). In modular networks, interactions are insular, occurring within separate groups or "mod-235 ules" more often than between modules. Modules in the network may fragment as the network 236 assembles, enhancing modularity. Conversely, nested networks are like a pyramid of interactions, 237 where there are some species that interact with many species, other species that interact with a sub-238 set of those species, and so on. If species entering the network tend to interact with the generalist base of the network pyramid as would be expected with preferential attachment, nestedness would increase through time. The connectance — the proportion of observed out of possible interactions — is also expected to increase as species preferentially attach to the core. Is this true? Lastly, the network specialization may change as the community assembles. If specialist species colo-243 nize the network or species begin to limit their interaction niche breath as the network assembles, 244 network-level specialization will increase (Blüthgen et al., 2006). 245 To evaluate network nestedness, we used the estimator weighted NODF (Almeida-Neto et al., 2008). NODF evaluates whether species with fewer partners interact with subsets of partners with which more connected species interact (Almeida-Neto et al., 2008). To estimate modularity, we 248 use a hierarchical clustering algorithm (Newman & Girvan, 2004; Csardi & Nepusz, 2006). We 249 calculated standardized z-scores so that nestedness and modularity metrics could be compared be-250 tween communities. The z-scores were calculated by generating an ensemble of 999 randomly 251 assembled communities, subtracting the mean of the statistic calculated across these communities 252 from the observed value, and then dividing by the standard deviation. To assemble random communities, we reshuffled the interactions between species but fixed the total number of interactions, species and interaction frequency distributions (Galeano *et al.*, 2009). We evaluate network specialization with the metric H2, which estimates the deviation of the observed interaction frequency between plants and pollinators from a null expectation where all partners interact in proportion to their abundances (Blüthgen *et al.*, 2006). It ranges from zero for generalized networks to one for specialized networks.

To test whether network modularity, nestedness, connectance or specialization changed linearly with assembly, we use linear mixed models with the descriptive network metrics as the response variable, year of assembly as the explanatory variable, and random effects of site and year. The number of species in a network affects affects the patterns of interaction possible, so we also examined the change in plant and pollinator species richness through time. We employ generalized linear mixed models with Poisson error to model richness. We scaled explanatory variables.

### 266 Network robustness

Lastly, we test whether the changes in interaction patterns associated with network assembly affect 267 the robustness of the network to species loss and to cascading perturbations. Following Memmott 268 et al. (2004), we simulate plant species extinction and the subsequent extinction cascades of polli-269 nator species. Because the reproduction of plant species if facilitated by active restoration efforts, it is unlikely the extinction of pollinator species would affect plant populations in the hedgerows. However, plants ceasing to bloom, for example in response to drought, will likely affect the pollina-272 tors that depend on them. We eliminate plants species based on their degree or abundance, and then 273 calculate the number of pollinators that secondarily went extinct. The area below the extinction 274 curve is an estimate of network robustness (Memmott et al., 2004; Dormann et al., 2008). 275

We also explored how the robustness to cascading perturbations changed as community assembled,

using algebraic connectivity — the second smallest eigenvalue of the Laplacian matrix (Fiedler, 1973) — as a proxy for network robustness (Gaiarsa et al., submitted). Algebraic connectivity relates to how difficult it is to turn a network into completely disconnected groups of species (Costa et al., 2007). The larger the algebraic connectivity, the more robust a network is to cascading perturbations, and the harder it is to break the community into isolated groups of species (Gaiarsa et al., submitted).

# Results

Over eight years and 747 samples, we collected and identified 19,547 wild bees and syrphids comprising 173 species from 50 genera. We observed 1,521 unique interactions between plants and pollinators.

# 287 Change point analysis

## 288 Identifying change points

The majority (76%) of the sites underwent at least one significant interaction reorganization (Fig. 1). 289 All five of the assembling hedgerows experienced network changing points, whereas only 40% 290 and 81% of non-assembling hedgerows and field margins, respectively, underwent significant in-291 teraction reorganizations. Assembling hedgerows had significantly more changing points than the non-assembling networks (estimate of the difference in the odds ratios between assembling and non-assembling networks, 3.316, 95% CI [1.314, 8.572], p-value= 0.0117). Network assembly 294 following restoration is thus punctuated by more interaction reorganizations than would be ex-295 pected by environmental shifts alone that would effect assembling and non-assembling networks 296 equally. 297

### **Characteristics of species that contribute to change points**

In contradiction to the predictions of assembly by preferential attachment, both pollinator persis-299 tence and degree were positively related to network position variability (estimate of the slope of 300 closeness variability and persistence  $\pm$  standard error of the estimate, 0.653  $\pm$  0.225, p-value=0.009; slope of closeness variability and degree,  $0.008 \pm 0.002$ , p-value=0.002). In addition, plant persistence and degree were not significantly related to network position variability (estimate of the 303 slope of closeness variability and persistence  $\pm$  standard error of the estimate,  $-2.063 \pm 3.091$ , p-304 value=0.5; slope of closeness variability and degree, 0.0018  $\pm$  0.002, p-value=0.3). The variability 305 of species network position was not the result of closeness linearly increasing through time, and, in 306 fact, plant and pollinator closeness decreased slightly through time (Fig. S1, estimate of the slope 307 of closeness through time  $\pm$  standard error of the estimate, pollinators:  $-0.0003 \pm 0.00005$ , p-308 value= $2.7 * 10^{-12}$ ; plants  $-0.007 \pm 0.001$ , p-value= $1.4 * 10^{-6}$ ). Through statistically significant, 309 the slopes are so slight they may not me biologically significant. 310

# Species and interaction turnover

The rates of plant, pollinator and interaction temporal turnover were similar across assembling hedgerows, non-assembling hedgerows and field margins, though mature hedgerows has marginally significantly less pollinator turnover than field margins (estimate  $\pm$  standard error of the estimate of the difference in turnover between field margins and mature hedgerows,  $-0.0498 \pm 0.026$ , p-value=0.058). When interactions where weighted by their similarity, both assembling and mature hedgerows had higher rates of turnover than field margins (estimate  $\pm$  standard error of the estimate of the difference in turnover between field margins and assembling hedgerows, 0.115  $\pm$  0.027, p-value=0.0002; field margins and mature hedgerows, 0.082  $\pm$  0.024, p-value=0.002). The weighted interaction turnover at assembling hedgerows, however, was not significantly higher than

in non-assembling, mature hedgerows.

# Temporal changes in interaction patterns

### Network structure

Network nestedness significantly increased with assembly (Fig. 5, estimate of the slope of nestedness through time  $\pm$  standard error of the estimate, 1.834  $\pm$  0.6142, p-value=0.022). Modularity 325 decreased (Fig. 5), though the slope was not significantly different from zero (estimate of the slope 326 of modularity through time  $\pm$  standard error of the estimate,  $-0.524 \pm 0.295$ , p-value=0.124). 327 Connectance decreased as the community assembled (Fig. 5, estimate of the slope of connectance 328 through time  $\pm$  standard error of the estimate,  $-0.0434 \pm 0.0152$ , p-value=0.03). In contrast, 329 specialization remained constant (estimate of the slope of specialization through time  $\pm$  standard 330 error of the estimate,  $0.003 \pm 0.015$ , p-value=0.827). 331 Both plant and pollinator species richness increased through time (Fig. 5, estimate of the slope of 332 richness through time  $\pm$  standard error of the estimate, pollinators: 0.193  $\pm$  0.0729, p-value=0.008; 333 plants:  $0.212 \pm 0.0653$ , p-value=0.001). Unsurprisingly, pollinator species are colonizing and per-334 sisting at the assembling hedgerows. Though some new plant species may establish themselves in 335 the hedgerows, because the plant species richness is based on the flowers in the network and not 336 the presence of a plant at a site, the increase in plant richness is likely due to plants attracting 337 visitors as they offer better rewards.

### Network robustness

Assembly did not effect the robustness of the networks to species extinction when species where removed incrementally by degree (estimate of the slope of robustness through time  $\pm$  standard error of the estimate,  $6*10^{-5} \pm 4*10^{-3}$ , p-value=0.987) or abundance (0.001  $\pm$  0.003, p-value=0.65, Fig. 6).

In contrast, the robustness of networks to cascading perturbations, as measured by the algebraic connectivity of the network, increased as the network assembled (estimate of the slope of robustness to cascading perturbations through time  $\pm$  standard error of the estimate, 0.6814  $\pm$  0.272, p-value=0.042, Fig. 6).

# Discussion

We show that the temporal assembly of plant-pollinator networks following restoration is a highly dynamic process where interactions often undergo significant reorganizations. These network or-350 ganizations are unlikely a product of environmental forces alone because the network changing 351 points in non-assembling communities are less frequent, and few consistent trends in when change 352 points occurred across all sites. Several sites had network changing points between years 2009 and 353 2011 (Fig. 1). In California, 2011 marked the beginning of a multi-year drought. The assembling 354 hedgerows were not sampled in 2010, so disentangling whether the changing points are due to skip-355 ping a year of assembly or the drought is not possible. Interestingly, most assembling hedgerows 356 did not undergo a significant interaction reorganization immediately after a hedgerow was planted 357 (i.e., the transition from weedy field margin to hedgerow). This result is consistent with the find-358 ing that in our study system, hedgerow restoration takes several years to have an impact on the 359 plant-pollinator communities (Kremen and M'Gonigle, in prep). 360

In addition to finding multiple network organization changing points during assembly, the species that contribute most to those reorganizations were not what is expected by preferential attachment. 362 Based on a preferential attachment-like mechanism, we expect that the most persistent and high 363 degree species would remain stable in the network core during assembly. Surprisingly, however, 364 we encountered the opposite pattern. For example, the five most ubiquitous species in our study 365 landscape — Halictus ligatus, Halictus tripartitus, Lasioglossum (Dialictus) incompletum, and 366 Toxomerus marginatus — were the only species that changed what module (i.e., community) of 367 which they were a member across years in all the assembling hedgerows. Because species degree 368 and persistence were strongly correlated, it is difficult to disentangle the causal mechanism for why 369 species with those characteristics are so variable in their network position. Generalized species 370 may be able to better exploit the limited floral resources in the intensively managed agriculture 371 landscape, and thus also most persistent. More persistent species also have longer phenologies, so 372 they can visit many different flowers, resulting in a higher degree. Either way, our result suggests 373 that adaptable species can change their network position to utilize the most advantageous floral 374 resources available, which may depend on the other pollinator species that are present and the state 375 of the plant community. Thus given the opportunity and ability to use different resources, species 376 will often change their network positions. 377

Interestingly, though assembling hedgerows had more network reorganizations than non-assembling communities, pollinator species and interaction turnover occurred at similar rates across site types.

Assembling hedgerows have higher turnover than non-assembling field margins only when interactions were weighted by their similarity. This is likely because though species and interactions are turning over at the field margins, species and interactions that fill similar roles in the network are replacing each other. In contrast, at the assembling hedgerows, unique interactions are turning over as the networks continually reorganize. Non-assembling mature hedgerow communities, however, had similar rates of weighted interaction turnover as assembling hedgerows but also the lowest pollinator turnover. Pollinator communities at mature hedgerows may be generally more

stable, but rare and/or specialized pollinators could generate this pattern if they entered a community, formed unique interactions with plants that did not previously share pollinators, but did
not persist in the networks. These species would not contribute strongly to network reorganization or species turnover, but would enhance weighted interaction turnover. Mature hedgerows may
thus both support more stable pollination communities and likely ecosystem services, while also
providing resources for rare and/or specialized species (Kremen & M'Gonigle, 2015; M'Gonigle
et al., 2015).

The frequent changing points in network organization, dynamic nature of species positions in the 394 networks, and turnover of species and interactions all point to an assembly mechanism other than 395 preferential attachment. Nestedness did increase with years post-restoration, as would be expected 396 if colonizing, specialist species attached to a central, generalist core (Albrecht et al., 2010). With 397 preferential attachment, however, we would also expect connectance and specialization to increase, 398 and we found no such trends. Wait if connectance increases doesn't that mean specialization de-399 creases? Which would we expect to change with preferential attachment? The stable level of 400 network-level specialization through the assembly process may be due to the increased coloniza-401 tion of specialized species (M'Gonigle et al., 2015) accompanied by an increase in the diet breath 402 of resident species. This would be expected if resident species were able to minimize their foraging 403 time by expanding their diet breath as plant diversity increases with hedgerow maturation (Waser 404 et al., 1996; Pyke, 1984; Blüthgen et al., 2007; Albrecht et al., 2010). Such a change in pollinator 405 behavior would also explain the increase in network nestedness. 406

The changes in network patterns with assembly also increased the robustness of the networks to cascading perturbations – providing further evidence that hedgerows are a valuable tool for promoting species conservation and ecosystem provision in agricultural areas (M'Gonigle *et al.*, 2015; Ponisio *et al.*, 2015; Kremen & M'Gonigle, 2015). Because network vulnerability to cascading perturbations, as measured by algebraic connectivity, is correlated with species richness, the increase

and plant and pollinator richness following restoration is at least partially responsible for enhancing network robustness to cascading effects. The relationship between diversity and stability in net-413 works has been the subject of considerable debate (e.g., May, 1972; Pimm, 1984; Montoya et al., 2006). Our results provide one of the few empirical examples of how restoring species diversity 415 contributes to enhancing network stability. Though these hedgerows were designed to promote 416 floral resources across the growing season and not specifically to promote any specific network 417 properties (Menz et al., 2010), the pollinators assembled into resilient interaction patterns. With 418 prior knowledge of the floral preferences of pollinators, future restoration efforts may also be able 419 to incorporate promoting network stability into planning efforts (M'Gonigle et al., 2016) 420

Interestingly, however, the robustness of hedgerow communities to species loss did not change with assembly. This is particularly surprising given the observed increase in nestedness, which is often associated with an enhanced in robustness to extinction (Memmott *et al.*, 2004). Assembling hedgerows have yet to reach sufficient levels of nestedness to realize the benefits nestedness confers. Nestedness of the assembling hedgerows, however, did not asymptote within the eight years following restoration that the sites were surveyed, so hedgerow networks may eventually reach sufficient levels of nestedness to gain the robustness advantage.

Plant-pollinator networks, in general, are highly dynamic, with high turnover of species and inter-428 actions both within and between seasons (Burkle & Alarcón, 2011). Though our non-assembling 429 communities experience fewer network reorganizations than the assembling hedgerows, 82% of 430 field margins and 40% of mature hedgerows underwent at least one changing point in network 431 structure. Pollinators are also highly opportunistic (Petanidou et al., 2008; Vázquez, 2005; Al-432 brecht et al., 2010), though trait complementarity such as tongue length and corolla depth impose 433 some biophysical limits to the interactions between plants and pollinators (Vázquez et al., 2009; 434 Vázquez et al., 2009; Stang et al., 2009, 2006; Santamaría & Rodríguez-Gironés, 2007). Such 435 opportunism may buffer plant-pollinator communities from global change (e.g., Ramos-Jiliberto

et al., 2012; Kaiser-Bunbury et al., 2010), but our limited understanding of the assembly of these communities impedes making such predictions (Vázquez et al., 2009; Burkle & Alarcón, 2011). 438 Unlike in the broader food web literature, we have few mechanistic models of mutualistic network 439 assembly (Valdovinos et al., 2013). In addition, the few developed often borrow their mechanisms 440 from competitive interactions, leading to inaccurate biological assumptions (Holland et al., 2006). 441 We need further development of mechanistic models of mutualistic system to generate testable pre-442 dictions, along with empirical exploration of network assembly. Plant-pollinator communities and 443 mutualisms broadly are vital for biodiversity maintenance and essential ecosystem service provi-444 sion. We must therefore understand the processes underlying their assembly to facilitate restoration 445 and conservation.

# 447 Acknowledgments

We would like to thank Paulo Guimarães and Aaron Clauset for their invaluable discussions, and Leto Peel for help with the change point analysis. We thank the growers and land owners that 449 allowed us to work on their property. We also greatly appreciate the identification assistance of 450 expert taxonomists Martin Hauser, Robbin Thorp and Jason Gibbs. This work was supported by 451 funding from the Army Research Office (W911NF-11-1-0361 to CK), the Natural Resources Con-452 servation Service (CIG-69-3A75-12-253, CIG-69-3A75-9-142, CIG-68-9104-6-101 and WLF-69-7482-6-277 to The Xerces Society), the National Science Foundation (DEB-0919128 to CK), The 454 U.S. Department of Agriculture (USDA-NIFA 2012-51181-20105 to Michigan State University). Funding for LCP was provided by an NSF Graduate Research Fellowship, the USDA NIFA Graduate Fellowship, and the Berkeley Institute for Data Science. Funding for MPG was provided by 457 São Paulo Research Foundation (FAPESP, grant 2013/13319-5). We also appreciate the Santa Fe 458 Institute for faciliting this international collaboration. 459

# References

- Ahn, Y.Y., Bagrow, J.P. & Lehmann, S. (2010). Link communities reveal multiscale complexity in networks. *Nature*, 466, 761–764.
- Alarcón, R., Waser, N.M. & Ollerton, J. (2008). Year-to-year variation in the topology of a plant–pollinator interaction network. *Oikos*, 117, 1796–1807.
- Albrecht, M., Riesen, M. & Schmid, B. (2010). Plant–pollinator network assembly along the chronosequence of a glacier foreland. *Oikos*, 119, 1610–1624.
- Almeida-Neto, M., Gumarães, P., Gumarães, P.R., Loyola, R.D. & Ulrich, W. (2008). A consistent metric for nestedness analysis in ecological systems: reconciling concept and measurement.
- *Oikos*, 117, 1227–1239.
- 470 Anderson, M.J., Crist, T.O., Chase, J.M., Vellend, M., Inouye, B.D., Freestone, A.L., Sanders,
- N.J., Cornell, H.V., Comita, L.S., Davies, K.F. et al. (2011). Navigating the multiple meanings
- of  $\beta$  diversity: a roadmap for the practicing ecologist. *Ecol. Lett.*, 14, 19–28.
- Barabási, A.L. & Albert, R. (1999). Emergence of scaling in random networks. *science*, 286, 509–512.
- Barnosky, A.D., Matzke, N., Tomiya, S., Wogan, G.O., Swartz, B., Quental, T.B., Marshall, C.,
- McGuire, J.L., Lindsey, E.L., Maguire, K.C. et al. (2011). Has the earth/'s sixth mass extinction
- already arrived? *Nature*, 471, 51–57.
- Bascompte, J., Jordano, P., Melián, C.J. & Olesen, J.M. (2003). The nested assembly of plant-
- animal mutualistic networks. *Proc. Natl. Acad. Sci. USA*, 100, 9383–9387.
- Bascompte, J., Jordano, P. & Olesen, J.M. (2006). Asymmetric coevolutionary networks facilitate biodiversity maintenance. *Science*, 312, 431–433.

- Bascompte, J. & Stouffer, D.B. (2009). The assembly and disassembly of ecological networks.
- <sup>483</sup> *Phil. Trans. R. Soc. B*, 364, 1781.
- Bates, D., Maechler, M., Bolker, B. & Walker, S. (2014). lme4: Linear mixed-effects models using
- Eigen and S4. R package version 1.1-7.
- Blüthgen, N., Menzel, F. & Blüthgen, N. (2006). Measuring specialization in species interaction
- networks. *BMC Ecol.*, 6, 9.
- Blüthgen, N., Menzel, F., Hovestadt, T., Fiala, B. & Blüthgen, N. (2007). Specialization, con-
- straints, and conflicting interests in mutualistic networks. *Curr. Biol.*, 17, 341–346.
- Burkle, L.A. & Alarcón, R. (2011). The future of plant-pollinator diversity: Understanding in-
- teraction networks across time, space, and global change. American Journal of Botany, 98,
- 492 528.
- <sup>493</sup> Chao, A., Chazdon, R.L., Colwell, R.K. & Shen, T.J. (2005). A new statistical approach for
- assessing similarity of species composition with incidence and abundance data. *Ecol. Lett.*, 8,
- 495 148–159.
- Costa, L.d.F., Rodrigues, F.A., Travieso, G. & Villas Boas, P.R. (2007). Characterization of com-
- plex networks: A survey of measurements. Advances in physics, 56, 167–242.
- 498 Csardi, G. & Nepusz, T. (2006). The igraph software package for complex network research.
- 499 *InterJournal*, Complex Systems, 1695.
- Devoto, M., Bailey, S., Craze, P. & Memmott, J. (2012). Understanding and planning ecological
- restoration of plant–pollinator networks. *Ecology letters*, 15, 319–328.
- Díaz-Castelazo, C., Guimaraes Jr, P.R., Jordano, P., Thompson, J.N., Marquis, R.J. & Rico-Gray,
- V. (2010). Changes of a mutualistic network over time: reanalysis over a 10-year period. *Ecol-*
- ogy, 91, 793–801.

- Dormann, C., Gruber, B. & Fründ, J. (2008). Introducing the bipartite package: analysing ecological networks. *R News*, 8, 8.
- Dunn, R.R., Harris, N.C., Colwell, R.K., Koh, L.P. & Sodhi, N.S. (2009). The sixth mass coextinction: are most endangered species parasites and mutualists? *Proceedings of the Royal Society*
- of London B: Biological Sciences, 276, 3037–3045.
- van Engelsdorp, D., Evans, J., Saegerman, C., Mullin, C., Haubruge, E., Nguyen, B., Frazier, M.,
- Frazier, J., Cox-Foster, D., Chen, Y., Underwood, R., Tarpy, D. & Pettis, J. (2009). Colony
- collapse disorder: A descriptive study. *Plos One*, 4, 1–17.
- Fang, Q. & Huang, S.Q. (2012). Relative stability of core groups in pollination networks in a biodiversity hotspot over four years. *PloS one*, 7, e32663.
- Fiedler, M. (1973). Algebraic connectivity of graphs. *Czechoslovak mathematical journal*, 23, 298–305.
- Forup, M., Henson, K., Craze, P. & Memmott, J. (2008a). The restoration of ecological interactions: plant-pollinator networks on ancient and restored heathlands. *J. Appl. Ecol.*, 45, 742–752.
- Forup, M.L., Henson, K.S., Craze, P.G. & Memmott, J. (2008b). The restoration of ecological interactions: plant–pollinator networks on ancient and restored heathlands. *Journal of Applied Ecology*, 45, 742–752.
- Galeano, J., Pastor, J.M. & Iriondo, J.M. (2009). Weighted-interaction nestedness estimator (wine):
  a new estimator to calculate over frequency matrices. *Environmental Modeling & Software*, 24,
  1342–1346.
- Graham, C.H. & Fine, P.V. (2008). Phylogenetic beta diversity: linking ecological and evolutionary processes across space in time. *Ecology letters*, 11, 1265–1277.

- Holland, J.N., Okuyama, T. & DeAngelis, D.L. (2006). Comment on asymmetric coevolutionary networks facilitate biodiversity maintenance. *Science*, 313, 1887b–1887b.
- Kaiser-Bunbury, C.N., Muff, S., Memmott, J., Müller, C.B. & Caflisch, A. (2010). The robustness of pollination networks to the loss of species and interactions: a quantitative approach incorporating pollinator behaviour. *Ecology Letters*, 13, 442–452.
- Kalinka, A.T. & Tomancak, P. (2011). linkcomm: an r package for the generation, visualization, and analysis of link communities in networks of arbitrary size and type. *Bioinformatics*, 27.
- Kembel, S., Cowan, P., Helmus, M., Cornwell, W., Morlon, H., Ackerly, D., Blomberg, S. & Webb, C. (2010). Picante: R tools for integrating phylogenies and ecology. *Bioinformatics*, 26, 1463–1464.
- Klein, A., Vaissière, B., Cane, J., Steffan-Dewenter, I., Cunningham, S., Kremen, C. & Tscharntke,
  T. (2007). Importance of pollinators in changing landscapes for world crops. *Proc. R. Soc. B*,
  274, 303–313.
- Kremen, C. (2008). *Bee Pollinators in Agricultural Ecosystems*, New York: Oxford University
   Press, chap. Crop Pollination Services From Wild Bees, pp. 10–26.
- Kremen, C. & M'Gonigle, L.K. (2015). Small-scale restoration in intensive agricultural landscapes supports more specialized and less mobile pollinator species. *J. Appl. Ecol.*, 52, 602–610.
- Kremen, C., Williams, N., Bugg, R., Fay, J. & Thorp, R. (2004). The area requirements of an ecosystem service: crop pollination by native bee communities in California. *Ecol. Lett.*, 7, 1109–1119.
- Kremen, C., Williams, N. & Thorp, R. (2002). Crop pollination from native bees at risk from agricultural intensification. *Proc. Natl. Acad. Sci. U.S.A.*, 99, 16812–16816.

- Kuznetsova, A., Bruun Brockhoff, P. & Haubo Bojesen Christensen, R. (2014). *ImerTest: Tests*
- for random and fixed effects for linear mixed effect models (lmer objects of lme4 package). R
- package version 2.0-11.
- May, R.M. (1972). Will a large complex system be stable? *Nature*, 238, 413–414.
- Memmott, J., Waser, N.M. & Price, M.V. (2004). Tolerance of pollination networks to species
- extinctions. *Proc. R. Soc. Lond. B*, 271, 2605–2611.
- Menz, M., Phillips, R., Winfree, R., Kremen, C., Aizen, M., Johnson, S. & Dixon, K. (2010).
- Reconnecting plants and pollinators: challenges in the restoration of pollination mutualisms.
- 557 *Trends Plant Sci.*, 16, 4–12.
- <sup>558</sup> M'Gonigle, L., Ponisio, L., Cutler, K. & Kremen, C. (2015). Habitat restoration promotes pollina-
- tor persistence and colonization in intensively-managed agriculture. *Ecol. Appl.*, 25, 1557–1565.
- <sup>560</sup> M'Gonigle, L.K., Williams, N.M., Lonsdorf, E. & Kremen, C. (2016). A tool for selecting plants
- when restoring habitat for pollinators. *Conservation Letters*.
- Montoya, J.M., Pimm, S.L. & Solé, R.V. (2006). Ecological networks and their fragility. *Nature*,
- 563 442, 259–264.
- Morandin, L. & Kremen, C. (2013). Hedgerow restoration promotes pollinator populations and
- exports native bees to adjacent fields. *Ecol. Appl.*, 23, 829–839.
- Neumann, P. & Carreck, N.L. (2010). Honey bee colony losses. J. Api. Res., 49, 1-6.
- Newman, M.E.J. & Girvan, M. (2004). Finding and evaluating community structure in networks.
- 568 Phys. Rev. E, 69, 026113.
- Olesen, J.M., Bascompte, J., Dupont, Y. & Jordano, P. (2007). The modularity of pollination
- networks. *Proc. Natl. Acad. Sci. USA*, 104, 19891–19896.

- Olesen, J.M., Bascompte, J., Elberling, H. & Jordano, P. (2008). Temporal dynamics in a pollination network. *Ecology*, 89, 1573.
- Ollerton, J., Winfree, R. & Tarrant, S. (2011). How many flowering plants are pollinated by animals? *Oikos*, 120, 321–326.
- Parker, V.T. (1997). The scale of successional models and restoration objectives. *Restoration ecology*, 5, 301–306.
- Peel, L. & Clauset, A. (2014). Detecting change points in the large-scale structure of evolving networks. *arXiv preprint arXiv:1403.0989*.
- Petanidou, T., Kallimanis, S., Tzanopoulos, J., Sgardelis, S. & Pantis, J. (2008). Long-term observation of a pollination network: fluctuation in species and interactions, relative invariance of network structure and implications for estimates of specialization. *Ecol. Lett.*, 11, 564–575.
- Pimm, S.L. (1984). The complexity and stability of ecosystems. *Nature*, 307, 321–326.
- Ponisio, L.C., M'Gonigle, L.K. & Kremen, C. (2015). On-farm habitat restoration counters biotic homogenization in intensively managed agriculture. *Global change biology*.
- Pyke, G.H. (1984). Optimal foraging theory: a critical review. *Annual review of ecology and*systematics, pp. 523–575.
- R Core Team (2015). *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Vienna, Austria.
- Ramos-Jiliberto, R., Valdovinos, F.S., Moisset de Espanés, P. & Flores, J.D. (2012). Topological plasticity increases robustness of mutualistic networks. *Journal of Animal Ecology*, 81, 896–904.

- Rezende, E.L., Lavabre, J.E., Guimarães, P.R., Jordano, P. & Bascompte, J. (2007). Non-random coextinctions in phylogenetically structured mutualistic networks. *Nature*, 448, 925–928.
- Santamaría, L. & Rodríguez-Gironés, M.A. (2007). Linkage rules for plant–pollinator networks:

  Trait complementarity or exploitation barriers? *PLoS Biol.*, 5, e31.
- Stang, M., Klinkhamer, P., Waser, N.M., Stang, I. & van der, M.E. (2009). Size-specific interaction patterns and size matching in a plant-pollinator interaction web. *Ann. Bot.*, 103.
- Stang, M., Klinkhamer, P.G. & Van Der Meijden, E. (2006). Size constraints and flower abundance determine the number of interactions in a plant–flower visitor web. *Oikos*, 112, 111–121.
- Valdovinos, F.S., Moisset de Espanés, P., Flores, J.D. & Ramos-Jiliberto, R. (2013). Adaptive foraging allows the maintenance of biodiversity of pollination networks. *Oikos*, 122, 907–917.
- Valdovinos, F.S., Ramos-Jiliberto, R., Garay-Narváez, L., Urbani, P. & Dunne, J.A. (2010). Consequences of adaptive behaviour for the structure and dynamics of food webs. *Ecology Letters*, 13, 1546–1559.
- Vázquez, D.P. (2005). Degree distribution in plant-animal mutualistic networks: forbidden links or random interactions? *Oikos*, 108, 421–426.
- Vázquez, D.P., Blüthgen, N., Cagnolo, L. & Chacoff, N.P. (2009). Uniting pattern and process in plant-animal mutualistic networks: a review. *Annals of Botany*, 103, 1445–1457.
- Vázquez, D.P., Chacoff, N.P. & Cagnolo, L. (2009). Evaluating multiple determinants of the structure of plant-animal mutualistic networks. *Ecology*, 90, 2039–2046.
- Waser, N.M., Chittka, L., Price, M.V., Williams, N.M. & Ollerton, J. (1996). Generalization in pollination systems, and why it matters. *Ecology*, 77, 1043.

Winfree, R., Williams, N., Dushoff, J. & Kremen, C. (2007). Native bees provide insurance against ongoing honey bee losses. *Ecol. Lett.*, 10, 1105–1113.

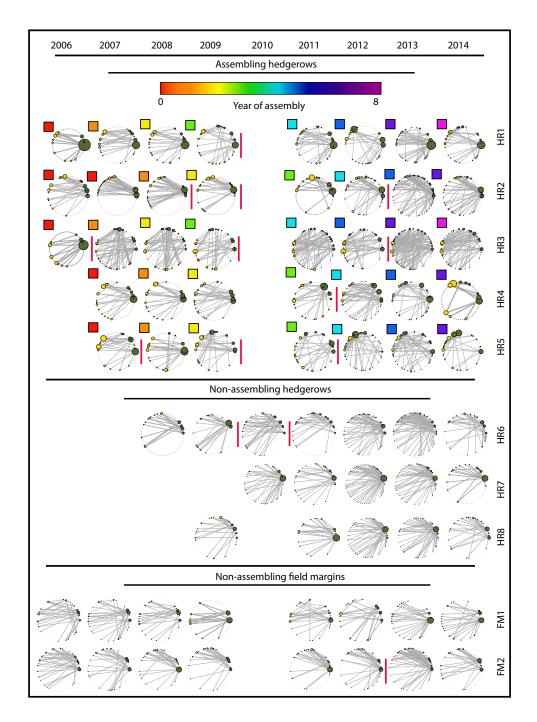


Figure 1: Assembling hedgerow networks had more changing points (vertical red lines) than non-assembling hedgerows and weedy field margins (a representative sample of non-assembling sites are depicted here). In each network, plants and pollinators are represented by green and yellow circles, respectively, weighted by their degree. Each species has a consistent position in the network across years. In the assembling hedgerows, colored squares in the corner of each network represent the year of assembly. Before hedgerows are installed (when they are still field margins) the year of assembly is zero.

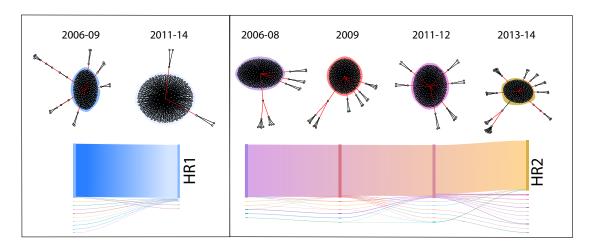


Figure 2: The community (i.e., module or compartment) membership between network changing points. Two representative assembling hedgerows are depicted. In the top panel, species are grouped by community. The bottom panels visualize the flow of species between communities between changing points.

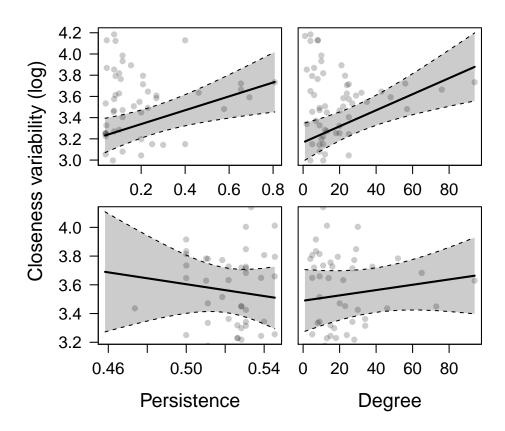


Figure 3: The coefficient of variation of network position, as represented by closeness, plotted against pollinator persistence and degree. Persistence and degree were positively related to network position variability in pollinators, but unrelated in plants. Points represent means for each species across sites. The solid line indicates the mean slope estimate and the dashed lines are the 95% confidence intervals around the estimate.

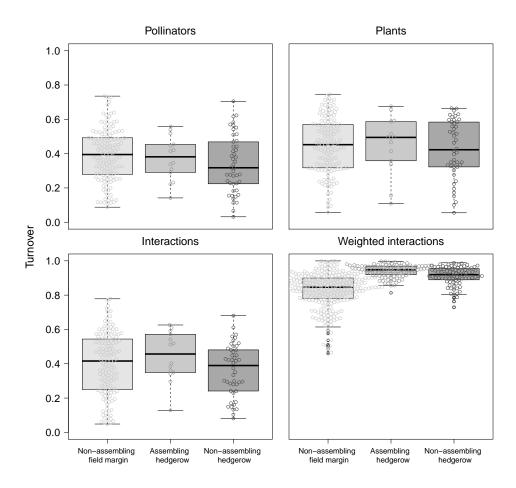


Figure 4: The species, interaction and weighted interactions turnover of plant-pollinator networks at non-assembling field margins sites, assembling hedgerows, and non-assembling, mature hedgerows. Rates of species and interaction turnover were similar between site types, though mature hedgerows has marginally significantly less pollinator turnover. However, when interactions where weighted by their similarity, both hedgerow types had higher turnover that unrestored field margins. Boxplots represent medians (black horizontal line) first and third quartiles (box perimeter) and extremes (whiskers).

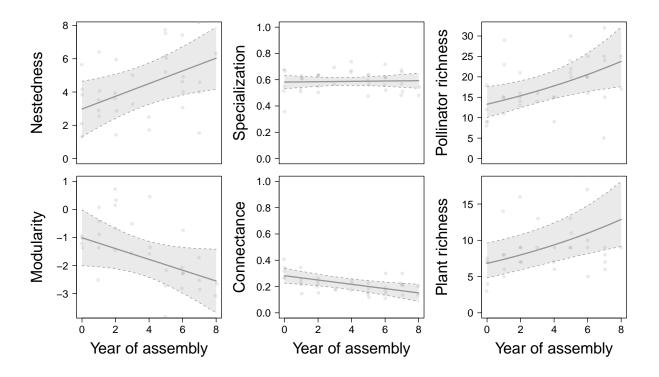


Figure 5: The nestedness, plant richness and pollinator richness increased as the networks assembled. Specialization and modularity remained consistent across years, while connectance decreased. The left panels represent z-scores. Scores greater than  $\sim 2$  or less than  $\sim -2$  are significantly more or less structured than randomly assembled networks. Points are the metric value for each site at each year of assembly. The solid line indicates the mean slope estimate and the dashed lines are the 95% confidence intervals around the estimate.

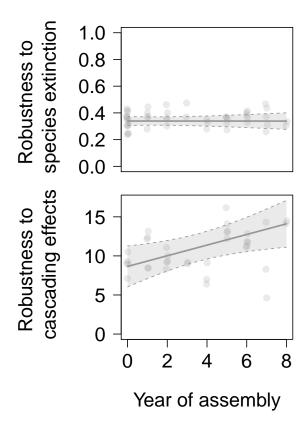


Figure 6: The robustness of networks to species extinction did not change with network assembly, but the resilience to to cascading perturbation increased. The robustness to species extinction is measured by incrementally removing species by degree, through removing species by abundance did not yield qualitatively different results. The robustness of networks to cascading perturbations is measured as the algebraic connectivity, the second smallest eigenvalue of the Laplacian matrix. Points are the value for each site at each year of assembly. The solid line indicates the mean slope estimate and the dashed lines are the 95% confidence intervals around the estimate.