## Mutations\_explorev2

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## Mutagenic data of spike protein

This is a notebook exploring the SARS-CoV-2 mutagenic data. The first step is to import the tidyverse library which will be used to conduct exploratory data analysis, and import the data.

```
library(tidyverse)
## -- Attaching packages ----- tidyverse 1.3.0 --
## v ggplot2 3.3.1
                   v purrr
                            0.3.4
## v tibble 3.0.1
                   v dplyr
                            1.0.0
## v tidyr 1.1.0
                   v stringr 1.4.0
         1.3.1
## v readr
                   v forcats 0.5.0
## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()
                  masks stats::lag()
file<-read.csv("data.csv",header= TRUE, sep=",")</pre>
dim(file)
## [1] 2340
head(file, N=6L)
    Residue.. Substitution
                          Top_rep1
                                   Top_rep2 Bottom_rep1 Bottom_rep2
## 1
                      A -0.1764999 0.2978451 -0.2039027 -0.24789819
          19
## 2
          19
                      ## 3
          19
                      D -1.4911309 -1.2155133
                                             0.9672447 1.27736869
## 4
          19
                      E -1.0613240 -1.8116679
                                            1.4735906 1.77240860
## 5
          19
                      F 1.2298997 1.0344321 -1.0662619 -1.18149682
## 6
                      G -1.2820811 -1.4374131
                                             0.9239352 1.21642268
          19
tail(file, N=-6L)
```

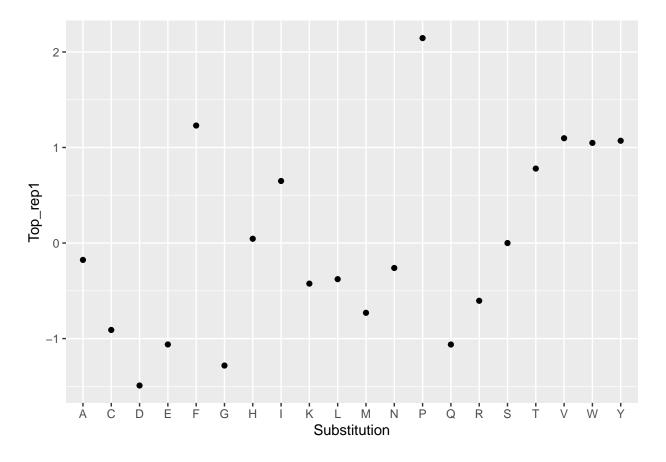
```
##
        Residue.. Substitution
                                  Top_rep1
                                              Top_rep2 Bottom_rep1 Bottom_rep2
## 2335
              518
                             R 0.0000000 0.00000000
                                                         0.0000000
                                                                     0.0000000
                                                                    -0.9474402
## 2336
              518
                                0.11209716 0.02784838
                                                       -0.3241399
## 2337
                                0.09676146 -0.77840324
                                                        -0.6421838
              518
                                                                    -0.5398540
## 2338
              518
                             V -0.48007793 -0.35046816
                                                        -0.5480825
                                                                    -0.4105959
## 2339
              518
                             W -0.14315194 -0.68674607
                                                        -0.4058743
                                                                    -0.3200862
## 2340
              518
                             Y -1.91622539 -0.05349618 -0.3532171
                                                                    -0.9761237
```

```
fin_res=file[2340,"Residue.."]
```

Then, the data can be compartmentalized into each residue. Residue 19 has been used as an example

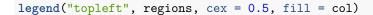
```
bool<-file[file$Residue..==19, ]
```

```
ggplot(data=bool)+geom_point(mapping=aes(x=Substitution,y=Top_rep1))
```

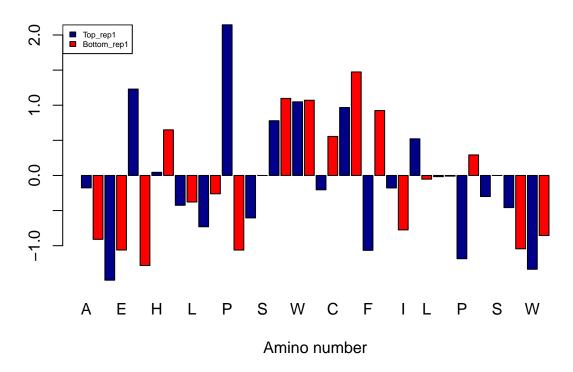


```
counts <- c(bool$Top_rep1,bool$Bottom_rep1)
x <- c(bool$Substitution, bool$Substitution)
regions <- c("Top_rep1", "Bottom_rep1")
col <- c("darkblue", "red")

barplot(counts, names.arg=x, main="Spike plot for '19' residue rep1",
    xlab="Amino number", col=c("darkblue", "red"),
    legend = rownames(counts), beside=TRUE)</pre>
```



## Spike plot for '19' residue rep1



Now, it is likely that the amino acid with the highest top\_rep value and the least bottom\_rep value is the mutagen with the highest affinity for the reactive binding domain.

```
tdiff1=abs(bool$Top_rep1-bool$Bottom_rep1)
mut1=bool$Substitution[which.max(tdiff1)]
```

#for