Alicia Breanne Dagle

520 W 114th St, Apt 24• New York, NY 10025 • (781) 686-0272 alicia.dagle@columbia.edu • www.linkedin.com/in/AliciaDagle

EDUCATION

Graduate:

Columbia University, New York, NY

Master of Science in Mechanical Engineering, GPA: 3.70/4.00

Master of Philosophy in Mechanical Engineering

Doctor of Philosophy in Mechanical Engineering

Expected Dec 2019

Expected May 2021

Expected May 2023

Relevant Coursework: Data Science; Deep Learning in Biomedical Engineering; Biostatistics; Intro to Robotics; Anatomy for Physicists & Engineers; Principles of Magnetic Resonance Imaging; Biomedical Imaging; Biophotonics; Computer Aided Design; Modern Control Systems; Advanced Fluid Mechanics; Advanced Thermodynamics

Undergraduate:

Columbia University, New York, NY

Bachelor of Science in Mechanical Engineering, GPA: 3.57/4.00 Combined Plan Program with **Clark University**, Worcester, MA

Bachelor of Arts in Physics, GPA: 3.94/4.00

May 2018 May 2018

Relevant Coursework: ; Mechanics of Solids; Heat Transfer; Electronics; Electricity & Magnetism; Classical Mechanics; Oscillations, Waves & Optics; Intro Chemistry I-II; Computer Simulations Lab; Honors Tutorial in Mechanical Engineering (Research in Robotics Rehabilitation Lab); Biomedical Ethics.

PROFESSIONAL EXPERIENCE

Siemens Healthineers – **Business Programs Intern** (Point of Care Division)

June 2017 – August 2017

- Assisted Sustaining Engineering R&D team to improve and support continued success of current products.
- Pursued design changes including design-to-cost savings, updated CAD drawings, managed project schedules, communicated and coordinated with business partners on-site and overseas, including manufacturing and vendors.
- Managed multiple projects, released documents for review and implementation.
- Performed quality investigations using LabView to measure pressures in RAPIDPoint 405/500 measurement cartridges.

Clark University – Teaching Assistant (Honors Calculus)

2014 - 2016

• Led review sessions, graded homework on an ongoing basis, provided students with detailed feedback.

RESEARCH EXPERIENCE

Living Materials Lab (Columbia University, Dr. Karen Kasza)

August 2018 – Present

- Investigating role of mechanical forces during embryonic development in model organism, Drosphila Melanogaster.
- Employing optogentic tools to apply external force, imaging with conofocal microscopy, processing using cell segmentation and particle image velocimetry.
- Analyzing cell position changes, rearrangements, and velocities using Tissue Analyzer, Segga, and PIV Lab.

Biomedical Optics REU (Wellman Center for Photomedicine, Dr. Seok-Hyun Yun)

June 2016 –August 2016

- Operated laser and Brillouin microscopy equipment, cultured cells, performed MATLAB analysis.
- Investigated the Brillouin stiffness measurements of cells under varying osmotic compression conditions.
- Successfully visualized intracellular components by mapping Brillouin shift.

Condensed Matter Research (Clark University, Dr. Michael Boyer)

January 2015 – May 2016

- Examined the surface properties of cuprous oxide nanocubes to further understand their catalytic behavior.
- Employed atomic force microscopy (AFM), scanning tunneling microscopy (STM), and scanning electron microscopy (SEM) to determine band-gap and visualize surface features.

Computational Sensing & Medical Robotics REU (Johns Hopkins University, Dr. Muyinatu Bell) June 2015 – August 2015

- Researched energy safety limitations and feasibility of implementing photoacoustic imaging to visualize the internal carotid artery during endonasal transsphenoidal surgery for pituitary tumor resection.
- Constructed phantom for testing, collected photoacoustic data, performed MATLAB analysis, determined the energy required for vessel visualization, and compared the required energy to fluence safety limits.
- Coauthored a publication, submitted an abstract and presented poster.

Bioengineering REU (Worcester Polytechnic Institute, Dr. Qi Wen)

June 2014 – August 2014

- Researched the effect of vimentin on cellular traction force, spreading area, and stiffness.
- Cultured cells, prepared polyacrylamide gels with fluorescent beads, captured images with a fluorescent microscope, and performed traction force microscopy (TFM) with a 3-D finite element model utilizing MATLAB and ANSYS. Applied atomic force microscopy to examine and plot stiffness of cells, submitted abstract and presented poster.

National Research Foundation Gradruate Research Fellowship (NSF GRFP) Pi Tau Sigma (International Mechanical Engineering Honors Society) nomination Coauthored publication (Bell et al. 2016, Proceedings of SPIE Photonics West) 1st Place Presentation Award for Computational Sensing & Medical Robotics REU Presentation at Council of Undergraduate Research (CUR) Symposium (Arlington, VA) Presentation at Biomedical Engineering Society (BMES) Conference (San Antonio, TX)

COMPUTER/LAB SKILLS

Computer: Microsoft Office, SolidWorks, MATLAB, LaTeX, LabView, Java, and R.

Lab: Finite element analysis, photoacoustic imaging, ultrasound, Brillouin Microscopy, TFM, AFM, STM, and SEM.

ACTIVITIES & VOLUNTEER EXPERIENCE

Active: Competative ballroom dance; MyNYC Mentor Program; ENG mentor for high school students.

Past: Competitive gymnastics; YouthLEAD interfaith organization; Best Buddies; Unified Sports; BMES volunteer.