Do Alternate Wetting and Drying Irrigation Technology and Nitrogen Rates Affect Rice Sheath Blight?

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Water and nitrogen management play vital roles in rice production. However, the mismanagement of these two crop inputs may trigger the development of sheath blight of rice, caused by *Thanatephorus cucumeris* (strain AG1-IA), which is favored by wet conditions, high relative humidity, and high nitrogen fertilizer levels. To understand how different combinations of water and nitrogen management affect sheath blight epidemics, we conducted two separate split-plot experiments with a water-saving (alternate wetting and drying) regime and a traditional puddled regime combined with differing nitrogen treatments in the dry seasons of 2015 and 2016. Disease was scored using the same methodology in both experiments using a sheath blight assessment scale for field evaluation developed at the International Rice Research Institute to assess the severity on infected sheaths and leaves while sheath blight incidence on tillers was counted per hill. We were unable to detect any clear differences in the incidence or severity of tiller sheath blight due to irrigation treatment. Leaf severity for the AWD treatment was clearly lower in the 2015 trial but higher in 2016. However, the level of disease severity remained low in both years, < 1 %. Our findings indicate that farmers can adopt water-saving technologies without risking increased sheath blight incidence. We suggest that further cross-disciplinary research in this area is warranted.

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# Introduction

Rice is mainly produced in Asia with over 90 % of production occurring there (Global Rice Science Partnership (GRiSP) 2013) and uses the largest amount of any agricultural commodity (Dawe 2005; Global Rice Science Partnership (GRiSP) 2013).

Climate change will bring changes to the environments in which our food crops are produced in (Reyer et al. 2013). Rice (*Oryza sativa*) is the most widely grown cereal crop in a diverse set of environments globally and will have to adapt to changing environments where it is grown as a result.

Rice’s reliance on freely available water, coupled with the effects of climate change mean that rice production practices will have to adapt to use new practices to produce more units of food with fewer water inputs.

Sheath blight disease, caused by *Thanatephorus cucumeris* (A.B. Frank) Donk. anastomosis group 1 IA, of rice, is an economically important disease worldwide throughout tropical and temperate production areas. Under conducive conditions, the disease can cause up to 50 % yield losses (Marchetti and Bollich 1991). Studies by Savary *et al.* (2000) estimated normal yield losses in tropical irrigated lowland rice in Asia at 5 to 10 % annually. The disease is best managed through an integrated disease management approach, which includes resistance (Kumar et al. 2009; Srinivasachary et al. 2011), fungicide applications where affordable (Rush and Lee 1983; Groth 2008), nitrogen (N) management (Slaton et al. 2003; Castilla et al. 1996; Tang et al. 2007) and water management (Castilla et al. 1996).

Alternate wetting and drying (AWD) is an irrigation technique for rice (*Oryza sativa* L.) developed by the International Rice Research Institute (IRRI) and its partners that saves about 15 to 40 % of irrigation water while not affecting yield (Bouman and Tuong 2001; Liping et al. 2007). In AWD rice, fields are exposed to several dry phases during the growing season without exposing the plants to water stress. To avoid yield decline under AWD, “safe” thresholds have been developed. Under safe AWD irrigation water is applied when the field water level reaches 15 cm below the soil surface (Richards and Sander 2014). Fields are furthermore kept puddled with water standing during the flowering period to avoid plant stress. As an added benefit to saving water, AWD also reduces greenhouse gas (GHG) emissions of rice fields, which are a substantial factor in the GHG budget of rice-producing countries, by around 50  % (Yan et al. 2005; Sander et al. 2016).

The AWD technology has been identified as promising climate-smart practice for different rice-growing regions that can stabilize rice production in water-scarce areas, as well as help, reduce the carbon footprint of rice production. Various countries, *e.g.* Bangladesh, Vietnam, Thailand and Philippines, plan to widely apply AWD to rice production (Ministry of Environment and Forests (MOEF) Government of the People’s Republic of Bangladesh 2015). However, a change like this in the water regime in rice fields on a large scale could trigger unintended effects, for example, changes in disease incidence or severity.

Many studies have been conducted *in silico* that examine how diseases may change with a changing climate. Rice blast is perhaps the most frequently studied rice disease in regards to the effects of climate change (Luo et al. 1995, 1998; Kim et al. 2015; Duku et al. 2016; Viswanath et al. 2017). However, Kim *et al.* (2015) have modelled the effects of climate change on sheath blight with epidemics predicted to decrease by 2100 in their study, which focused on the Korean peninsula.

Some *in-situ* studies of the effects of climate change on rice diseases have focused on the effects of temperature on bacterial blight (Webb et al. 2010) and the effects of drought (Dossa et al. 2017).

While the effects of climate change directly on rice disease have been studied, the effects of mitigation practices have not. Rice farmers may be hesitant to adopt water-saving technologies because changing rice production practices could have unintended effects on the pests and diseases that occur in the rice crop. This is a reasonable concern because of the well-documented effects of N (Slaton et al. 2003; Tang et al. 2007) and the effects of alternating wet and dry periods (Castilla et al. 1996) on sheath blight. To gain a better understanding of what the effects of adopting AWD on rice sheath blight could have, we established field experiments that incorporated irrigation technology and N fertilisation to create a conducive environment for the disease to develop.

We hypothesized that AWD would not have a positive effect on sheath blight incidence and severity in tropical rice production that would deter farmers from adopting it.

# Materials and Methods

## Experimental Site

Two experiments were conducted at the International Rice Research Institute’s (IRRI) Ziegler Experiment Station in Los Baños, Calabarzon, Philippines (latitude 14° 11’ N, longitude 121° 15’ E). The climate is classified as humid-tropical with the wet season starting in early June to December and dry season between December and late May. The experiments were both established in the dry seasons of 2015 and 2016. In the 2016 experiment changes were made in inoculation techniques, fertilisation rates and plot sizes to optimize the experiment based on findings from the 2015 experiment. The differences are detailed in the following sections.

## Pathogen

An isolate of *Thanatephorus cucumeris* (strain AG1-IA) infected lowland rice was maintained on potato dextrose agar (PDA) medium slants. The isolate was transferred to 90 mm Petri dishes containing PDA and incubated a room temperature (20 to 27°C) for five days. Glass bottles of autoclaved rice grain and hull substrate (1/5, w/w) were prepared and plugs of the culture were transferred from Petri dishes to the autoclaved substrate and incubated at room temperature for two weeks and used to inoculate the plots (Castilla et al. 1996).

## Host Plant

NSIC Rc222, an inbred semi-dwarf high-yielding variety, released by the Philippine Rice Research Institute (PhilRice), with a 114-day maturity when transplanted was used in both experiments. The variety is commonly grown by farmers in the area, having good yields and moderate resistance to brown plant hopper, green leaf hopper and yellow stem borer, but susceptibility to tungro and no known resistance to sheath blight.

## 2015 Experimental Design

The plot design was a split-plot randomised complete block design with four replicates where irrigation was the main plot treatment and N rate was the split-plot treatment.

The main plot treatments were alternate wetting and drying (AWD) and continuously puddled or farmers’ practice as the control treatment. The main plot size was 12 m x 12 m (144 m2), with a split-plot size of 5 m x 5 m (25 m2). Replication size was 12 m x 24 m (288 m2) with a buffer of 1 m per split-plot for a whole experiment size of 1,152 m2.

Irrigation in AWD plots was determined by the water level in plots, *i.e.* when the water level reached 15 cm below the soil surface irrigation water was applied to a level of 5 cm. In puddled plots, a standing water layer of 3 to 5 cm was maintained throughout the growing season.

The split-plot treatment rates of N were N0 (no N supply), N100 (100 kg per ha applied as urea in three splits) and N120 (120 kg per ha applied as urea in three splits) (Table 1).

Trays of NSIC Rc222 were seeded on 19 December 2014. Plots were established by manually transplanting seedlings on 9 January 2015 in hills with six to eight seedlings per hill and a distance of 20 cm within and between rows.

The plots were inoculated 20 days after transplanting by applying 151 g of inoculum over the split-plot area with a 1 m buffer on either end (4 m x 11 m).

## 2016 Experimental Design

The 2016 experiment followed the same split-plot design as described for the 2015 experiment. However, the plot size was increased and due to these changes, the replicate sizes are not equal as necessitated by the use of a larger area for the experiment due to differences in the blocks. The main plot sizes were: Block 1 (B1) 21 m x 20.5 m (412.5 sq m) and Block 2 (B2) 20.25 m x 21.6 m (437.4 m2). The split-plot sizes were B1 21 m x 10.25 m (215.25 m2), B2 20.25 m x 10.8 m (218.7 m2). The replication sizes were B1 – 42 m x 20.5 m (861 m2) and B2 – 40.5 m x 21.6 m (874.8 m2). A buffer 0.5 m per split-plot was used and the overall experiment size was 3471.6 m2.

split-plot N rates differed from the 2015 rates with only two N treatments, N60 (60 kg/ha as urea split into two applications) and N180 (180 kg/ha in three splits) being applied (Table 1).

Nurseries of NSIC Rc222 were established on 7 January 2016. Seedlings were manually transplanted from 20 to 22 January 2016 in hills with six to eight seedlings per hill with a distance of 20 cm within and between rows.

Based on the 2015 results, the inoculum application methods were modified in 2016 to increase the amount of inoculum and applied to a smaller area. Plots were inoculated 41 days after transplanting using ten bottles per one sampling area (1 m x 1 m) per plot, where one bottle contained 151 g inoculum. A total amount of 1,510 g of inoculum was applied to each 1 m x 1 m area.

## Data Collection

In both experiments, two sample areas, sized 1 m x 1 m, per plot were assessed. The total number of tillers per hill and number of tillers with sheath blight (incidence) was measured for nine hills per sample area. Tiller sheath severity was measured for four tillers per hill and six leaves tiller using a disease assessment rating scale for field evaluation developed at IRRI. The scale is an unequally spaced categorical scale where (i) 0 – no disease; (ii) 1 – trace to 1 % severity; (iii) 2 – 1 to 5 % severity; (iv) 3 – 5 to 15 % severity; (v) 4 – 15 to 50 % severity; and (vi) 5 – 50 to 100 % severity. Severity was considered to be the amount of leaf or sheath tissue covered by sheath blight lesions. A leaf was considered living if at least 50 % of its area was not brown or dark brown due to natural senescence. A leaf that was yellow-green or light green due to a disease, such as tungro or N deficiency was still considered a living leaf. The same rating methodology was used in both experiments. Five disease assessments were made in the 2015 experiment and four disease assessments were made in the 2016 experiment, respectively.

## Statistical Analysis

The area under the disease progress stairs (AUDPS) (Simko and Piepho 2012) was calculated for both disease incidence and severity using R (version 4.0.0), (R Core Team 2020), package agricolae, (version 1.3-3), (Mendiburu 2020).

The contributed R package, rstanarm (version 2.19.3), (Brilleman et al. 2018; Goodrich et al. 2020), was used to fit Bayesian generalized linear models with water management and N fertilisation as fixed effects and replicate as a random effect. Six models were created where the dependent variables were: 2015 tiller sheath blight incidence, 2015 tiller sheath blight severity, 2015 leaf sheath blight severity; 2016 tiller sheath blight incidence, 2016 tiller sheath blight severity; 2016 leaf sheath blight severity, respectively. The independent variables were the irrigation management (WMGT) and N rate (NRTE) and the interaction of WMGT and NRTE with replicate treated as a random variable. The base levels (control treatments) for the analyses for 2015 were: N rate – N0, irrigation management – puddled. For for 2016 the base level values were: N rate – N60, irrigation management – puddled. The same weakly informative priors were set for the fixed and random effects in all models, location = 0, scale = 10. Model fitness was evaluated using mcmc\_trace() and pp\_check() from the contributed package, bayesplot (version 1.7.2), (Gabry et al. 2019) as well as summary information.

The resulting models were tested using a test for equivalency using equivalence\_test() from the contributed package, bayestestR (version 0.7.0), (Makowski et al. 2019) to evaluate whether differences between the sheath blight responses varied between the water and N management treatments. The “Test for Practical Equivalence” is based on the Highest Density Interval (HDI) plus the Region of Practical Equivalence (ROPE) decision rule (Kruschke 2014, 2018), *HDI+ROPE decision rule* and checks whether parameter values should be accepted or rejected against the null hypothesis, the ROPE. To do this, it checks the percentage of the 89 % HDI that is in the null region ROPE. If this amount is sufficiently low, the null hypothesis is rejected. If the amount is large enough, the null hypothesis is accepted.

## Hypothesis

We hypothesised that higher amounts of nitrogen would increase the incidence and severity of sheath blight, while alternate wetting and drying would decrease the disease incidence and severity.

# Results

## 2015 Experiment

In 2015 the incidence of tiller sheath blight remained low throughout the growing season (Figure 1a; Table 2). Water management was undecided in the test for equivalency (Figure 3a; Table 2) between the AWD and puddled treatments. However, the effect of the N treatments of both N100 and N120 on tiller incidence were higher and clearly different when compared with the control N0 treatment (Figure 4a, 2).

Tiller sheath blight severity remained below 2 % (Figure 1c, 2c). Both the N100 and N120 treatment effects caused the tiller severity to be higher and were clearly different than the control N0 treatment (Figure 4c; Table 2). However, the AWD water management was not clearly the same or different from the puddled treatment (Figure 3c; Table 2).

The severity of leaf sheath blight remained low, less than 0.4 % across all treatments (Figure 1e, 2e). As with the tiller incidence and severity, the effects of both the N100 and N120 treatments on leaf severity resulted in higher values and were and clearly different than the N0 treatment (Figure 4e; Table 2). The AWD water management treatment was again undecided (Figure 3e; Table 2).

## 2016 Experiment

The changes to the inoculation methodology resulted in a higher rate of infection in 2016, with the N180 treatment reaching a maximum value of 98 % incidence at the third observation (Figure 2b). The N treatment N180 value was higher and clearly different than the control N60 treatment (Figure 4b; Table 3. As in the 2015 study, water management treatment was undecided (Figure 3b; Table 3).

As with the tiller incidence, the tiller severity increased with the changed inoculation methods with a maximum of 7.6% for the puddled treatment (Figure 2d). In 2016 the N180 treatment was again higher and clearly different from the N60 treatment, with N180 severity being higher (Figure 4d; Table 3). The AWD water management, which was lower than the puddled treatment, was also clearly different (Figure 3d; Table 3).

In 2016 the higher N rate, N180, were clearly different, higher, than the control treatments, N60. However the water management treatment was undecided (Figure 3f, 4b; Table 2).

# Discussion

In both experiments, we were unable to detect any statistically clear effect of AWD on sheath blight that led to increases in the disease that could hinder adoption of the technology, or decreases that would favour it except in one case. In the 2016, experiment the tiller sheath blight severity was clearly lower for the AWD treatment than puddled treatment (Figure 3d; Table 2), indicating a possible adverse effect of using AWD on tiller sheath blight severity under high sheath blight disease pressure. In all other cases, the difference was unclear.

The findings of the effects of N rates on sheath blight were as expected in both experiments. Higher rates of N caused an increase in disease incidence and severity in all of the cases examined.

By increasing the plot size and increasing inoculum amount applied to a smaller area, the changes made for the 2016 experiment improved the experiment. The sheath blight incidence increased and the variability of sheath blight in the plots decreased (Figure 1a:b, 2c:d, 3a:b, 4c:d). Based on these results, it is suggested that the methods used in the 2016 experiment should be adopted in any further studies of this kind.

It should be noted that the levels of leaf severity remained low (< 1 %) throughout the growing season in all treatments for both years. Therefore, the sheath blight disease levels in these studies may be lower than they would be if conducted in the wet season. However, it should be noted that normally it is only practical to implement AWD in the dry season due to the inability to completely drain the paddy in the wet season. Still, sheath blight remains an issue in both seasons and the use of AWD would not appear to increase the disease but may be effective in reducing it in some cases because of reduced water in the paddy reducing the moisture for the fungus to infect and disperse the infective propagules.

The rice establishment method is known to affect the spread of sheath blight (Willocquet et al. 2000). Manually transplanted rice has been found to have higher apparent infection rates than direct-seeded rice. Therefore, as manual transplanting becomes less common due to labor and water availability constraints (Toriyama et al. 2005), sheath blight risk may be decreased due to changes in establishment practices. A combination of AWD and direct-seeded rice could be beneficial for both saving water and reducing the risk of sheath blight.

Diseases are the only issue that farmers face when growing rice. Recently, Lorica *et al.* (2020) have also found that AWD did not affect rodent damage in crops. The findings our research along with those of Lorica *et al.* (2020) highlight the need for cross-disciplinary research that examines what the effects of new technologies that are being developed to adapt to and mitigate climate change on the pests and diseases that occur in rice. Ultimately these findings are useful as IRRI promotes AWD as water-saving technologies because farmers can be assured that the changes will not increase the risks from these pests and pathogens.

Climate change is predicted to decrease sheath blight risk over the longer term (Kim et al. 2015). However, over the next 10 to 20 years the risk is predicted to remain the same as from 2000 to 2010 in research that has been conducted on the matter to date (Kim et al. 2015). Therefore, it is likely to remain an important disease into the near future with little resistance available in currently available varieties. Given the results of the N treatments showing clear effects, in most cases increasing the amount and severity of the disease, we would argue that this suggests that there is no reason AWD could not be adopted as we were unable to detect any clear differences due to irrigation management in most of the cases. Therefore, based on these studies the use of AWD technologies would not appear to be a factor that increases the incidence or severity of rice sheath blight disease and should be adopted given the other benefits that it provides.

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AHS and BOS conceived the topic. AHS and NPC conceived the research. NPC assisted in the field work in 2015 and conducted the field work in 2016. AHS conducted the data analysis. All the authors contributed to the writing of the manuscript.

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# Literature Cited

Bouman, B. A. M., and Tuong, T. P. 2001. Field water management to save water and increase its productivity in irrigated lowland rice. Agricultural Water Management. 49:11–30 Available at: <http://www.sciencedirect.com/science/article/pii/S0378377400001281>.

Brilleman, S. L., Crowther, M. J., Moreno-Betancur, M., Buros Novik, J., and Wolfe, R. 2018. Joint longitudinal and time-to-event models via Stan. Available at: <https://github.com/stan-dev/stancon_talks/>.

Castilla, N. P., Leano, R. M., Elazhour, F. A., Teng, P. S., and Savary, S. 1996. Effects of plant contact, inoculation pattern, leaf wetness regime, and nitrogen supply on inoculum efficiency in rice sheath blight. Journal of Phytopathology. 144:187–192.

Dawe, D. 2005. Increasing water productivity in rice-based systems in Asia–past trends, current problems, and future prospects. Plant Production Science. 8:221–230.

Dossa, G. S., Torres, R., Henry, A., Oliva, R., Maiss, E., Cruz, C. V., et al. 2017. Rice response to simultaneous bacterial blight and drought stress during compatible and incompatible interactions. European Journal of Plant Pathology. 147:115–127.

Duku, C., Sparks, A. H., and Zwart, S. J. 2016. Spatial modelling of rice yield losses in Tanzania due to bacterial leaf blight and leaf blast in a changing climate. Climatic Change. 135:569–583.

Gabry, J., Simpson, D., Vehtari, A., Betancourt, M., and Gelman, A. 2019. Visualization in Bayesian workflow. J. R. Stat. Soc. A. 182:389–402.

Global Rice Science Partnership (GRiSP). 2013. *Rice Almanac: Source book for one of the most important economic activities on earth*. eds. Jay Maclean, Bill Hardy, and Gene Hettel. IRRI.

Goodrich, B., Gabry, J., Ali, I., and Brilleman, S. 2020. Rstanarm: Bayesian applied regression modeling via Stan. Available at: <https://mc-stan.org/rstanarm>.

Groth, D. E. 2008. Effects of cultivar resistance and single fungicide application on rice sheath blight, yield, and quality. Crop Protection. 27:1125–1130.

Kim, K.-H., Cho, J., Lee, Y. H., and Lee, W.-S. 2015. Predicting potential epidemics of rice leaf blast and sheath blight in South Korea under the RCP 4.5 and RCP 8.5 climate change scenarios using a rice disease epidemiology model, EPIRICE. Agricultural and forest meteorology. 203:191–207.

Kruschke, J. 2014. *Doing Bayesian data analysis: A tutorial with R, JAGS, and Stan*. Academic Press.

Kruschke, J. K. 2018. Rejecting or accepting parameter values in Bayesian estimation. Advances in Methods and Practices in Psychological Science. 1:270–280.

Kumar, K. V. K., Reddy, M. S., Kloepper, J. W., Lawrence, K. S., Groth, D. E., and Miller, M. E. 2009. Sheath blight disease of rice (*Oryza sativa* L.) - an overview. Biosciences, Biotechnology Research Asia. 6:465–480.

Liping, F., Bouman, B. A. M., Tuong, T. P., Cabangon, R. J., Li, Y., Lu, G., et al. 2007. Exploring options to grow rice using less water in northern China using a modelling approach: I. Field experiments and model evaluation. Agricultural Water Management. 88:1–13 Available at: <http://www.sciencedirect.com/science/article/pii/S0378377406002630>.

Lorica, R. P., Singleton, G. R., Stuart, A. M., and Belmain, S. R. 2020. Rodent damage to rice crops is not affected by the water-saving technique, alternate wetting and drying. Journal of Pest Science.

Luo, Y., TeBeest, D. O., Teng, P. S., and Fabellar, N. G. 1995. Simulation studies on risk analysis of rice leaf blast epidemics associated with global climate change in several asian countries. Journal of Biogeography.:673–678.

Luo, Y., Teng, P. S., Fabellar, N. G., and TeBeest, D. O. 1998. The effects of global temperature change on rice leaf blast epidemics: A simulation study in three agroecological zones. Agriculture, Ecosystems & Environment. 68:187–196.

Makowski, D., Ben-Shachar, M. S., and Lüdecke, D. 2019. bayestestR: Describing effects and their uncertainty, existence and significance within the Bayesian framework. Journal of Open Source Software. 4:1541.

Marchetti, M. A., and Bollich, C. N. 1991. Quantification of the relationship between sheath blight severity and yield loss in rice. Plant Disease.

Mendiburu, F. de. 2020. *Agricolae: Statistical procedures for agricultural research*. Available at: <https://CRAN.R-project.org/package=agricolae>.

Ministry of Environment and Forests (MOEF) Government of the People’s Republic of Bangladesh. 2015. Intended nationally determined contributions (INDC). Available at: <http://www4.unfccc.int/ndcregistry/PublishedDocuments/Bangladesh%20First/INDC_2015_of_Bangladesh.pdf>.

R Core Team. 2020. *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing. Available at: <https://www.R-project.org/>.

Reyer, C. P. O., Leuzinger, S., Rammig, A., Wolf, A., Bartholomeus, R. P., Bonfante, A., et al. 2013. A plant’s perspective of extremes: Terrestrial plant responses to changing climatic variability. Global Change Biology. 19:75–89.

Richards, M., and Sander, B. O. 2014. *Alternate wetting and drying in irrigated rice*. Copenhagen, Denmark: CGIAR Research Program on Climate Change, Agriculture; Food Security (CCAFS).

Rush, M. C., and Lee, F. N. 1983. Rice sheath blight: A major rice disease. Plant Disease. 67:829–832.

Sander, B. O., Wassmann, R., and Siopongco, J. D. L. C. 2016. Mitigating greenhouse gas emissions from rice production through water-saving techniques: Potential, adoption and empirical evidence. In eds. C. T. Hoanh, R. Johnston, and V. Smakhtin. Centre for Agriculture; Biosciences International, p. 193.

Savary, S., Willocquet, L., Elazegui, F. A., Castilla, N. P., and Teng, P. S. 2000. Rice pest constraints in tropical Asia: Quantification of yield losses due to rice pests in a range of production situations. Plant Disease. 84:357–369.

Simko, I., and Piepho, H.-P. 2012. The area under the disease progress stairs: Calculation, advantage, and application. Phytopathology. 102:381–389.

Slaton, N. A., Cartwright, R. D., Meng, J., Gbur Jr., E. E., and Norman, R. J. 2003. Sheath blight severity and rice yield as affected by nitrogen fertilizer rate, application method, and fungicide. Agronomy Journal. 95:1489–1496.

Sparks, A. H., Castilla, N. P., and Sander, B. O. 2020. Data for "do alternate wetting and drying irrigation technology and nitrogen rates affect rice sheath blight?".

Sparks, A. H., Castilla, N. P., and Sander, B. O. 2020. Reproducible research compendium for analysing effects of water management and nitrogen on rice sheath blight.

Srinivasachary, Willocquet, L., and Savary, S. 2011. Resistance to rice sheath blight (*Rhizoctonia solani* Kühn) [(teleomorph: *Thanatephorus cucumeris* (A.B. Frank) Donk.] Disease: Current status and perspectives. Euphytica. 178:1–22.

Tang, Q., Peng, S., Buresh, R. J., Zou, Y., Castilla, N. P., Mew, T. W., et al. 2007. Rice varietal difference in sheath blight development and its association with yield loss at different levels of N fertilization. Field Crops Research. 102:219–227.

Toriyama, K., Heong, K. L., Hardy, B., and others. 2005. Rice is life: Scientific perspectives for the 21st century. Proceedings of the World Rice Research Conference held in Tsukuba, Japan, 4-7 November 2004. In *Rice is life: scientific perspectives for the 21st century. Proceedings of the World Rice Research Conference held in Tsukuba, Japan, 4-7 November 2004.*, International Rice Research Institute (IRRI).

Viswanath, K., Sinha, P., Kumar, S. N., Sharma, T., Saxena, S., Panjwani, S., et al. 2017. Simulation of leaf blast infection in tropical rice agro-ecology under climate change scenario. Climatic change. 142:155–167.

Webb, K. M., Oña, I., Bai, J., Garrett, K. A., Mew, T., Cruz, C. M. V., et al. 2010. A benefit of high temperature: Increased effectiveness of a rice bacterial blight disease resistance gene. New Phytologist. 185:568–576.

Willocquet, L., Fernandez, L., and Savary, S. 2000. Effect of various crop establishment methods practised by Asian farmers on epidemics of rice sheath blight caused by *Rhizoctonia solani*. Plant Pathology. 49:346–354.

Yan, X., Yagi, K., Akiyama, H., and Akimoto, H. 2005. Statistical analysis of the major variables controlling methane emission from rice fields. Global Change Biology. 11:1131–1141.

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# Tables

(#tab:Table-1) Nitrogen application rates for 2015 and 2016 experiments in kilograms per hectare. Five different nitrogen treatments were applied depending on the experiment. In the 2015 experiment three nitrogen rate treatments were applied: no N (N0), 100 kg/ha N (N100) and 180 kg/ha N (N120). In the 2016 experiment, two nitrogen rate treatments were applied: 60 kg/ha N (N60) and 180 kg/ha N (N180). Treatments in both years were applied in splits at: basal, tillering growth stages for all treatments and panicle initiation growth stage for treatments with three applications.

| Total | Basal | Tillering | Panicle Initiation |
| --- | --- | --- | --- |
| 0 | 0 | 0 | 0 |
| 100 | 60 | 20 | 20 |
| 120 | 60 | 30 | 30 |
| 60 | 30 | 30 | 0 |
| 180 | 60 | 60 | 60 |

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(#tab:Table-2) An 89 % test for practical equivalence for 2015 experiment. Response variables are shown as T. Inc., Tiller Incidence; T. Sev., Tiller Severity; L. Sev., Leaf Severity. The parameters are WMGTAWD, irrigation treatment AWD; NRTEN100, nitrogen rate of 100 k/ha; NRTEN120 kg/ha. ROPE low and ROPE high provide the limits of the Region of Practical Equivalence (ROPE). ROPE Percentage is the proportion of the Highest Density Interval (HDI) that lies inside the ROPE. All points within this interval have a higher probability of density than points outside the interval. ROPE Equivalence is the test result, either “rejected”, “accepted” or “undecided”. “HDI Low” and “HDI High” are the lower and upper HDI limits for the parameters.

| Response | Parameter | ROPE low | ROPE high | ROPE Percentage | ROPE Equivalence | HDI low | HDI high |
| --- | --- | --- | --- | --- | --- | --- | --- |
| T. Inc. | (Intercept) | -0.1 | 0.1 | 0.00 | Rejected | 0.6393 | 1.6486 |
| T. Inc. | WMGTAWD | -0.1 | 0.1 | 38.95 | Undecided | -0.1648 | 0.3823 |
| T. Inc. | NRTEN100 | -0.1 | 0.1 | 0.00 | Rejected | 0.3000 | 0.9781 |
| T. Inc. | NRTEN120 | -0.1 | 0.1 | 0.00 | Rejected | 0.6588 | 1.3369 |
| T. Sev. | (Intercept) | -0.1 | 0.1 | 0.00 | Rejected | 4.5539 | 12.1202 |
| T. Sev. | WMGTAWD | -0.1 | 0.1 | 3.51 | Undecided | -1.5077 | 4.4971 |
| T. Sev. | NRTEN100 | -0.1 | 0.1 | 0.00 | Rejected | 5.6870 | 12.9153 |
| T. Sev. | NRTEN120 | -0.1 | 0.1 | 0.00 | Rejected | 14.2522 | 21.4325 |
| L. Sev. | (Intercept) | -0.1 | 0.1 | 3.59 | Undecided | -0.2022 | 2.6889 |
| L. Sev. | WMGTAWD | -0.1 | 0.1 | 0.00 | Rejected | -0.8055 | -0.1254 |
| L. Sev. | NRTEN100 | -0.1 | 0.1 | 0.00 | Rejected | 1.1034 | 1.9276 |
| L. Sev. | NRTEN120 | -0.1 | 0.1 | 0.00 | Rejected | 1.3633 | 2.1932 |

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(#tab:Table-3) An 89 % test for practical equivalence for 2015 experiment. Response variables are shown as T. Inc., Tiller Incidence; T. Sev., Tiller Severity; L. Sev., Leaf Severity. The parameters are WMGTAWD, irrigation treatment AWD; and NRTEN180, nitrogen rate of 180 k/ha. ROPE low and ROPE high provide the limits of the Region of Practical Equivalence (ROPE). ROPE Percentage is the proportion of the Highest Density Interval (HDI) that lies inside the ROPE. All points within this interval have a higher probability of density than points outside the interval. ROPE Equivalence is the test result, either “rejected”, “accepted” or “undecided”. “HDI Low” and “HDI High” are the lower and upper HDI limits for the parameters.

| Response | Parameter | ROPE low | ROPE high | ROPE Percentage | ROPE Equivalence | HDI low | HDI high |
| --- | --- | --- | --- | --- | --- | --- | --- |
| T. Inc. | (Intercept) | -0.16088 | 0.16088 | 0.00 | Rejected | 9.58152 | 11.5138 |
| T. Inc. | WMGTAWD | -0.16088 | 0.16088 | 16.43 | Undecided | -0.02809 | 0.7564 |
| T. Inc. | NRTEN180 | -0.16088 | 0.16088 | 0.00 | Rejected | 1.56226 | 2.3446 |
| T. Sev. | (Intercept) | -0.49405 | 0.49405 | 0.00 | Rejected | 17.67145 | 25.2787 |
| T. Sev. | WMGTAWD | -0.49405 | 0.49405 | 0.00 | Rejected | -3.79478 | -1.2082 |
| T. Sev. | NRTEN180 | -0.49405 | 0.49405 | 0.00 | Rejected | 0.56555 | 3.1147 |
| L. Sev. | (Intercept) | -0.05534 | 0.05534 | 0.00 | Rejected | 0.73125 | 1.4633 |
| L. Sev. | WMGTAWD | -0.05534 | 0.05534 | 42.04 | Undecided | -0.19451 | 0.1681 |
| L. Sev. | NRTEN180 | -0.05534 | 0.05534 | 0.00 | Rejected | 0.12556 | 0.5014 |

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# Figures

Figure 1: Sheath blight disease progress curves for 2015 and 2016 experiments. Disease progress was measured in the 2015 experiment at five assessment points and 2016 experiment at four assessment points during the dry season of both years. Main plot irrigation treatments were permanently puddled (PDL) and alternate wetting and drying (AWD). Points represent the mean observations of four replications.

Figure 1: Sheath blight disease progress curves for 2015 and 2016 experiments. Disease progress was measured in the 2015 experiment at five assessment points and 2016 experiment at four assessment points during the dry season of both years. Main plot irrigation treatments were permanently puddled (PDL) and alternate wetting and drying (AWD). Points represent the mean observations of four replications.

Figure 2: Sheath blight incidence progress was measured in the 2015 experiment at five assessment points and 2016 experiment at four assessment points during the dry season both years. Five different N treatments were applied to split-plots depending on the experiment. In the 2015 experiment three N rate treatments were applied: no N (N0), 100 kg/ha N (N100) and 180 kg/ha N (N120). In the 2016 experiment, two N rate treatments were applied: 60 kg/ha N (N60) and 180 kg/ha N (N180). Points represent the mean observations of four replications.

Figure 2: Sheath blight incidence progress was measured in the 2015 experiment at five assessment points and 2016 experiment at four assessment points during the dry season both years. Five different N treatments were applied to split-plots depending on the experiment. In the 2015 experiment three N rate treatments were applied: no N (N0), 100 kg/ha N (N100) and 180 kg/ha N (N120). In the 2016 experiment, two N rate treatments were applied: 60 kg/ha N (N60) and 180 kg/ha N (N180). Points represent the mean observations of four replications.

Figure 3: Sheath blight progress was measured in the 2015 at five assessment points and 2016 experiment at four points. Sheath blight incidence was rated as the number of infected tillers divided by the total number of tillers per hill and used to calculate the area under the disease progress stairs (AUDPS). Sheath blight severity was rated on an unevenly spaced ordinal scale from zero to five. Main plot irrigation treatments were permanently puddled (PDL) and alternate wetting and drying (AWD).

Figure 3: Sheath blight progress was measured in the 2015 at five assessment points and 2016 experiment at four points. Sheath blight incidence was rated as the number of infected tillers divided by the total number of tillers per hill and used to calculate the area under the disease progress stairs (AUDPS). Sheath blight severity was rated on an unevenly spaced ordinal scale from zero to five. Main plot irrigation treatments were permanently puddled (PDL) and alternate wetting and drying (AWD).

Figure 4: Sheath blight severity progress was measured in the 2015 at five assessment points and 2016 experiment at four points. Sheath blight incidence was rated as the number of infected tillers divided by the total number of tillers per hill and used to calculate the area under the disease progress stairs (AUDPS). Sheath blight severity was rated on an unevenly spaced categorical scale and converted to the mid-point percentage value to calculate the AUDPS. Five different N treatments were applied to split-plots depending on the experiment. In the 2015 experiment three N rate treatments were applied: no N (N0), 100 kg/ha N (N100) and 180 kg/ha N (N120). In the 2016 experiment, two N rate treatments were applied: 60 kg/ha N (N60) and 180 kg/ha N (N180).

Figure 4: Sheath blight severity progress was measured in the 2015 at five assessment points and 2016 experiment at four points. Sheath blight incidence was rated as the number of infected tillers divided by the total number of tillers per hill and used to calculate the area under the disease progress stairs (AUDPS). Sheath blight severity was rated on an unevenly spaced categorical scale and converted to the mid-point percentage value to calculate the AUDPS. Five different N treatments were applied to split-plots depending on the experiment. In the 2015 experiment three N rate treatments were applied: no N (N0), 100 kg/ha N (N100) and 180 kg/ha N (N120). In the 2016 experiment, two N rate treatments were applied: 60 kg/ha N (N60) and 180 kg/ha N (N180).

# Colophon

This report was generated on 2020-07-10 13:33:32 using the following computational environment and dependencies:

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The current Git commit details are:

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#> Remote: main @ origin (git@github.com:adamhsparks/rice-awd-shb.git)  
#> Head: [5fd5512] 2020-07-10: Add acknowledgements for Nick