Dynamic Optimization with Particle Swarms (DOPS): A metaheuristic for parameter estimation in biochemical models

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Abstract

Mathematical modeling is a powerful tool to analyze, and ultimately design biochemical networks. However, the estimation of biochemical model parameters is a significant challenge. Parameter estimation in biochemical models typically involves expensive function evaluations and noisy data, making it difficult to quickly obtain optimal solutions. Biochemical models often also have many local extrema which further complicates parameter estimation. Toward these challenges, we developed Dynamic Optimization with Particle Swarms (DOPS), a novel global meta-heuristic that combined features of multi-swarm particle swarm optimization with dynamically dimensioned search (DDS). DOPS uses a multi-swarm particle swarm optimization technique to generate candidate solution vectors, the best of which is greedily updated using dynamically dimensioned search. We tested the performance of DOPS on a model of human coagulation cascade. We performed \mathcal{T} = 25 trials with \mathcal{N} = 4000 function evaluations per trial, and compared the performance of DOPS with other commonly used meta-heuristics such as differential evolution (DE), simulated annealing (SA) and dynamically dimensioned search (DDS). We further tested the predictive power of the coagulation model parameters against data not used in training, and found good agreement between simulations and experimental measurements. Lastly, we tested the performance of DOPS on commonly used test functions for global optimization and on published biochemical parameter estimation benchmark problems. For the wide range of problems that we considered, DOPS outperformed other metaheuristic approaches despite a limited number of function evaluations. Taken together, DOPS is a promising meta-heuristic approach for the estimation of biochemical model parameters in relatively few function evaluations.

Keywords: Parameter identification, Meta-heuristic optimization, Biochemical modeling

Introduction

Cells process nutrients and respond to changes in their environment using complex biochemical networks. These networks contain thousands of components interconnected
through nonlinear enzyme catalyzed reactions. Mathematical modeling has evolved as a
powerful paradigm to analyze, and ultimately design these complex networks [1–5]. Mathematical modeling of biochemical networks is often an iterative process. First, models are
formulated from biochemical knowledge, and then model parameters are estimated using
experimental data [6–8]. Parameter estimation is typically framed as a non-linear optimization problem wherein the residual (or objective function) between experimental data
and model simulations is minimized using an optimization strategy [9]. Optimal parameters obtained from model training are then used to validate the model on unseen experimental data. If validation fails, model construction and calibration are repeated iteratively
until satisfactory results are obtained.

Parameter estimation is a major challenge in the iterative development of biochemical 14 models. Although parameter estimation has been a well studied problem in engineering 15 for decades [10-13], the complex dynamics of large biological systems and noisy, often 16 incomplete experimental data pose a unique estimation challenge. Often optimization 17 problems involving biological systems are non-linear and multi-modal i.e. typical mod-18 els often have multiple local minima or maxima [7, 9]. Non-linearity coupled with multimodality generally renders local optimization techniques such as pattern search [14], 20 Nelder-Mead simplex methods [15], steepest descent methods or Levenberg-Marquardt 21 [16] incapable of reliably obtaining optimal solutions as they generally terminate at local minimum. Though deterministic global optimization techniques (for example algorithms based on branch and bound) can handle non-linearity and multi-modality [17, 18], the absence of derivative information, discontinuity of the objective functions, non-smooth regions or the lack of knowledge about the objective function hampers these techniques.

Meta-heuristic stochastic optimization approaches like Genetic Algorithms (GAs) [ADI-27 REF], Simulated Annealing (SA) [19], Evolutionary Programming [ADI-REF] and population based searches like Differential Evolution (DE) [20] have shown promise on nonlinear multi-modal problems [21]. These techniques do not make any assumptions about the 30 structure of objective function nor do they require a priori information about the objec-31 tive function. Though they do not guarantee strong convergence, these approaches are effective in finding near optimal solutions. Mendes et al. [22] used simulated annealing 33 to estimate rate constants for the irreversible inhibition of HIV proteinase, Modchang et 34 al. [23] used a genetic algorithm to estimate parameters for a model of signal transduc-35 tion [ADI-EXPAND], differential evolution approaches have also been effective on various biological problems [24-26]. Tashkova et al. compared different meta-heuristics for pa-37 rameter estimation on a dynamic model of endocytosis and showed that DE was the 38 most effective [27]. Banga and co-workers have also successfully applied scatter-search methods to estimate parameters on non-linear biological processes [28-30]. Hybrid approaches that combine a meta-heuristic with a local optimization search, wherein a near 41 globally optimal solution obtained using a meta-heuristic is refined using a local search have also become popular. Villaverde et al. combined scatter search with local search methods for parameter estimation in a collection of systems biology models [31]. Fan et al. recently showed that population based meta-heuristics along with decomposition based methods can be also used to model gene circuits from mRNA data [32]. Despite these successes, a major drawback of most metaheuristic approaches is the large number of function evaluations required to explore the parameter space. Typically as models grow in size and complexity, evaluation of the objective function becomes computationally 49 expensive. Thus performing a large number of function evaluations is not desirable (and 50 perhaps not feasible). 51

In this study, we developed Dynamic Optimization with Particle Swarms (DOPS), a

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novel meta-heuristic that combines the global search capability of multi-swarm particle swarm optimization and dynamically dimensioned search (DDS). The objective of DOPS is to obtain near optimal parameter estimates for large biochemical models within a relatively few function evaluations. DOPS uses a multi-swarm particle swarm optimization 56 technique to generate candidate solution vectors which are then greedily updated us-57 ing dynamically dimensioned search. We first considered a model of human coagulation 58 cascade to test the performance of DOPS. Coagulation is a large, complex biochemical 59 network involving strong positive feedback. We then tested the performance of DOPS on 60 commonly used test functions for global optimization (Ackley and Rosenbrock) and pub-61 lished biochemical parameter estimation benchmark problems [31]. DOPS outperformed 62 common meta-heuristic approaches like Differential Evolution (DE), Simulated Anneal-63 ing (SA) and dynamically dimensioned search (DDS) on both the test functions and the 64 coagulation model. DOPS also performed well on the benchmark problems where it outperformed enhanced scatter search (eSS) and recovered the nominal parameters with only 4000 function evaluations ([ADI-compared with what?]) across all the benchmark 67 problems considered. Taken together, these studies suggest DOPS is a promising metaheuristic approach for the estimation of biochemical model parameters in relatively few function evaluations.

71 Results

Optimization problem formulation. The problem of parameter estimation in a dynamic biological model consists of finding an optimal parameter vector which minimizes the difference between model simulations and \mathcal{E} experimental measurements. This difference is quantified by an objective function $K(\mathbf{p})$ which is typically the Euclidean norm of the simulation error subject to problem specific and parameter bounds constraints:

minimize
$$K(\mathbf{p}) = \sum_{i=1}^{\mathcal{E}} (g_i(t_i, \mathbf{x}, \mathbf{p}, \mathbf{u}) - y_i)^2$$

subject to $\dot{\mathbf{x}} = \mathbf{f}(t, \mathbf{x}(t, \mathbf{p}), \mathbf{u}(t), \mathbf{p})$
 $\mathbf{x}(t_0) = \mathbf{x}_0$
 $\mathbf{c}(t, \mathbf{x}, \mathbf{p}, \mathbf{u}) \geqslant \mathbf{0}$
 $\mathbf{p}^L \leqslant \mathbf{p} \leqslant \mathbf{p}^U$ (1)

where t is time, $\mathbf{x}(t, \mathbf{p})$ is the state variable vector with an initial state \mathbf{x}_0 , $\mathbf{u}(t)$ is a model input vector, \mathbf{f} is the system of model equations (e.g., differential equations or algebraic constraints) and \mathbf{p} is the model parameter vector. The parameter search (or model simulations) can be subject to \mathbf{c} linear or non-linear constraints, and parameter bound constraints where \mathbf{p}^L and \mathbf{p}^U denote the lower and upper parameter bounds, respectively. The problem eventually is to find:

$$\mathbf{p}^* = \arg\min_{\mathbf{p}} K(\mathbf{p}) \tag{2}$$

Dynamic optimization with particle swarms (DOPS). DOPS is a novel meta-heuristic
which combines multi-swarm particle swarm methods with the dynamically dimensioned
search approach of Shoemaker and colleagues (Fig. 1). The goal of DOPS is to estimate
optimal or near optimal parameter vectors for high-dimensional biological models within a
specified number of function evaluations. Toward this objective, DOPS begins by using a

particle swarm search and then dynamically switches, using an adaptive switching criteria,
 to the DDS search phase.

Phase 1: Swarm phase. We began the particle swarm phase of DOPS by randomly initializing a swarm of \mathcal{K} -dimensional particles (represented as z_i), wherein each particle corresponded to a \mathcal{K} -dimensional parameter vector. After initialization, particles were randomly partitioned into k equal sized sub-swarms $\mathcal{S}_1, \ldots, \mathcal{S}_k$. Thereafter within each sub-swarm \mathcal{S}_k , particles were updated according to the rule:

$$\mathbf{z}_{i,j} = \theta_{1,j-1} \mathbf{z}_{i,j-1} + \theta_2 \mathbf{r}_1 \left(\mathcal{L}_i - \mathbf{z}_{i,j-1} \right) + \theta_3 \mathbf{r}_2 \left(\mathcal{G}_k - \mathbf{z}_{i,j-1} \right)$$
(3)

where $(\theta_1,\theta_2,\theta_3)$ were adjustable parameters, \mathcal{L}_i denotes the best solution found by particle i within sub-swarm k for function evaluation $1 \to j-1$, and \mathcal{G}_k denotes the best solution found over all particles within sub-swarm \mathcal{S}_k . The quantities r_1 and r_2 denote uniform random vectors with the same dimension as the number of unknown model parameters $(\mathcal{K} \times 1)$. Equation (3) is similar to the general particle swarm update rule, however, it does not contain velocity terms. In DOPS the parameter $\theta_{1,j-1}$ depends upon the function evaluations and is updated according to:

$$\theta_{1,j} = \frac{(\mathcal{N} - j) * (\mathbf{w}_{max} - \mathbf{w}_{min})}{(\mathcal{N} - 1)} + \mathbf{w}_{min}$$
(4)

where \mathcal{N} represents the total number of function evaluations, \mathbf{w}_{max} and \mathbf{w}_{min} are the maximum and minimum inertia weights, respectively. While updating the particles, we made sure all dimensions of the solution represented by the particle were within bounds using a set of reflection boundary conditions:

$$\begin{array}{l} \text{if } z_{i,j}^{old} < z_i^{min} \text{ then} \\ \mid z_{i,j}^{new} = z_{i,j}^{old} + (z_i^{min} - z_{i,j}^{old}) \text{ if } z_{i,j}^{new} > z_i^{max} \text{ then} \\ \mid z_{i,j}^{new} = z_i^{max} \\ \mid \text{ end} \\ \end{array} \\ \begin{array}{l} \text{end} \\ \text{if } z_{i,j}^{old} > z_i^{max} \text{ then} \\ \mid z_{i,j}^{new} = z_{i,j}^{old} + (z_{i,j}^{old} - z_i^{max}) \text{ if } z_{i,j}^{new} < z_i^{min} \text{ then} \\ \mid z_{i,j}^{new} = z_i^{min} \\ \mid \text{ end} \\ \end{array} \\ \begin{array}{l} \text{end} \\ \end{array} \\ \text{end} \end{array}$$

After every \mathcal{M} function evaluations, particles were randomly redistributed to a new sub-swarm, and updated according to Eqn. (3). This process continued for $\mathcal{F}*\mathcal{N}$ functions evaluations, where \mathcal{F} is the fraction of evaluations in the particle swarm phase of DOPS.

If the simulation error stagnates for example, does not change by more than 1% for a specified number of evaluations, the swarm phase is terminated and DOPS switches to exploring parameter space using a DDS approach.

Phase 2: DDS phase. At the conclusion of the swarm phase, the overall best particle, \mathcal{G}_k , over the k sub-swarms was to initalize the DDS phase. DOPS takes $(1-\mathcal{F})*\mathcal{N}$ function evaluations in the DDS phase and then terminates the search. For the DDS phase, the best parameter estimate was updated using the rule:

$$\mathcal{G}_{new}(\mathbf{J}) = \begin{cases}
\mathcal{G}(\mathbf{J}) + \mathbf{r}_{normal}(\mathbf{J})\sigma(\mathbf{J}), & \text{if } \mathcal{G}_{new}(\mathbf{J}) < \mathcal{G}(\mathbf{J}). \\
\mathcal{G}(\mathbf{J}), & \text{otherwise.}
\end{cases} \tag{5}$$

where J represents the set containing the specific dimensions being perturbed, r_{normal} denotes a normal random vector of the same dimensions as \mathcal{G} , and σ denotes the perturbation amplitude:

$$\sigma = \mathbf{R}(\mathcal{MAX} - \mathcal{MIN}) \tag{6}$$

where ${\bf R}$ is the scalar perturbation size parameter, ${\cal MAX}$ and ${\cal MIN}$ are $({\cal K}\times 1)$ vectors that represent the maximum and minimum bounds on each dimension. The set ${\bf J}$ was constructed using a probability function ${\cal P}_j$ that represented the probability whether a specific dimension j was perturbed or not. This function is a monotonically decreasing function that decreases with the number of function evaluations. ${\cal P}_j$ can be any monotonically decreasing function, in our approach we used the following function:

$$\mathcal{P}_{i} = 1 - \log(j/((1 - \mathcal{F}\mathcal{R}) * \mathbf{N})) \tag{7}$$

Thus the number of dimensions of the candidate vector that are updated or perturbed 125 decreases with the as the number of function evaluations increase. These updates are 126 greedy in nature, so \mathcal{G}_{new} becomes the new solution vector only if it is better than the old 127 one \mathcal{G} . The decrease of weight function in equation 4, reflection boundary conditions in 128 equation 5, the update function described in equations 6 and 7, the selection probability in 129 equation 8 are the means by which DDS ideas are incorporated into DOPS. The fraction 130 of evaluations \mathcal{FR} within the swarm phase is based on a switching strategy wherein the 131 switch from swarm phase to DDS phase happens when the error due to the best solution 132 does not drop more than 1% of the original error, continuously for more than a prescribed 133 number of function evaluations. This allows the solution to quickly jump out of local optima and avoid any convergence issues that are generally associated with swarm based searches.

Performance of DOPS on a model of the human coagulation cascade. We compared the performance of DOPS with simulated annealing (SA), differential evolution (DE), and dynamically dimensioned search (DDS) for a model of the human coagulation cascade. Coagulation is an archetype biochemical network that is highly interconnected, containing both positive and negative feedback (Fig. 2). The biochemistry of coagulation, though complex, has been well studied [33-39], and reliable experimental protocols have been developed to interrogate the system [40-43]. Thus, the human coagulation cascade is an ideal test case; coagulation is challenging because it contains both fast and slow dynamics, but also accessible because of the availability of comprehensive data sets for model identification and validation. In this study, we used the coagulation model of Luan et al [43]. Luan and coworkers modeled coagulation as a system of coupled non-linear ordinary differential equations. The Luan model contained 148 parameters and 92 species and has been validated using 21 published datasets.

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Coagulation is mediated by a family proteases in the circulation, called factors and a 150 key group of blood cells, called platelets. The central process in coagulation is the conversion of prothrombin (fII), an inactive coagulation factor, to the master protease thrombin (FIIa). Thrombin generation involves three phases, initiation, amplification and termination [13,14]. Initiation requires a trigger event, for example a vessel injury which exposes tissue factor (TF), which leads to the activation of factor VII (FVIIa) and the formation of the TF/FVIIa complex. Two converging pathways, the extrinsic and intrinsic cascades, then process and amplify this initial coagulation signal. There are several control points in the cascade that inhibit thrombin formation, and eventually terminate thrombin generation. Tissue Factor Pathway Inhibitor (TFPI) inhibits upstream activation events, while antithrombin III (ATIII) neutralizes several of the proteases generated during coagulation, including thrombin. Thrombin itself also inadvertently plays a role in its own inhibition; thrombin, through interaction with thrombomodulin, protein C and endothelial cell protein C receptor (EPCR), converts protein C to activated protein C (APC) which attenuates the coagulation response by proteolytic cleavage of amplification complexes. Termination occurs after either prothrombin is consumed, or thrombin formation is neutralized by inhibitors such as APC or ATIII.

To estimate the coagulation model parameters, we used data sets from TF/VIIa initiated coagulation in the absence of anticoagulants. The objective function was a linear combination of two error functions, representing coagulation initiated with different concentrations of TF-VIIa (5pM, 5nM) [40]. We restricted the number of function evaluations to $\mathcal{N}=4000$ for each algorithm we tested, and performed 25 trials of each experiment to collect average performance data (Table ZZ).DOPS converged faster and had a lower final error compared to the other algorithms (Fig. 3). Within the first 1000 function evaluations of DOPS, there was a rapid drop in error. Approximately between 500-1000 function evaluations a switch to dynamically dimensioned search phase happens (this switch varied from trial to trial since the switch was based on the error from the swarm phase). Overall at the end of 4000 function evaluations DOPS minimized the error (final objective error is 5.5916e+04) to a greater extent than the other algorithms.

Using the parameters obtained at the end of 4000 function evaluations we examined the 'fits' between models predictions and experimental data (Fig.4). The solid lines represent the mean value of prediction over 25 trials and the shaded region represents the 99% confidence interval. Subsequently we used these optimal parameters to make model predictions that was compared against completely 'unseen' or untrained experimental data where coagulation was initiated with 500pM,50pM,10pM concentrations of TF-VIIa respectively (Fig.5).

Performance of DOPS on benchmark problems. Villaverde and co-workers recently published a set of benchmark biochemical problems to evaluate parameter estimation methods [31]. From a computational cost perspective problems they categorized the

problems as most expensive, intermediate and least expensive. We evaluated the performance of our algorithms on a problem from the most expensive and least expensive categories. The first problem (most expensive) is a genome wide kinetic model of *Saccharomyces cerevisiae* with 261 reactions, 262 variables and 1759 parameters (henceforth referred to as problem B1). The reactions were modeled using modular rate law and generalized form of Michaelis-Menten kinetics. The second problem (henceforth referred to as problem B4, least expensive) is a metabolic model of Chinese Hamster Ovary (CHO) with 35 metabolites, 32 reactions and 117 parameters. In both cases pseudo time series data was generated by the authors. For problem B1, the time series data consisted of 44 observables and for problem B4 the data corresponded to 13 different metabolites. For a detailed description about the model architectures please refer to the work of Villaverde et al. [31]

We fixed the number of function evaluations at 4000 for DOPS and trained both the models against the pseudo experimental data. In both cases we found good fits (Fig. 8 and Fig. 9) to the problems within 4000 evaluations. We recaptured the 'nominal' parameters' in both cases within 4000 evaluations (Fig. 7). The final objective function value (Table 2) is an order smaller than nominal error for B1 possibly due to overfitting against the noise that was added to the synthetic data. Having obtained good fits on the coagulation problem and the benchmarks we proceeded to compare the performance of DOPS with other algorithms on commonly used test functions for global optimization. We used a 300 dimensional Rastrigin function and 300 dimensional Ackley function. Both Ackley and Rastrigin have multiple local minima and maxima and attain a global minimum value of 0. We tested the performance of the 4 different heuristic approaches on these two functions. In each experiment we again fixed the number of function evaluations at 4000 and ran 25 experiments. In both cases (Fig. 6) we see that the error convergence rate for DOPS is much faster as compared to the other three heuristics and it finds the

- 215 global minimum of 0 in both cases. In (Table 2) we summarize the results obtained on the
- coagulation model, the benchmark problems and the test functions.
- 217 Influence of the number of sub-swarms on DOPS performance.

18 Discussion

Our study presents a novel approach for high-dimensional parameter estimation in com-219 plex biological systems with relatively few function evaluations. In this approach we com-220 bined a variant of a well known meta heuristic particle swarm optimization with Dynam-221 ically Dimensioned Search (DDS). We tested our approach on an ODE model of coag-222 ulation with 148 parameters and 92 species. Coagulation is an ideal system to test our 223 approach since the biology is well known and complex, with multiple feed back loops 224 that are tightly regulated. We used experimental data under different conditions to obtain 225 optimal parameters and used these parameters to make predictions against unseen experimental data. We obtained good fits and made sufficiently accurate enough predictions using parameters obtained from 4000 function evaluations. Further, we also used high-228 dimensional forms of commonly used test functions of global optimization and showed that we were able to find the global minimum for 300 dimensional Ackley and Rastrigin func-230 tions faster than other meta-heuristics. We also considered two recently published bench-231 mark problems to test parameter optimization approaches and showed that we were able 232 to retrieve the nominal parameter vector within 4000 function evaluations. Meta-heuristic 233 approaches are generally effective in finding close to optimum solutions of complex, multi-234 modal functions. In addition, they generally obviate the need for any a priori knowledge 235 (like function derivative). However they take an exorbitant number of objective function 236 evaluations to come close to an optimum. When the objective function evaluations tend 237 to become expensive it is infeasible to take up a large number of evaluations. As the 238 dimensionality of parameter space increases, the search region gets widened and thus 239 the problem becomes more challenging. In addition, most of these approaches require 240 optimization of 'algorithm parameters' before the actual optimization and also involve computationally expensive update operations. Tolson and Shoemaker, through DDS, showed 242 that randomly perturbing a subset of dimensions in high dimensional parameter space is an effective way to obtain near optimal solutions with few function evaluations. Though their approach is based on a single solution, the decision vector carries forward from one iteration to the next. Hence it behaves similar to a population based search. In our approach, we tried to eliminate the probability of starting from a bad region of search using a variant of particle swarm optimization. Particle Swarm Optimization (PSO) is a population based meta heuristic which does not have any complex operations like recombination, mutation or selection that are associated with other population based meta-heuristics like Differential Evolution (DE) or Genetic Algorithm (GA). Several particle swarm variants have been proposed to improve the search ability and rate of convergence, that involve different neighborhood structures, multi-swarms or adaptive parameters. Multi-swarm PSO with small particle neighborhoods have been shown to better in searching on complex multi-modal solutions [44]. Multi swarm methods, in addition, avoid rapid convergence to a local optimum or stable point and are able to generate diverse solutions. Generation of diverse solutions in the early stage gives a better exploratory capability and thus converge of upon multiple optima.

We utilized this capacity of multi swarms to generate diverse candidate solutions which can be used as initial solutions for a DDS like search. Thus the solution vectors obtained at the end of multi swarm search phase have a better propensity to not get stuck in bad regions in DDS phase as opposed to starting with a totally random initial solution. Given a fixed number of function evaluations, we have been able to show that we were able to obtain better solutions for the coagulation model and the test functions faster that other commonly known meta-heuristics and also DDS alone. The computational cost was prohibitively high for parameter estimation on this problem using a standard PSO. Thus we used SA, DE and DDS for the purpose of comparison. Choosing the number of function evaluations is largely a function of cost and complexity of the objective function. Traditionally the stopping condition for a parameter optimization problem can be the number

of function evaluations, percentage of initial error achieved or an absolute error threshold. However in case of complex, expensive functions where we desire a value within a certain period of time, the number of evaluations are used as a stopping criterion. In our current study we used a value of 4000 which we based upon the time taken (approximately 8-10 seconds) for a single objective function evaluation in the coagulation case. We used the same value of 4000 for benchmarks published by Villaverde and co-workers [31]. In this work by Villaverde et al. they used a population based search enhanced Scatter Search (eSS) to estimate the biochemical model parameters. Quite surprisingly we took a couple of orders lesser number of function evaluations 10³ to obtain the optimal parameter vector as compared to the enhanced Scatter Search (eSS) with a local optimizer, which took around the order of 10⁵ number of evaluations. The amount of CPU time taken (on an Intel Xeon processor 2.4 GHz) is lesser as compared to eSS on a similar architecture.

A surprisingly remarkable aspect about our algorithm has been that we did not 'pre optimize' any parameters of the algorithm to suit a specific problem. In the swarm phase this includes the number of particles, number of sub-swarms, acceleration constants or the number of generations after which the particles are redistributed and the neighborhood perturbation parameter in DDS phase. We used the same parameters for all the problems. The same rule was applied to the rest of the meta-heuristics barring Simulated Annealing. For SA, we optimized the cooling schedule for the coagulation model. Thus, in this approach any overhead that usually comes with additional function evaluations to pre optimize parameters was avoided.

The performance of our approach seems impressive given that it performed well on different complex systems with no pre optimization of algorithm parameters being required. We comfortably outperformed existing, widely used meta-heuristics and were also able to find minima of high dimensional global optimization test functions. Thus this approach may be well suited to large scale global optimization. In addition, surprisingly, we were

able to obtain optimal parameter vectors for two different large scale systems biology models with a couple of orders fewer number of function evaluations as compared to en-297 hanced Scatter Search (eSS). However it is guite possible that highly optimized versions 298 of common meta heuristics may outperform us on these systems. This aspect is currently 299 beyond the scope of this study. Our approach can also be combined with local derivative 300 based searches to improve upon the accuracy of the solutions. In addition, the current 301 implementation of the algorithm is designed to switch only once from the swarm phase to 302 the DDS. In the DDS phase, the search uses only one candidate vector although there is 303 a provision to start with a population of candidate vectors. Incorporating a more intelligent 304 switching strategy that can do switch from swarm phase to DDS phase multiple times and 305 having a population of candidate vectors are some aspects of the algorithm that can be 306 studied further. 307

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Table 1: Error Analysis.

TF/FVIIa concentration	S.E.	
5 nM	0.0376	
500 pM	0.0564	
50 pM	0.1125	
10 pM	0.0823	
5 pM	0.0338	

Table 2: Table with optimization settings and results for the coagulation problem, the benchmarks and test functions using DOPS. For each problem the bounds on the parameter vector, the total number of function evaluations, the best initial objective value and the best final objective value are specified. Nominal objective value represents the objective value using the true parameter vector or the nominal parameter vector. The CPU time is the time taken for the problem on a 2.4GHz Intel Xeon Architecture running Matlab 2014b.

	Coagulation	B1	B4	Ackley	Rastrigin
Evaluations	4000	4000	4000	4000	4000
Lower Bound	0.001.pnom	5.pnom	5.pnom	30	5.12
Upper Bound	1000.pnom	0.2.pnom	0.2.pnom	-15	-5.12
CPU Time	10.1 hrs	38.3 hrs	6.2 min	2.8 s	2.6 s
Scaled initial error	1.0	1.0	1.0	1.0	1.0
Scaled final error	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
Scaled nominal error	0.42	0.1	< 0.01	0	0

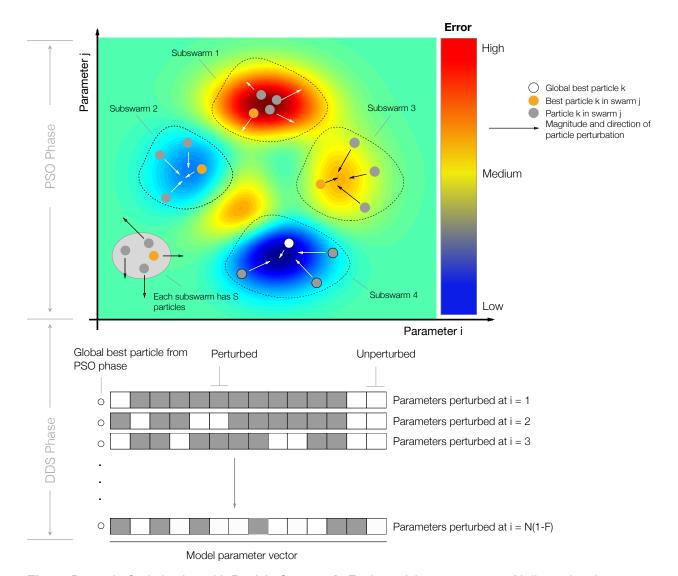


Fig. 1: Dynamic Optimization with Particle Swarms.**A**: Each particle represents an N dimensional parameter vector. Particles are given randomly generated initial solutions and grouped into different sub-swarms. Within each swarm the magnitude and direction of the movement a particle is influenced by the position of the best particle and also by its own experience. After every g number of function evaluations the particles are mixed and randomly assigned to different swarms. When the error due to the global best particle (best particle amongst all the sub-swarms - orange color) does not drop over a certain number of function evaluations, the swarm search is stopped and the search switches to a Dynamically Dimensioned Search with global best particle as the initial solution vector or candidate vector. **B**: The candidate vector performs a greedy global search for the remaining number of function evaluations. The search neighborhood is dynamically adjusted by varying the number of dimensions that are perturbed (in black) in each evaluation step. The probability that a dimension is perturbed decreases as the number of function evaluations increase. Thus as the evaluations increase the optimality of the solution is preserved.

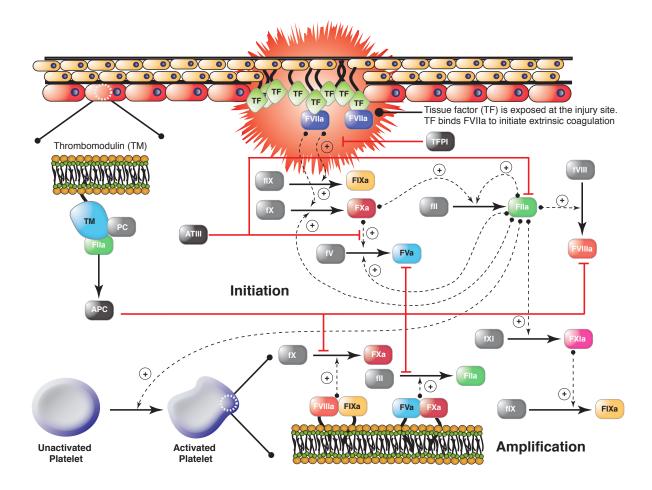


Fig. 2: Schematic of the extrinsic and intrinsic coagulation cascade[43]. Inactive zymogens upstream (grey) are activated by exposure to tissue factor (TF) following vessel injury. Tissue factor and activated factor VIIa (FVIIa) form a complex that activates factor X (fX) and IX (fIX). FXa activates downstream factors including factor VIII (fVIII) and fIX. Factor V (fV) is primarily activated by thrombin (FIIa). In addition, we included a secondary fV activation route involving FXa. FXa and FVa form a complex (prothrombinase) on activated platelets that converts prothrombin (fII) to FIIa. FIXa and FVIIIa can also form a complex (tenase) on activated platelets which catalyzes FXa formation. Thrombin also activates upstream coagulation factors, forming a strong positive feedback ensuring rapid activation. Tissue factor pathway inhibitor (TFPI) downregulates FXa formation and activity by sequestering free FXa and TF-FVIIa in a FXa-dependent manner. Antithrombin III (ATIII) inhibits all proteases. Thrombin inhibits itself binding the surface protein thrombomodulin (TM). The IIa-TM complex catalyzes the conversion of protein C (PC) to activated protein C (APC), which attenuates the coagulation response by the proteolytic cleavage of fV/FVa and fVIII/FVIIIa.

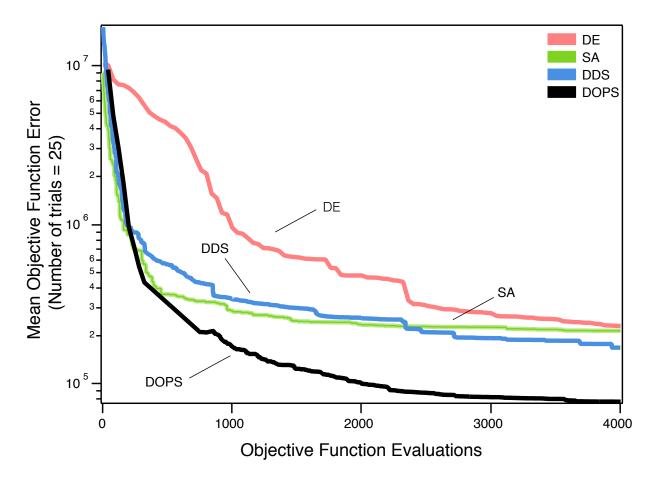


Fig. 3: Error convergence rates of the 5 different algorithms on the coagulation model. The objective error is the mean over N= 25 trials. DOPS, DDS and SA have the steepest drop in error during first 300 function evaluations. Thereafter the error drop in DDS and SA remains nearly constant whereas DOPS continues to drops further. At the end of 4000 function evaluations DOPS attains the lowest error. The next best estimate using DDS is nearly 3 times greater than the lowest error using DDS.

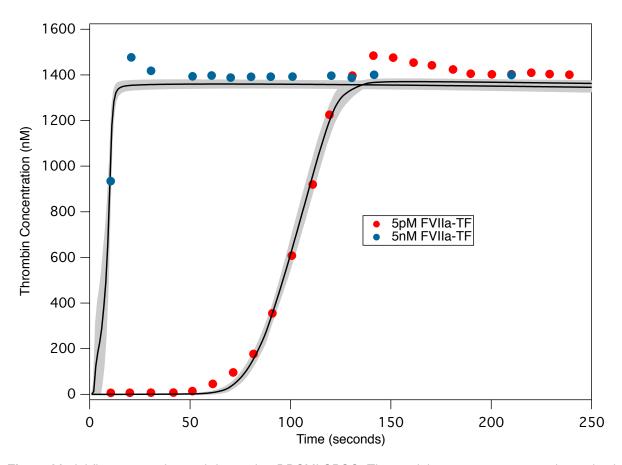


Fig. 4: Model fits on experimental data using DDSMLSPSO. The model parameters were estimated using DOPS. Solid black lines indicate the simulated mean thrombin concentration using parameter vectors from N= 25 trials. The grey shaded region represents the 99% confidence estimate of the mean simulated thrombin concentration. The experimental data is reproduced from the synthetic plasma assays of Mann and co-workers. Thrombin generation is initiated by adding Factor VIIa-TF (5nM - Red and 5pM - Green) to synthetic plasma containing 200 μ mol/L of phospholipid vesicles (PCPS) and a mixture of coagulation factors (II,V,VII,VIII,IX,X and XI) at their mean plasma concentrations.

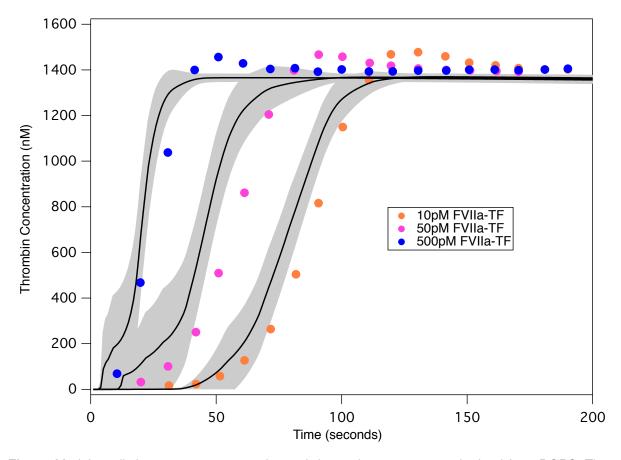


Fig. 5: Model predictions on unseen experimental data using parameters obtained from DOPS. The parameter estimates that were obtained using DOPS were tested against data that was not used in the model training. Solid black lines indicate the simulated mean thrombin concentration using parameter vectors from N=25 trials. The grey shaded region represents the 99% confidence estimate of the mean simulated thrombin concentration. The experimental data is reproduced from the synthetic plasma assays of Mann and co-workers. Thrombin generation is initiated by adding Factor VIIa-TF (500pM - Blue, 50pM - Pink and 10pM - Orange respectively) to synthetic plasma containing 200 μ mol/L of phospholipid vesicles (PCPS) and a mixture of coagulation factors (II,V,VII,VIII,IX,X and XI) at their mean plasma concentrations.

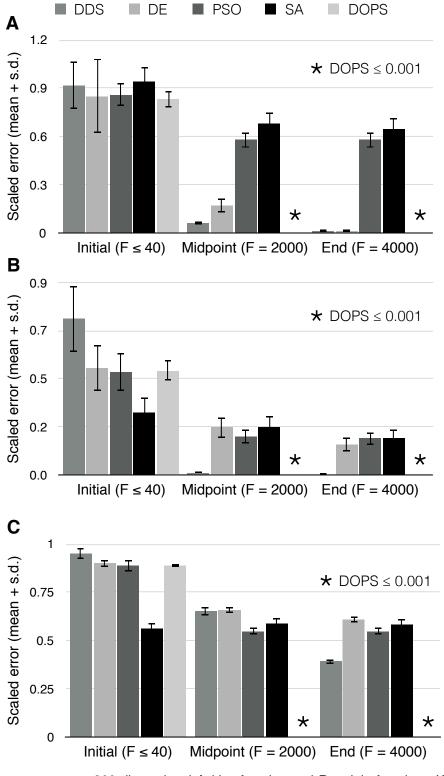


Fig. 6: Error convergence on 300 dimensional Ackley function and Rastrigin function. **(A)** Error convergence of 4 different meta-heuristics on 300 dimensional Ackley function **(B)** Error convergence of 4 different meta-heuristics on 300 dimensional Rastrigin function

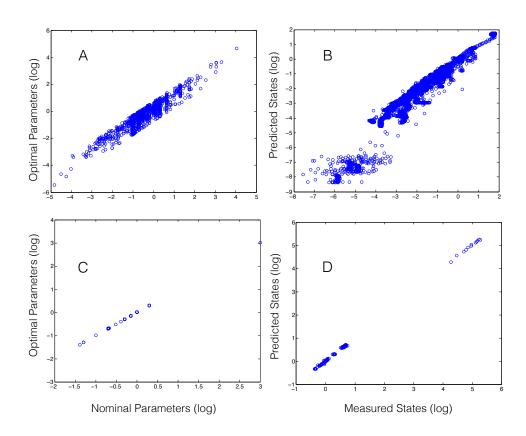


Fig. 7: (A) Difference between nominal and optimal parameters for problem B1: Genome wide kinetic model of *S.cerevisiae* with 1759 unknown parameters. **(B)** Difference between experimental (measured) data and data simulated with optimal parameters for problem B1: Genome wide kinetic model of *S.cerevisiae* with 1759 unknown parameters. **(C)** Difference between nominal and optimal parameters for problem B4: Metabolic model of Chinese Hamster Ovary Cells (CHO) cells with 117 parameters. **(D)** Difference between experimental (measured) data and data simulated with optimal parameters for problem B4: Metabolic model of Chinese Hamster Ovary Cells (CHO) cells with 117 parameters.

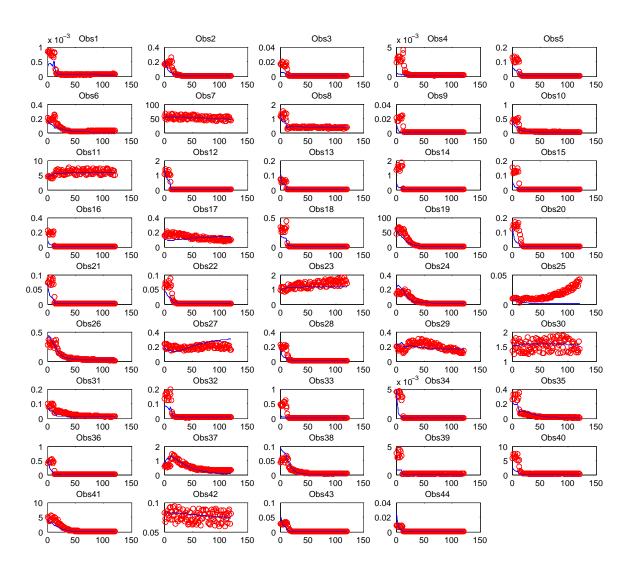


Fig. 8: (Data fits for Problem B1) Pseudo-experimental data (red circles) vs. optimal solution obtained using DOPS (solid blue lines) for the 44 observed states. X axis: time [s]; Y axis: metabolite concentrations [mM].

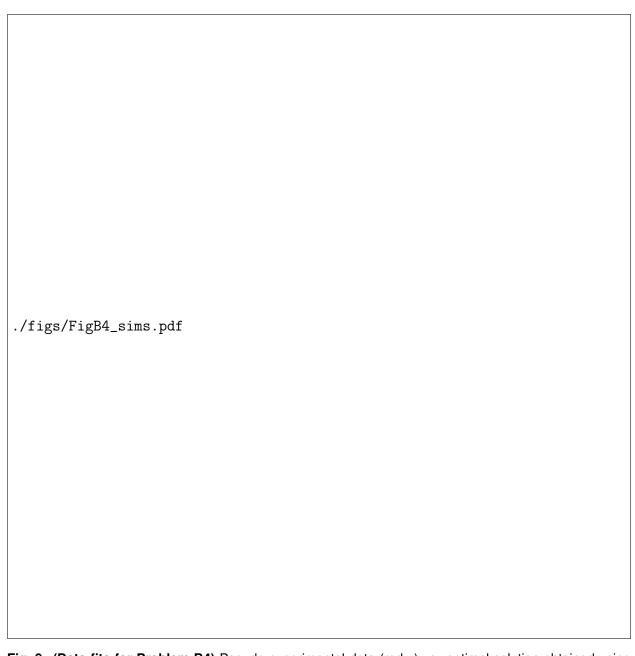


Fig. 9: (Data fits for Problem B4) Pseudo-experimental data (red x) vs. optimal solution obtained using DOPS (solid blue lines) for the 13 observed states. X axis: time [s]; Y axis: metabolite concentrations [mM].