Date: 27.06.21

Name: Zviki/Hodaya

Exp no. **13**

A. Experiment type: (Partial reprogramming, Comparative expression etc.)

Comparative expression

- B. Experiment goal: to compare the effect of cleaning out dsRNA with cellulose on a different protocols for IVT using GFP.
- C. Experimental variables:
- 1. GFP (jvr) Cellulose/no Cellulose
- 2. ModGFP (jvr) Cellulose/no Cellulose
- 3. GFP AG (txk) Cellulose/no Cellulose
- 4. ModGFP AG (txk) Cellulose/no Cellulose
- 5. GFP (jvr)+EKB Cellulose/no Cellulose
- 6. GFP (jvr)+ModEKB Cellulose/no Cellulose
- 7. ModGFP (jvr)+EKB Cellulose/no Cellulose
- 8. ModGFP (jvr)+ModEKB Cellulose/no Cellulose
- 9. GFP AG (txk)+EKB Cellulose/no Cellulose
- 10. ModGFP AG (txk)+ModEKB Cellulose/no Cellulose
- 11. NC

IVT preparation:

		GFP AG	ModGFP	ModGFP			Lipofectamin
	GFP (jvr)	(txk)	(jvr)	AG (txk)	EKB	ModEKB	(ul)
GFP (jvr)	1200						24 (6ul x 4)

ModGFP (jvr)		1200	1200				24 (6ul x 4)
GFP AG (txk)							24 (6ul x 4)
ModGFP AG (txk)				1200			24 (6ul x 4)
GFP (jvr)+EKB	1200				200		24 (6ul x 4)
GFP							
(jvr)+ModEKB	1200					200	24 (6ul x 4)
ModGFP							
(jvr)+EKB			1200		200		24 (6ul x 4)
ModGFP							
(jvr)+ModEKB			1200			200	24 (6ul x 4)
GFP AG (txk)+EKB		1200			200		24 (6ul x 4)
ModGFP AG							
(txk)+ModEKB				1200		200	24 (6ul x 4)
ModGFP (jvr)							24 (6ul x 4)
+EKB-SG			1200				
	3600	2400	3600	2400	600	600	240
x2 duplicate	7200	4800	7200	4800	1200	1200	
x3 transfections	21600	14400	21600	14400	3600	3600	720
for cellulose							
cleaning	54000	36000	54000	36000	9000	9000	
all together	75600	50400	75600	50400	12600	12600	720

D. Starting conditions

1. Cells type: (type, age, gender) NHDF73M

2. Cells passage: p6

3. No. of replications: 2

4. Size of plate: 6 Well

5. No. of wells/plates: 42 wells/7 plates

6. No. of cells per well: 150,000

7. No. of cells total: 6,300,000

8. Confluency et starting exp: 60%

- 9. Medium(s): rpmi
- 10. Medium volume (per well): 2.5
- 11. Medium replacement: (every other day etc.)
 - Before the first transfection
 - 3hr after each transfection.
- 12. Splitting during the experiment? (when, how many?) no
- 13. Incubator general/low oxygen
 - E. The methods of the experiment (transfections, staining, sort, qPCR etc.)

Method	Link for	No. of	Step in the	Done	Link for
	detailed	cells	experiment	yes/no	results
	protocol				
Seeding		150,000cells		yes	
27.06.21		per well (42			
		wells)			
Mixes	Transfection			yes	
preparation	calculation				
27.06.21	exp.13 27.6.21				
1 st transfection	Transfection			yes	
(morning)	calculation				
28.06.21	exp.13 27.6.21				
Medium			3hr post	yes	
replacement			transfection		

28.06.21				
Pictures 1 st			yes	<u>06.07.21 new</u>
transfection				
29.06.21				
2 nd transfection	<u>Transfection</u>		yes	
(morning)	calculation			
29.06.21	exp.13 27.6.21			
Medium		3hr post	yes	
replacement		transfection		
29.06.21				
Pictures 2 nd			yes	<u>06.07.21 new</u>
transfection				
30.06.21				
3 rd transfection	<u>Transfection</u>		yes	
(morning)	calculation			
30.06.21	exp.13 27.6.21			
Medium		3hr post	yes	
replacement		transfection		
30.06.21				
Pictures			yes	<u>06.07.21 new</u>
3 rd transfection				
01.07.21				
Cells counting			yes	cells counting
01.07.21				<u>& FACS</u>
				<u>results</u>
				01.07.21
GFP intensity			yes	cells counting
(FACS)				<u>& FACS</u>
01.07.21				<u>results</u>
				01.07.21
RNA extraction			yes	<u>06.07.21 new</u>

for qPCR			
01.07.21			

F. Experiment end date: 01.07.21

G. Conclusion: (With reference to experiment goal)

Cells viability (cells counting):

- Cellulose cleaning improves the cells viability in the most samples.
- The best cells viability improvement is in non-modified samples (mostly).

GFP expression (FACS analysis):

- ModGFP with cellulose cleaning presents the highest expression of GFP.
- ModGFP / modGFP+EKB / modGFP+modEKB with cellulose cleaning present the highest improvement (GFP expression) in compared to the same samples W/O cellulose cleaning.
- Addition of EKB/modEKB to modGFP did not improve the cells viability and GFP expression of modGFP.