**The list of protocol files in the runset file**

1. V2\_2\_96\_SPRI\_33\_purification\_AliquotBackupLibraries.pro
2. V2\_2\_96\_SPRI\_33\_purification\_V2.pro
3. V2\_2\_ProtA\_10x\_dilution\_AddEBT.pro
4. V2\_2\_ProtB\_10x\_dilution\_AddPurifLibr.pro
5. V2\_NanoDrop3xDilution.pro

Bravo procedure details

* Starts with aliquot 5ul of unpurified indexing libraries into a backup plate.
* Robot will calculates SPRI beads volume based on selected volume of sample volume to be purified.
* Automatically calculated volume of SPRI beads from the SPRI beads reservoir will be added into the samples plate.
* Then continued with adding selected sample volume from sample plate to the reaction plate.
* Same volume of SPRI beads from the SPRI beads reservoir will be added into the samples plate.
* Incubate the sample/SPRI beads for 10 minutes by mixing gently, and incubate additional 10 minutes without mixing.
* The reaction plate will be move to magnetic rack to pellet the beads for 10 minutes.
* Supernatant will be removed and discarded.
* Bead pellets will be washed twice with 80% EtOH without re-suspending the bead pellet.
* After removing the wash solution, bead pellet will be air dry on the magnetic rack for 15 minutes.
* Selected elution volume of the elution buffer will be added into air dried beads on the magnetic rack. Bead pellet will be incubated on the magnet for 3 minutes.
* Gently aspirate purified libraries, transfer into a fresh plate.
* 120ul of elution buffer will be added into the reaction plate after elution step is completed.
* Then will do 10x dilution of final purified libraries.
* Then will do 2.5x dilution of final purified libraries for NanoDrop measurement.
* Seal the purified library plate and the reaction plate.
* Store the sealed purified plate for further application and sealed reaction plate can be discarded.

