# Assignment 13.a for **STATGR6103**

submitted to Professor Andrew Gelman

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# 1 Question 1

Millions of people in rural Bangladesh are exposed to dangerous levels of arsenic in their drinking water which they get from home wells. Several years ago a survey was conducted in small area of Bangladesh to see if people with high arsenic levels would be willing to switch to a neighbor's well. File http://www.stat.columbia.edu/gelman/bda.course/wells.dat has the data; all you need here are the variables switch (1 if the respondent said he or she would switch, 0 otherwise) and arsenic (the concentration in the respondent's home well, with anything over 0.5 considered dangerous). Apply the probit model described in exercise 1 above to predict the probability of switching given arsenic level. The goal is inference for the coefficients a and b.

Given that the goal of the survey is inference about the parameters 'a' and 'b', I first examine the data in order to understand the predictor and the dependent variable. able 1 displays summary statistics of these two variables.

Table 1: Data summary

stat	switch	arsenic		
min	0	0.51		
mean	0.57	1.66		
max	1	9.65		

The dependent variable (switch) is a binary response variable, where people indicate their willingness to switch to a neighbor's well. The predictor (arsenic) is the level of arsenic in the drinking water. We are asked to implement a probit model to predict probability of switching given arsenic levels. We are asked to motivate the probit model using a latent variable formulation with  $z_i \sim N(\alpha + \beta x_i, 1)$  such that;

$$y_i = \begin{cases} 1, z_i > 0 \\ 0, \text{ otherwise} \end{cases}$$

The probability that  $y_i = 1$  is given by the probit regression below;

$$Pr(y_i = 1|x_i) = \Phi(z_i)$$
$$= \Phi(\alpha + \beta x_i)$$

Where  $\Phi$  is the cumulative distribution function (CDF) of the standard normal distribution and our goal is to estimate coefficients or parameters  $\alpha$  and  $\beta$  using a probit regression, with three techniques. I program an HMC and allow it to estimate the parameters of the probit model, then I run the same model in Stan and finally estimate the parameters using a variational inference algorithm and compare the results.

Since the outcome variable is binary, I use a binomial likelihood function with the following analytical form;

$$p(y|\alpha, \beta, x) \propto \prod_{i=1}^{N} (\Phi(\alpha + \beta x_i))^{y_i} (1 - \Phi(\alpha + \beta x_i))^{1-y_i}$$

I also consider noninformative priors on  $\alpha$  and  $\beta$  such that;

$$p(\alpha, \beta) \propto 1$$

Thus the posterior density is given by;

$$p(\alpha, \beta|y, x) \propto p(\alpha, \beta)p(y|\alpha, \beta, x)$$

$$= \prod_{i=1}^{N} (\Phi(\alpha + \beta x_i))^{y_i} (1 - \Phi(\alpha + \beta x_i))^{1-y_i}$$

#### 1.1 Part A

Program Hamiltonian Monte Carlo for the probit model described in the previous exercise, again using the latent-variable formulation (so you are jumping in a space of n+2 dimensions). Tune the algorithm and run to approximate convergence.

I program an HMC and run 4 chains with 1000 iterations. I set the mass matrix to scale with the variance of the posterior density and tune the  $\epsilon$  and L parameters in order to get approximately 65% convergence. Table 2 shows the posterior intervals as estimated by the HMC algorithm

	mean	se.mean	$\operatorname{sd}$	2.5%	25%	50%	75%	97.5%	n.eff	Rhat
$\alpha$	-0.2	0	0	-0.3	-0.2	-0.2	-0.2	-0.1	572	1
β	0.2	0	0	0.2	0.2	0.2	0.2	0.3	714	1

Table 2: Posterior Distributions: HMC in R

#### 1.2 Part B

#### Check your results by running Stan.

I fit a probit model in Stan and get the results summarized in Table 3.

	Postorior	Distributions:	Ston
Table 5.	I OSCETIOI	בווטווווטוווטוווסוום.	Stan

	mean	se.mean	$\operatorname{sd}$	2.5%	25%	50%	75%	97.5%	n.eff	Rhat
$\alpha$	-0.18	0.00	0.04	-0.26	-0.21	-0.18	-0.15	-0.11	839	1
β	0.23	0.00	0.02	0.19	0.21	0.23	0.24	0.27	869	1

## 1.3 Part C

#### Compare to the variational Bayes inferences for a and b from the previous above.

I program a variational Bayes algorithm to estimate the parameters  $\alpha$  and  $\beta$ . The posterior intervals of the parameters are displayed in Table 4. I make some plots to compare the inferences from the

Table 4: Posterior Distributions: Stan

	mean	se.mean	$\operatorname{sd}$	2.5%	25%	50%	75%	97.5%
$\alpha$	-0.178	0.00	0.03	-0.29	-0.21	-0.18	-0.16	-0.11
β	0.227	0.00	0.03	0.20	0.21	0.23	0.25	0.27

3 approaches, namely HMC, Stan and variational inference. Figure 1 shows a scatterplot of samples drawn from the posterior densities for each of the 3 approaches.

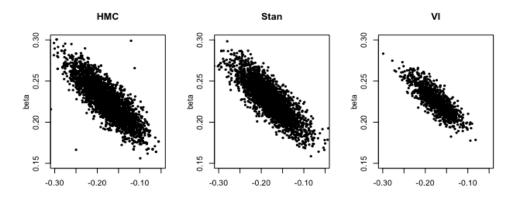


Figure 1: The results from all the approaches are quite similar.

Figure 2 and 3 show the marginal posterior density of the parameters  $\alpha$  and  $\beta$  as estimated by the 3 approaches, respectively.

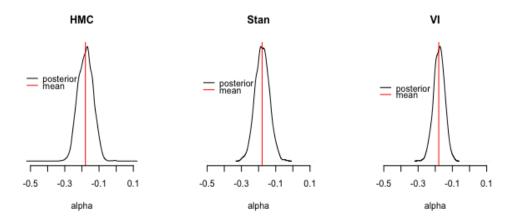


Figure 2: All 3 approaches estimate a similar posterior density for  $\alpha$ , the mean of the marginal posterior distribution is marked by the red line.

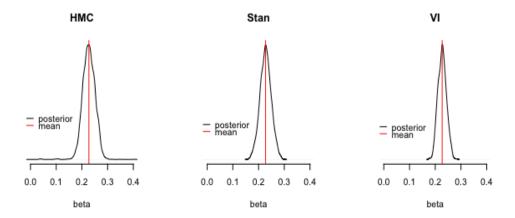


Figure 3: All 3 approaches estimate a similar posterior density for  $\beta$ , the mean of the marginal posterior distribution is marked by the red line.

We see that the results from HMC, Stan and variational Bayes algorithm are very similar and comparable.

## 2 Code

#### 2.1 R code

```
Listing 1: R code
rm(list = ls())
setwd("/Users/Advait/Desktop/New_School/Fall16/BDA/Class24")
library(rstan)
rstan_options(auto_write = TRUE)
options(mc.cores = parallel::detectCores())
install.packages("data.table", dependencies = T)
library(data.table)
library ( plyr )
library (arm)
library (msm)
library (mvtnorm)
library(LearnBayes)
df <- fread("http://www.stat.columbia.edu/</pre>
~gelman/bda.course/wells.dat")
str(df)
names(df) <- c("s.no","switch", "arsenic", "dist", "assoc", "educ")</pre>
summary(df)
###
df_final <- data.frame(df$switch, df$arsenic)</pre>
class(df_final)
str(df_final)
##
#Stan
y <- df_final$df.switch
x <- df_final$df.arsenic
N <- length(df_final$df.switch)</pre>
stanc("13a.stan")
fit_stan <- stan("13a.stan", data = list("N", "y", "x"),</pre>
                 iter = 2000, chains = 3)
print(fit_stan)
# plot(ext_stan$y_rep)
# points(ext_stan$y_rep, col = "red")
# plot(colMeans(ext_stan$y_rep), y, pch = 16, cex = 0.7)
# abline(0,1)
```

#### Listing 2: R code contd.

```
##
##PartA
#HMC
#log posterior function
n <- 1
log_post <- function(param, x , y){</pre>
  param_prior <- dunif(param, -100, 100, log = T)</pre>
  log_prior <- sum(param_prior)</pre>
  log_likelihood <- sum(dbinom(y, n,</pre>
  prob = pnorm(param[1] + param[2]*x),log = T))
  return(log_prior + log_likelihood)
}
log_post(c(0.1, -0.3), x, y)
#Numerical gradient
gradient_num <- function(param, x, y){</pre>
  d <- length(param)</pre>
  e <- 0.0001
  diff <- rep(NA,d)
  for(k in 1:d){
    th_hi <- param
    th_lo <- param
    th_hi[k] \leftarrow param[k] + e
    th_lo[k] <- param[k] - e
    diff[k] \leftarrow (log_post(th_hi,x,y) - log_post(th_lo,x,y))/(2*e)
  return(diff)
}
gradient_num(c(-0.001,-.04),x,y)
###
#HMC iter
hmc_iteration <- function(param,x,y, epsilon,L,M){</pre>
  M_inv <- 1/M
  d <- length(param)</pre>
  # Sample 10 points randomly from a normal
  \#distribution with mean = 0 and standard deviation = sqrt(M)
  phi <- rnorm(d , 0, sqrt(M))</pre>
  param_old <- param</pre>
  log_p_old <- log_post(param,x,y) - 0.5*sum(M_inv*phi^2)</pre>
  phi <- phi + 0.5*epsilon*gradient_num(c(param), x, y)</pre>
```

#### Listing 3: R code contd.

```
for (1 in 1:L){
    param <- param + epsilon*M_inv*phi</pre>
    phi <- phi + (if (l==L)0.5 else 1)*epsilon*
    gradient_num(c(param),x,y)
  phi <- -phi
  log_p_star <- log_post(c(param),x,y) - 0.5*sum(M_inv*phi^2)</pre>
  r <- exp(log_p_star - log_p_old)
  if(is.nan(r)) r <- 0</pre>
  p_{jump} \leftarrow min(r,1)
  param_new <- if(runif(1) < p_jump) param else param_old</pre>
  return (list (param = param_new, p_jump = p_jump))
}
##
##HMC run
hmc_run <- function (starting_values, iter, epsilon_0, L_0, M) {</pre>
  # Get the number of rows and store in chains
  chains <- nrow (starting_values)</pre>
  # The number of parameters that you have in the starting values
  d <- ncol (starting_values)</pre>
  # Create space to store iterations of the parameters
  sims <- array (NA, c(iter, chains, d),</pre>
                  dimnames=list (NULL, NULL,
                  colnames (starting_values)))
  warmup <- 0.5*iter
  p_jump <- array (NA, c(iter, chains))</pre>
  for (j in 1:chains){
    param <- starting_values[j,]</pre>
    for (t in 1:iter){
      epsilon <- runif (1, 0, 2*epsilon_0)
      L <- ceiling (2*L_0*runif(1))
      temp <- hmc_iteration (param,x,y, epsilon,L,M)</pre>
      p_jump[t,j] <- temp$p_jump</pre>
      sims[t,j,] <- temp$param</pre>
      param <- temp$param
    } }
  monitor (sims, warmup)
  cat ("Avg_acceptance_probs:",
       fround(colMeans(p_jump[(warmup+1):iter,]),2),"\n")
  return (list (sims=sims, p_jump=p_jump))
}
                                    7
```

#### Listing 4: R code contd.

```
##RUN it
parameter_names <- c (paste ("beta[",1:2,"]",sep=""))</pre>
chains <- 4
log_post_interim <- function(param){</pre>
  log_prior <- 0</pre>
  log_likelihood <- sum(dbinom(y, n,</pre>
  invlogit(param[1] + param[2]*x),log = T))
  return(log_prior + log_likelihood)
}
lat <- laplace(log_post_interim, c(0.5,-0.5))</pre>
M <- ginv(lat$var)</pre>
mass_vector <- c(0.005, 0.001)
#Starts
starts <- array (NA,c(chains,d),</pre>
            dimnames=list(NULL,parameter_names))
for (j in 1:chains){
  starts[j,1] \leftarrow rnorm (1,0,0.5)
  starts[j,2] <- rnorm (1,0,0.5)
fit_hmc1 <- hmc_run(starting_values = starts,</pre>
                      iter = 1000,
                      epsilon_0 = 0.0005,
                      L_0 = 10,
                      M = mass_vector)
```

# Listing 5: R code contd. ############### ##Variational Bayes N <- 3020 $X \leftarrow matrix(c(rep(1, N), x), nrow = N, ncol = 2)$ #Lower bound function lower.bound2 <- function(param, X, stars, sigs, draws){</pre> ab <- t(X)\%\*\%X\%\*\%(param\%\*\%t(param) + sigs) part1 <- sum(diag(ab))/2</pre> part2 <- t(param)\%\*\%param + sum(diag(sigs))</pre> part2 <- part2\*(beta.mat.prior[1,1])</pre> part2 <- part2/2</pre> + (1/2)\*log(det(solve(beta.mat.prior))) + (length(param)/2)\*log(2\*pi) part3 <- t(stars)\%\*\%stars</pre> part4 <- length(param)/2</pre> + (1/2)\*log(det(sigs)) + (length(param)/2)\*log(2\*pi) bounds <- part1 + part2 + part3/2 + part4 parts <- c(-part1, -part2, part3/2, part4)</pre> bounds<- list(bounds, parts)</pre> names(bounds)<- c("bounds", "parts")</pre> return(bounds) ##we will use the lower-bound to monitor convergence, ##stopping the model when the lower-bound drops below 1e-8 func.reg<- function(X, draws){</pre> beta.mat.prior <<- diag(1/10, ncol(X))</pre> beta.VA <- matrix(NA, nrow=1000, ncol=ncol(X)) ystars <- rep(0, nrow(X))</pre> **for**(j in 1:nrow(X)){ ystars[j] <- ifelse(draws[j]==1,</pre> rtnorm(1, mean=0.5, sd=1, lower=0, upper=Inf), rtnorm(1, mean=-0.5, sd=1, lower=-Inf, upper=0)) } ## we will store the progress of the lower bound on the model bounds<- c() zz <- 0

# ##this stores the parts of the lower bound parts<- matrix(NA, nrow=1000, ncol=4) j <- 0 ##creating a while loop while(zz == 0){ j <- j + 1</pre>

beta.VA[j,] <- solve(t(X)\%\*\%X + beta.mat.prior)</pre>

##covariates current estimates of the coefficients

sigs <- solve(t(X)\%\*\%X + beta.mat.prior)</pre>

##now, computing the expected value for each

##computing the inner product of the

##individual's\_augmented\_data,\_given
\_\_\_\_##current\_estimates\_of\_the\_approximating

\_\_\_\_ystars[which(draws==0)]\_<-\_stars[draws==0]\_+
\_\_\_\_\_num1[which(draws==0)]/denom1[which(draws==0)]
\_\_\_\_ystars[which(draws==1)]\_<-\_stars[draws==1]\_+</pre>

\_\_\_\_num1[which(draws==1)]/(1\_-\_denom1[which(draws==1)])

\_\_##the\_information\_to\_be\_returned,\_after\_convergence

\_\_\_\_trial\_<-\_lower.bound2(beta.VA[j,],\_X,\_ystars,\_sigs,\_draws)</pre>

\_\_\_\_##distribution\_on\_the\_coefficients

\_\_\_\_##calculating\_the\_lower\_bound

\_\_\_\_ab\_<-\_abs(bounds[j]\_-\_bounds[j-1])

\_\_stuff<-\_list(bounds,\_beta.VA[j,],\_sigs)
\_\_names(stuff)<-\_c("bound","param",\_"sigma")</pre>

\_\_\_\_bounds[j]\_<-\_trial\$bounds
\_\_\_\_parts[j,]\_<-\_trial\$parts</pre>

\_\_\_\_##monitor\_convergence

\_\_\_\_if(ab\_<\_1e-8){
\_\_\_\_zz\_<-\_1}

\_\_return(stuff)

\_\_\_if(j>1){

\_\_\_}

}

##updating the beta parameters

stars <- X\%\*\%beta.VA[j,]
denom1 <- pnorm(-stars)
num1 <- dnorm(-stars)</pre>

\%\*\%t(X)\%\*\%ystars

### Listing 7: R code contd.

```
example.run<- func.reg(X, y)</pre>
example.run
betas <- rmvnorm(10<sup>3</sup>,
                 mean = example.run$param,
                 sigma = example.run$sigma)
#############
par(mfrow = c(1, 3),
    mgp = c(2,1,0),
    mar = c(2,3,4,2))
plot(fit_hmc1$sims[,,1],fit_hmc1$sims[,,2], ylim = c(0.15,0.30),
     xlim = c(-0.3, -0.05), pch = 16, cex = 0.7,
     main = "HMC", xlab = "alpha", ylab = "beta")
plot(ext_stan$alpha, ext_stan$beta, pch = 16, cex = 0.7,
     main = "Stan", xlab = "alpha", ylab = "beta", ylim = c(0.15,0.30),
     xlim = c(-0.3, -0.05))
plot(betas[,1], betas[,2], pch = 16, cex = 0.7,
     main = "VI", xlab = "alpha", ylab = "beta", ylim = c(0.15,0.30),
     xlim = c(-0.3, -0.05))
############
par(mfrow = c(1, 3))
plot(density(fit_hmc1$sims[,,1]),main = "HMC",
     xlab = "alpha", ylab = NA,
     bty = "n", yaxt = "n", xlim = c(-0.5, 0.1))
abline(v = mean(fit_hmc1$sims[,,1]), col = "red")
legend(-0.55,7, legend = c("posterior", "mean"), lty = c(1,1),
       col = c("black", "red"), bty = "n")
##
plot(density(ext_stan$alpha), main = "Stan",
     xlab = "alpha", ylab = NA,
     bty = "n", yaxt = "n", xlim = c(-0.5, 0.1))
abline(v = mean(fit_hmc1$sims[,,1]), col = "red")
legend(-0.55,7, legend = c("posterior", "mean"), lty = c(1,1),
       col = c("black", "red"), bty = "n")
plot(density(betas[,1]), main = "VI",
     xlab = "alpha", ylab = NA,
     bty = "n", yaxt = "n", xlim = c(-0.5, 0.1))
abline(v = mean(fit_hmc1$sims[,,1]), col = "red")
legend(-0.55, 8.5, legend = c("posterior", "mean"), lty = c(1,1),
       col = c("black", "red"), bty = "n")
```

#### Listing 8: R code contd.

```
##################
par(mfrow = c(1, 3))
plot(density(fit_hmc1$sims[,,2]),main = "HMC",
     xlab = "beta", ylab = NA,
     bty = "n", yaxt = "n", xlim = c(0, 0.4))
abline(v = mean(fit_hmc1$sims[,,2]), col = "red")
legend(-0.05,7, legend = c("posterior", "mean"), lty = c(1,1),
       col = c("black", "red"), bty = "n")
##
plot(density(ext_stan$beta), main = "Stan",
     xlab = "beta", ylab = NA,
     bty = "n", yaxt = "n", xlim = c(0, 0.4))
abline(v = mean(fit_hmc1$sims[,,2]), col = "red")
legend(-0.05,7, legend = c("posterior", "mean"), lty = c(1,1),
       col = c("black", "red"), bty = "n")
plot(density(betas[,2]),main = "VI",
     xlab = "beta", ylab = NA,
     bty = "n", yaxt = "n", xlim = c(0, 0.4))
abline(v = mean(fit_hmc1$sims[,,2]), col = "red")
legend(-0.05, 8.5, legend = c("posterior", "mean"), lty = c(1,1),
       col = c("black", "red"), bty = "n")
```

# 2.2 Stan Code

```
Listing 9: Stan Code
data{
int N;
int y[N];
real x[N];
}
parameters{
  real alpha;
  real beta;
}
model{
 for(i in 1:N)
  y[i] ~ bernoulli(Phi(alpha + beta * x[i]));
// }
// generated quantities{
//
    real y_rep[N];
//
    for(i in 1:N)
     y_rep[i] = bernoulli_rng(Phi(alpha + beta * x[i]));
//
}
```