

Signatures of non–small-cell lung cancer relapse patients: differential expression and gene network analysis

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Introduction

Lung cancer is both the second most represented cancer diagnosis and the leading cause of cancer death within the United States¹. Despite the high occurrence of non–small-cell lung cancer (NSCLC), 30 to 55% of patients relapse after curative resection, and the 5-year relative survival rate is 15 to 21%¹. Biomarker programs help select patients who may benefit from a given drug, thereby high costs of cancer medication and cancer drug failures may be reduced^{2,3}. We identified differentially expressed genes that are significantly linked to disease outcomes, which could lead to investigations of how these candidate biomarkers function to impact risk of relapse.

Methods

- NSCLC RNA samples were taken from 38 patients,
 - clinical outcomes were determined by the American College of Surgery Oncology Group ⁴;
 - 20 were diagnosed as disease-free, and 18 as relapse patients.
- RSEM⁵ was used to assemble and quantify the abundance of transcripts, which produced gene–count-based expression.
- EBSeq⁶ was used to perform median normalization and differential expression analysis with a threshold of false discovery rate (FDR) < 0.05.
- Differentially expressed genes were analyzed for over-representation of protein complexes, gene ontology terms and pathways with ConsensusPathDB⁷.

Results

Empirical Bayesian methods⁶ identified 122 differentially expressed genes (FDR < 0.05). Among these genes, only one protein complex-based set was over-represented, G protein-coupled receptor ligand. This included 7 of our differentially expressed genes, 4 of which have suggested implications in NSCLC prognosis: Existing research indicates that overexpression of TGF- α , IGF1, or IGF2 predicts poor survival, which is consistent with the gene expression of our relapse group⁸⁻¹⁰. Interestingly, while overexpression of HGF is also associated with poor survival, we saw higher expression of HGF in the disease-free patients¹¹⁻¹³.

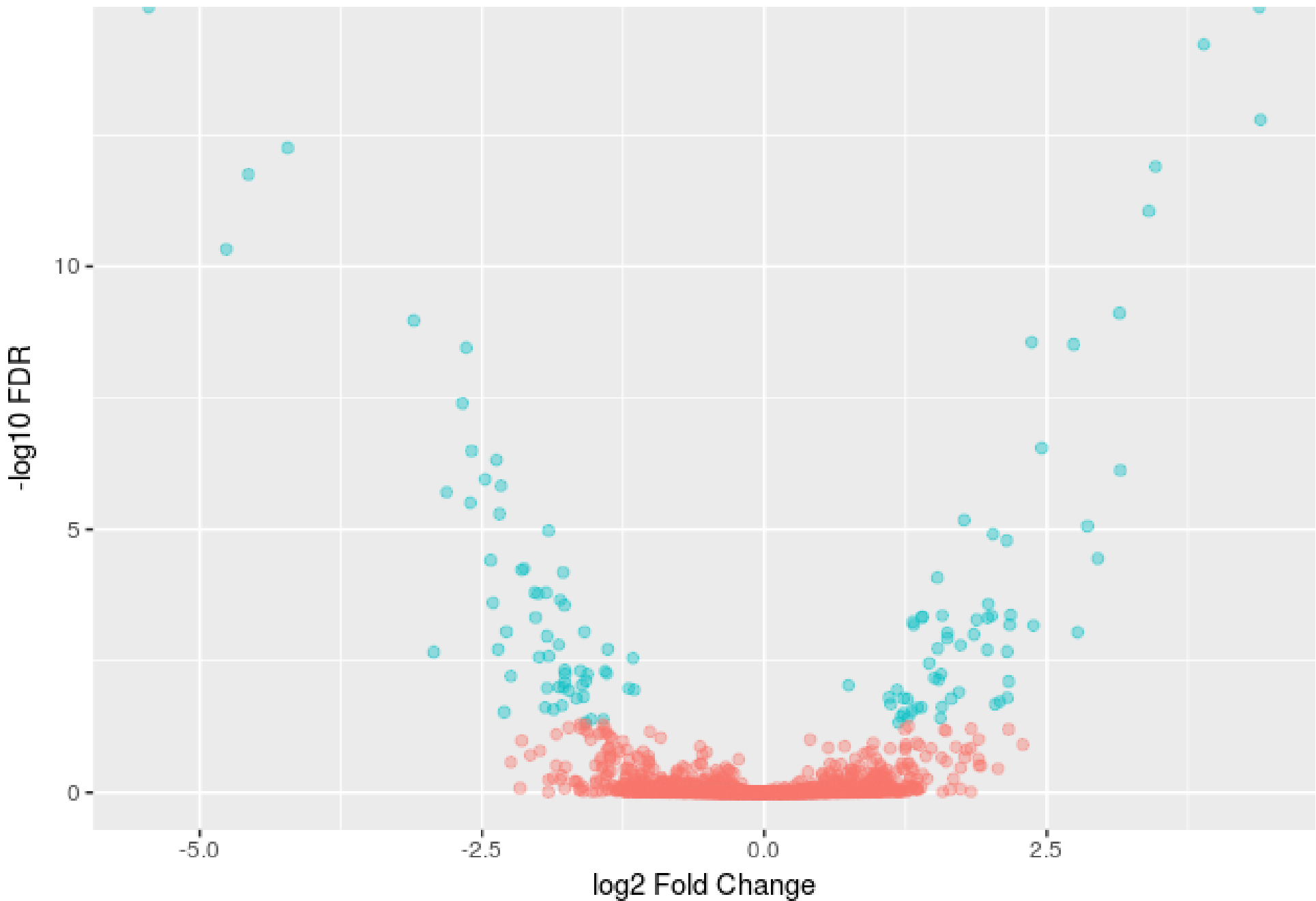


Figure 1 | Volcano plot of differentially expressed genes (blue) and similarly expressed genes (red).

| Gene | PPDE | Real FC | R Mean | DF Mean |
|----------|------|------------|-------------|------------|
| CPS1 | 1 | 21.0382203 | 3861.452147 | 183.53510 |
| UPK1B | 1 | 20.8719792 | 1640.955927 | 78.61052 |
| KLK11 | 1 | 14.8335518 | 1476.094288 | 99.50118 |
| CD70 | 1 | 11.0330937 | 161.379705 | 14.61778 |
| PI3 | 1 | 10.5927592 | 2248.496323 | 212.25824 |
| ADGRB1 | 1 | 8.8462140 | 885.679398 | 100.11073 |
| KIAA0087 | 1 | 6.6737762 | 118.581664 | 17.75980 |
| COMP | 1 | 5.1584334 | 2448.075614 | 474.56928 |
| SLC7A14 | 1 | 0.1605313 | 29.495441 | 183.78865 |
| C11orf70 | 1 | 0.1568151 | 55.741074 | 355.51117 |
| WNT11 | 1 | 0.1163370 | 63.640695 | 547.11328 |
| SHISA3 | 1 | 0.0536560 | 55.093038 | 1026.95836 |
| PRR4 | 1 | 0.0421482 | 48.838449 | 1158.95959 |
| SEC14L5 | 1 | 0.0367918 | 9.453563 | 257.20964 |
| LGALS4 | 1 | 0.0228392 | 28.253722 | 1237.50056 |

Table 1 | Top 15 differentially expressed genes (FDR < 0.05). Posterior probability of differential expression (PPDE); fold change (FC); relapse and disease-free normalized expression (R & DF means).

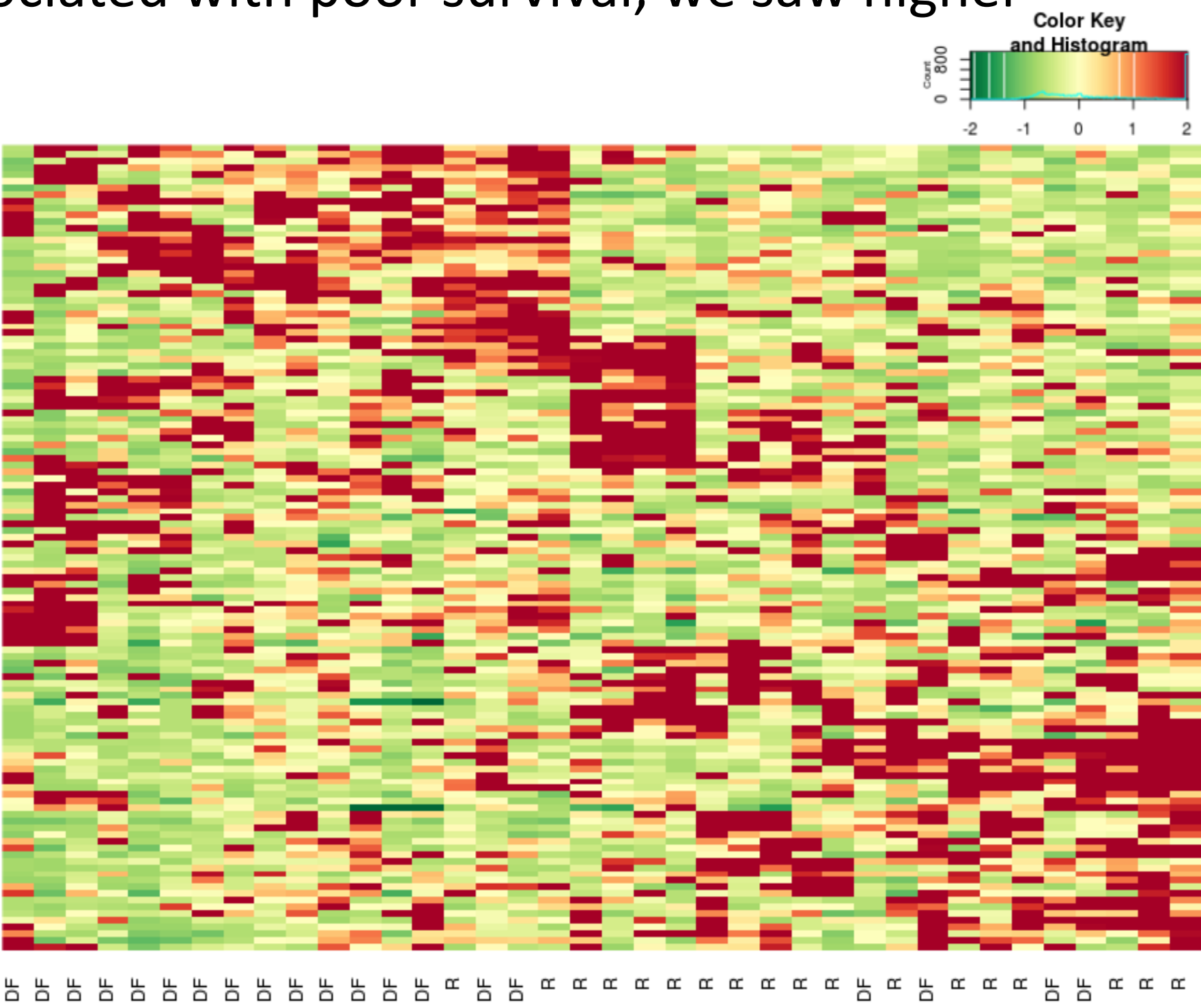
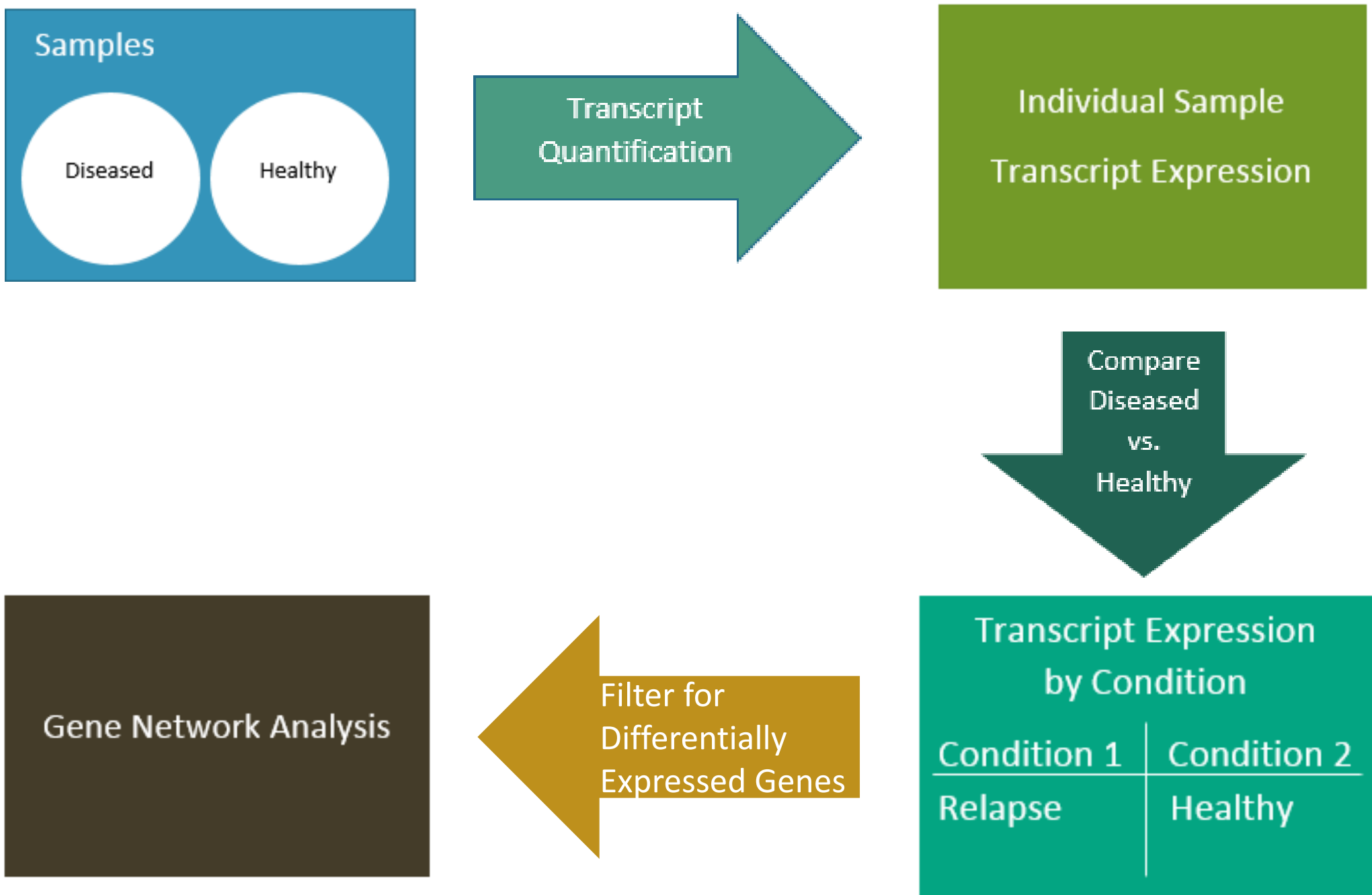


Figure 2 | Heatmap of normalized gene–count-based expression. Disease-free (DF); relapse (R). 122 differentially expressed genes on y-axis.

| GO term | Category, level | P-value | Q-value |
|--|-----------------|----------|---------|
| GO:0008284 positive regulation of cell proliferation | BP 5 | 4.66e-06 | 0.00142 |
| GO:0019372 lipoxygenase pathway | BP 5 | 6.95e-05 | 0.0106 |
| GO:0099503 secretory vesicle | CC 5 | 0.000236 | 0.00189 |
| GO:0033293 monocarboxylic acid binding | MF 5 | 0.000267 | 0.00479 |
| GO:0004252 serine-type endopeptidase activity | MF 5 | 0.000274 | 0.00479 |

| Pathway | Source | P-value | Q-value |
|--|--------------|----------|---------|
| GPCR signaling-G alpha i | INOH | 6.77e-05 | 0.00636 |
| GPCR signaling-pertussis toxin | INOH | 6.77e-05 | 0.00636 |
| Prostaglandin Synthesis and Regulation | Wikipathways | 8.8e-05 | 0.00636 |
| GPCR signaling-cholera toxin | INOH | 9.73e-05 | 0.00636 |
| GPCR signaling-G alpha s Epac and ERK | INOH | 0.000113 | 0.00636 |

Table 2 | (A) Top five level five gene ontology (GO) terms. **(B)** Top five pathways. Biological process (BP); cellular component (CC); molecular function (MF); Integrating Network Objects with Hierarchies database (INOH).



Conclusions

Identifying NSCLC patients at risk of recurrence is crucial in cancer research. Our analyses identified 122 differentially expressed genes among disease-free and relapse NSCLC patients, including known lung cancer-related genes and new candidate biomarker genes that are involved in the diverse processes related to NSCLC development. Future research in alternative splicing and the development of a predictive model based on our results could support methods of identifying individual recurrence risk.

References

- American Cancer Society. (2017) Cancer Facts & Figures 2017. Atlanta: American Cancer Society
- Goossens, N., Nakagawa, S., Sun, X., & Hoshida, Y. (2015). Cancer biomarker discovery and validation. *Translational Cancer Research*, 4(3), 256–269.
- Workman, P., Draetta, G. Schellens, J., Bernards, R. (2017). How Much Longer Will We Put Up With \$100,000 Cancer Drugs? *Cell*. 168(4):579-583.
- Anderson, P. *et al.* (2014). Predictive modeling of lung cancer recurrence using alternative splicing events versus differential expression data. *IEEE Conference on Computational Intelligence in Bioinformatics and Computational Biology*, 1–8.
- Li, B., & Dewey, C. N. (2011). RSEM: accurate transcript quantification from RNA-Seq data with or without a reference genome. *BMC Bioinformatics*, 12, 323.
- Leng, N., *et al.* (2013). EBSeq: an empirical Bayes hierarchical model for inference in RNA-seq experiments. *Bioinformatics*, 29(8), 1035–1043.
- Herwig, Ralf, *et al.* “Analyzing and interpreting genome data at the network level with ConsensusPathDB.” (2016). *Nature Protocols*, vol. 11, no. 10, pp. 1889–1907.
- Wu, H., Liu, Y., Shu, X. O., & Cai, Q. (2016). MiR-374a suppresses lung adenocarcinoma cell proliferation and invasion by targeting TGFA gene expression. *Carcinogenesis*, 37(6), 567–575.
- Kim, J.-S., Kim, E. S., Liu, D., Lee, J. J., Solis, L., Behrens, C., ... Lee, H.-Y. (2014). Prognostic Implications of Tumoral Expression of Insulin-like Growth Factor (IGF)-1 and -2 in Patients with Non-small Cell Lung Cancer. *Clinical Lung Cancer*, 15(3), 213–221.
- Velcheti, V., Ramaswamy, G. Insulin-Like Growth Factor and Lung Cancer. (2006) *Journal of Thoracic Oncology*, 1(7). Pp. 607 – 610.
- Ichimura, E., Maeshima, A., Nakajima, T., & Nakamura, T. (1996). Expression of c-met/HGF Receptor in Human Non-small Cell Lung Carcinomas in vitro and in vivo and Its Prognostic Significance. *Japanese Journal of Cancer Research*, 87(10), 1063-1069.
- Olivero, M., Rizzo, M., Madeddu, R., Casadio, C., Pennacchietti, S., Nicotra, M. R., ... Di Renzo, M. F. (1996). Overexpression and activation of hepatocyte growth factor/scatter factor in human non-small-cell lung carcinomas. *British Journal of Cancer*, 74(12), 1862–1868.
- Siegfried, J., Weissfeld, L., Singh-Kaw, P., Weyant, R., Testa, J., Landreneau, R. (1997). Association of Immunoreactive Hepatocyte Growth Factor with Poor Survival in Resectable Non-Small Cell Lung Cancer. *Cancer Res* 57(3), 433-439

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