

Genetics and Physiology of a *tolE* Mutant of *Escherichia coli* K-12 and Phenotypic Suppression of Its Phenotype by Galactose

KERSTIN G. ERIKSSON-GRENNBERG AND KURT NORDSTRÖM

Department of Microbiology, University of Umeå, S-901 87 Umeå, Sweden

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The *tolE* mutation causes tolerance to colicins E2 and E3 as well as other effects on the phenotype of *Escherichia coli* K-12. The lipopolysaccharide of the mutant shows a reduction in the content of galactose, glucose, and rhamnose. The phenotype of the mutant, including the composition of the lipopolysaccharide, is suppressed by galactose. The map position is shown by the gene order *trp-purB-tolE-tolD-galKETO*.

In a study of ampicillin-resistant mutants of *Escherichia coli* K-12, we found a class of mutants that were changed in the properties of the outer envelope. These mutants show a pleiotropic phenotype, and they can be divided into many subclasses (5). Two of these contain mutants (*tolD* and *tolE*) that are tolerant to colicins E2 and E3. The *tolD* type has been described before (3). In this paper we describe the *tolE* mutant.

The strains used in this study are listed in Table 1. The *tolE* mutant G11e6 differs in many respects from its parent strain G11a1, but the main features are tolerance to colicins E2 and E3, sensitivity to phage C21 (but resistance in the presence of galactose), and that the mutant is galactose negative when tested on purple base agar after pregrowth in the presence of glucose (5) (see Table 4). The *tolE* mutant was found to produce uridine 5'-diphosphate galactose-4-epimerase activity (tested as described in ref. 7).

The *tolE* mutation is transferred as a late marker in conjugations with HfrC. Since the mutant phenotype cannot be directly selected for, all genetic data are based on linkage to other genes. Strain G11e6 (*tolE*) was conjugated with strain MS31 (*galE*). Out of 92 Trp⁺ recombinants, 6 had obtained the *tolE* gene, whereas only one had lost the *galE* gene. Hence, the gene *tolE* cannot be closely linked to *galE*. The gene is transferred between *trp* and *galE*. Strain G11e6 (*tolE*) was conjugated with strain X195. The gene order was found to be *pyrF-trp-purB-tolE* (Table 2).

To map the *tolE* mutation in relation to *tolD*,

the conjugation reported in Table 3 was performed. Trp⁺ recombinants were selected, and four different phenotypes were obtained. Two of these were identical to the parental phenotypes and one was wild type with respect to *tolD* and *tolE*. This suggests that the fourth phenotype has the genotype *tolD, tolE*. Since wild-type recombinants were obtained, *tolE* cannot be allelic to *tolD*. Wild-type recombinants were much less frequent than the double mutants, which locates *tolE* between *trp* and *tolD*. Thus the gene order is *trp-purB-tolE-tolE-galKETO*. The locus *tolE*, as well as *tolD*, is genetically

TABLE 1. *E. coli* K-12 strains used and their relevant characters^a

Strain	Parent strain	Origin (reference no.)	Sex	Genotype ^b
G11a1	G11	4	HfrC	<i>ilv, metB, ampA1</i>
G11e1	G11a1	4	HfrC	<i>ilv, metB, ampA1, tolD</i>
G11e6	G11a1	5	HfrC	<i>ilv, metB, ampA1, tolE</i>
MS31	MS3	3	F ⁻	<i>pyrD, trp, galE, strA</i>
RE103	RC711	12	F ⁻	<i>proA, trp, his, cmlA, strA</i>
AS2	c	c	F ⁻	<i>proA, trp, tolD, strA</i>
X195	?	11	F ⁻	<i>his, met, pro, purB, pyrF, trp, tyr, strA</i>

^a All strains carry prophage λ.

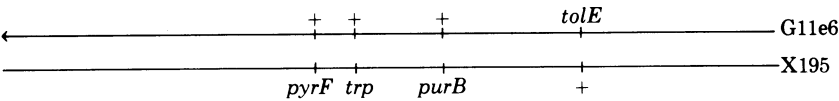
^b Abbreviations: *amp*, ampicillin; *cml*, chloramphenicol; *gal*, galactose; *his*, histidine; *ilv*, isoleucine-valine; *met*, methionine; *pro*, proline; *pur*, purine; *pyr*, pyrimidine; *str*, streptomycin; *tol*, tolerance to colicins E2 and E3; *trp*, tryptophan; *tyr*, tyrosine. The capital letters after some of the symbols refer to the genetic map of Taylor and Trotter (14).

^c A His⁺/Str^r-recombinant from a cross between G11e1 (*tolD*) and RE103 (*cmlA*). The strain AS2 is Cml⁺ and contains the gene *tolD*.

TABLE 2. Genotypes of recombinants from a cross between G11e6 and X195^a

Donor gene selected	Genotype of recombinant				% of recombinants
	<i>pyrF</i>	<i>trp</i>	<i>purB</i>	<i>tolE</i>	
<i>pyrF</i> ⁺	1	1	1	1	25
	1	1	1	0	33
	1	1	0	1	1
	1	1	0	0	27
	1	0	1	0	1
	1	0	0	0	13
<i>trp</i> ⁺	1	1	1	1	28
	1	1	1	0	46
	1	1	0	1	1
	1	1	0	0	17
	0	1	1	1	1
	0	1	1	0	4
	0	1	0	0	3
<i>purB</i> ⁺	1	1	1	1	29
	1	1	1	0	34
	1	0	1	0	2
	0	1	1	1	1
	0	1	1	0	3
	0	0	1	1	12
	0	0	1	0	19

^a Conjugations were performed as described before (5). In each case, 115 recombinants were tested. Streptomycin was used as counter selection. The genotypes of the parents are as follows:



In the body of the table, the donor allele is always labelled “1” and the recipient “0.”

TABLE 3. Results of conjugations between the donor strain G11e6 (*tolE*) and the recipient strain AS2 (*tolD*)^a

Recombinant class	Phenotype			Geno- type	% of Trp ⁺ recom- binants
	Resistance to:		Growth on ga- lactose ^b		
	Cholate (mg/ml)	Chlor- amphen- icol (μg/ml)			
Recipient type	< 10	5	+	<i>tolD</i>	46
Donor type	< 10	2	—	<i>tolE</i>	30
Wild type	50	2	+	+	2
Double re- combinant	50	5	+	<i>tolD</i> , <i>tolE</i>	22

^a Conjugations were performed as described before (5). Trp⁺-recombinants (175) were tested. Streptomycin was used as counter selection.

^b Tested on purple base agar containing galactose.

distinct from other loci, giving tolerance to colicins of the E group (3).

Sensitivity to phage C21 is due to a reduction of the galactose content of the lipopolysaccha-

ride (13). The lipopolysaccharide of strain G11e6 (*tolE*) contained a reduced amount of galactose, glucose, and rhamnose (Table 4). Sensitivity to phage C21 was lost when the cells were pregrown in the presence of galactose. This loss was accompanied by a restoration of the carbohydrate composition of the lipopolysaccharide. Furthermore, all other phenotypic properties were suppressed by galactose. This sugar had no effect on the *tolD* mutant G11e1.

Mutations in the *galU*, *lpsA*, or *lpsB* genes result in a reduction of the glucose, galactose, and rhamnose content of the lipopolysaccharide (5). However, none of these three mutations affects the response to colicins of the E group. To our knowledge, it has never been reported that colicin tolerance can be due to changes in the outer membrane. This strengthens the view that colicin tolerance can be a nonspecific consequence of changes in the cell envelope and due to impaired diffusion of colicin molecules through the cell wall into their targets (3). This is even more likely after the demonstration that colicin E3 acts as an enzyme on the ribosomes

TABLE 4. Effect of galactose on the carbohydrate composition of the lipopolysaccharide (LPS) and on some phenotypic properties of *tolE* and *tolD* mutants

Strain and carbon source added to LB medium	Lipopolysaccharide ($\mu\text{g}/\text{mg}$ of LPS) ^a					Survival in presence of colicin E2 ^b	Uptake of gentian violet (%) ^c	Resistance to sodium cholate ($\mu\text{g}/\text{ml}$) ^d	Response to phage C21
	Rhamnose	Galactose	Glucose	Heptose	Hexosamine				
G11a1 (wild type)									
Glucose	12	32	52	108	59	3.5×10^{-4}	51	37.5	R
Galactose	12	30	53	97	65	5.9×10^{-4}	43	37.5	R
G11e6 (<i>tolE</i>)									
Glucose	6	16	28	76	70	0.040	72	7.5	S
Galactose	11	32	69	105	59	8×10^{-4}	32	22.5	R
G11e1 (<i>tolD</i>)									
Glucose	19	27	62	95	56	0.58	77	10	R
Galactose	8	24	58	85	58	0.30	76	15	R

^a Lipopolysaccharide was determined as described before (5). Hexosamine was determined as described by Ghuyssen et al. (6).

^b The bacteria were grown in LB medium (1) or in LB medium in which glucose was replaced by galactose. In both cases the media were supplemented with medium E (15) and glucose, 0.2% (wt/vol). At a cell density of 4×10^8 cells per ml, colicin E2 was added at a multiplicity of 8 killing units per bacterium. After 10 min at 37 C, samples were taken, diluted 100 times in ice-cold LB, and viable count was determined. Survival was defined as the ratio between viable count of the sample and that of a parallel sample to which no colicin was added. Colicins were prepared by the method of Nagel de Zwaig and Luria (8).

^c The bacteria were grown as in footnote b. At a cell density of 2×10^8 cells per ml, they were chilled on ice, centrifuged, and resuspended in glucose minimal medium (15) containing 0.2% glucose and 10 μg of gentian violet. After 10 min on a rotary shaker at 37 C, the bacteria were removed by centrifugation, and the remaining gentian violet was determined at 590 nm (10).

^d Overnight cultures in minimal medium E (15) with either glucose or galactose as carbon sources were incubated in fresh media, and optical density was followed until 100 Klett units. The cells were diluted and 100 to 200 cells were spread on plates containing different concentrations (every 2.5 mg/ml from 0–55 mg/ml) of sodium cholate (single cell test) and the same medium as in the pregrowth medium. Resistance is given as the highest concentration at which the cells plated gave rise to the same size of colonies as on the control plates without cholate (9).

(2). Furthermore, survival of the *tolE* strain in presence of colicin E2 was far from complete (Table 4).

The gene product of the *tolE* gene is not known. However, one tempting possibility is that *tolE* is involved in the regulation of the *galU* and *pgm* genes. These genes are concerned with the biosynthesis of galactose uridine 5'-diphosphate and uridine 5'-diphosphate galactose, which are substrates for the biosynthesis of lipopolysaccharide. It is suggested that galactose acts by derepressing the *galU* and *pgm* genes to normal activity.

In conclusion, we have discovered a previously undescribed gene, *tolE*, which genetically and phenotypically is distinct from other genes causing tolerance to colicins of the E group (4). The effect of the mutation is phenotypically suppressed by galactose, which indicates that the *tolE* mutant is defective in the regulation of some reaction(s) that is (are) involved both in

galactose catabolism and in the biosynthesis of lipopolysaccharide.

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LITERATURE CITED

- Bertani, G. 1951. Studies on lysogenesis. I. The mode of phage liberation by lysogenic *Escherichia coli*. J. Bacteriol. 62:293–300.
- Bowman, C. M., J. Sidiharo, and M. Nomura. 1971. Specific inactivation of ribosomes by colicin E3 *in vitro* and mechanism of immunity in colicinogenic cells. Nature N. Biol. 234:133–137.
- Burman, L. G., and K. Nordström. 1971. Colicin tolerance induced by ampicillin or mutation to ampicillin resistance in a strain of *Escherichia coli* K-12. J. Bacteriol. 106:1–13.
- Eriksson-Grennberg, K. G., H. G. Boman, J. A. T. Jansson, and S. Thorén. 1965. Resistance of *Escherichia coli* to penicillins. I. Genetic study of some

- ampicillin-resistant mutants. *J. Bacteriol.* **90**:54-62.
5. Eriksson-Grennberg, K. G., K. Nordström, and P. Englund. 1971. Resistance of *Escherichia coli* to penicillins. IX. Genetics and physiology of class II ampicillin-resistant mutants that are galactose negative or sensitive to bacteriophage C21, or both. *J. Bacteriol.* **108**:1210-1223.
 6. Ghuysen, J.-M., D. J. Tipper, and J. L. Strominger. 1966. Enzymes that degrade bacterial cell walls, p. 685-699. In E. F. Neufeld and V. Ginsburg (ed.), *Methods in enzymology*, vol. 8. Academic Press Inc., New York.
 7. Maxwell, E. S., K. Kurahashi, and H. M. Kalckar. 1962. Enzymes of the Leloir pathway, p. 178. In S. P. Colowick and N. O. Kaplan (ed.), *Methods in enzymology*, vol. 5. Academic Press Inc., New York.
 8. Nagel de Zwaig, R., and S. E. Luria. 1967. Genetics and physiology of colicin-tolerant mutants of *Escherichia coli*. *J. Bacteriol.* **94**:112-113.
 9. Nordström, K., K. G. Eriksson-Grennberg, and H. G. Boman. 1968. Resistance of *Escherichia coli* to penicillins. III. Amp^B, a locus affecting episomally and chromosomally mediated resistance to ampicillin and chloramphenicol. *Genet. Res.* **12**:157-168.
 10. Normark, S., and B. Westling. 1971. Nature of the penetration barrier in *Escherichia coli* K-12: effect of macromolecular inhibition on penetrability in strains containing the *envA* gene. *J. Bacteriol.* **108**:45-50.
 11. Pearce, L. E., and E. Meynell. 1968. Specific chromosomal affinity of a resistance factor. *J. Gen. Microbiol.* **50**:159-172.
 12. Reeve, E. C. R. 1968. Genetical analysis of some mutations causing resistance to tetracycline in *Escherichia coli* K12. *Genet. Res.* **11**:303-309.
 13. Shedlovsky, A., and S. Brenner. 1963. A chemical basis for the host-induced modification of T-even bacteriophages. *Proc. Nat. Acad. Sci. U.S.A.* **50**:300-305.
 14. Taylor, A. L., and C. D. Trotter. 1972. Linkage map of *Escherichia coli* strain K-12. *Bacteriol. Rev.* **36**:504-524.
 15. Vogel, H. J., and D. M. Bonner. 1956. Acetylornithinase of *Escherichia coli*: partial purification and some properties. *J. Biol. Chem.* **218**:97-106.