

# HuBMAP + HPA – Hacking the Human Body

**Progress Meeting 7 Group A** 

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Stain Normalization

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In histopathology, stains are used to increase the contrast between different structures for their manual examination. These two chemicals typically stain: the nuclei a dark purple (Hematoxylin) and the cytoplasm a light pink (Eosin). Thus all pixels in an idealized histology image are principally composed of two colors.

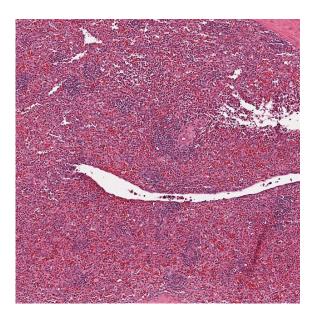
## **common problem**

One common problem in analysis of tissue samples is undesirable variation in color due to differences in color responses of slide scanners (as shown in Fig. 1), raw materials and manufacturing techniques of stain vendors, and staining protocols across different pathology labs.

This competition uses data from two different consortia, the **Human Protein Atlas (HPA)** and **Human BioMolecular Atlas Program (HuBMAP)**.



Training-bluish-gray



Testing-red or purple

## A METHOD FOR NORMALIZING HISTOLOGY SLIDES FOR QUANTITATIVE ANALYSIS

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## Structure-Preserving Color Normalization and Sparse Stain Separation for Histological Images

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1. Convert RGB to OD

$$OD = -\log\left(\frac{I}{255}\right)$$

2. Two vertical color matrices are obtained by SVD decomposition

stain matrix S concentration matrix C

3. Normalize the color matrix to Target (testing)

## Vahadane method

• SNMF——sparse non-negative matrix factorization (function: seperate the stains)

sparse: a given location(pixel) is likely to be occupied with just one material, not a mixture of two

non-negativity: make the optical density be zero or positive, optical density is meaningless if the value is negative

• SPCN——structure-preserving color normalization

changes color of one image (source) to match that of another (target) while reliably keeping source structural information intact

## StainTools

```
!pip install staintools
!pip install spams
```

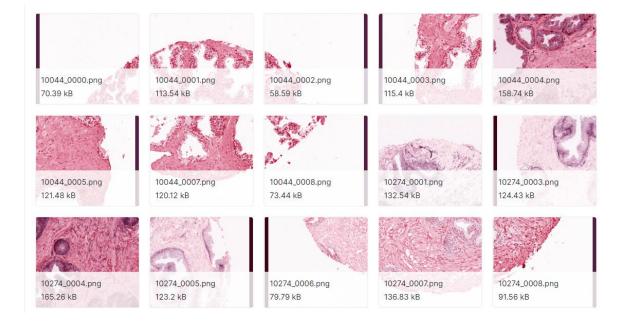
```
# Read data
target = staintools.read_image("./data/my_target_image.png")
to_transform = staintools.read_image("./data/my_image_to_transform.png")

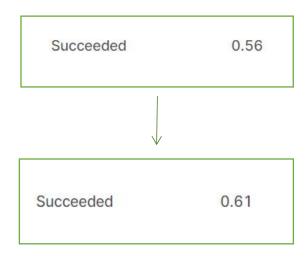
# Standardize brightness (This step is optional but can improve the tissue mask calculation
target = staintools.LuminosityStandardizer.standardize(target)
to_transform = staintools.LuminosityStandardizer.standardize(to_transform)

# Stain normalize
normalizer = staintools.StainNormalizer(method='vahadane')
normalizer.fit(target)
transformed = normalizer.transform(to_transform)
```

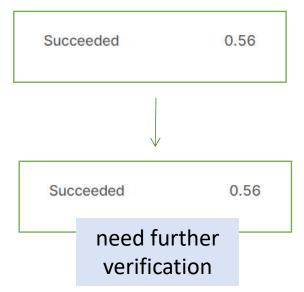
#### 10044\_0000.png 10044\_0001.png 10044\_0002.png 10044\_0003.png 10044\_0004.png 57.56 kB 62.73 kB 97.36 kB 96.25 kB 159.5 kB 10044\_0005.png 10044\_0007.png 10044\_0008.png 10274\_0001.png 10274\_0003.png 95.7 kB 89.81 kB 58.92 kB 103.87 kB 115.18 kB 10274\_0004.png 10274\_0005.png 10274\_0006.png 10274\_0007.png 10274\_0008.png 166.21 kB 115.19 kB 78.13 kB 112.4 kB 72.93 kB

## Vahadane method



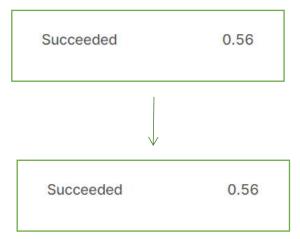


## Vahadane method





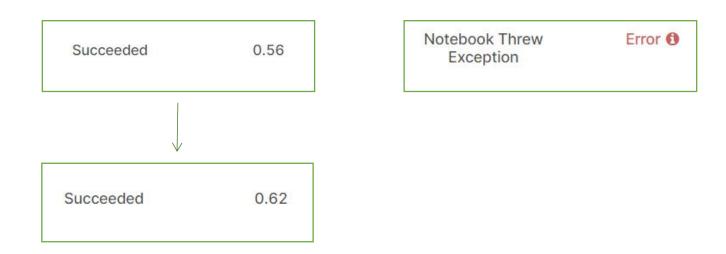
## Vahadane method



Only did colour normalization for the training sets

If we want to do colour normalization to testing sets, several problems...

- 1. Fail to pip install *spams* without Internet
- 2. We try to use *torchstain* instead of *staintools*, it has good effect as well, and it can be installed without Internet, while some errors happen during scoring



- 1. The cause of Vahadane's poor performance is being investigated: Is Vahadane a bad fit for this dataset or is it a problem with the dataset handling.
- 2. Try to resolve the error message
- 3. GAN-stain normalization for testing image
- 4. Find the final effect of combining the previous improvements with the normalization method that is known to be useful

## Thanks for listening