



Identification of GPCR Transition Pathways Using Gō Models

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Abstract

One caveat of standard all-atom molecular dynamics (MD) simulations is that they are resource-intensive, particularly when inspecting state transition pathways. To date, the timescales involved in processes such as G protein-coupled receptor (GPCR) activation make it largely impossible to study them using standard all-atom MD under equilibrium conditions. Our group previously showed that simpler structure-based (Gō-like) potentials could be used to study GPCR state transitions, yielding results in qualitative agreement with those obtained from standard all-atom simulations but with negligible computational cost. These potentials are energetically smooth by design, with few non-native traps and a single global energy minimum corresponding to the end-point of the transition of interest. In the case of rhodopsin, a prototypical GPCR, two distinct but reproducible pathways were observed during activation and deactivation. While it is possible that a separation of pathways is mechanistically required for function, it could also be the result of simulating state transitions in two separate energy surfaces out of equilibrium (one for activation and one for deactivation). To study state interconversion in equilibrium, we have implemented a strategy for modeling Gō-like potentials with multiple energy minima at all-heavy-atom resolution. This approach allows for extensive sampling still at a low computational cost and can be extrapolated to almost any biomolecular system in a straightforward manner to complement current standard all-atom MD strategies.

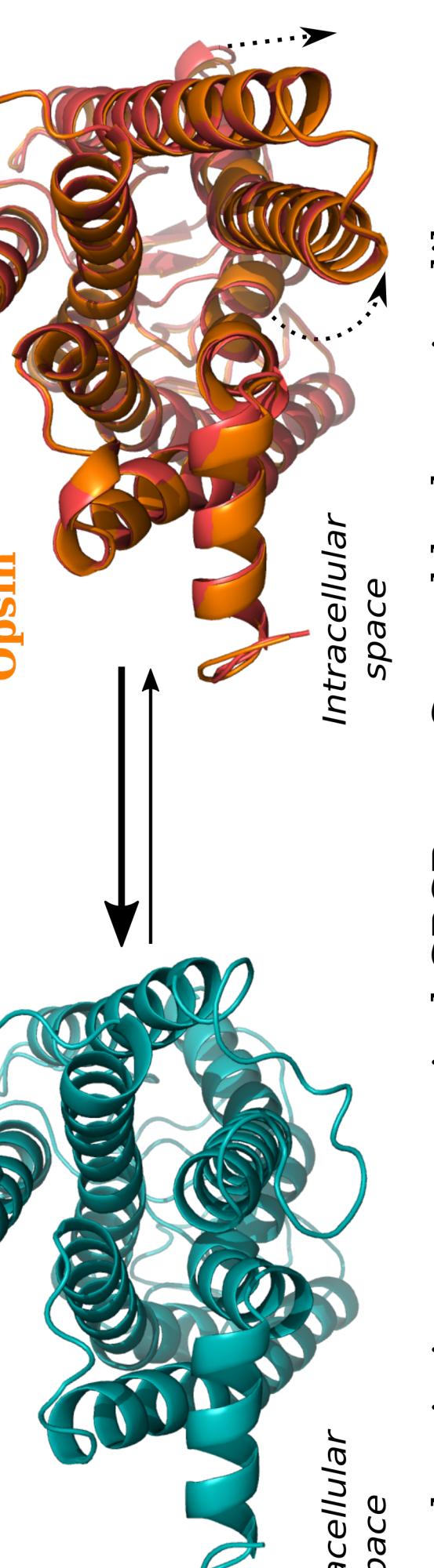


G Protein-Coupled Receptors

- Ubiquitous 7 transmembrane (7 TM) α -helical proteins
- Highly conserved topology
- Functional diversity and specificity
- 826 distinct members in humans
- Target of 40% small-molecule drugs
- Transduce information across membranes in response to stimuli
- Conformational changes mediate downstream signaling

Opsin Exists in Equilibrium Between Inactive-Like and Active-Like Conformations

Inactive-like
Active-like



- Rhodopsin is a prototypical GPCR
- Mediates visual signaling cascade
- Opsin: ligand-free (apo) rhodopsin
- Extremely low signaling
- Crystal looks active-like
- Opsin/Meta II RMSD is 0.51 Å
- Inactive ensemble more populated at physiological pH
- FTIR spectra inactive-like

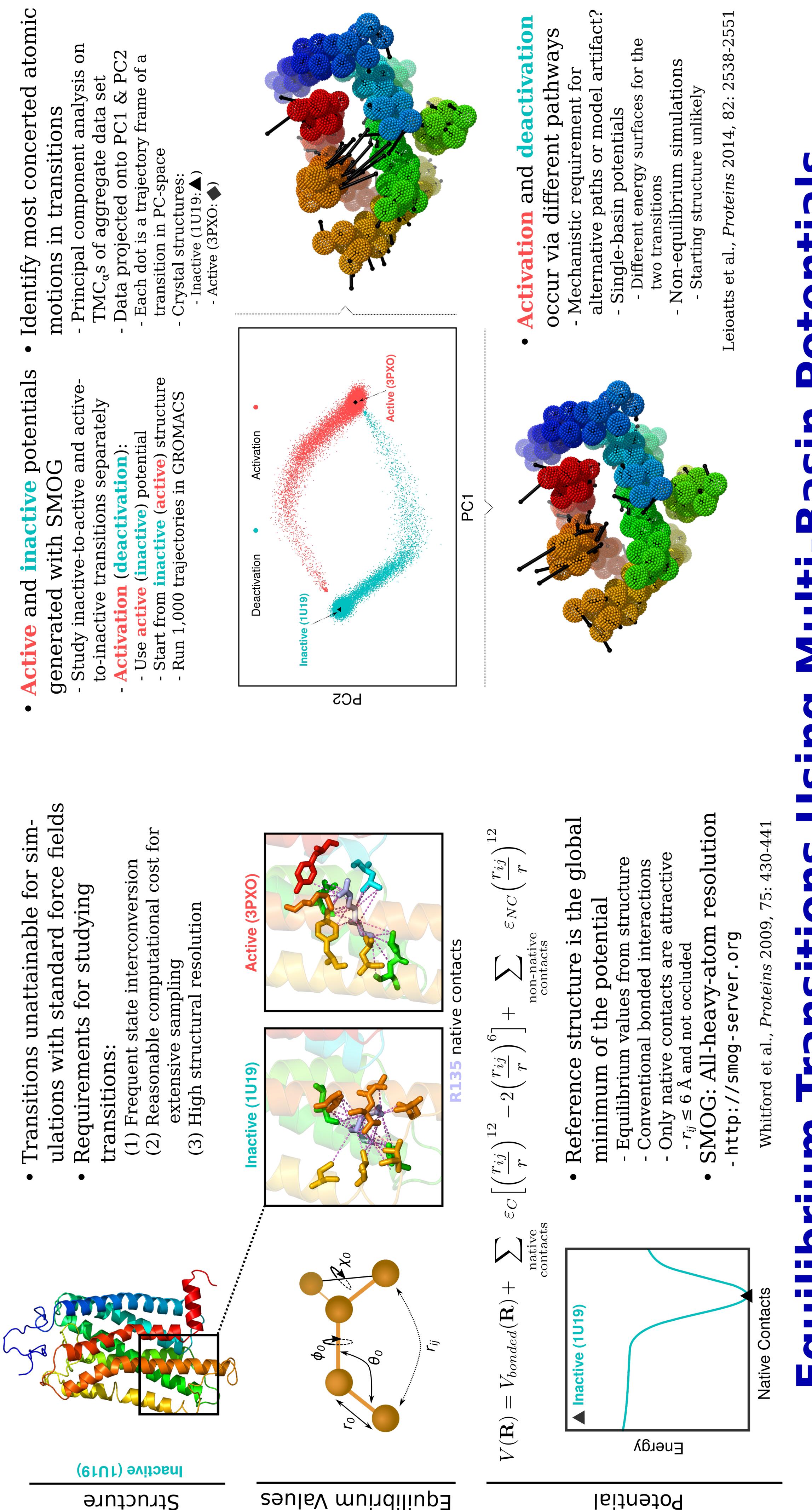
Standard MD Undersamples

Functional Transitions

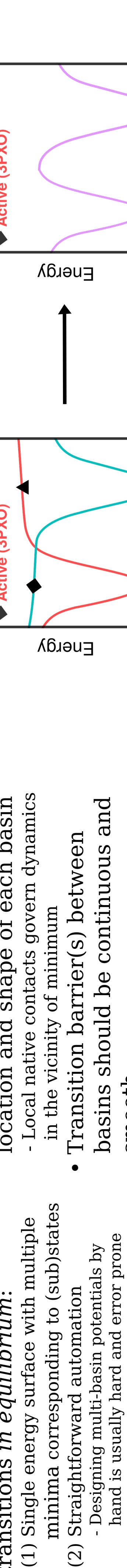
- Inactive-like and active-like opsins
- Starting structures:
 - Inactive-like: 1U19 (apo)
 - Active-like: 3CAP
- Standard all-atom MD simulations
- Aggregate simulation time: 55.2 μs
- 1.5 years of supercomputer time
- Ionic lock: R135(N)-E247(O)
- Distance: 4.7 Å
- Active-like: 15.8 Å
- Standard MD captures fluctuations within a state
- Transitions are rare

Opsin Transition Pathways from Single-Basin Gō Models

Gō-Like Models Are Structure-Based Potentials



Equilibrium Transitions Using Multi-Basin Potentials



- Requirements for studying state transitions in equilibrium:
 - Single energy surface with multiple minima corresponding to (sub)states
 - Straightforward automation
 - Designing multi-basin potentials by hand is usually hard and error-prone
 - Use global mixing schemes
 - Compute energies of each basin locally and then mix them macroscopically
- Mixed potential should preserve location and shape of each basin
 - Local native contacts govern dynamics in the vicinity of minimum
 - Transition barrier(s) between basins should be continuous and smooth
 - Dynamics propagation requires differentiable energies



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 - Compute energies of each basin locally and then mix them macroscopically
- Computationally convenient
 - Natural extension to multi-basin potentials
 - Each new potential adds an exponential term
 - Relative stability of each basin can be controlled individually

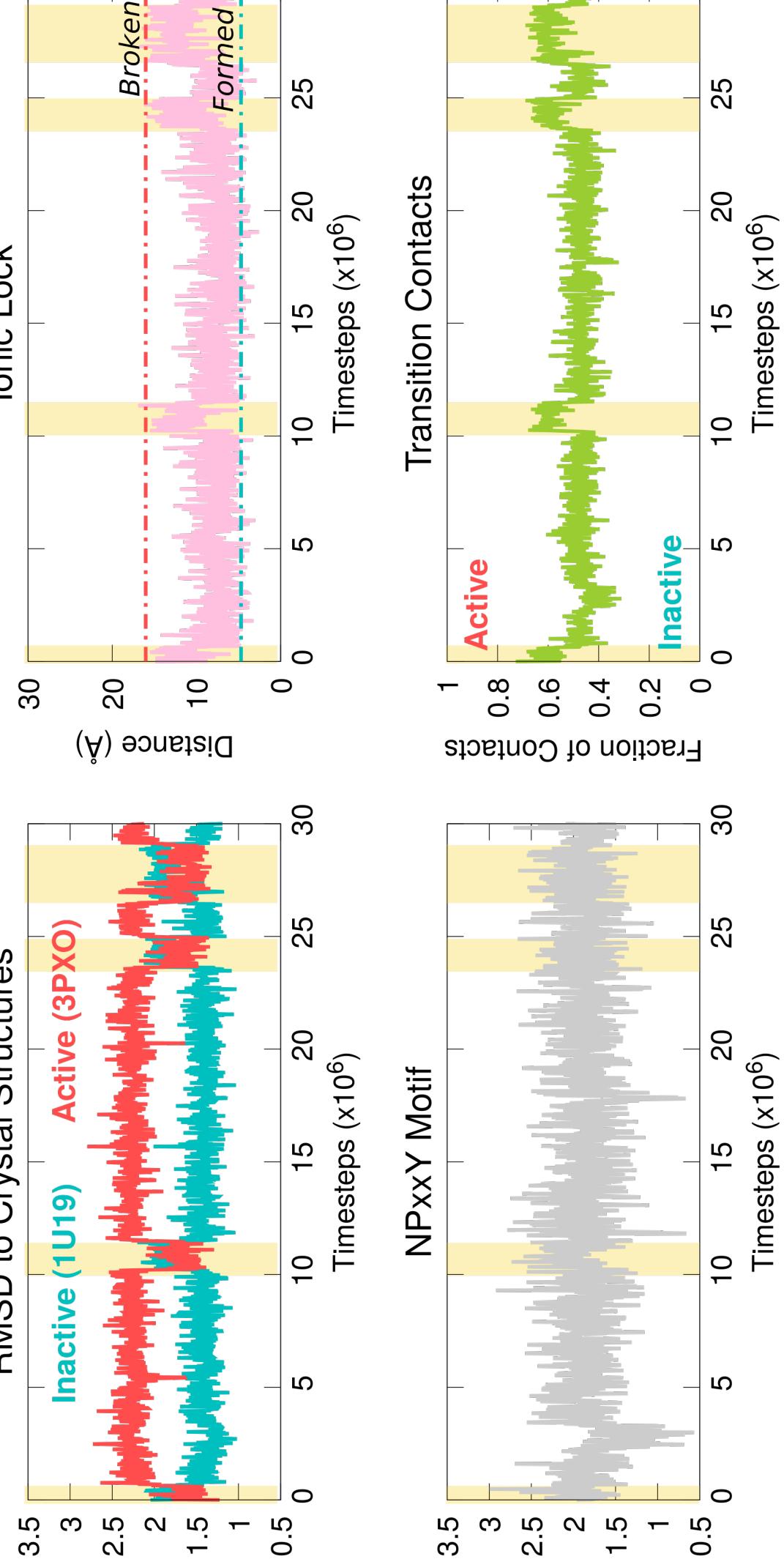


- Initial implementation in CHARMM
- Dynamics propagation is slow
- Current implementation in OpenMM
- Patch to support SMOC input files in OpenMM Github as of April 2017
- Multi-basin potentials implemented as a Python module
- Interpolation and Boltzmann-weighted formalisms implemented as CustomIntegrators
 - Mixed potential energy accessible as a StateData reporter
- SHAKE requires matching bond lengths among sets of potentials
- Bonds averaged over all potentials

Simulation Details

Starting Structure	PDB Accession Codes	Potentials	Resources
Inactive	1U19 (apo)	Dual-basin using Boltzmann-Weighting and Interpolation	10 ⁶ timesteps 1 CPU 5.8 hours
Active	3PXO (apo)		

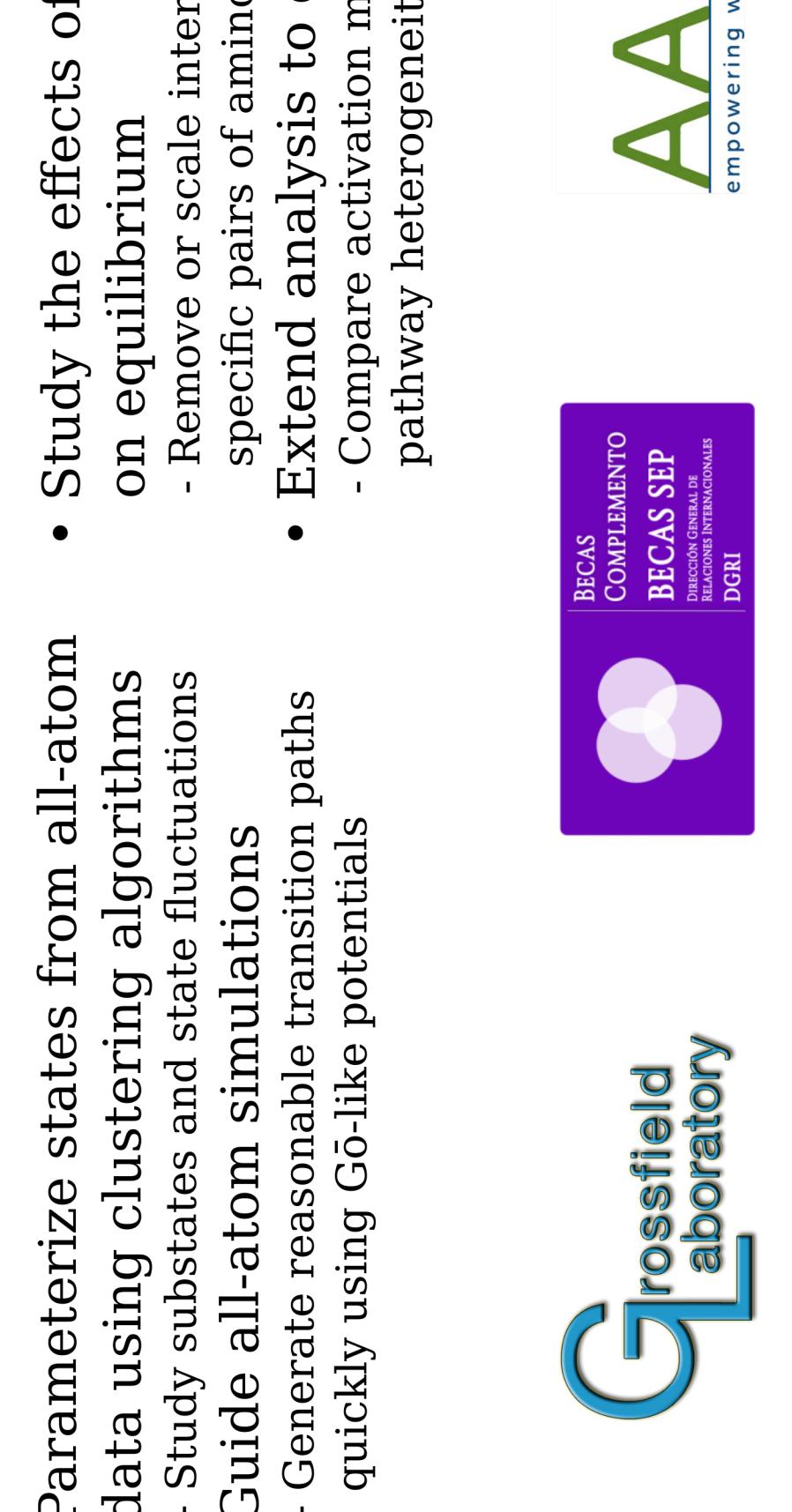
Opsin Transitions in Equilibrium



- System size: ~2,600 atoms
 - Protein stable in the absence of a lipid bilayer and water
- Force field: SMOC1/2
- Ensemble: NVT
 - T = 50 K in reduced units
- Langevin dynamics
 - Collision frequency = 0.1 ps⁻¹
 - VDW cutoff: 10 Å
 - No electrostatics
 - Timestep: 2 fs
- Software: OpenMM compiled with Clang and OpenCL

Conclusions

- Transition contacts
 - Set of contacts derived from transition end-points: unique active (inactive)
 - Evolution of state-specific contacts Side-chain centroids within 8 Å transition region
 - Transition = native broken + active / native + active
 - 1. All unique active (inactive) formed (broken) 0. No unique active (inactive) formed (broken)
- Leioatts et al., *Proteins* 2014, 82: 2538-2551
- Whitford et al., *Proteins* 2009, 75: 430-441
- Double-basin interpolation
 - Starting from **inactive** structure
 - $\Delta V_{\text{active}} = 0 \text{ kJ/mol}$ and $\Delta V_{\text{inactive}} = 0 \text{ kJ/mol}$
 - Single-basin potentials
 - Different energy surfaces for the two transitions
 - Non-equilibrium simulations
 - Starting structure unlikely
 - Double-basin interpolation trajectory
 - Starting from **active** structure
 - $\Delta V_{\text{active}} = 0 \text{ kJ/mol}$ and $\Delta V_{\text{inactive}} = 0 \text{ kJ/mol}$
 - Gō-like models can capture state transitions
 - Study state interconversion at negligible computational cost
 - Retain all-heavy-atom resolution
- Standard MD undersamples state transitions in complex systems
 - Model multiple states in the same energy surface
 - Study transitions in equilibrium
 - Formalism is systematic-independent
 - Readily extrapolated to biomolecular systems with two or more known states
- Leioatts et al., *Biophys. J.* 2015, 109: 608-617



Future Directions

- Study the effects of "mutations" on equilibrium
 - Remove or scale interaction between specific pairs of amino acids
 - Extend analysis to other GPCRs
 - Compare activation mechanism and pathway heterogeneity
- Data analysis was performed using LOOS (Lightweight Object-Oriented Structure library, an open source C++ and Python library for MD analysis developed by the Grossfield lab)
 - AAUW: empowering women since 1881
 - https://github.com/grossfield/loos



AAUW
empowering women since 1881
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Equation for Native Contacts:
$$V_{\text{mix}}(\mathbf{R}) = \frac{V_1(\mathbf{R}) + V_2(\mathbf{R}) + \Delta V_1}{2} - \sqrt{\left(\frac{V_1(\mathbf{R}) + V_2(\mathbf{R}) + \Delta V_2}{2} \right)^2 + \Delta^2}$$

Equation for Interpolation:

$V_{\text{mix}}(\mathbf{R}) = \frac{V_1(\mathbf{R}) + \Delta V_1 + V_2(\mathbf{R}) + \Delta V_2}{2} - \sqrt{\left(\frac{V_1(\mathbf{R}) + V_2(\mathbf{R}) + \Delta V_2}{2} \right)^2 + \Delta^2}$

Equation for Boltzmann-Weighting:

$V_{\text{mix}}(\mathbf{R}) = -\frac{1}{\beta_{\text{mix}}} \ln \left(\exp(-\beta_{\text{mix}}(V_1(\mathbf{R}) + \Delta V_1)) + \exp(-\beta_{\text{mix}}(V_2(\mathbf{R}) + \Delta V_2)) \right)$

Equation for Local native contacts:

$V^*(r) = \varepsilon_{ij} \left(1 + \left(\frac{\varepsilon_o}{\varepsilon_{ij}} \right) \left(\frac{r_{\text{rep}}}{r} \right)^{12} \right) \left(1 - e^{-\frac{(r-r_{ij})^2}{2r_{ij}^2}} \right) - 1$

where,

- r_{ij} : Critical distance

- ε_{ij} : Basin depth

- r_{rep} : Excluded volume

- ε_o : 1 kJ/mol

Okazaki et al., *PNAS* 2006, 103: 11844-11849

Noel and Onuchic, *Springer US* 2012, 31:54