LAB 2 BLOCK 2 - Group A15

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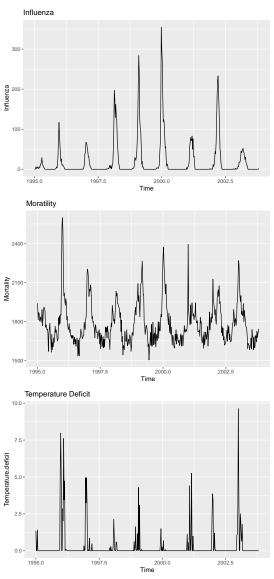
16 December 2019

Assignment 1. Using GAM and GLM to examine mortality rates

The Excel document influenza.xlsx contains weekly data on the mortality and the number of laboratory-confirmed cases of influenza in Sweden. In addition, there is information about population-weighted temperature anomalies (temperature deficits).

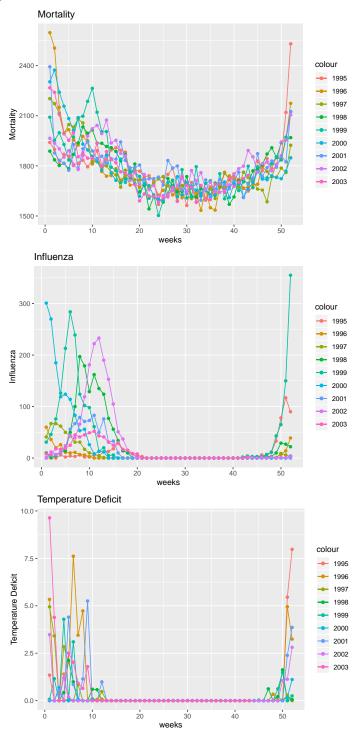
1. Use time series plots to visually inspect how the mortality and influenza number vary with time (use Time as X axis). By using this plot, comment how the amounts of influenza cases are related to mortality rates.

Plotting the data against the time:



It seems to be a modality in the data. It can be seen that Influenza peaks seems to have a correlation with peaks in mortality and temperature deficit

As the model is meant to learn from weeks variations along the year, it is better to separate the data into several time series per year and see how the data looks now.



Now it is quite clear that the data shows that during winter weeks Influenza cases and Mortality related to increases, also the temperature deficit seems to follow that trend.

2. Use gam() function from mgcv package to fit a GAM model in which Mortality is normally distributed and modelled as a linear function of Year and spline function of Week, and make sure that the model parameters are selected by the generalized cross-validation. Report the underlying probabilistic model.

It is fitted one model with the following formula:

```
Mortality \approx Year + spline(Week)
```

The summary of the obtained probabilistic model is:

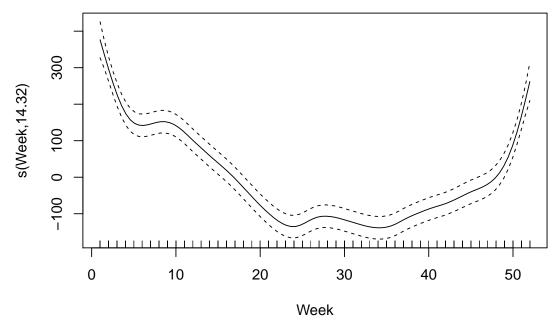
```
##
## Family: gaussian
## Link function: identity
##
## Formula:
## Mortality ~ Year + s(Week, k = length(unique(data$Week)))
##
## Parametric coefficients:
##
               Estimate Std. Error t value Pr(>|t|)
## (Intercept) -680.598
                          3367.760
                                    -0.202
                                              0.840
                             1.685
## Year
                  1.233
                                     0.732
                                              0.465
##
## Approximate significance of smooth terms:
             edf Ref.df
                            F p-value
## s(Week) 14.32 17.87 53.86 <2e-16 ***
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## Rank: 52/53
## R-sq.(adj) = 0.677
                         Deviance explained = 68.8%
## GCV = 8708.6 Scale est. = 8398.9
The model Mean and variance are :
## [1] "Train predictions mean:"
## [1] 1783.765
## [1] "Train predictions variance:"
## [1] 25981.62
```

Thus as the summary states that the model family is Gaussian, it can be said that:

$$Mortality \sim Year + spline(Week)$$
 $Mortality \sim N(\mu = 1783.765, \sigma = 25981.62)$

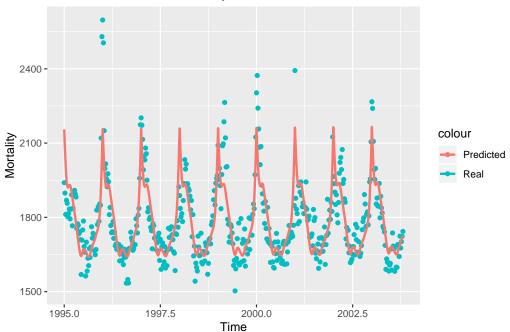
The smoother curve approximated by the GAM model is:

Smooth Approximation of Mortality



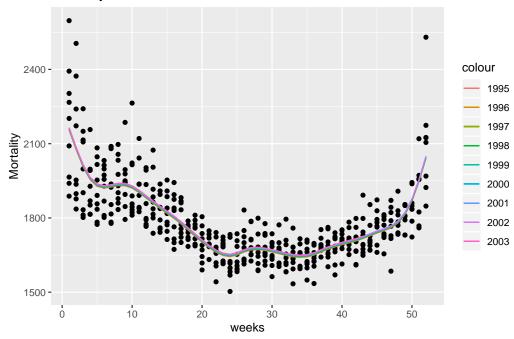
3. Plot predicted and observed mortality against time for the fitted model and comment on the quality of the fit. Investigate the output of the GAM model and report which terms appear to be significant in the model. Is there a trend in mortality change from one year to another? Plot the spline component and interpret the plot.





Although it seems to be a fair general approximation it seems to be missing some abnormal positive and negative peaks.

Mortality Overall Real and Predictions



It can be seen that given the variability of mortality per week each year, the GAM fit acceptably well the data, though still too general, thus not as good as we would like to.

The 10 most significant terms in the model are:

```
## [1] "The 10 most important coefficients:"

## s(Week).26 s(Week).36 s(Week).25 s(Week).20 s(Week).22 s(Week).39 s(Week).18

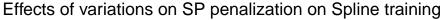
## 6962.767 6367.790 6224.860 3619.116 2193.093 2038.671 1588.407

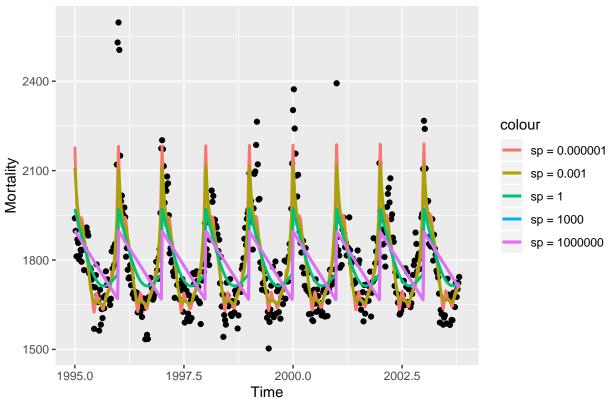
## s(Week).16 s(Week).40 s(Week).12

## 1478.685 1468.449 1406.516
```

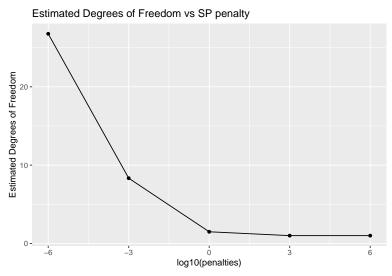
4. Examine how the penalty factor of the spline function in the GAM model from step 2 influences the estimated deviance of the model. Make plots of the predicted and observed mortality against time for cases of very high and very low penalty factors. What is the relation of the penalty factor to the degrees of freedom? Do your results confirm this relationship?

The smoothing penalty in gam() is given by the sp parameter sent to the constructor to be used in training. For this testing puropose it have been used the following penalties $\{10^{-6}, 10^{-3}, 1, 10^{3}, 10^{6}\}$





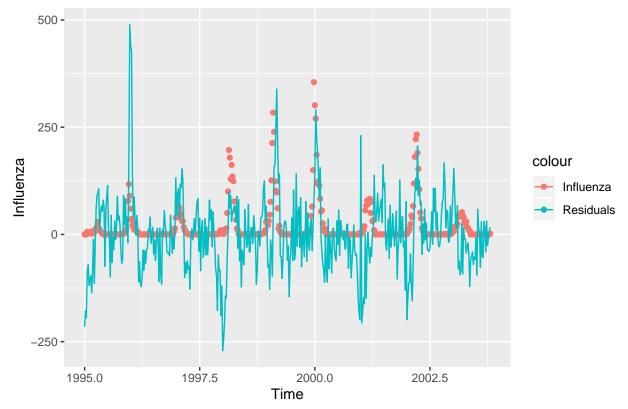
It can be seen that the lower the penalty the better the fit. In this case it looks good to have such a low penalty, though in other cases it might overfit the data.



Extracting the degrees of freedom from each penalization iteration it can be seen that the lower the penalization the more relaxed is the model, thus, the higher the degrees of freedom. The penalty factor is inversely proportional to the degrees of freedom. Indeed, by looking at the above plots, this relationship is confirmed. A high penalty essentially drops a higher number of parameters in the linear smoother matrix S_{λ} (by approximating them to zero).

5. Use the model obtained in step 2 and plot the residuals and the influenza values against time (in one plot). Is the temporal pattern in the residuals correlated to the outbreaks of influenza?

At a first glance Residuals seem to be noise compared to Influenza behavior.



Nonetheless visual assumptions might be incorrect. Testing numerically by getting their correlation statistic is low, thus there is no evidence to say that the residuals from the model and the influenza measures are correlated.

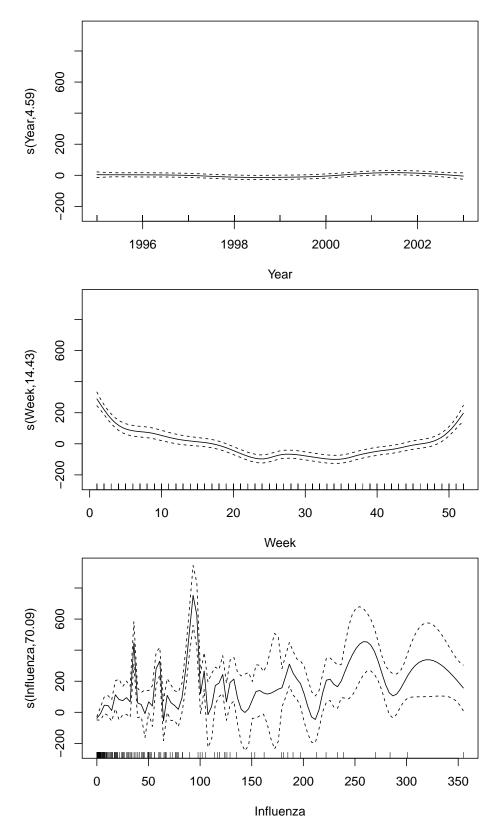
[1] 0.3397395

6. Fit a GAM model in R in which mortality is be modelled as an additive function of the spline functions of year, week, and the number of confirmed cases of influenza. Use the output of this GAM function to conclude whether or not the mortality is influenced by the outbreaks of influenza. Provide the plot of the original and fitted Mortality against Time and comment whether the model seems to be better than the previous GAM models.

It has been trained a GAM using the following formula:

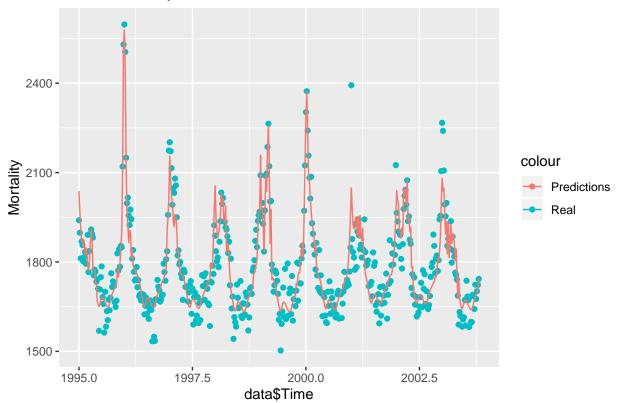
$$Mortality \approx spline(Year) + spline(Week) + spline(Influenza)$$

The obtained smoothers are the following.



Now, using the model to predict the Mortality :

Second GAM predictions



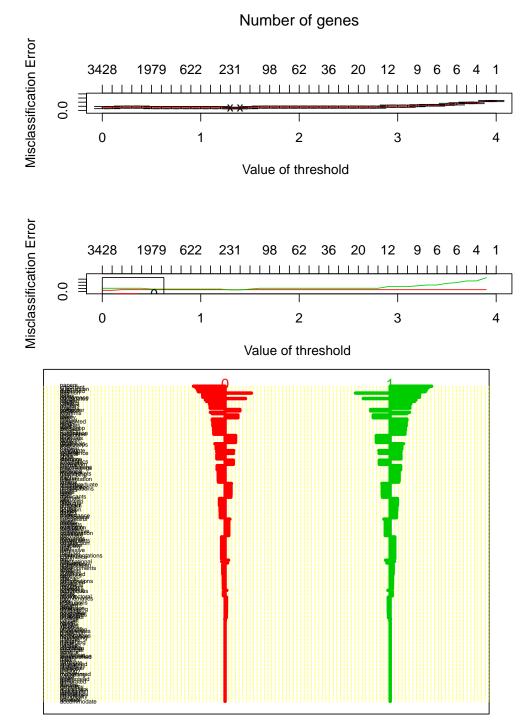
The model seems to fit better the data, though there are still some data points not being well approximatted. This is sign that the predictor is not overfitted, so compared against the previous predictors, this is the best one.

Assignment 2. High-dimensional methods

The data file data.csv contains information about 64 e-mails which were manually collected from DBWorld mailing list. They were classified as: 'announces of conferences' (1) and 'everything else' (0) (variable Conference)

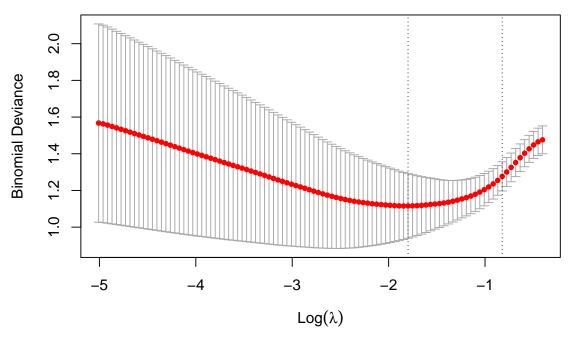
```
## Warning in RNGkind("Mersenne-Twister", "Inversion", "Rounding"): non-uniform
## 'Rounding' sampler used
```

1. Divide data into training and test sets (70/30) without scaling. Perform nearest shrunken centroid classification of training data in which the threshold is chosen by cross-validation. Provide a centroid plot and interpret it. How many features were selected by the method? List the names of the 10 most contributing features and comment whether it is reasonable that they have strong effect on the discrimination between the conference mails and other mails? Report the test error.



We get the threshold is 1.3 with using cross-validation. From the centroid plot we can see the contribution of each word made to the result(conference or not). There are 693 features selected in total. The 10 most contributing features are "papers", "important", "submission", "due", "published", "position", "call", "conference", "dates", "candidates". It's clear that these word have a strong connection to conference. The test error is 5%. 2. Compute the test error and the number of the contributing features for the following methods fitted to the training data:

a. Elastic net with the binomial response and $\alpha = 0.5$ in which penalty is selected by the cross-validation



10 %

b. Support vector machine with "vanilladot" kernel.

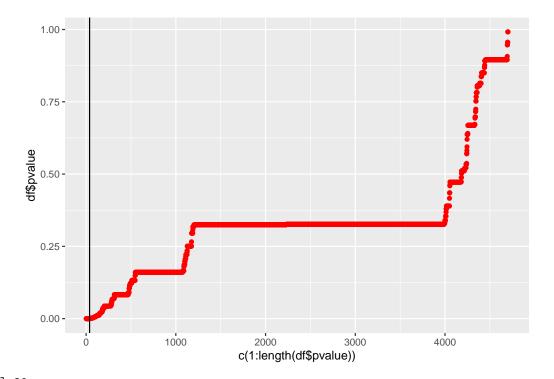
Setting default kernel parameters

5 %

Compare the results of these models with the results of the nearest shrunken centroids (make a comparative table). Which model would you prefer and why?

Error rate for Elastic net is 10% and for SVM is 5%. In this case we prefer to use SVM since it ignores the effect of high-dimensional data and it provides the lowest misclassification rate.

3. Implement Benjamini-Hochberg method for the original data, and use t.test() for computing p-values. Which features correspond to the rejected hypotheses? Interpret the result.



```
## [1] 39
##
                             pvalue
                 name
               papers 1.116910e-10
## 3036
## 4060
           submission 7.949969e-10
             position 8.219362e-09
## 3187
## 3364
            published 1.835157e-07
## 2049
            important 3.040833e-07
## 596
                  call 3.983540e-07
## 869
           conference 5.091970e-07
## 607
           candidates 8.612259e-07
## 1045
                dates 1.398619e-06
## 3035
                paper 1.398619e-06
## 4282
               topics 5.068373e-06
## 2463
              limited 7.907976e-06
## 606
            candidate 1.190607e-05
## 599
               camera 2.099119e-05
## 3433
                ready 2.099119e-05
## 389
              authors 2.154461e-05
## 3125
                  phd 3.382671e-05
## 3312
             projects 3.499123e-05
## 2974
                  org 3.742010e-05
## 681
               chairs 5.860175e-05
## 1262
                  due 6.488781e-05
## 2990
             original 6.488781e-05
## 2889
         notification 6.882210e-05
## 3671
               salary 7.971981e-05
## 3458
               record 9.090038e-05
## 3891
               skills 9.090038e-05
## 1891
                 held 1.529174e-04
## 4177
                 team 1.757570e-04
## 3022
                pages 2.007353e-04
```

```
## 4628
         worksnop 2.00.00
committee 2.117020e-04
           workshop 2.007353e-04
## 810
## 3285
        proceedings 2.117020e-04
## 272
               apply 2.166414e-04
## 4039
               strong 2.246309e-04
## 2175 international 2.295684e-04
              degree 3.762328e-04
## 1088
## 1477
           excellent 3.762328e-04
                post 3.762328e-04
## 3191
## 3243
           presented 3.765147e-04
```

There are 39 features correspond to the rejected hypotheses with $\alpha=0.05$.

Appendix A: Code for Assignment 1

```
Assignment 1
# Import data
dataPath <- "data/influenza.xlsx"</pre>
data <- read.xlsx(dataPath)
# Time Series plotting
mortPlot <- ggplot(data) +</pre>
    geom_line(aes(x=Time, y=Mortality), color="black") + ggtitle("Moratility")
infPlot <- ggplot(data) +</pre>
    geom_line(aes(x=Time, y=Influenza), color="black") + ggtitle("Influenza")
tempPlot <- ggplot(data) +</pre>
   geom_line(aes(x=Time, y=Temperature.deficit), color="black") +
   ggtitle("Temperature Deficit")
infPlot
mortPlot
tempPlot
# Time series per year
years <- unique(data$Year)</pre>
weeks <- unique(data$Week)</pre>
mortData <- list()</pre>
infData <- list()</pre>
tempData <- list()</pre>
for(i in 1:length(years)) {
   year <- years[i]</pre>
   mortData[[i]] <- data$Mortality[which(data$Year == year)]</pre>
   infData[[i]] <- data$Influenza[which(data$Year == year)]</pre>
   tempData[[i]] <- data$Temperature.deficit[which(data$Year == year)]</pre>
}
names(mortData) <- years</pre>
names(infData) <- years</pre>
names(tempData) <- years</pre>
# create data.frames for ggplot
plotData <- function(d, title) {</pre>
   shortWeeks <- 1:length(d$'2003')</pre>
   p <- ggplot() +
       geom_line(aes(x=weeks, y=d$'1995', color="1995")) +
       geom_line(aes(x=weeks, y=d$'1996', color="1996")) +
       geom_line(aes(x=weeks, y=d$'1997', color="1997")) +
       geom_line(aes(x=weeks, y=d$'1998', color="1998")) +
       geom_line(aes(x=weeks, y=d$'1999', color="1999")) +
       geom_line(aes(x=weeks, y=d$'2000', color="2000")) +
       geom_line(aes(x=weeks, y=d$'2001', color="2001")) +
       geom_line(aes(x=weeks, y=d$'2002', color="2002")) +
       geom_line(aes(x=shortWeeks, y=d$'2003', color="2003")) +
       geom_point(aes(x=weeks, y=d$'1995', color="1995")) +
       geom_point(aes(x=weeks, y=d$'1996', color="1996")) +
       geom_point(aes(x=weeks, y=d$'1997', color="1997")) +
```

```
geom_point(aes(x=weeks, y=d$'1998', color="1998")) +
        geom_point(aes(x=weeks, y=d$'1999', color="1999")) +
        geom_point(aes(x=weeks, y=d$'2000', color="2000")) +
        geom_point(aes(x=weeks, y=d$'2001', color="2001")) +
        geom_point(aes(x=weeks, y=d$'2002', color="2002")) +
        geom_point(aes(x=shortWeeks, y=d$'2003', color="2003")) +
        ggtitle(title) + ylab(title)
    return(p)
}
mortPlot <- plotData(mortData, "Mortality")</pre>
infPlot <- plotData(infData, "Influenza")</pre>
tempPlot <- plotData(tempData, "Temperature Deficit")</pre>
mortPlot
infPlot
tempPlot
# Training GAM
model <- gam (
            Mortality ~ Year +
            s(Week, k = length(unique(data$Week))),
            data=data,
            method = "GCV.Cp"
summary(model)
modelMean <- mean(model$y)</pre>
modelVar <- var(model$y)</pre>
print("Train predictions mean:")
modelMean
print("Train predictions variance:")
modelVar
plot(model, main="Smooth Approximation of Mortality")
# Predictions
predictions <- predict(model, data)</pre>
ggplot(data) +
    geom_point(aes(x=Time, y=Mortality, color="Real")) +
    geom_line(aes(x=Time, y=predictions, color="Predicted"), size=1) +
    ggtitle("Real and Predicted Mortality")
# Plot GAM predictions per year
plotAll <- function(d, p, title) {</pre>
    shortWeeks <- 1:length(d$'2003')</pre>
    a <- ggplot() +
        geom_point(aes(x=weeks, y=d$'1995')) +
        geom_point(aes(x=weeks, y=d$'1996')) +
        geom_point(aes(x=weeks, y=d$'1997')) +
        geom_point(aes(x=weeks, y=d$'1998')) +
        geom_point(aes(x=weeks, y=d$'1999')) +
        geom_point(aes(x=weeks, y=d$'2000')) +
        geom_point(aes(x=weeks, y=d$'2001')) +
        geom_point(aes(x=weeks, y=d$'2002')) +
        geom_point(aes(x=shortWeeks, y=d$'2003')) +
        geom_line(aes(x=weeks, y=p$'1995', color="1995")) +
```

```
geom_line(aes(x=weeks, y=p$'1996', color="1996")) +
        geom_line(aes(x=weeks, y=p$'1997', color="1997")) +
        geom_line(aes(x=weeks, y=p$'1998', color="1998")) +
        geom_line(aes(x=weeks, y=p$'1999', color="1999")) +
        geom_line(aes(x=weeks, y=p$'2000', color="2000")) +
        geom_line(aes(x=weeks, y=p$'2001', color="2001")) +
        geom_line(aes(x=weeks, y=p$'2002', color="2002")) +
        geom line(aes(x=shortWeeks, y=p$'2003', color="2003")) +
        ggtitle(title)
    return(a)
}
preds <- list()</pre>
for (i in 1:length(years)) {
    year <- years[i]</pre>
    d <- data[which(data$Year == year),]</pre>
    preds[[i]] <- predict(model, d)</pre>
}
names(preds) <- years</pre>
p <- plotAll(mortData, preds, "Mortality Overall Real and Predictions")</pre>
p <- p + ylab("Mortality")</pre>
# significant terms.
orderedCoefficients <- model$coefficients[order(model$coefficients, decreasing = TRUE)]
print("The 10 most important coefficients:")
orderedCoefficients[1:10]
# Penalty factor analysis
penalties <- c(1e-6, 1e-3, 1, 1e3, 1e6)
predictions <- list()</pre>
estDegFreedom <- vector(length = length(penalties))</pre>
for (i in 1:length(penalties)) {
    model <- gam ( Mortality ~ Year +</pre>
                     s(Week, k = length(unique(data$Week))),
                     data=data,
                     method = "GCV.Cp",
                     sp = penalties[i]
    predictions[[i]] <- predict(model, data)</pre>
    estDegFreedom[i] <- summary(model)$edf</pre>
df <- data.frame(</pre>
   time = data$Time,
    real = data$Mortality,
    sp_1u = predictions[[1]],
    sp_1m = predictions[[2]],
    sp_1 = predictions[[3]],
    sp_1k = predictions[[4]],
    sp_1M = predictions[[5]]
)
```

```
ggplot(df) +
    geom_point(aes(x=time, y=real), color="black") +
    geom\_line(aes(x=time, y=sp\_1u, color="sp = 0.000001"), size=1) +
    geom\_line(aes(x=time, y=sp_1m, color="sp = 0.001"), size=1) +
    geom_line(aes(x=time, y=sp_1 , color="sp = 1" ), size=1) +
    geom_line(aes(x=time, y=sp_1k, color="sp = 1000"), size=1) +
    geom_line(aes(x=time, y=sp_1M, color="sp = 1000000"), size=1) +
   ggtitle("Effects of variations on SP penalization on Spline training") +
   xlab("Time") + ylab("Mortality")
# Relation between penalty factors and degrees of freedom
ggplot() +
   geom_point(aes(x=log10(penalties), y=estDegFreedom)) +
    geom_line(aes(x=log10(penalties), y=estDegFreedom)) +
   ggtitle("Estimated Degrees of Freedom vs SP penalty") +
   ylab("Estimated Degrees of Freedom")
# Comparing model residuals against influenza cases
model <- gam (
                Mortality ~ Year +
                s(Week, k = length(unique(data$Week))),
                data=data,
                method = "GCV.Cp"
ggplot(data) +
    geom_point(aes(x=Time, y=Influenza, color="Influenza")) +
    geom_line(aes(x=Time, y=model$residuals, color="Residuals")) +
    ggtitle("")
# Correlation measurement
corr <- cor(model$residuals, data$Influenza)</pre>
print(corr)
# GAM 2 training
model <- gam( formula = Mortality ~ s(Year, k = length(unique(data$Year))) +</pre>
                                     s(Week, k = length(unique(data$Week))) +
                                s(Influenza, k=length(unique(data$Influenza))),
              data = data,
              method = "GCV.Cp"
            )
plot(model)
# GAM 2 predictions
predictions <- predict(model, data)</pre>
ggplot() +
   geom_point(aes(x=data$Time, y=data$Mortality, color="Real")) +
    geom_line(aes(x=data$Time, y=predictions, color="Predictions")) +
   ggtitle("Second GAM predictions") + ylab("Mortality")
```

Appendix B: Code for Assignment 2

```
##
                               Assignment 2
data<-read.csv2("data/data.csv",check.names = FALSE)</pre>
names(data)<-iconv(names(data),to="ASCII")</pre>
RNGversion("3.5.1")
n=dim(data)[1]
set.seed(12345)
id=sample(1:n, floor(n*0.7))
train=data[id,]
test=data[-id,]
x<-t(train[,-4703])
y<-train[[4703]]
x_test<-t(test[,-4703])
y test<-test[[4703]]
my data<-list(x=x,y=as.factor(y),geneid=as.character(1:nrow(x)),genenames=rownames(x))
my_data_test<-list(x=x_test,y=as.factor(y_test),geneid=as.character(1:nrow(x_test)),genenames=rownames(x_test)
mod<-pamr.train(my_data,threshold = seq(0,4,0.1))</pre>
cvmodel<-pamr.cv(mod,my data)</pre>
thr<-cvmodel$threshold[which.min(cvmodel$error)]</pre>
pamr.plotcv(cvmodel)
pred<-pamr.predict(mod,my_data_test$x,threshold = thr,type="class")</pre>
pamr.plotcen(mod,my_data,thr)
#res<-as.data.frame(pamr.listgenes(mod,my_data,thr,genenames = TRUE))</pre>
#listgene
#my_data$genenames[as.numeric(listgene)]
library(glmnet)
x < -train[, -4703]
y<-train[[4703]]
x_test<-test[,-4703]
y test<-test[[4703]]</pre>
mod<-cv.glmnet(as.matrix(x),y,alpha=0.5,family="binomial")</pre>
plot(mod)
penalty_min<-mod$lambda.min</pre>
real_mod<-glmnet(as.matrix(x),y,alpha=0.5,lambda = penalty_min,family="binomial")
pred<-predict(real_mod,as.matrix(x_test),type="class")</pre>
cft<-table(pred,y_test)</pre>
mis_rate < -1-(cft[1,1]+cft[2,2])/sum(cft)
```

```
cat(mis_rate*100,"%")
fit<-ksvm(as.matrix(x),y,data=train,kernel="vanilladot",type="C-svc",scale=FALSE)</pre>
pred<-predict(fit,x_test,type="response")</pre>
cft<-table(pred,y_test)</pre>
mis_rate < -1 - (cft[1,1] + cft[2,2]) / sum(cft)
cat(mis rate*100,"%")
x=as.matrix(data[,-4703])
y=as.factor(data[[4703]])
df<-data.frame(name=c(),pvalue=c())</pre>
for(i in 1:ncol(x)){
  tmpv<-t.test(x[,i]~y,alternative="two.sided",conf.level=0.95)$p.value</pre>
  tdf<-data.frame(name=colnames(x)[i],pvalue=tmpv)</pre>
  df<-rbind(df,tdf)</pre>
df<-df[order(df$pvalue),]</pre>
a=0.05
max_i=1
for(i in 1:length(df$pvalue)){
  if(df$pvalue[i] <= a*i/length(df$pvalue)){</pre>
    max i=i
  }
}
ggplot()+geom_point(aes(x=c(1:length(df$pvalue)),y=df$pvalue),col="red")+geom_vline(xintercept = 39)
print(max_i)
df[1:39,]
```

Appendix C : Environment setup Code

```
knitr::opts_chunk$set(echo = FALSE)
library(openxlsx)
library(ggplot2)
library(mgcv)
library(pamr)
library(glmnet)
library(kernlab)
library(readr)
RNGversion('3.5.1')
set.seed(12345)
```