Metadata template[[1]](#footnote-1) for datasets of *L&O-Letters* articles

**Instructions:**

Metadata provides enough structured information for other scientists to understand and use your data. To prepare your metadata, you will need to fill in the information in the tables below and take the followings steps:

1. Fill in the tables below for your dataset that you will be making available. If you have more than one dataset, then fill in information requested for Table 2 (the data dictionary) for each dataset.
2. Save this file in this RTF format and upload your metadata to the *L&O-Letters* website when you submit your manuscript.
3. Timing of depositing your data in a repository: You should submit your data to a repository at the time of submission, however, you do not need to provide the link to the data until the manuscript has received a decision of major or minor revision. During the review process, we will review your metadata. In some cases, reviewers may ask for the data during the review stage, at which point you need to make it available.

[PLEASE DELETE THESE INSTRUCTIONS ONCE YOU FILL THIS FORM IN]

**Table 1.** Description of the fields needed to describe the creation of your dataset.

|  |  |
| --- | --- |
| **Title of dataset** | Phytoplankton pigment composition and pigment-specific growth and microzooplankton grazing rates subantarctic waters of the Campbell Plateau region SE of New Zealand |
| **URL of dataset** | The metadata tables are being uploaded to PANGAEA repository archive data sets while this process is finalized, the metadata including HPLC pigment concentrations from CTD profiles, serial dilution experiments and apparent growth rates measured in each incubation bottle can be found at the following github address:  https://github.com/agutierrez2001/CampbellPlateau |
| **Abstract** | The main objective of this study is to determine whether phytoplankton dynamics in subantarctic waters are consistent with the current understanding of iron-limited HNLC w=systems globally. To do so we assessed the phytoplankton taxon-specific growth and microzooplankton grazing rates during austral summer-autumn transition in subantarctic waters on and off Campbell Plateau characterized by contrasting HNLC conditions and productivity.  The data generated include High-pressure liquid chromatography (HPLC) pigment concentration data and flow-cytometry picophytoplankton abundance and fluorescence data from a) depth-resolved CTD profiles at ‘Biomass stations’, b) serial dilution experiments conducted with surface mixed-layer water in ‘Experimental stations’, c) apparent growth rates estimated in each incubation bottle of dilution experiments, and d) photoacclimation correction index (Phi) assessed from changes in cell fluorescence measured by flow-cytometry during 24-h dilution experiments. |
| **Keywords** | Phytoplankton growth rate, microzooplankton grazing rate, taxon-specific, HPLC pigment, subantarctic, HNLC regions, prasinophytes, diatoms, haptophytes, Ecumenical hypothesis |
| **Lead author for the dataset** | Andrés Gutiérrez Rodríguez |
| **Title and position of lead author** | Staff researcher, principal investigator. |
| **Organization and address of lead author** | National Institute of Water and Atmospheric research (NIWA) 301 Evans Bay Parade, Hataitai, Wellington, 6021, New Zealand |
| **Email address of lead author** | andres.gutierrez@ieo.csic.es |
| **Additional authors or contributors to the dataset** | Mikel Latasa, Karl Safi, Scott D Nodder |
| **Organization associated with the data** | National Institute of Water and Atmospheric research (NIWA) |
| **Funding** | Scott D Nodder - New Zealand Ministry of Business, Innovation and Employment that funded this research through the Strategic Science Investment Funding to the National Coasts Oceans Centre |
| **License** | [CCO](https://creativecommons.org/publicdomain/zero/1.0/) |
| **Geographic location – verbal description** | Campbell Plateau region southeast of New Zealand (SW Pacific). |
| **Geographic coverage bounding coordinates** | Campbell Plateau and the Subantarctic Basin - Latitude -49,20 S to -54,25 S; Longitude 166,76 E to 176,82 E |
| **Time frame - Begin date** | 19/03/2017 |
| **Time frame - End date** | 29/03/2017 |
| **General study design** | In this field study we present results of observations and experiments carried out along two open-ocean transects in Subantarctic waters across the Campbell Plateau, southeast of New Zealand during the Campbell Plateau physical oceanography voyage (TAN1702) in austral autumn of 2017. CTD casts were conducted at 56 stations distributed along two north-south transects crossing the Campbell Plateau (<500 m isobath) and the Subantarctic Basin (>1000 m) to the south (<https://doi.pangaea.de/10.1594/PANGAEA.904458>).  In the present study we distinguish between a subset of ‘Experimental’ stations (n = 8), where serial dilution experiments coupled to high-performance liquid chromatography (HPLC) and flow-cytometry (FCM) analysis were carried out with surface mixed-layer water; and a subset of additional ‘Biomass’ stations (n = 7), where depth-resolved physico-chemical and biological measurements (e.g., nutrients, size-fractionated chlorophyll a (SF-Chla), optical microscopy, FCM and HPLC) were also undertaken throughout the euphotic zone. ‘Experimental’ stations were used to characterize phytoplankton group-specific growth and grazing rates and associated carbon production and microzooplankton consumption dynamics in surface-mixed layer waters (10 m) On-plateau (n = 5) and Off-plateau (n = 3) with ‘Biomass’ stations providing a broader context of the environmental conditions and phytoplankton biomass and taxonomic composition across the region. |
| **Methods description** | Water column temperature, salinity, and dissolved oxygen were measured using a Seabird Electronics (SBE) 911plus CTD and a 24 x 10‐L SBE32 rosette water sampler. The CTD sensor was configured with SBE 3plus, SBE 4, and SBE 43 dual sensors for the parameters above, a Seapoint fluorescence sensor, and a photosynthetically active radiation (PAR) sensor (Biospherical Instruments QCP‐2300L‐HP).  Serial dilution experiments were prepared and incubated as described in Gutiérrez-Rodríguez et al. (2020) (<https://doi.org/10.1029/2019JC015550>). Bottles were incubated for 24-hr under simulated surface mixed-layer temperature and irradiance levels. Initial and final samples were taken for HPLC and FCM analysis following the procedure described above. Initial samples were taken in duplicate and triplicate, respectively. |
| **Laboratory, field, or other analytical methods** | **HPLC analysis** - For HPLC, 1.7-2-L seawater samples were vacuum-filtered onto 25 mm GFF filters. Filters were dry-blotted, flash-frozen and kept at -80 ◦C until HPLC analysis in the lab following details in Latasa et al. (2022). Pigments were extracted in 90% acetone, and injected onto an Agilent (Waldbronn, Germany) HPLC 1200 system. Absorption signals were recorded at 474 nm and 664 nm for carotenoids and chlorophylls, respectively. The separation method was based on Zapata et al. (2000) with minor modifications.  **FCM analysis**- For FCM, duplicated 1.5 mL seawater samples were preserved with a solution containing glutaraldehyde (25% sigma) and pluronic acid (10% Sigma) at a 9:1 proportion (Marie et al. 2014), flash-frozen in liquid nitrogen and stored at -80ÅãC. Samples were analyzed with a BD FACSCalibur instrument and Synechococcus (SYN) and picoeukaryotes (PEUK) cells quantified using TrucountTM beads (Becton Dickinson, Mountain View, CA) as described in Hall and Safi (2001).  **Apparent growth rates (k, day-1)** - Phytoplankton apparent growth rates k in each incubation bottle were estimated from changes between the initial (N0) and final (Nt) pigment concentration and cell abundance assuming exponential growth and loss processes during the incubation time (t), following the equation: k (day−1) = (1/t)x ln(Nt/N0) x D where D is the dilution factor. Nutrient-amended intrinsic growth rate (μnut ) was estimated as the y intercept of the linear regression between k and D in the dilution series (n = 5). Microzooplankton grazing rate (m, day−1) was estimated as the difference between μnut and the mean apparent growth rate (μnutnet) in non-diluted, nutrient-amended bottles (m = μnut −μnutnet). Intrinsic growth rate (μ, day−1) was then calculated as the sum between the mean apparent growth rate in non-diluted, unamended bottles (μnet) and grazing) (μ = m + μnet). Regional averages of the m : μ ratio were calculated first using the arctangent transformed values of the individual experiments and then using the inverse tangent function to transform back the average values (Calbet and Landry 2004).  **Photoacclimation correction factor (Phi)** **and corrected apparent growth rates (k)** - The photoacclimation factor (Phi) was estimated using cell red fluorescence (FL3) and side scatter (SSC) from FCM initial and final measurements to account for photoacclimation-related changes in cell pigment content during incubations. The photoacclimation factor (Phi = FL3/SSCfinal / FL3/SSCinitial) was calculated for PEUK and used to correct the pigment-based apparent growth rates estimated in each bottle following the procedure described in Gutierrez-Rodriguez et al. (2020). Estimated Phi values and corrected apparent growth rates for each group/pigment can be found in https://github.com/agutierrez2001/CampbellPlateau. These corrected k values were then used to calculate instantaneous growth and grazing coefficients from linear regressions and for each group. |
| **Taxonomic species or groups** | Diatoms, Prasinophytes, Haptophytes, Pelagophytes, Dinoflagellates, Cryptophytes, Synechococcus, Picoeucaryotes |
| **Quality control** | Photoacclimation-related changes in cell pigment content were corrected using a FCM assessed photoacclimation correction factor. |
| **Additional information** |  |
|  |  |

**Table 2.** Data dictionary: description of the variables (i.e., columns) in EACH dataset. You must provide sufficient detail for another user to understand and use the data. If there are 10 variables (i.e., columns) in the dataset, then there should be 10 rows in this table that describe each column. Be sure to include all relevant information for your dataset, including the unique identifiers for your dataset or system, dates, replicate numbers, latitude and longitude of sampling locations, etc.

Dataset filename: *GitHub\_CTD\_HPLC\_FCM\_metadata.xlsx*

Dataset description:*Phytoplankton HPLC pigment concentrations and FCM picophytoplankton cell abundance from CTD profiles and surface-mixed layer in ‘Biomass’ and ‘Experimental’ stations, respectively.*

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Column name** | **Description** | **Units** | **Code explanation** | **Data format** | **Missing data code** |
| *The name of the variable in the dataset; avoid special characters, dashes and spaces* | *A detailed description of the variable* | *Units the variable is measured in* | *If you use codes in your column, please explain each code, such as: LR = Little Rock Lake; A=sample; etc.* | *State exactly how the data are stored; for dates, state how it is formatted, including time zone, etc.* | *If data are missing, indicate how they are stored, such as NULL, NA, blank cell, etc.* |
| Station | Station number | na |  |  |  |
| Cast | CTD Cast number | na |  |  |  |
| Latitude | Latitude South | Decimal degrees |  |  |  |
| Longitude | Longitude East | Decimal degrees |  |  |  |
| Date | Date of sample collection | New Zealand Standard Time |  | dd/mm/yyyy |  |
| Region | On vs Off Campbell Plateau | na |  |  |  |
| SeafloorDepth | Depth of the seafloor in meters | m |  |  |  |
| Per | Peridinin | ng/L |  |  | nd |
| 19But | 19'butanoyloxifucoxanthin | ng/L |  |  | nd |
| Fuco | Fucoxanthin | ng/L |  |  | nd |
| Neox | Neoxanthin | ng/L |  |  | nd |
| Pras | Prasinoxanthin | ng/L |  |  | nd |
| Viol | Violaxanthin | ng/L |  |  | nd |
| 19Hex | 19'hexanoyloxifucoxanthin | ng/L |  |  | nd |
| Ddx | Diadinoxanthin | ng/L |  |  | nd |
| Allo | Alloxanthin | ng/L |  |  | nd |
| Zea | Zeaxanthin | ng/L |  |  | nd |
| Lut | Lutein | ng/L |  |  | nd |
| Chlc2\_MGDG18\_14 | Chlorophyll c2 monogalactosyldiacylglyceride ester [18:14/14:0] | ng/L |  |  | nd |
| Chlc2\_MGDG14\_14 | Chlorophyll c2 monogalactosyldiacylglyceride ester [14:0/14:0] | ng/L |  |  | nd |
| Chlb | Chlorophyll b | ng/L |  |  | nd |
| DVChla | Divinyl chlorophyll a | ng/L |  |  | nd |
| MVChla | Monovinyl chlorophyll a | ng/L |  |  | nd |
| TChla | Sum of MVChla+DVChla | ng/L |  |  | nd |
| SYN | Synechococcus FCM cell abundance | Cell/mL |  |  | nd |
| PEUK | Picoeukaryotes FCM cells abundance | Cell/mL |  |  | nd |

Dataset filename: *GitHubTable\_DILEX\_HPLC\_FCM\_metadata.xlsx*

Dataset description: *HPLC phytoplankton pigment concentrations and FCM picophytoplankton cell abundance measured in each initials and finals (from each incubation bottle) of serial dilution experiments.*

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Column name** | **Description** | **Units** | **Code explanation** | **Data format** | **Missing data code** |
| *The name of the variable in the dataset; avoid special characters, dashes and spaces* | *A detailed description of the variable* | *Units the variable is measured in* | *If you use codes in your column, please explain each code, such as: LR = Little Rock Lake; A=sample; etc.* | *State exactly how the data are stored; for dates, state how it is formatted, including time zone, etc.* | *If data are missing, indicate how they are stored, such as NULL, NA, blank cell, etc.* |
| Sample\_ID | Sample identification |  |  |  |  |
| Station | Station number |  |  |  |  |
| Cast | CTD Cast number |  |  |  |  |
| Date | Date of sample collection | Decimal degrees |  |  |  |
| Latitude\_South | Latitude South | Decimal degrees |  |  |  |
| Latitude\_East | Longitude East | New Zealand Standard Time |  | dd/mm/yyyy |  |
| Depth | Depth of sample collection | m |  |  |  |
| IncubationTime | Initials or finals (24h) samples |  |  |  |  |
| NutrientTreatment | Nutrient amended or unamended experimental bottle |  |  |  |  |
| ExpBottleNumber | Experimental incubation bottle labelling (1-11) |  |  |  |  |
| DilutionFactor | Fraction of whole seawater in each incubation bottle | proportion |  |  |  |
| Volume\_filtered | Volume of seawater filtered for HPLC analysis | L |  |  |  |
| Notes | Notes from lab notebook indicating possible problems during sampling/filtering |  |  |  |  |
| QC | Quality control based on notebook annotations | '‘0’ retain ‘1’ removed from rate calculation |  |  |  |
| Per | Peridinin | ng/L |  |  | nd |
| But | 19'butanoyloxifucoxanthin | ng/L |  |  | nd |
| Fuco | Fucoxanthin | ng/L |  |  | nd |
| Neox | Neoxanthin | ng/L |  |  | nd |
| Pras | Prasinoxanthin | ng/L |  |  | nd |
| Viol | Violaxanthin | ng/L |  |  | nd |
| Hex | 19'hexanoyloxifucoxanthin | ng/L |  |  | nd |
| Ddx | Diadinoxanthin | ng/L |  |  | nd |
| Allo | Alloxanthin | ng/L |  |  | nd |
| Zea | Zeaxanthin | ng/L |  |  | nd |
| Chlc2\_MGDG18\_14 | Chlorophyll c2 monogalactosyldiacylglyceride ester [18:14/14:0] | ng/L |  |  | nd |
| Chlc2\_MGDG14\_14 | Chlorophyll c2 monogalactosyldiacylglyceride ester [14:0/14:0] | ng/L |  |  | nd |
| Chlb | Chlorophyll b | ng/L |  |  | nd |
| DVChla | Divinyl chlorophyll a | ng/L |  |  | nd |
| MVChla | Monovinyl chlorophyll a | ng/L |  |  | nd |
| TChla | Sum of MVChla+DVChla | ng/L |  |  | nd |
| SYN | Synechococcus FCM cell abundance | Cell/mL |  |  | nd |
| PEUK | Picoeukaryotes FCM cells abundance | Cell/mL |  |  | nd |

Dataset filename: *GitHubTable.APPARENT\_GROWTHRATES\_metadata.xlsx*

Dataset description: *Contains taxon-specific apparent growth rates estimated for each incubation bottle in each serial dilution experiment estimated from changes in cell abundance or pigment concentration between initial and final samples. The dataset also contains FCM cell fluorescence (FL3) and size (SSC) data and derived photoacclimation correction factor (Phi) used to correct pigment based pigment-based apparent growth rates for photoacclimation-related cell pigment content during the incubation.*

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Column name** | **Description** | **Units** | **Code explanation** | **Data format** | **Missing data code** |
| *The name of the variable in the dataset; avoid special characters, dashes and spaces* | *A detailed description of the variable* | *Units the variable is measured in* | *If you use codes in your column, please explain each code, such as: LR = Little Rock Lake; A=sample; etc.* | *State exactly how the data are stored; for dates, state how it is formatted, including time zone, etc.* | *If data are missing, indicate how they are stored, such as NULL, NA, blank cell, etc.* |
| Station | Station number |  |  |  |  |
| Cast | CTD Cast number |  |  |  |  |
| IncubationTime | Initials or finals (24h) samples |  |  |  |  |
| ExpBottleNumber | Experimental incubation  bottle labelling (1-11) |  |  |  |  |
| DilutionFactor | Fraction of whole seawater  in each incubation bottle |  |  |  |  |
| SYN\_abundance | Synechococcus cell  abundance from FCM counts |  |  |  |  |
| SYN\_k | Synechococcus apparent  growth rates estimated in each incubation bottle |  |  |  |  |
| PEUK\_abundance | Picoeukaryotes cell abundance from FCM counts |  |  |  |  |
| PEUK\_k | Picoeukaryotes apparent growth rates  estimated in each incubation bottle |  |  |  |  |
| FL3\_SSC\_PEUK | Red fluorescence (FL3) to side scatter ratio (SSC)  for PEUK in each incubation bottle |  |  |  |  |
| Phi\_FL3\_PEUK | Photoacclimation index estimated from changes in FL3/SSC of PEUK in each incubation bottle between start and end of experiment - Phi-FL3= ln((FL3/SSC\_t24)/(FL3/SSC\_T0)) |  |  |  |  |
| Phi\_nut\_PEUK | Photoacclimation index estimated from changes in FL3/SSC of PEUK in non-diluted (wsw) nutrient-amended (nut) bottles between start and end of experiment - Phi nut= ln(mean((FL3/SSC\_t24\_wsw\_nut)/(FL3/SSC\_T0\_wsw\_nut))) |  |  |  |  |
| Phi\_nonut\_PEUK | Photoacclimation index estimated from changes in FL3/SSC of PEUK in non-diluted (wsw) nutrient-amended (nonut) bottles between start and end of experiment - Phi nonut= ln(mean((FL3/SSC\_t24\_wsw\_nonut)/(FL3/SSC\_T0\_wsw\_nonut))) |  |  |  |  |
| Notes | Notes from lab notebook indicating problems during sampling/filtering |  |  |  |  |
| TChla\_k | TChla Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the Phi-FL3 estimated for each incubation bottle. |  |  |  |  |
| TChla\_k\_Phi\_ind\_corr | TChla Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the Phi-FL3 estimated for each incubation bottle. |  |  |  |  |
| TChla\_k\_Phi\_mean\_corr | TChla Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the mean Phi-nut and Phi-nonut correction factors for nutrient-amended (bottles 1-8) and unamended (9-11) bottles. |  |  |  |  |
| Fuco\_k | Fucoxanthin Apparent growth rates estimated from changes in cell abundance or pigment concentrations without photoacclimation correction |  |  |  |  |
| Fuco\_k\_Phi\_ind\_corr | Fucoxanthin Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the Phi-FL3 estimated for each incubation bottle. |  |  |  |  |
| Fuco\_k\_Phi\_mean\_corr | Fucoxanthin Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the mean Phi-nut and Phi-nonut correction factors for nutrient-amended (bottles 1-8) and unamended (9-11) bottles. |  |  |  |  |
| Pras\_k | Prasinoxanthin Apparent growth rates estimated from changes in cell abundance or pigment concentrations without photoacclimation correction |  |  |  |  |
| Pras\_k\_Phi\_ind\_corr | Prasinoxanthin Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the Phi-FL3 estimated for each incubation bottle. |  |  |  |  |
| Pras\_k\_Phi\_mean\_corr | Prasinoxanthin Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the mean Phi-nut and Phi-nonut correction factors for nutrient-amended (bottles 1-8) and unamended (9-11) bottles. |  |  |  |  |
| Chlb\_k | Chlorophyll b Apparent growth rates estimated from changes in cell abundance or pigment concentrations without photoacclimation correction |  |  |  |  |
| Chlb\_k\_Phi\_ind\_corr | Chlorophyll b Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the Phi-FL3 estimated for each incubation bottle. |  |  |  |  |
| Chlb\_k\_Phi\_mean\_corr | Chlorophyll b Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the mean Phi-nut and Phi-nonut correction factors for nutrient-amended (bottles 1-8) and unamended (9-11) bottles. |  |  |  |  |
| 19But\_k | 19'butanoyloxifucoxanhtin Apparent growth rates estimated from changes in cell abundance or pigment concentrations without photoacclimation correction |  |  |  |  |
| 19But\_k\_Phi\_ind\_corr | 19'butanoyloxifucoxanhtin Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the Phi-FL3 estimated for each incubation bottle. |  |  |  |  |
| 19But\_k\_Phi\_mean\_corr | 19'butanoyloxifucoxanhtin Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the mean Phi-nut and Phi-nonut correction factors for nutrient-amended (bottles 1-8) and unamended (9-11) bottles. |  |  |  |  |
| Per\_k | Peridinin Apparent growth rates estimated from changes in cell abundance or pigment concentrations without photoacclimation correction |  |  |  |  |
| Per\_k\_Phi\_ind\_corr | Peridinin Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the Phi-FL3 estimated for each incubation bottle. |  |  |  |  |
| Per\_k\_Phi\_mean\_corr | Peridinin Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the mean Phi-nut and Phi-nonut correction factors for nutrient-amended (bottles 1-8) and unamended (9-11) bottles. |  |  |  |  |
| Allox\_k | Alloxanthin Apparent growth rates estimated from changes in cell abundance or pigment concentrations without photoacclimation correction |  |  |  |  |
| Allox\_k\_Phi\_ind\_corr | Alloxanthin Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the Phi-FL3 estimated for each incubation bottle. |  |  |  |  |
| Allox\_k\_Phi\_mean\_corr | Alloxanthin Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the mean Phi-nut and Phi-nonut correction factors for nutrient-amended (bottles 1-8) and unamended (9-11) bottles. |  |  |  |  |
| 19Hex\_k | 19'hexanoyloxifucoxanhtin Apparent growth rates estimated from changes in cell abundance or pigment concentrations without photoacclimation correction |  |  |  |  |
| 19Hex\_k\_Phi\_ind\_corr | 19'hexanoyloxifucoxanhtin Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the Phi-FL3 estimated for each incubation bottle. |  |  |  |  |
| 19Hex\_k\_Phi\_mean\_corr | 19'hexanoyloxifucoxanhtin Apparent growth rates estimated from changes in cell abundance or pigment concentrations and corrected for photoacclimation using the mean Phi-nut and Phi-nonut correction factors for nutrient-amended (bottles 1-8) and unamended (9-11) bottles. |  |  |  |  |
|  |  |  |  |  |  |
|  |  |  |  |  |  |
|  |  |  |  |  |  |

**Table 3. Data provenance**

If you used data derived from other sources, provide the information here so future users know where the data came from.

|  |  |  |  |
| --- | --- | --- | --- |
| **Dataset title** | **Dataset DOI or URL** | **Creator (name & email)** | **Contact (name & email)** |
|  |  |  |  |
|  |  |  |  |

**Scripts/code (software) –** *OPTIONAL*

It is recommended that you also provide your scripts along with your data, although it is not required at this time in our journal.

|  |  |  |
| --- | --- | --- |
| **File name** | **Description** | **Scripting language** |
|  |  |  |
|  |  |  |

**Notes and Comments:**

1. *This document liberally borrows from a similar document provided by the Environmental Data Initiative* [↑](#footnote-ref-1)