# bcb546x\_r\_assignments

Zihao Zheng 10/9/2018

#### Part I

#### 1. Data inspection

```
setwd("~/Documents/BCB546X-Fall2018/assignments/UNIX_Assignment/")
library(tidyverse)
library(reshape2)
library(ggrepel)
Check the size of files to be loaded
file.size("fang_et_al_genotypes.txt") %>% utils:::format.object_size("auto")
## [1] "10.5 Mb"
file.size("snp_position.txt") %>% utils:::format.object_size("auto")
## [1] "80.8 Kb"
load files to R
fang_et_al <- read_delim("fang_et_al_genotypes.txt",delim = "\t",col_names = T)</pre>
snp_position <- read_delim("snp_position.txt",delim = "\t",col_names = T)</pre>
number of columns and rows for each file
# fang_et_al column#
ncol(fang_et_al)
## [1] 986
# fang et al row#
nrow(fang_et_al)
## [1] 2782
head(fang_et_al)
## # A tibble: 6 x 986
##
     Sample_ID JG_OTU
                         Group abph1.20 abph1.22 ae1.3 ae1.4 ae1.5 an1.4 ba1.6
     <chr>>
               <chr>>
                         <chr> <chr>
                                         <chr>>
                                                  <chr> <chr> <chr> <chr> <chr> <chr>
##
                                                                     C/C
## 1 SL-15
               T-aust-1 TRIPS ?/?
                                         ?/?
                                                  T/T
                                                        G/G
                                                               T/T
                                                                           ?/?
## 2 SL-16
               T-aust-2 TRIPS ?/?
                                         ?/?
                                                  T/T
                                                        ?/?
                                                               T/T
                                                                     C/C
                                                                           A/G
## 3 SL-11
               T-brav-1 TRIPS ?/?
                                         ?/?
                                                  T/T
                                                        G/G
                                                               T/T
                                                                     ?/?
                                                                           G/G
                                         ?/?
                                                  T/T
## 4 SL-12
               T-brav-2 TRIPS ?/?
                                                        G/G
                                                               T/T
                                                                     C/C
                                                                           G/G
## 5 SL-18
               T-cund
                       TRIPS ?/?
                                         ?/?
                                                  T/T
                                                        G/G
                                                               T/T
                                                                     C/C
                                                                           ?/?
## 6 SL-2
               T-dact-1 TRIPS ?/?
                                         ?/?
                                                  T/T
                                                        G/G
                                                               T/T
                                                                     C/C
                                                                           A/G
## # ... with 976 more variables: ba1.9 <chr>, bt2.5 <chr>, bt2.7 <chr>,
       bt2.8 <chr>, Fea2.1 <chr>, Fea2.5 <chr>, id1.3 <chr>, lg2.11 <chr>,
       lg2.2 <chr>, pbf1.1 <chr>, pbf1.2 <chr>, pbf1.3 <chr>, pbf1.5 <chr>,
## #
## #
       pbf1.6 <chr>, pbf1.7 <chr>, pbf1.8 <chr>, PZA00003.11 <chr>,
```

```
## #
       PZA00004.2 <chr>, PZA00005.8 <chr>, PZA00005.9 <chr>,
## #
       PZA00006.13 <chr>, PZA00006.14 <chr>, PZA00008.1 <chr>,
       PZA00010.5 <chr>, PZA00013.10 <chr>, PZA00013.11 <chr>,
## #
       PZA00013.9 <chr>, PZA00015.4 <chr>, PZA00017.1 <chr>,
## #
## #
       PZA00018.5 <chr>, PZA00029.11 <chr>, PZA00029.12 <chr>,
## #
       PZA00030.11 <chr>, PZA00031.5 <chr>, PZA00041.3 <chr>,
## #
       PZA00042.2 <chr>, PZA00042.5 <chr>, PZA00043.7 <chr>,
       PZA00045.1 <chr>, PZA00047.2 <chr>, PZA00049.12 <chr>,
## #
## #
       PZA00050.9 <chr>, PZA00051.2 <chr>, PZA00058.5 <chr>,
       PZA00058.6 <chr>, PZA00060.2 <chr>, PZA00061.1 <chr>,
## #
## #
       PZA00065.2 <chr>, PZA00069.4 <chr>, PZA00070.5 <chr>,
       PZA00078.2 <chr>, PZA00079.1 <chr>, PZA00081.17 <chr>,
## #
## #
       PZA00084.2 <chr>, PZA00084.3 <chr>, PZA00086.8 <chr>,
## #
       PZA00088.3 <chr>, PZA00090.2 <chr>, PZA00092.1 <chr>,
## #
       PZA00092.5 <chr>, PZA00093.2 <chr>, PZA00096.26 <chr>,
## #
       PZA00097.13 <chr>, PZA00098.14 <chr>, PZA00100.10 <chr>,
## #
       PZA00100.12 <chr>, PZA00100.14 <chr>, PZA00100.9 <chr>,
## #
       PZA00103.20 <chr>, PZA00106.9 <chr>, PZA00107.18 <chr>,
       PZA00108.12 <chr>, PZA00108.14 <chr>, PZA00108.15 <chr>,
## #
## #
       PZA00109.3 <chr>, PZA00109.5 <chr>, PZA00111.2 <chr>,
## #
       PZA00111.4 <chr>, PZA00111.5 <chr>, PZA00111.6 <chr>,
       PZA00111.8 <chr>, PZA00114.3 <chr>, PZA00116.2 <chr>,
## #
       PZA00119.4 <chr>, PZA00120.4 <chr>, PZA00123.1 <chr>,
## #
       PZA00125.2 <chr>, PZA00131.14 <chr>, PZA00132.17 <chr>,
## #
## #
       PZA00132.18 <chr>, PZA00132.3 <chr>, PZA00135.6 <chr>,
       PZA00137.2 <chr>, PZA00139.14 <chr>, PZA00140.10 <chr>,
## #
       PZA00140.6 <chr>, PZA00140.9 <chr>, PZA00142.6 <chr>,
       PZA00148.2 <chr>, PZA00153.3 <chr>, ...
# snp_position column#
ncol(snp_position)
## [1] 15
# snp_position row#
nrow(snp_position)
## [1] 983
head(snp_position)
## # A tibble: 6 x 15
##
     SNP_ID cdv_marker_id Chromosome Position alt_pos mult_positions amplicon
##
     <chr>>
                    <int> <chr>
                                      <chr>>
                                               <chr>>
                                                       <chr>>
                                                                       <chr>>
## 1 abph1~
                     5976 2
                                      27403404 <NA>
                                                       <NA>
                                                                       abph1
## 2 abph1~
                                      27403892 <NA>
                     5978 2
                                                       <NA>
                                                                       abph1
## 3 ae1.3
                     6605 5
                                      1678897~ <NA>
                                                       <NA>
                                                                       ae1
## 4 ae1.4
                     6606 5
                                      1678896~ <NA>
                                                       <NA>
                                                                       ae1
## 5 ae1.5
                     6607 5
                                      1678898~ <NA>
                                                       <NA>
                                                                       ae1
## 6 an1.4
                     5982 1
                                      2404985~ <NA>
                                                       <NA>
                                                                       an1
## # ... with 8 more variables: cdv_map_feature.name <chr>, gene <chr>,
       `candidate/random` <chr>, Genaissance_daa_id <int>,
       Sequenom_daa_id <int>, count_amplicons <int>, count_cmf <int>,
## #
## #
       count_gene <int>
```

#### 2. Data processing

```
split fang et al into maize group and teosinte group
```

```
maize.snp <- fang_et_al %>% filter(Group %in% c("ZMMIL","ZMMLR","ZMMMR"))
teosinte.snp <- fang_et_al %>% filter(Group %in% c("ZMPBA","ZMPIL","ZMPJA"))
```

transpose genotypic data, and merge with the snp\_position

```
maize group

maize.tmp <- data.frame(t(maize.snp)) %>% tibble::rownames_to_column()
colnames(maize.tmp) <- maize.tmp[1, ]
maize.tmp <- maize.tmp[-(1:3),]
colnames(maize.tmp)[1] <- "SNP_ID"
merged.maize.genotype <- merge(snp_position,maize.tmp,by = "SNP_ID")

# teosinte group
teosinte.tmp <- data.frame(t(teosinte.snp)) %>% tibble::rownames_to_column()
colnames(teosinte.tmp) <- teosinte.tmp[1, ]
teosinte.tmp <- teosinte.tmp[-(1:3),]
colnames(teosinte.tmp)[1] <- "SNP_ID"
merged.teosinte.genotype <- merge(snp_position,teosinte.tmp,by = "SNP_ID")</pre>
```

For maize group, generate 10 files (1 for each chromosome) with SNPs ordered based on increasing position values and with missing data encoded by this symbol: ?

For maize group, generate 10 files (1 for each chromosome) with SNPs ordered based on decreasing position values and with missing data encoded by this symbol: -

```
merged.maize.genotype[merged.maize.genotype=== "?/?"]<- "-/-"
maize_split_decr <- split(merged.maize.genotype, merged.maize.genotype$Chromosome)

for (i in 1:10) {
   maize_split_decr[[i]]<-
    maize_split_decr[[i]][order(as.numeric(maize_split_decr[[i]]$Position)),]
   write_delim(maize_split_decr[[i]],paste0(new_names[i],"_decr.txt"),delim = "\t")
}</pre>
```

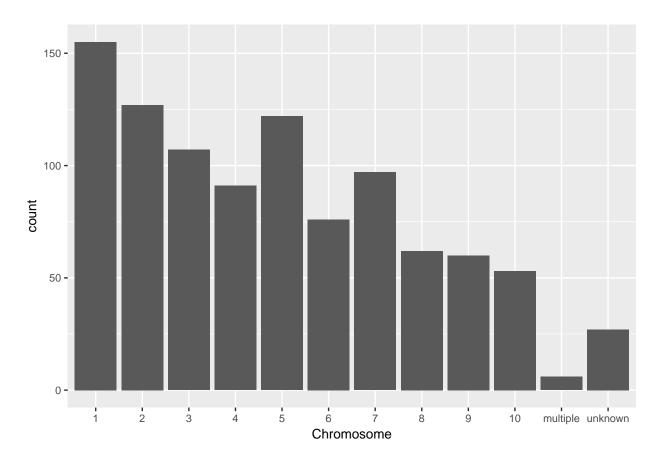
For teosinte group, generate 10 files (1 for each chromosome) with SNPs ordered based on increasing position values and with missing data encoded by this symbol: ?

## Part II

Merge fang\_et\_al and snp\_position to get a master file

```
genotype.tmp <- data.frame(t(fang_et_al)) %>% tibble::rownames_to_column()
colnames(genotype.tmp) <- genotype.tmp[1, ]
genotype.tmp <- genotype.tmp[-(1:3),]
colnames(genotype.tmp)[1] <- "SNP_ID"
merged.genotype <- merge(snp_position,genotype.tmp,by = "SNP_ID")</pre>
```

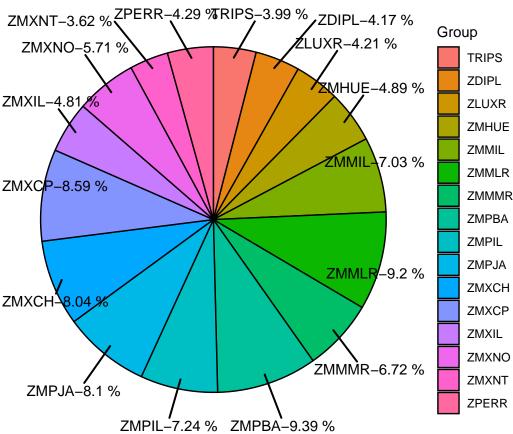
1(a). Plot the total number of SNPs in our dataset on each chromosome.



## 1(b). What groups contribute most of these SNPs?

```
fang_et_al_short <- fang_et_al %>% select(-c(Sample_ID, JG_OTU)) %>%
  melt(id.vars = "Group", variable.name = "SNP_ID") %>% unique()
snp_position.tmp <- snp_position %>% select(SNP_ID,Chromosome) %>%
  merge(fang_et_al_short,by = "SNP_ID")
snp_position.tmp[snp_position.tmp=="?/?"] <- NA</pre>
snp_position.tmp <- na.omit(snp_position.tmp)</pre>
snp_group_stat <- snp_position.tmp %>% count(Group)
snp_group_stat$Label <- paste(snp_group_stat$Group,</pre>
                        paste(round(((snp_group_stat$n/sum(snp_group_stat$n))*100),2),"%"), sep="-")
snp_group_stat$pos = (cumsum(c(0, snp_group_stat$n))
                       + c(snp_group_stat$n / 2, .01))[1:nrow(snp_group_stat)]
p2 <- ggplot(snp_group_stat, aes(1, n, fill = Group)) +</pre>
    geom_col(color = 'black',
             position = position_stack(reverse = TRUE),
             show.legend = TRUE) +
    geom_text_repel(aes(x = 1.4, y = pos, label = Label),
                    nudge_x = .3,
                    segment.size = .7,
```

```
show.legend = FALSE) +
coord_polar('y') +
theme_void()
p2
```

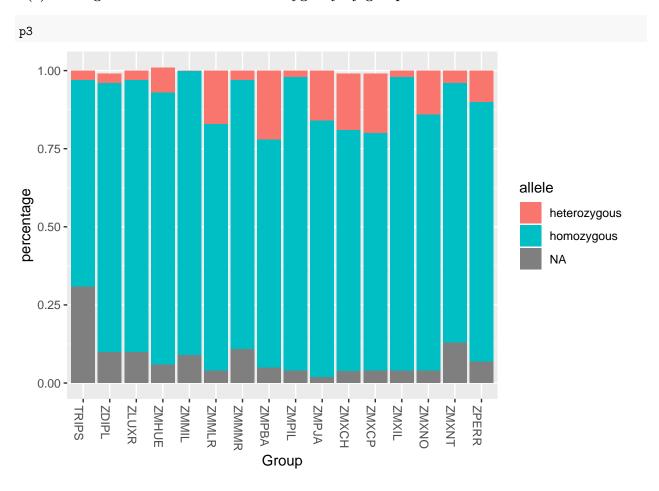


## 2. Missing data and amount of heterozygosity

```
allele_stat_sample <- fang_et_al_short2 %>% group_by(Sample_ID,allele) %>%
    summarise(n = n()) %>% mutate(countT= sum(n)) %>%
    mutate(percentage=round(n/countT,2))

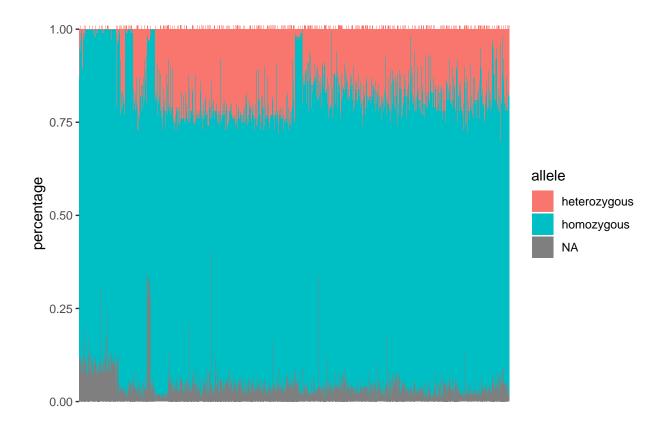
p4 <- ggplot(allele_stat_sample) +
    geom_bar(aes(y = percentage, x = Sample_ID, fill = allele), stat="identity") +
    theme(axis.title.x=element_blank(),
        axis.text.x=element_blank(),
        axis.ticks.x=element_blank())</pre>
```

## 2(a). Missing data and amount of heterozygosity by group



## 2(b). Missing data and amount of heterozygosity by individual

p4



#### 3. Visualization of Minor Allele Frequency (MAF) of maize group and teosinte group

Calculate MAF for each group

```
MAF.tmp <- fang_et_al_short2 %>% filter(allele=="homozygous") %>%
  group_by(Group,SNP_ID,value) %>%
  summarise(n = n()) %>% mutate(countT= sum(n)) %>%
  mutate(allele_freq=round(n/countT,3)) %>%
  filter(allele_freq < 0.5) %>%
  select(Group,SNP_ID,allele_freq) %>%
  group_by(Group,SNP_ID) %>% top_n(-1)
colnames(MAF.tmp)[3] <- "MAF"</pre>
MAF.tmp$species <- NA
MAF.tmp$species[MAF.tmp$Group %in% c("ZMMIL","ZMMLR","ZMMMR")] <- "maize"
MAF.tmp$species[MAF.tmp$Group %in% c("ZMPBA","ZMPIL","ZMPJA")] <- "teosinte"
MAF.tmp <- na.omit(MAF.tmp)</pre>
p5<- ggplot(MAF.tmp, aes(x=MAF, fill=Group)) +
    geom_histogram(binwidth=0.01, alpha=.5, position="identity") +
    facet_grid(species ~ .)
p5
```

