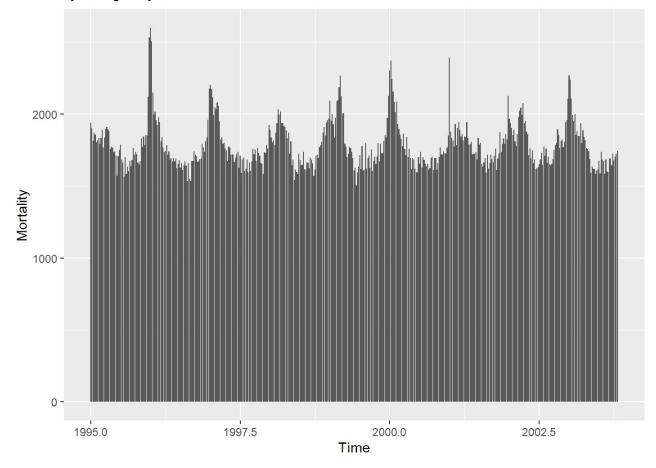
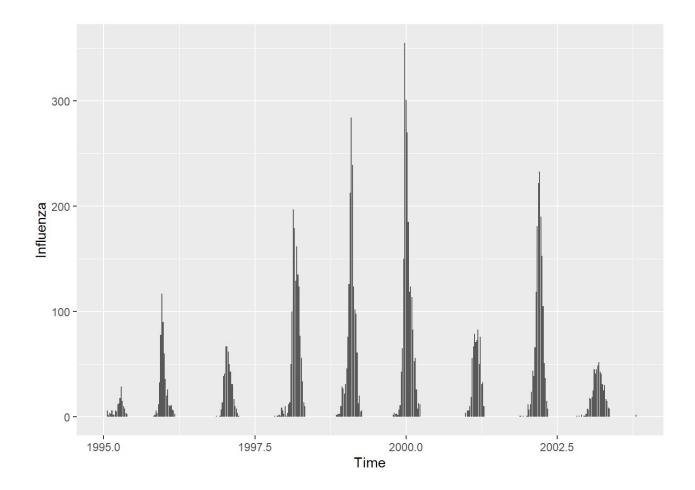
lab2-block2_group5_report

Bjorn_Hansen, Erik Anders, Ahmed Alhasan 12/16/2019

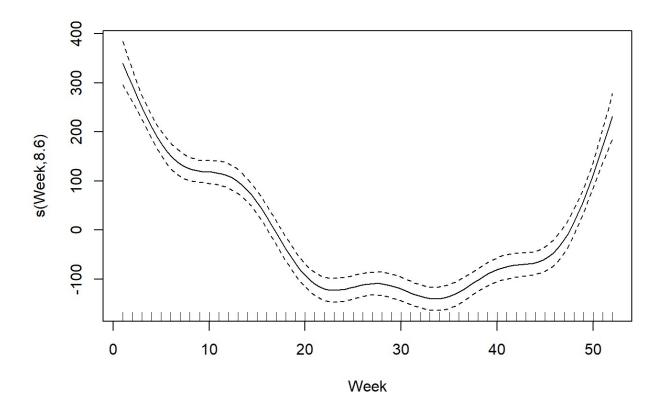
Assignment 1. Using GAM and GLM to examine the mortality rates

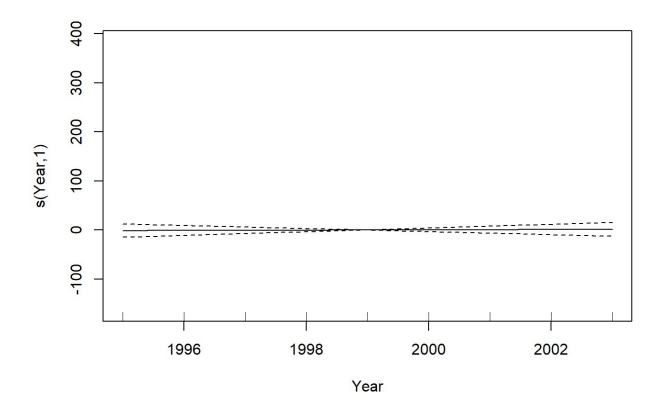
As can be seen from the plots below there does not seem to be a clear relationship between influenza and mortality by viewing the plots. There are perhaps peaks of influenza when there are relative peaks of mortality during the year of 2000.





```
## Family: gaussian
## Link function: identity
## Formula:
## Mortality ~ Year + s(Week, k = length(unique(data$Week)))
## Parametric coefficients:
              Estimate Std. Error t value Pr(>|t|)
## (Intercept) -680.598 3367.760 -0.202
                                         0.840
## Year
               1.233
                        1.685 0.732
                                           0.465
##
## Approximate significance of smooth terms:
## edf Ref.df F p-value
## s(Week) 14.32 17.87 53.86 <2e-16 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## Rank: 52/53
## R-sq.(adj) = 0.677 Deviance explained = 68.8\%
## GCV = 8708.6 Scale est. = 8398.9 n = 459
```

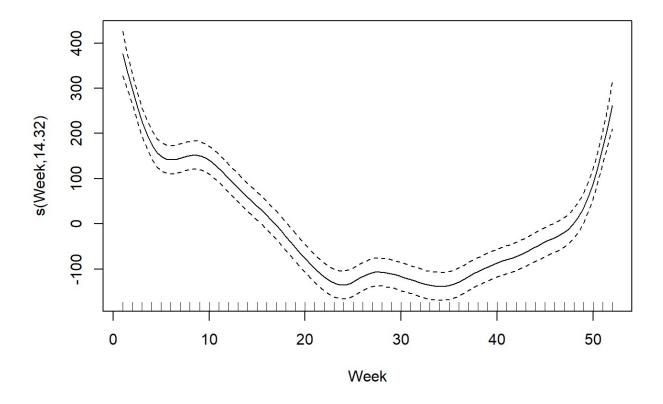


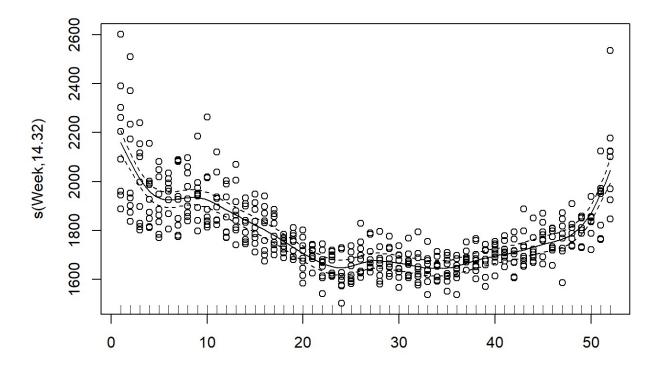


3

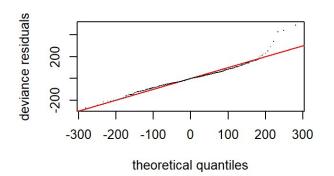
As is seen in the plots below, the histogram of the residuals seem to be normaly distributed and the model seems fit the data well. The week of the year is the most significant feature. The year is not of importance as mortality does not change much between years. It can also be seen from the previous summary of the model that the year has a large p-value.

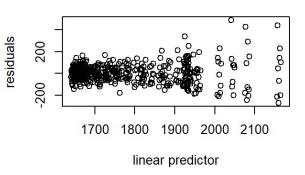
```
## s(Week)
## 0.0001131932
```





Resids vs. linear pred.

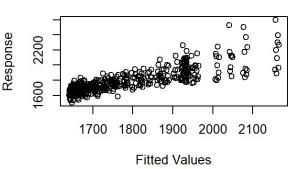




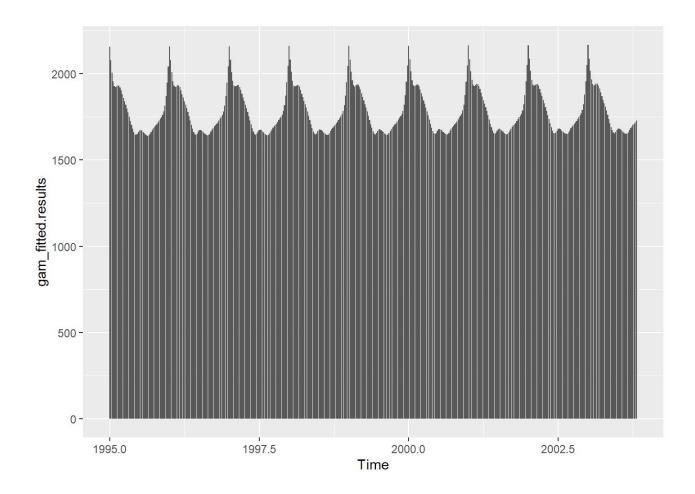
Histogram of residuals

-200 0 200 400 Residuals

Response vs. Fitted Values

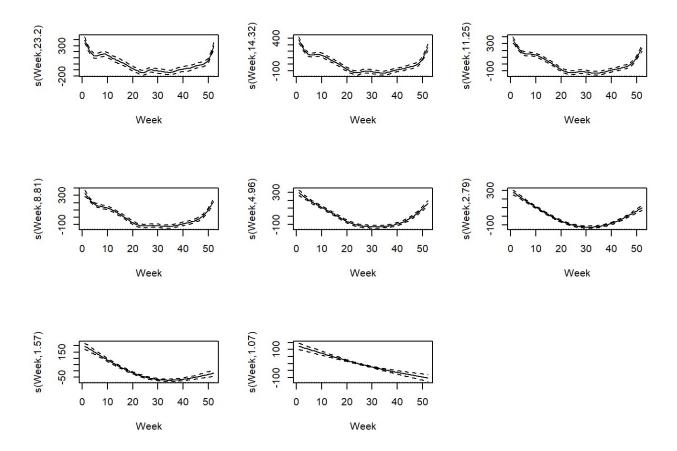


```
##
## Method: GCV Optimizer: magic
## Smoothing parameter selection converged after 9 iterations by steepest
## descent step failure.
## The RMS GCV score gradient at convergence was 0.00106719 .
## The Hessian was positive definite.
## Model rank = 52 / 53
##
## Basis dimension (k) checking results. Low p-value (k-index<1) may
## indicate that k is too low, especially if edf is close to k'.
##
## k' edf k-index p-value
## s(Week) 51.0 14.3 1.09 0.98</pre>
```



4

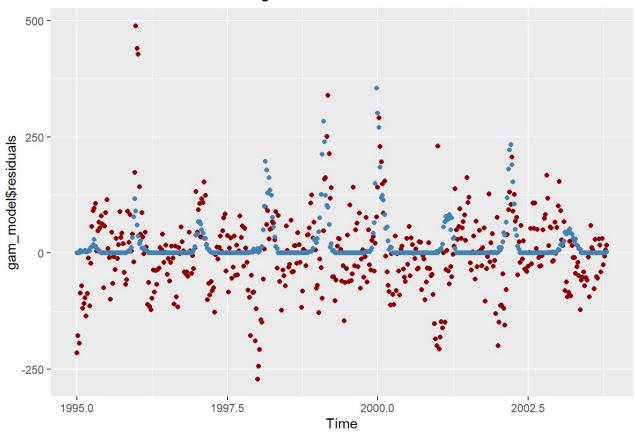
As can be seen from the output below, the lower penalty factor makes the model fit the data more loosly whereas a larger penalty factor gives the model a exact fit. The fit of the model does however not change much after a penalty factor of 13 is introduced.



5

Viewing the plot bleow it would seem viable to say that the temporal pattern in the residuals correlate to the outbreaks of influenza.

Residuals/ Influenza values against time

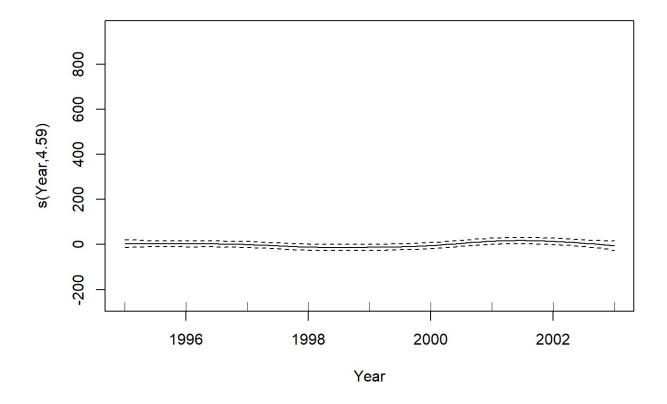


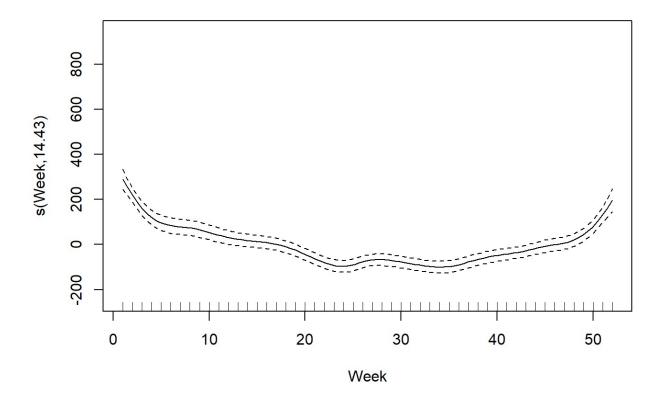
6

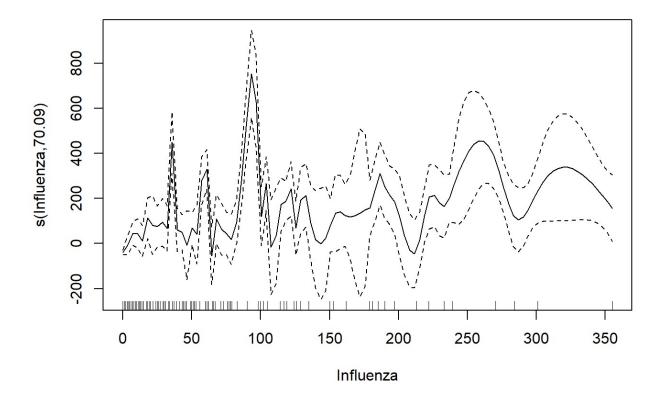
Plot shows that the fitted values are good aproximations of the mortality values. Using the summary function it is seen from the p- values that influenza and week are significant contributers of the mortality rate. The year is however not a significant contributer.

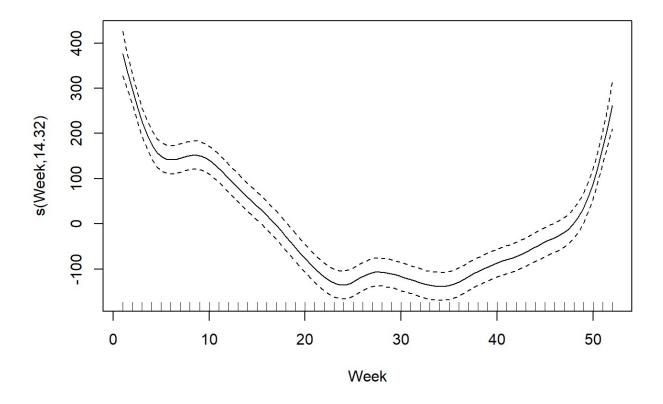
The first plot below is from the original (previous) model and the second from the spline. The spline model is a better model than the previous GAM model. The last output plot shows that the fitted values match the data very well.

```
## Family: gaussian
## Link function: identity
## Formula:
## Mortality ~ s(Year, k = length(unique(data$Year))) + s(Week,
      k = length(unique(data$Week))) + s(Influenza, k = length(unique(data$Influenz
a)))
##
## Parametric coefficients:
              Estimate Std. Error t value Pr(>|t|)
## (Intercept) 1783.765 3.198 557.8 <2e-16 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## Approximate significance of smooth terms:
                 edf Ref.df
##
                                 F p-value
              4.587 5.592 1.500 0.178
## s(Year)
## s(Week) 14.431 17.990 18.763 <2e-16 ***
## s(Influenza) 70.094 72.998 5.622 <2e-16 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
## Rank: 134/144
## R-sq.(adj) = 0.819 Deviance explained = 85.4%
## GCV = 5840.5 Scale est. = 4693.7 n = 459
```

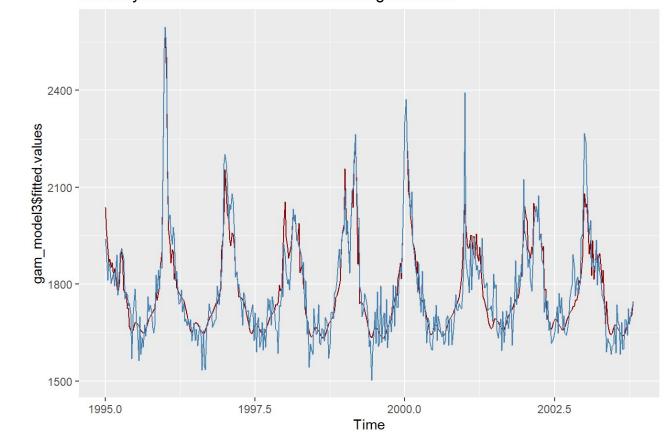








Mortality/ Fitted Influenza values values against time



Assignment 2. High-dimensional methods

1.

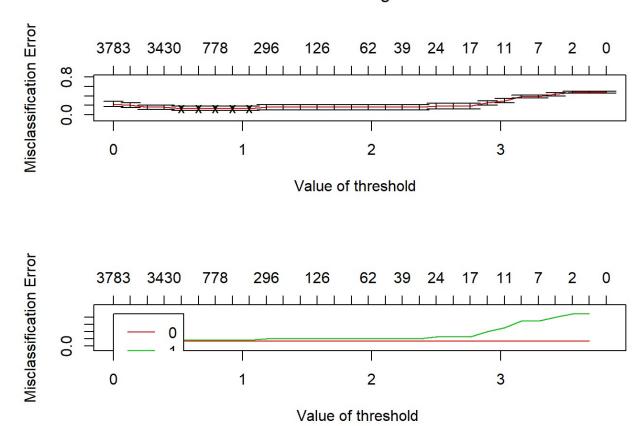
The nearest squished centroids model using cross validation had at lowest 6 errors, the largest threshold with 6 errors was 2.757. This threshold was then used in the output plots below. Using the cntroid plot, the 12 most contributing features for making predictions both negative (0) and positive (1).

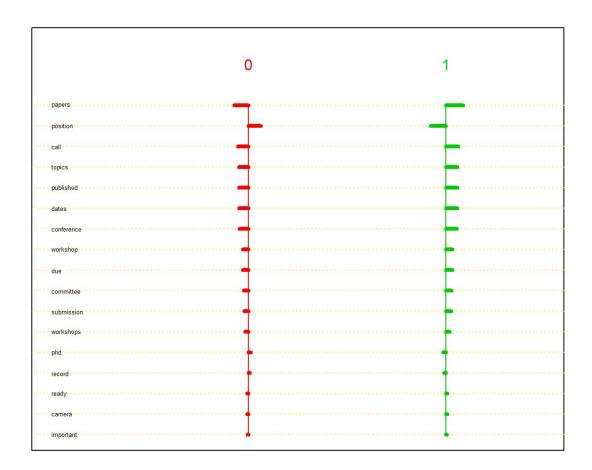
```
## 123456789101112131415161718192021222324252627282930
```

```
## 12Fold 1 :123456789101112131415161718192021222324252627282930
## Fold 2 :123456789101112131415161718192021222324252627282930
## Fold 3 :123456789101112131415161718192021222324252627282930
## Fold 4 :123456789101112131415161718192021222324252627282930
## Fold 5 :123456789101112131415161718192021222324252627282930
## Fold 6 :123456789101112131415161718192021222324252627282930
## Fold 7 :123456789101112131415161718192021222324252627282930
## Fold 8 :123456789101112131415161718192021222324252627282930
## Fold 9 :123456789101112131415161718192021222324252627282930
## Fold 10 :123456789101112131415161718192021222324252627282930
```

```
## Call:
## pamr.cv(fit = pamr_m, data = data2)
     threshold nonzero errors
## 1 0.000
               3783
                       10
## 2 0.132
               3548
                       9
               3499
                       7
## 3 0.263
## 4 0.395
                       7
               3430
## 5 0.527
               2368
                       6
## 6 0.658
                922
                       6
## 7 0.790
                778
                       6
## 8 0.922
                717
                       6
## 9 1.053
                339
                       6
## 10 1.185
                296
                       7
## 11 1.317
                188
                       7
## 12 1.448
                158
                       7
## 13 1.580
                126
                       7
## 14 1.712
                111
                       7
## 15 1.843
                       7
                 87
## 16 1.975
                 62
                       7
## 17 2.107
                 56
                       7
## 18 2.238
                 39
                       7
## 19 2.370
                 37
                       7
## 20 2.502
                 24
                       8
## 21 2.633
                 19
                       8
## 22 2.765
                       8
                 17
## 23 2.897
                 13
                       11
## 24 3.028
                 11
                       13
## 25 3.160
                  9
                       17
                  7
## 26 3.292
                       17
## 27 3.423
                  7
                       19
## 28 3.555
                  2
                       21
## 29 3.687
                  1
                       21
## 30 3.818
                       21
```

Number of genes





```
## Y hat
## Y 0 1
## 0 11 0
## 1 0 9
```

```
## [1] "NSC Error Rate with Test Data: 0"
```

```
##
        id
             0-score 1-score
## [1,] 3036 -0.146 0.1752
  [2,] 3187 0.1269 -0.1522
## [3,] 596 -0.1074 0.1289
## [4,] 4282 -0.0975 0.117
## [5,] 1045 -0.0968 0.1161
## [6,] 3364 -0.0968 0.1161
## [7,] 869 -0.0926 0.1111
## [8,] 4628 -0.0598 0.0717
## [9,] 1262 -0.0598 0.0717
## [10,] 810 -0.0488 0.0585
## [11,] 4060 -0.0455 0.0546
## [12,] 4629 -0.0343 0.0412
## [13,] 3125 0.0221 -0.0265
## [14,] 3458 0.0154 -0.0184
## [15,] 599 -0.0154 0.0184
## [16,] 3433 -0.0154 0.0184
## [17,] 2049 -0.011 0.0132
```

```
## [1] "Top 10 most contributing features:"
```

```
## papers
## position
## call
## topics
## dates
## published
## conference
## workshop
## due
## committee
```

2a,b

```
## Y hat
## Y 0 1
## 0 10 1
## 1 1 8
```

```
## [1] "Elastic net Error Rate with Test Data: 0.1"
```

```
## [1] "Number of contributing features: 64"
```

```
0-score 1-score
##
        id
             name
##
   [1,] 3036 papers
                        -0.146 0.1752
   [2,] 3187 position
                        0.1269 -0.1522
   [3,] 596 call
                        -0.1074 0.1289
##
##
  [4,] 4282 topics
                        -0.0975 0.117
## [5,] 1045 dates
                        -0.0968 0.1161
## [6,] 3364 published -0.0968 0.1161
## [7,] 869 conference -0.0926 0.1111
## [8,] 4628 workshop
                       -0.0598 0.0717
## [9,] 1262 due
                        -0.0598 0.0717
## [10,] 810 committee -0.0488 0.0585
## [11,] 4060 submission -0.0455 0.0546
## [12,] 4629 workshops -0.0343 0.0412
## [13,] 3125 phd
                        0.0221 -0.0265
## [14,] 3458 record
                        0.0154 -0.0184
## [15,] 599 camera
                        -0.0154 0.0184
## [16,] 3433 ready
                        -0.0154 0.0184
## [17,] 2049 important -0.011 0.0132
```

Error RateFeatures Selected

NSC 0 17 Elastic Net0.1 64 SVM 1 43

Which model would you prefer and why?

• The NSC gave the least error but used too many variables, and could have been a higher rate if set.seed was different. The Elastic Net and SVM perform very close to each other, however Elastic Net is more interpretable and preferable to the other two.

3. Benjamini-Hochberg

Which features correspond to the rejected hypotheses?

	P.Values T_F
	<dbl> <chr></chr></dbl>
papers	1.116910e-10 Rejecte
submission	7.949969e-10 Rejecte
position	8.219362e-09 Rejecte
published	1.835157e-07 Rejecte
mportant	3.040833e-07 Rejecte
call	3.983540e-07 Rejecte
conference	5.091970e-07 Rejecte

	P.Values T_F <dbl> <chr></chr></dbl>	
candidates	8.612259e-07 Rejected	
dates	1.398619e-06 Rejected	
paper	1.398619e-06 Rejected	
1-10 of 39 rows	Previous 1 2 3 4 Ne	xt

Interpret the result.

• The list of words selected by Benjamini-Hochberg method emphisize on lowering the false-discovery rate, meaning these words are the ones that give the least False Positive errors.

Appendix

```
knitr::opts_chunk$set(echo = TRUE)
library(readx1)
library(ggplot2)
setwd("C:/Users/Bjorn/Documents/LIU/machine learning/labs")
data = read_excel("Influenza.xlsx")
ggplot(data=data, aes(x=Time, y=Mortality))+
  geom_bar(stat = "identity")
ggplot(data=data, aes(x=Time, y=Influenza))+
  geom_bar(stat = "identity")
library(mgcv)
gam_model = gam(Mortality~Year+s(Week, k=length(unique(data$Week))), data=data,
                family = gaussian(link = "identity"), method="GCV.Cp")
gam_model2 = gam(Mortality~Year+s(Week)+s(Year, k=length(unique(data$Year))),
                 data=data, family = gaussian(link = "identity"))
summary(gam_model)
plot(gam_model2)
gam_model$sp
# s=interp(data$Year,data$Week, fitted(gam model))
# plot ly(x=\sim s x , y=\sim s y, z=\sim s z, type="surface")
plot(gam_model)
plot(gam model, shift=mean(data$Mortality), residuals=T, pch=1, xlab="") #plot with da
ta points included.
gam.check(gam model) #Gives some interesting inforamtion about the model.
gam fitted.results = predict(gam model, newdata=data)
ggplot(data=data, aes(x=Time, y=gam_fitted.results))+
  geom_bar(stat = "identity")
par(mfrow=c(3,3))
k=c(0.000011,0.0001131932,0.0003,0.0008,0.008,0.08,0.8,10)
for(i in k){
model = gam(Mortality~Year+s(Week, k=length(unique(data$Week))), data=data,
                        family = gaussian(link = "identity"), sp=i)
mod = model[i]
plot(model)
}
ggplot(data=data, aes(x=Time))+
  geom_point(aes(y=gam_model$residuals), color="darkred")+
  geom_point(aes(y=Influenza), color="steelblue")+
  ggtitle("Residuals/ Influenza values against time")
gam_model3 = gam(Mortality~ s(Year, k=length(unique(data$Year)))+s(Week, k=length(uniq
ue(data$Week)))+
                   s(Influenza, k=length(unique(data$Influenza))), data=data, family =
gaussian(link = "identity"))
summary(gam_model3)
plot(gam_model3) # plot new model.
plot(gam_model) # previous model.
ggplot(data=data, aes(x=Time))+
```

```
geom_line(aes(y=gam_model3$fitted.values), color="darkred")+
  geom_line(aes(y=Mortality), color="steelblue")+
  ggtitle("Mortality/ Fitted Influenza values values against time")
setwd("C:/Users/Bjorn/Documents/LIU/machine_learning/labs")
data<-read.csv2("data.csv", header = TRUE, sep = ";", check.names = FALSE ,encoding =</pre>
"latin1")
#names(data)<-iconv(names(data), to = "ASCII", sub = "")</pre>
#1.
n=dim(data)[1]
set.seed(12345)
id=sample(1:n, floor(n*0.7))
train=data[id,]
test=data[-id,]
train<-na.omit(train)</pre>
train2<-train[,-which(names(train) == "Conference")]</pre>
train2<-t(train2)</pre>
#data2<-list(x=train[,-which(names(train) == "Conference")], y=factor(train$Conferenc
e))
data2<-list(x=train2, y=as.factor(train$Conference),geneid=as.character(1:nrow(train</pre>
2)), genenames=rownames(train2))
library(pamr)
pamr_m<-pamr.train(data2)</pre>
pamr_cv<-pamr.cv(pamr_m, data2)</pre>
pamr_cv
pamr.plotcv(pamr_cv)
pamr.plotcen(pamr m, data2, threshold=2.757)
test2<-test[,-which(names(train) == "Conference")]</pre>
test2<-t(test2)
data3<-list(x=test2, y=test$Conference)</pre>
pred_nsc = pamr.predict(pamr_m, threshold = 2.757, newx=test2)
nsc_confmat = table("Y"=test$Conference,"Y hat"=pred_nsc)
nsc confmat
print(paste("NSC Error Rate with Test Data:", round(1-sum(diag(nsc_confmat))/sum(nsc_c
onfmat),4)))
a=pamr.listgenes(pamr_m,data2,threshold=2.757)
print("Top 10 most contributing features:")
cat( paste( colnames(data)[as.numeric(a[,1])][1:10], collapse='\n' ) ) #paste first 10
features.
library(glmnet)
```

```
library(pamr)
library(kableExtra)
library(kernlab)
xdata = as.matrix(train[,-ncol(train)])
ydata = train[,ncol(train)]
xtest = as.matrix(test[,-ncol(test)])
ytest = test[,ncol(test)]
elasticnet_model = cv.glmnet(x=xdata,y=ydata,alpha=0.5,family="binomial")
elasticnet model.predict = predict(elasticnet model, newx = as.matrix(test[,-4703]), t
ype = "class", s="lambda.min")
elasticnet confmat = table("Y"=test$Conference,"Y hat"=elasticnet model.predict)
elasticnet_confmat
print(paste("Elastic net Error Rate with Test Data:", round(1-sum(diag(elasticnet_conf
mat))/sum(elasticnet_confmat),4)))
coefs = as.matrix(coef(elasticnet_model, elasticnet_model$lambda.min))
elastic features <- length(names(coefs[coefs != 0,]))</pre>
print(paste("Number of contributing features:",elastic_features))
features = pamr.listgenes(pamr_m, data2, threshold = 2.757, genenames = TRUE)
res1 = list("Error Rate" = round(1-sum(diag(nsc_confmat))/sum(nsc_confmat),4), "Featur
es Selected" = nrow(features))
res2 = list("Error Rate" = round(1-sum(diag(elasticnet_confmat))/sum(elasticnet_confma
t),4), "Features Selected" = elastic_features)
invisible(capture.output(
  svm <- ksvm(Conference ~ .,</pre>
            data = train,
            kernel="vanilladot",
            scaled = FALSE)))
svm_pred <- predict(svm, newdata = test)</pre>
svm_mat <- table(ytest, svm_pred)</pre>
svm rate <- 1 - sum(diag(svm mat)) / sum(svm mat)</pre>
res3 <- list("Error Rate" = svm_rate, "Features Selected" = svm@nSV)</pre>
result = rbind("NSC" = res1, "Elastic Net" = res2, "SVM" = res3)
knitr::kable(result)
hochberg <- function(x, y, alpha) {</pre>
  p \leftarrow apply(x, 2, function(x_data)\{t.test(x_data \sim y, alternative = "two.sided") p.va
lue})
```

```
<- as.matrix(sort(p))
  rank
  1
          <- length(p)
  values <- (1:1/1) * alpha
           <- matrix(0,4702,1)
  T_F
  Z
           <- data.frame("P-Values" = rank, "T_F" = T_F)</pre>
  for(i in 1:4702){
   if(rank[i] <= values[i]){</pre>
      z[i,2] <- "Rejected"</pre>
   else{z[i,2] <- "Accepted"}</pre>
  lowest_p <- subset(z, T_F == "Rejected")</pre>
  return(lowest_p)
}
lowest_p <- hochberg(x = data[,-4703], y = data[,4703], alpha=0.05)
lowest_p
```