INP Data Analysis Bootcamp
Imaging Section: Mouse Data

Welcome! Your TAs for this session are:

Clayton Barnes, Graduate Student, Cardin Lab
Katie Ferguson, Postdoc, Cardin Lab
Antonio Oliveira Fonseca, Graduate Student, Van Dijk Lab

Data for this task are traces of the change in Ca^{2+} fluorescence from layer 2/3 somatostatin-positive (SOM+) GABAergic interneurons in V1 cortex of an awake mouse. These cells are cool because they are very visually responsive, even when the visual stimulus is large (this is not true for all cells). They inhibit the distal dendrites of neighboring pyramidal cells as well as other interneuron populations.

You will be given two data files. They contain the change in fluorescence of individual cells during the presentation of drifting grating visual stimuli, along with other information like the timing of the visual stimuli. The files are called:

'sampleCRFdata.mat' – here we have varied the contrast of the stimuli.
'sampleRFdata.mat' – here we have varied the size of the stimuli.

Each .mat file will load the following variables:

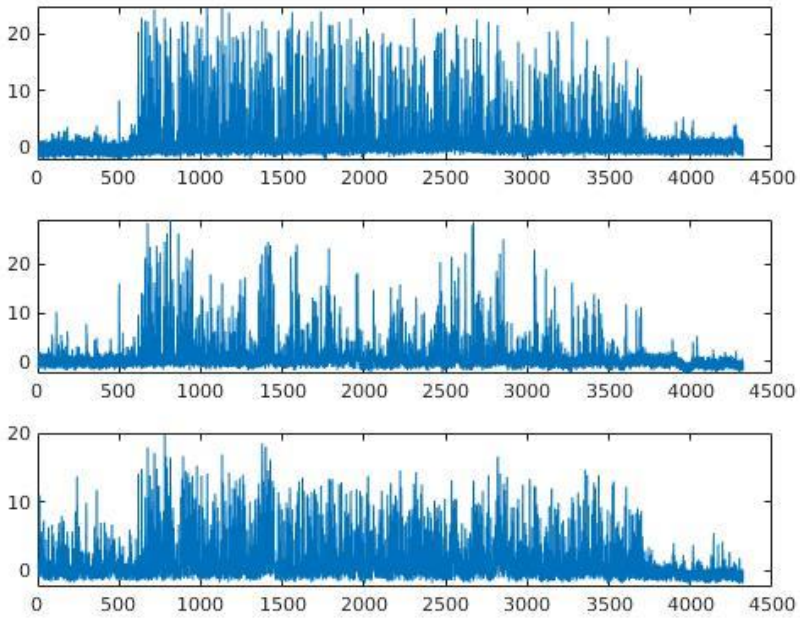
cellData: a matrix; the change in Ca^{2+} fluorescence (time points x neurons)
stimValue: a vector; the values of the various stimuli
time: a vector; time for the fluorescence trace
visOn: a vector; the indices of the visual stimuli onset
visOff: a vector; the indices of the visual stimuli offset

as well as the optional wheel variables (the mice are positioned on a wheel while they look at the visual stimuli):

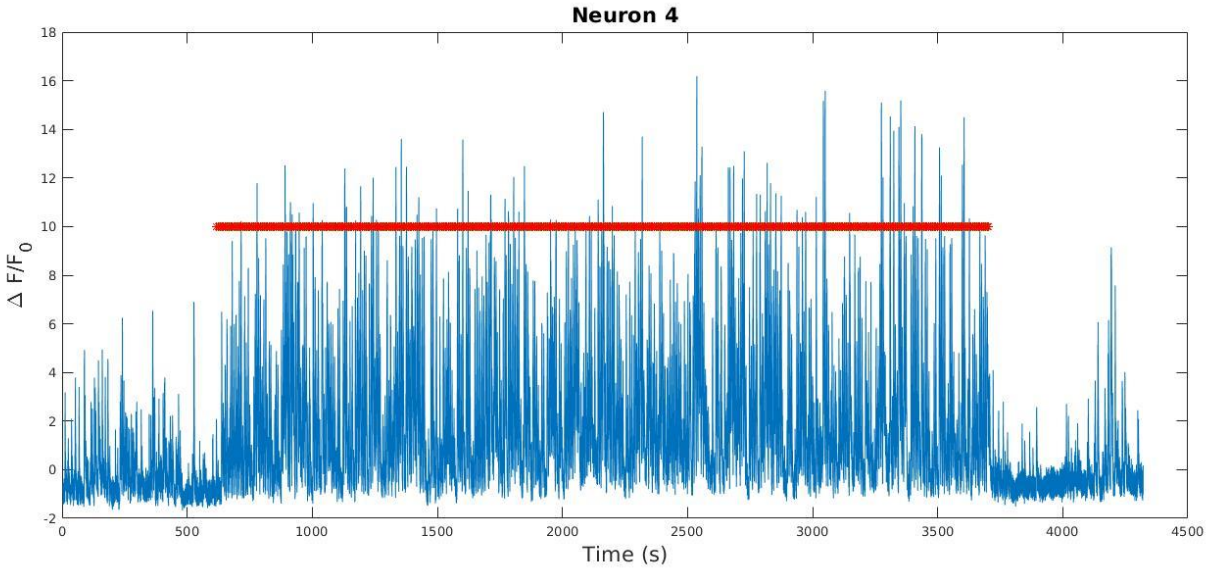
wheelOn: a vector; the indices of the onset of running
wheelOff: a vector; the indices of the offset of running (i.e., onset of sitting)

In each section of 'INP_imaging_mouse.m', use the suggestions given to fill in the rest of the necessary code.

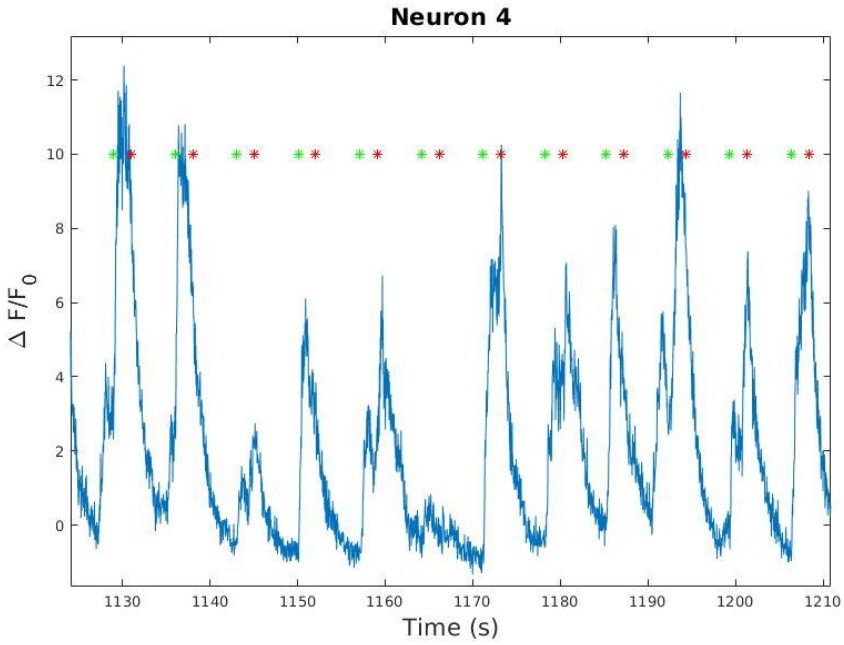
We will start by looking at the size tuning data ('sampleRFdata.mat '). Choose your favorite three neurons. Use the subplot function to plot their Ca2+ fluorescence over time. Your plot should look something like this (it will vary slightly depending on which neurons you choose):



Now, pick your favorite cell and plot its change in fluorescence across time. Add asterisks to tell you when the visual stim is turning on, and when it is turning off (get the timing of the visual responses from the variables visOn and visOff).

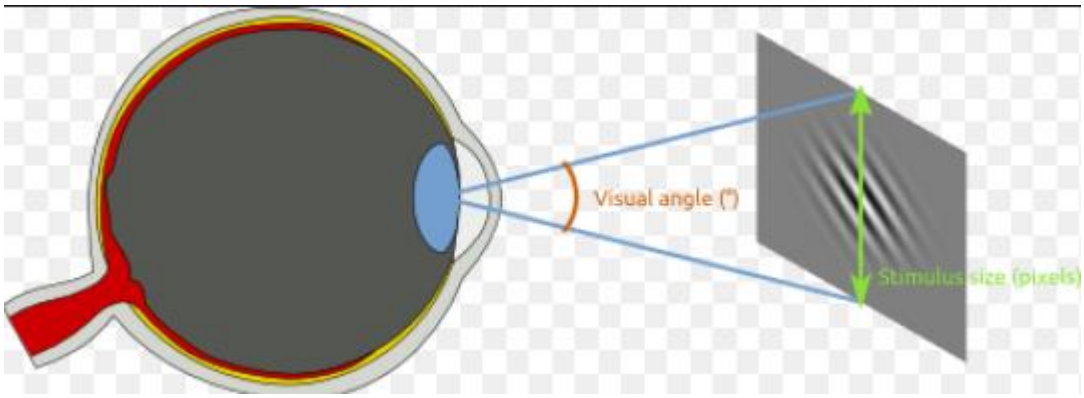


If you zoom into your Ca2+ traces, you should see something like this:



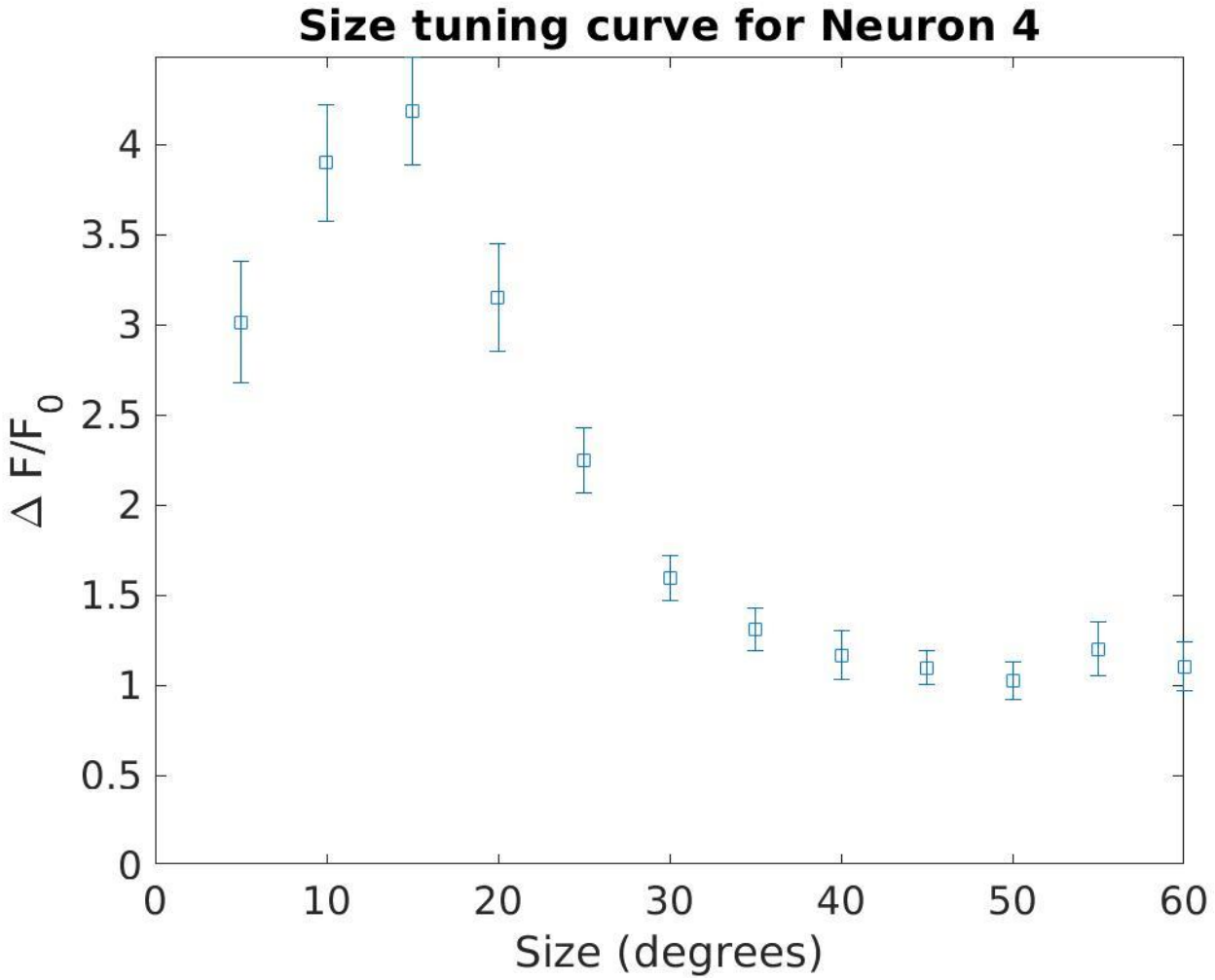
You can see how robustly these cells respond to the visual stimuli!

There is quite a bit of variability in the strength of response to each stimuli (you can see this easily in the zoomed in Ca^{2+} trace). This is because all the stimuli are not the same – we are varying the sizes of the stimui, as determined by the visual angle (in degrees), as depicted below.



https://osdoc.cogsci.nl/3.3/visualangle/

Let's find the mean and SEM of the responses for each stimulus, and create and error bar plot to visualize.



What can we see about how this cell responds to different sized stimuli? Why does this matter?

There are some exercises at the end. Don't do the 'bonus challenges' until you finish the exercises. You can choose the exercises that are most interesting to you. For example, how do these SOM+ cells respond to stimuli of varying contrasts? If you're interested in learning more about the wheel data, just ask!