## Cotiviti Healthcare

## 2024-09-18

```
# Libraries
library(caret)

## Loading required package: ggplot2

## Loading required package: lattice

library(randomForest)

## randomForest 4.7-1.1

## Type rfNews() to see new features/changes/bug fixes.

##
## Attaching package: 'randomForest'

## The following object is masked from 'package:ggplot2':
##
## margin
```

```
# Loading my data
data <- read.csv("C:\\Users\\jacob\\OneDrive\\Desktop\\R Studio Projects 2024\\Datasets\\healthc</pre>
are.csv")
# First I need to convert the categorical variables to factors to get it ready for training and
splitting the data
data$Gender <- as.factor(data$Gender)</pre>
data$Outcome <- as.factor(data$Outcome)</pre>
# Now it is time to split the data into training and testing sets
set.seed(42)
trainIndex <- createDataPartition(data$Outcome, p = 0.7, list = FALSE)</pre>
trainData <- data[trainIndex, ]</pre>
testData <- data[-trainIndex, ]</pre>
# Now lets train my Random Forest model of my healthcare data
set.seed(42)
rf_model <- train(Outcome ~ Age + Gender + Dosage_Med1_mg + Dosage_Med2_mg + Days_On_Treatment,
                   data = trainData,
                   method = "rf",
                   trControl = trainControl(method = "cv", number = 5))
# Now I need to predict on test data
rf_predictions <- predict(rf_model, testData)</pre>
# Lets take a look at the consfusion matrix
conf_matrix <- confusionMatrix(rf_predictions, testData$Outcome)</pre>
print(conf matrix)
```

```
## Confusion Matrix and Statistics
##
##
              Reference
## Prediction Improved No Change Worsened
     Improved
                    797
                               473
##
                                        337
##
     No Change
                     61
                                31
                                         22
     Worsened
                                 8
                                          7
##
                     13
##
## Overall Statistics
##
##
                  Accuracy : 0.4774
                    95% CI: (0.4538, 0.5011)
##
##
       No Information Rate : 0.498
##
       P-Value [Acc > NIR] : 0.9596
##
##
                     Kappa : -0.005
##
##
    Mcnemar's Test P-Value : <2e-16
##
## Statistics by Class:
##
##
                        Class: Improved Class: No Change Class: Worsened
## Sensitivity
                                                  0.06055
                                 0.91504
                                                                  0.019126
## Specificity
                                 0.07745
                                                  0.93290
                                                                  0.984816
## Pos Pred Value
                                 0.49596
                                                  0.27193
                                                                  0.250000
## Neg Pred Value
                                 0.47887
                                                  0.70581
                                                                  0.791400
## Prevalence
                                 0.49800
                                                  0.29274
                                                                  0.209262
## Detection Rate
                                 0.45569
                                                  0.01772
                                                                  0.004002
                                 0.91881
                                                  0.06518
                                                                  0.016009
## Detection Prevalence
## Balanced Accuracy
                                 0.49624
                                                  0.49672
                                                                  0.501971
```

```
# Now I want to see it in ggplot2 to get a better understanding
conf_matrix <- confusionMatrix(rf_predictions, testData$Outcome)
print(conf_matrix)</pre>
```

```
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                    797
                               473
##
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# I was having issues so I just load the library just in case for knitting purposes
library(ggplot2)
# Now I need to convert the confusion matrix to a data frame becauase it will not configure corr
```

```
# I was having issues so I just load the library just in case for knitting purposes
library(ggplot2)

# Now I need to convert the confusion matrix to a data frame becauase it will not configure corr
ectly
conf_df <- as.data.frame(conf_matrix$table)

# Time for a plot in ggplot2 with a level of frequency to show the variations in colors to make
the confusion matrix easier to visualize for novice users
ggplot(data = conf_df, aes(x = Reference, y = Prediction, fill = Freq)) +
    geom_tile() +
    geom_text(aes(label = Freq), vjust = 1) +
    scale_fill_gradient(low = "white", high = "blue") +
    labs(title = "Confusion Matrix Heatmap", x = "Actual", y = "Predicted")</pre>
```

## Confusion Matrix Heatmap



```
## RMSE Rsquared MAE
## 1.048782e+02 2.783348e-04 9.109412e+01
```

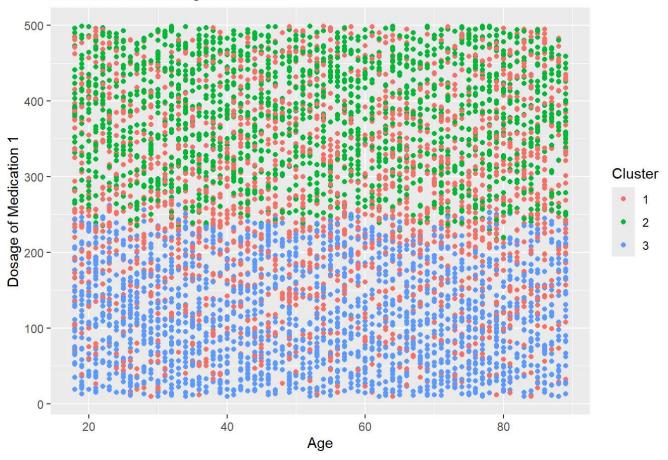
```
# Now it is time to see if the model is significant
model <- lm(Days_On_Treatment ~ Age + Dosage_Med1_mg + Dosage_Med2_mg + Social_Interaction_Scor
e, data = trainData)
# I begin with a summary of the model
summary(model)</pre>
```

```
##
## Call:
## lm(formula = Days On Treatment ~ Age + Dosage Med1 mg + Dosage Med2 mg +
       Social Interaction Score, data = trainData)
##
## Residuals:
##
       Min
                 10
                     Median
                                   3Q
                                           Max
## -189.721 -91.927
                       0.659
                               90.946 187.988
##
## Coefficients:
##
                             Estimate Std. Error t value Pr(>|t|)
## (Intercept)
                           173.841548 6.743021 25.781
                                                           <2e-16 ***
                                                   0.598 0.5498
## Age
                             0.047466
                                        0.079360
## Dosage Med1 mg
                            -0.002237 0.011629 -0.192
                                                           0.8475
                                                   1.653 0.0984 .
## Dosage Med2 mg
                             0.019421
                                        0.011748
## Social_Interaction_Score 0.082860 0.056731
                                                   1.461 0.1442
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 105.1 on 4080 degrees of freedom
## Multiple R-squared: 0.001314, Adjusted R-squared: 0.0003346
## F-statistic: 1.342 on 4 and 4080 DF, p-value: 0.2519
# Now lets scale the data down to size
scaled_data <- scale(data[, c("Age", "Dosage_Med1_mg", "Dosage_Med2_mg", "Social_Interaction_Sco</pre>
re")])
# I will utilize K-means clustering
set.seed(123)
kmeans_result <- kmeans(scaled_data, centers = 3)</pre>
# Next I need to add the cluster assignments to the data as a factor
data$Cluster <- factor(kmeans result$cluster)</pre>
# I was having problems knitting to pdf so I just used the ggplot2 library to make sure to Visua
lize the clusters from the data and I used geom point because the dots were to huge
library(ggplot2)
ggplot(data, aes(x = Age, y = Dosage_Med1_mg, color = Cluster)) +
```

labs(title = "K-means Clustering of Patients", x = "Age", y = "Dosage of Medication 1")

geom\_point() +

## K-means Clustering of Patients



```
# My libraries but I could haved used more but do to time constraints
library(readxl)
library(dplyr)
```

```
##
## Attaching package: 'dplyr'
```

```
## The following object is masked from 'package:randomForest':
##
## combine
```

```
## The following objects are masked from 'package:stats':
##
## filter, lag
```

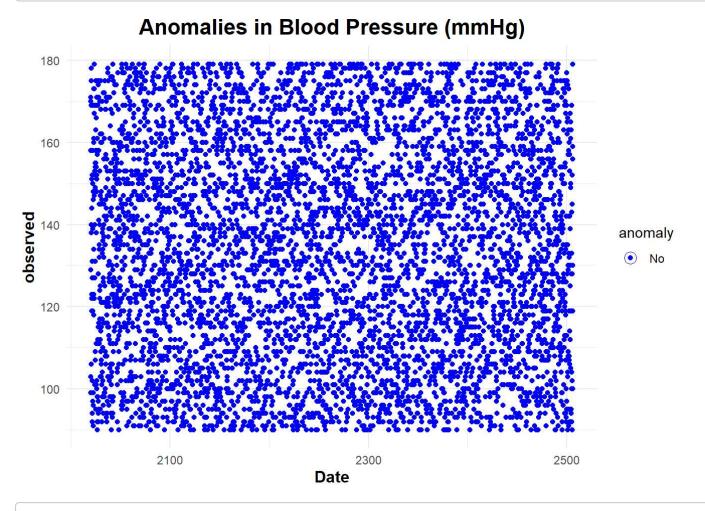
```
## The following objects are masked from 'package:base':
##
## intersect, setdiff, setequal, union
```

```
library(anomalize)
library(tibbletime)
```

```
##
## Attaching package: 'tibbletime'
## The following object is masked from 'package:stats':
##
##
       filter
library(ggplot2)
# Loading my data from my excel file but first I set the working directory
healthcare <- read excel("C:/Users/jacob/OneDrive/Desktop/R Studio Projects 2024/Datasets/health
care.xlsx")
# I decided to add the Date column by months in a numerical format as I was having trouble fitti
ng all the months on the plot so the data represents monthly records starting from January 2020
so each number represents a month, 12 moinths in a year is simply 12 counting from January 2020.
healthcare <- healthcare %>%
 mutate(Date = seq(as.Date("2020-01-01"), by = "month", length.out = n()))
# My table need to be converted using the tibble library packed I implemented prior to calculate
the time for the months.
healthcare_tbl <- as_tibble(healthcare)</pre>
# Convert to a time-based tibble
healthcare_time_tbl <- healthcare_tbl %>%
  as_tbl_time(index = Date)
# Now its is time to perform my anomaly detection on the 'Blood_Pressure_mmHg' column
anomaly detection <- healthcare time tbl %>%
 time_decompose(Blood_Pressure_mmHg, method = "stl", frequency = 12, trend = 60) %>%
  anomalize(remainder, method = "iqr") %>%
  time_recompose()
## frequency = 12 months
## trend = 60 months
## Registered S3 method overwritten by 'quantmod':
##
     method
```

##

as.zoo.data.frame zoo



#Extra credit: I was just curious what it would look like as a 3d plot: Here is the html link to
my plot ####https://plotly.com/~averymarcellusfalumni2022/3/#/
library(plotly) # 3D plotting

```
##
## Attaching package: 'plotly'
```

```
## The following object is masked from 'package:ggplot2':
##
##
       last_plot
## The following object is masked from 'package:stats':
##
##
       filter
## The following object is masked from 'package:graphics':
##
##
       layout
library(cluster) # clustering algorithms
library(ggplot2) # plotting
library(mclust)
                  # Gaussian Mixture Model and EM algorithm
## Package 'mclust' version 6.1.1
## Type 'citation("mclust")' for citing this R package in publications.
library(MASS)
                  # Bayes decision boundary
##
## Attaching package: 'MASS'
## The following object is masked from 'package:plotly':
##
##
       select
## The following object is masked from 'package:dplyr':
##
##
       select
```

```
# I had to first set Up my Plotly Credentials in R Studio
# By creating a Plotly username and API key from the website
Sys.setenv("plotly username" = "averymarcellusfalumni2022")
Sys.setenv("plotly_api_key" = "VYKPa2fBMcdjXY718C6d")
# Next I had to make sure my data was properly formatted. #'healthcare' with 'Blood Pressure mmH
g' and 'Cholesterol mg dL'
# My data is already loaded in the environment
# Now lets take a look at the K-means clustering based on my 'Blood Pressure mmHg' and 'Choleste
rol_mg_dL' data
set.seed(123)
kmeans result <- kmeans(healthcare[, c("Blood Pressure mmHg", "Cholesterol mg dL")], centers =</pre>
5)
# Now I need to fit my Gaussian Mixture Model (GMM)
gmm_result <- Mclust(healthcare[, c("Blood_Pressure_mmHg", "Cholesterol_mg_dL")], G = 5)</pre>
# Clustering time for plotting in 3D
healthcare$kmeans_cluster <- as.factor(kmeans_result$cluster)</pre>
healthcare$gmm cluster <- as.factor(gmm result$classification)
# 3D Plot for K-means clustering with Plotly but I need to make sure "z" is numeric because cate
rgorical variables is most of the time binary and will not work with my "x" and "y" numerical va
riables. I mean I could convert but I am running out of time researching the syntac errors on st
ackoverflow.com
kmeans_plot <- plot_ly(</pre>
 healthcare.
  x = ~Blood_Pressure_mmHg,
 y = ~Cholesterol mg dL,
  z = ~as.numeric(kmeans_cluster),
 type = 'scatter3d',
  mode = 'markers',
  marker = list(size = 4, color = ~as.numeric(kmeans_cluster), colorscale = 'Viridis')
) %>%
  layout(
    scene = list(
      xaxis = list(title = 'Blood Pressure (mmHg)'),
      yaxis = list(title = 'Cholesterol (mg/dL)'),
      zaxis = list(title = 'K-means Cluster')
    ),
    title = "3D K-means Clustering"
  )
# Now it is time to use 3D plotting for GMM clustering statistical analysis but once again "z" n
eeds to be numerical
gmm plot <- plot ly(</pre>
 healthcare,
  x = \sim Blood Pressure mmHg,
 y = ~Cholesterol_mg_dL,
  z = ~as.numeric(gmm cluster),
  type = 'scatter3d',
```

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```
Cotiviti Healthcare
 mode = 'markers',
  marker = list(size = 4, color = ~as.numeric(gmm cluster), colorscale = 'Viridis')
  layout(
    scene = list(
      xaxis = list(title = 'Blood Pressure (mmHg)'),
      yaxis = list(title = 'Cholesterol (mg/dL)'),
      zaxis = list(title = 'GMM Cluster')
    title = "3D GMM Clustering"
  )
# Next I created a "api create" function I researched online for my plot to be public and a url
link to access to play around with the plot
# Time to upload my K-means results to my plot with my Plotly account but I must set it to TRUE
(set public = TRUE) because setting it to FALSE will result in my plot being private and we do n
ot want that.
kmeans_link <- api_create(kmeans_plot, filename = "Kmeans_Clustering_3D", sharing = "public")</pre>
## Found a grid already named: 'Kmeans_Clustering_3D Grid'. Since fileopt='overwrite', I'll try
to update it
## Found a plot already named: 'Kmeans Clustering 3D'. Since fileopt='overwrite', I'll try to up
date it
# Time to upload my GMM plot to my Plotly account by once again setting it to TRUE (set public =
gmm link <- api create(gmm plot, filename = "GMM Clustering 3D", sharing = "public")</pre>
## Found a grid already named: 'GMM_Clustering_3D Grid'. Since fileopt='overwrite', I'll try to
update it
## Found a plot already named: 'GMM_Clustering_3D'. Since fileopt='overwrite', I'll try to updat
e it
# I am so excited I can't wait to see the results, Time to print the URL to access my plots to a
nalysis the results
print(kmeans_link)
```

print(gmm link)