Assignment 2 STATS 330

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Mean and Vairance Analysis

```
Visits.df <- read.csv("Visits.csv")</pre>
mean(Visits.df$visits)
## [1] 5.844076
var(Visits.df$visits)
## [1] 37.84379
observed=table(Visits.df$visits)
observed
##
##
             2
                  3
                          5
                               6
                                   7
                                               10
                                                            13
                                                                     15
                                                                         16
## 635 520 493 368 362 275 268 227 190 173 122 108 102
                                                                70
                                                                                      28
                                                                     61
                                                                         53
                                                                             49
                                                                                  26
                         25
                              26
                                      28
                                          29
                                                        32
                                                            33
                                                                34
                                                                     36
                                                                         37
                                                                             39
                                                                                  41
                                                                                      43
                      9 12
        26
            24
                 17
                            13
                                   7
                                     11
        45
                 48
##
            46
         1
     1
n=sum(observed)
```

As we can see by thoutput the variance is much larger than the mean, so there is over dispersion. Since the Poisson model assumes that the mean equals variance, this model is not a good fit for this set of data.

Fitting Poisson Model

```
fit1<- glm(visits~1,family=poisson,data=Visits.df)

The intercept is \beta_0 = log(\mu).
```

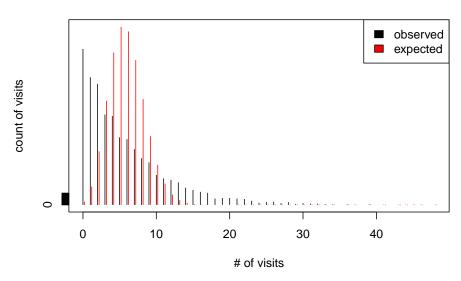
Poisson Graph

[1] 4406

```
x=as.numeric(names(observed))
expected.Pois=n*dpois(x, lambda=exp(coef(fit1)))
plot(x,observed, type="h",lwd=1, lend="butt", xlab="# of visits",
ylab="count of visits",
main="Observed vs Poisson expected counts",
```

```
xlim=range(x), ylim= c(0, max(observed,expected.Pois)))
lines(x+.2, expected.Pois,type="h",
lwd=1, lend="butt",col="red")
legend("topright", fill=c("black","red"),
legend=c("observed","expected"))
```

Observed vs Poisson expected counts



As you can see the red and blue bars don't match closely meaning that this model doesn't fit well.

Poisson Variance

```
poi_variance <- exp(coef(fit1))
poi_tail_probability <- ppois(12,lambda = poi_variance, lower.tail = FALSE)
poi_tail <- sum(Visits.df$visits >12)/n
poi_variance

## (Intercept)
## 5.844076
var(Visits.df$visits)

## [1] 37.84379
poi_tail_probability

## [1] 0.007203304
poi_tail
```

[1] 0.1277803

Since the sample variance is larger than the poisson variance this suggests over dispersion. Since the predicted tail larger than 12 is larger than the observed it shows that there is a lack of fit.

Expected Counts for Poisson Model

```
## inspecting the data>>=
Egt5=expected.Pois>=5
# note ! is R's way of saying NOT
E.Pois=c(expected.Pois[Egt5], sum(expected.Pois[!Egt5]))
O.Pois=c(observed[Egt5], sum(observed[!Egt5]))
E.Pois
## [1] 12.764224 74.595099 217.969724 424.610563 620.364128 725.091055
## [7] 706.247903 589.623801 430.725807 279.688274 163.451960 86.838702
## [13] 42.291000 19.011679 7.936122 4.789959

J <- length(E.Pois)
J</pre>
## [1] 16
```

Chi-Squared for Poisson

There is one parameter (the mean) so p = 1 and J = 16

```
chisq <- sum((0.Pois - E.Pois)^2/E.Pois)
p_value <- pchisq(chisq, df = J - 1, lower.tail = FALSE)
chisq

## [1] 68043.05
p_value

## [1] 0
p-value is less than 0.05 so the Poisson model doesn't fit very well.</pre>
```

Negative Binomial Fit

```
library(MASS)
fit2 <- glm.nb(visits ~ 1, data = Visits.df)</pre>
summary(fit2)
##
## Call:
## glm.nb(formula = visits ~ 1, data = Visits.df, init.theta = 1.037433909,
       link = log)
##
## Coefficients:
##
               Estimate Std. Error z value Pr(>|z|)
## (Intercept) 1.76543
                           0.01605
                                       110
                                             <2e-16 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## (Dispersion parameter for Negative Binomial(1.0374) family taken to be 1)
##
       Null deviance: 5029.2 on 4405 degrees of freedom
## Residual deviance: 5029.2 on 4405 degrees of freedom
## AIC: 25085
##
```

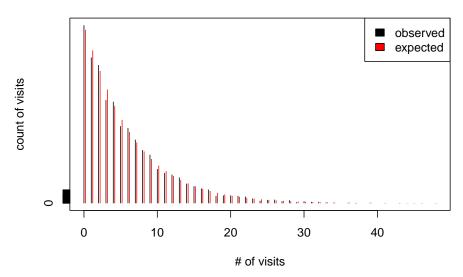
```
## Number of Fisher Scoring iterations: 1
##
##
##
                           1.0374
                  Theta:
##
              Std. Err.:
                           0.0272
##
    2 x log-likelihood: -25081.2590
mean_nb <- exp(coef(fit2))</pre>
mean_nb
## (Intercept)
##
      5.844076
```

Theta is small so overdispersion is high.

Negative Binomial Graph

```
x=as.numeric(names(observed))
theta_hat <- fit2$theta
expected.nb=n*dnbinom(x, mu = mean_nb, size = theta_hat)
plot(x,observed, type="h",lwd=1, lend="butt", xlab="# of visits",
ylab="count of visits",
main="Observed vs Negatvie Binomial expected counts",
xlim=range(x), ylim= c(0, max(observed,expected.nb)))
lines(x+.2, expected.nb,type="h",
lwd=1, lend="butt",col="red")
legend("topright", fill=c("black","red"),
legend=c("observed","expected"))</pre>
```

Observed vs Negatvie Binomial expected counts



The black and red bars in this model is very similar so the NB model is a much better fit.

Negtaive Binomial Variance

```
nb_variance <- mean_nb * (1+mean_nb/theta_hat)
nb_tail_probability <- pnbinom(12,mu = mean_nb,size = theta_hat ,lower.tail = FALSE)
nb_tail <- sum(Visits.df$visits >12)/n
nb_variance

## (Intercept)
## 38.76495
var(Visits.df$visits)

## [1] 37.84379
nb_tail_probability

## [1] 0.1267397
nb_tail
## [1] 0.1277803
```

The variances and tails are very similar to each other meaning that the NB model is much better fit

Expected Counts for NB Model

```
## inspecting the data>>=
Egt5_nb=expected.nb>=5
# note ! is R's way of saying NOT
E.nb=c(expected.nb[Egt5_nb], sum(expected.nb[!Egt5_nb]))
O.nb=c(observed[Egt5_nb], sum(observed[!Egt5_nb]))
E.nb
  [1] 618.815040 545.196950 471.670886 405.561547 347.643668 297.444402
## [7] 254.178643 217.013857 185.159931 157.899863 134.597369 114.694904
## [13] 97.707727 83.216567 70.860074 60.327620 51.352691 43.706959
## [19] 37.195034 31.649867 26.928745 22.909821 19.489117 16.577939
## [25] 14.100663 11.992824 10.199489 8.673856
                                                     7.376062
                                                                6.272157
## [31]
         5.333234 22.657596
J_nb <- length(E.nb)</pre>
J_nb
## [1] 32
p_nb = 2 cause we got mean and dispersion, and J = 32
```

Chi-Squared for NB

```
chisq_nb <- sum((0.nb - E.nb)^2/E.nb)
p_value_nb <- pchisq(chisq_nb, df = J_nb - 2, lower.tail = FALSE)
chisq_nb

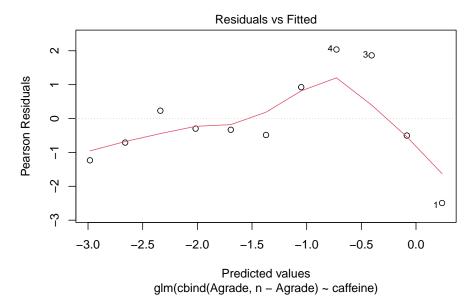
## [1] 26.35574
p_value_nb

## [1] 0.6568566</pre>
```

The p-value is very large so the NB model is a very good fit for this data and much better than the Poisson model.

Caffeine Model

```
Caffeine.df=read.csv("Caffeine.csv")
## null model
mod.null=glm(cbind(Agrade,n-Agrade)~1, family=binomial, data =Caffeine.df)
## linear log-odds model
mod1=glm(cbind(Agrade,n-Agrade)~caffeine, family=binomial, data =Caffeine.df)
summary(mod1)
##
## Call:
## glm(formula = cbind(Agrade, n - Agrade) ~ caffeine, family = binomial,
       data = Caffeine.df)
##
## Coefficients:
               Estimate Std. Error z value Pr(>|z|)
## (Intercept) 0.238469
                          0.226199
                                    1.054
                           0.001009 -6.381 1.75e-10 ***
## caffeine
              -0.006442
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## (Dispersion parameter for binomial family taken to be 1)
##
      Null deviance: 69.358 on 10 degrees of freedom
##
## Residual deviance: 18.625 on 9 degrees of freedom
## AIC: 55.87
## Number of Fisher Scoring iterations: 4
1-pchisq(deviance(mod1), df.residual(mod1))
## [1] 0.0285817
plot(mod1, which=1)
```



We have a model for the log-odds of getting an A-grade based on the amount of caffeine consumed. We have a small p-value (less than 0.05) so this model isn't a good fit. The residuals are not constant so no EOV.

Likelihood Function and Saturated Model

```
LLcaffeine=function(p,n=Caffeine.df$n, y=Caffeine.df$Agrade){
    out=y*log(p)+(n-y)*log(1-p)
# log(0) adds zero to LL
    out[is.na(out)]=0
    out
}

ps <- Caffeine.df$Agrade / Caffeine.df$n
LL_saturated <- LLcaffeine(ps)
LL_saturated
## [1] -19.095425 -20.526953 -20.526953 -20.794415 -19.095425 -13.516836
## [7] -11.780234 -9.752489 -9.752489 -4.384342 0.000000
```

Likelihhod Function in Null Model

Null Deviance

```
null_deviance <- (-2) * (sum(LL_null) - sum(LL_saturated))
null_deviance</pre>
```

```
## [1] 69.35772
mod1$null.deviance
## [1] 69.35772
```

Likelihood in Linear Caffeine Model

Residual Deviance

```
residual_deviance <- (-2) * (sum(LL_model)-sum(LL_saturated))
residual_deviance

## [1] 18.62452
mod1$deviance

## [1] 18.62452
```

Pearson Residuals

```
observed <- Caffeine.df$Agrade
expected <- preds * Caffeine.df$n</pre>
residual_pearson <- (observed - expected) / sqrt(expected * (1 - preds))
residual_pearson
                               3
## -2.4933594 -0.5018859
                       1.8640483
                                 2.0373760
                                           0.9259154 -0.4859299 -0.3314440
##
          8
                     9
                              10
residuals(mod1, type = "pearson")
##
          1
                     2
                               3
## -2.4933594 -0.5018859 1.8640483 2.0373760 0.9259154 -0.4859299 -0.3314440
## -0.2980178 0.2318156 -0.7097135 -1.2329671
```

Quasi-Binomial Model

```
mod2 <- glm(cbind(Agrade, n-Agrade) ~ caffeine, family = quasibinomial, data = Caffeine.df)
pearson_residuals <- residuals(mod1, type = "pearson")
dispersion_k <- sum(pearson_residuals^2) / df.residual(mod1)
summary(mod2)</pre>
```

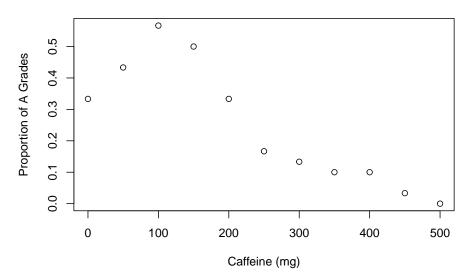
```
##
## Call:
  glm(formula = cbind(Agrade, n - Agrade) ~ caffeine, family = quasibinomial,
##
       data = Caffeine.df)
##
##
## Coefficients:
                Estimate Std. Error t value Pr(>|t|)
##
                           0.315095
                                      0.757 0.46851
## (Intercept)
               0.238469
  caffeine
               -0.006442
                           0.001406
                                    -4.581 0.00133 **
##
## Signif. codes:
                  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
  (Dispersion parameter for quasibinomial family taken to be 1.940452)
##
##
##
       Null deviance: 69.358 on 10 degrees of freedom
## Residual deviance: 18.625
                             on
                                 9
                                     degrees of freedom
## AIC: NA
##
## Number of Fisher Scoring iterations: 4
dispersion_k
```

Plot of Proportions of A-Grades

[1] 1.940452

```
A_grades <- Caffeine.df$Agrade / Caffeine.df$n
plot(Caffeine.df$caffeine, A_grades, xlab = "Caffeine (mg)", ylab = "Proportion of A Grades", main = "A
```

A Grade Proportion vs Caffeine



It seems that at around 100mg of Caffeine is the most optimal amount for getting an A Grade and once you start have more than this amount it gets lower and lower.

New Model

```
mod3 <- glm(cbind(Agrade, n-Agrade) ~ caffeine + I(caffeine^2), family = binomial, data = Caffeine.df)
summary(mod3)
##
## Call:
## glm(formula = cbind(Agrade, n - Agrade) ~ caffeine + I(caffeine^2),
##
      family = binomial, data = Caffeine.df)
##
## Coefficients:
##
                  Estimate Std. Error z value Pr(>|z|)
## (Intercept)
                -3.974e-01 3.021e-01 -1.315 0.18836
## caffeine
                 4.600e-03 3.633e-03
                                       1.266 0.20538
## I(caffeine^2) -2.762e-05 9.257e-06 -2.984 0.00285 **
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
## (Dispersion parameter for binomial family taken to be 1)
##
##
      Null deviance: 69.3577
                              on 10 degrees of freedom
## Residual deviance: 7.6639 on 8 degrees of freedom
## AIC: 46.909
##
## Number of Fisher Scoring iterations: 5
1 - pchisq(deviance(mod3), df.residual(mod3))
## [1] 0.4669742
anova(mod1, mod3, test = "Chisq")
## Analysis of Deviance Table
## Model 1: cbind(Agrade, n - Agrade) ~ caffeine
## Model 2: cbind(Agrade, n - Agrade) ~ caffeine + I(caffeine^2)
    Resid. Df Resid. Dev Df Deviance Pr(>Chi)
## 1
            9
                 18.6245
## 2
            8
                  7.6639 1
                              10.961 0.0009307 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
```

The p-value is very low for the squared term so it is significant and a better fit for this data. This allows for a parabola shaped graph to exist.

Executive Summary

The first model used was a linear model which seemed to have poor fit for the data given. There was over dispersion and lack of consistent residuals. After using a quasi-binomial model however we had a much better fit and could easily compare the A grades to the caffeine intake, we were able to see that at 100mg of caffeine, this was the most potimal point of getting an A grade.