

Cell-type specific enrichment analysis

Dexamethasone-Stimulated Human Array Project

```
library(data.table)
library(dplyr)
library(ggplot2)
library(corrplot)
library(factoextra)
library(viridis)
library(ggpubr)
library(UpSetR)
```

```
cbPalette <- c( "#0072B2", "#009E73", "#E69F00", "#F0E442", "#D55E00", "#CC79A7", "#56B4E9", "#999999")
separate <- ""
```

```
treatment <- "veh"
```

```
pval.veh.df <- fread(paste0("~/bio/code/mpip/dex-stim-human-array/output/data/integrative/cell_type_enr"))
```

```
pval.veh.bcc.df <- pval.veh.df[, 1:13]
```

```
fdr.bcc.df <- matrix(p.adjust(as.vector(as.matrix(pval.veh.bcc.df[, 2:13])), method='fdr'),
                    ncol=12) %>%
  data.frame()
```

```
fdr.bcc.df <- cbind(pval.veh.bcc.df$CpG_ID, fdr.bcc.df)
colnames(fdr.bcc.df) <- colnames(pval.veh.bcc.df)
```

```
sign.pval.df <- fdr.bcc.df %>% reshape2::melt(measure.vars = colnames(fdr.bcc.df)[2:13]) %>% setDT()
colnames(sign.pval.df) <- c("CpG_ID", "Type", "fdr")
```

```
sign.pval.df <- sign.pval.df[fdr <= 0.05]
sign.pval.df <- na.omit(sign.pval.df)
sign.pval.df
```

```
##           CpG_ID Type      fdr
##      1: cg16535257 Bas 0.024366508
##      2: cg13938959 Bas 0.011324577
##      3: cg06931612 Bas 0.031029976
##      4: cg00582671 Bas 0.010838442
##      5: cg11143486 Bas 0.028726078
##      ---
## 734223: cg08425796 Treg 0.003233155
## 734224: cg14496081 Treg 0.035331162
## 734225: cg01370805 Treg 0.003505756
## 734226: cg24849633 Treg 0.035063650
## 734227: cg12502079 Treg 0.005416987
```

```
veh.sign.pval.df <- sign.pval.df
veh.sign.pval.df[["Treatment"]] <- treatment
```

DEX

```
treatment <- "dex"
```

```
pval.dex.df <- fread(paste0("~/bio/code/mpip/dex-stim-human-array/output/data/integrative/cell_type_enr"))
```

```
pval.dex.bcc.df <- pval.dex.df[, 1:13]
```

```
fdr.bcc.df <- matrix(p.adjust(as.vector(as.matrix(pval.dex.bcc.df[, 2:13])), method='fdr'),
                    ncol=12) %>%
  data.frame()
```

```
fdr.bcc.df <- cbind(pval.dex.bcc.df$CpG_ID, fdr.bcc.df)
colnames(fdr.bcc.df) <- colnames(pval.dex.bcc.df)
```

```
sign.pval.df <- fdr.bcc.df %>% reshape2::melt(measure.vars = colnames(fdr.bcc.df)[2:13]) %>% setDT()
colnames(sign.pval.df) <- c("CpG_ID", "Type", "fdr")
sign.pval.df <- sign.pval.df[fdr <= 0.05]
```

```
sign.pval.df <- na.omit(sign.pval.df)
sign.pval.df
```

```
##           CpG_ID Type      fdr
##      1: cg21038584 Bas 0.036610119
##      2: cg07211239 Bas 0.038491071
##      3: cg04260676 Bas 0.002423863
##      4: cg02050917 Bas 0.039608066
##      5: cg26785499 Bas 0.024939802
##      ---
## 200548: cg00392007 Treg 0.048659128
## 200549: cg06665773 Treg 0.049944129
## 200550: cg01280327 Treg 0.022621541
## 200551: cg24322531 Treg 0.014209176
## 200552: cg01548456 Treg 0.049194224
```

```
dex.sign.pval.df <- sign.pval.df
dex.sign.pval.df[["Treatment"]] <- treatment
```

Distribution plots for Basleline and Dex together

```
sign.pval.df <- rbind(veh.sign.pval.df, dex.sign.pval.df)
intersect.cpgs <- intersect(dex.sign.pval.df$CpG_ID, veh.sign.pval.df$CpG_ID)

print(paste0("Number of unique dex CpGs: ", length(unique(dex.sign.pval.df$CpG_ID))))
```

```
## [1] "Number of unique dex CpGs: 118284"
```

```
print(paste0("Number of unique baseline CpGs: ", length(unique(veh.sign.pval.df$CpG_ID))))
```

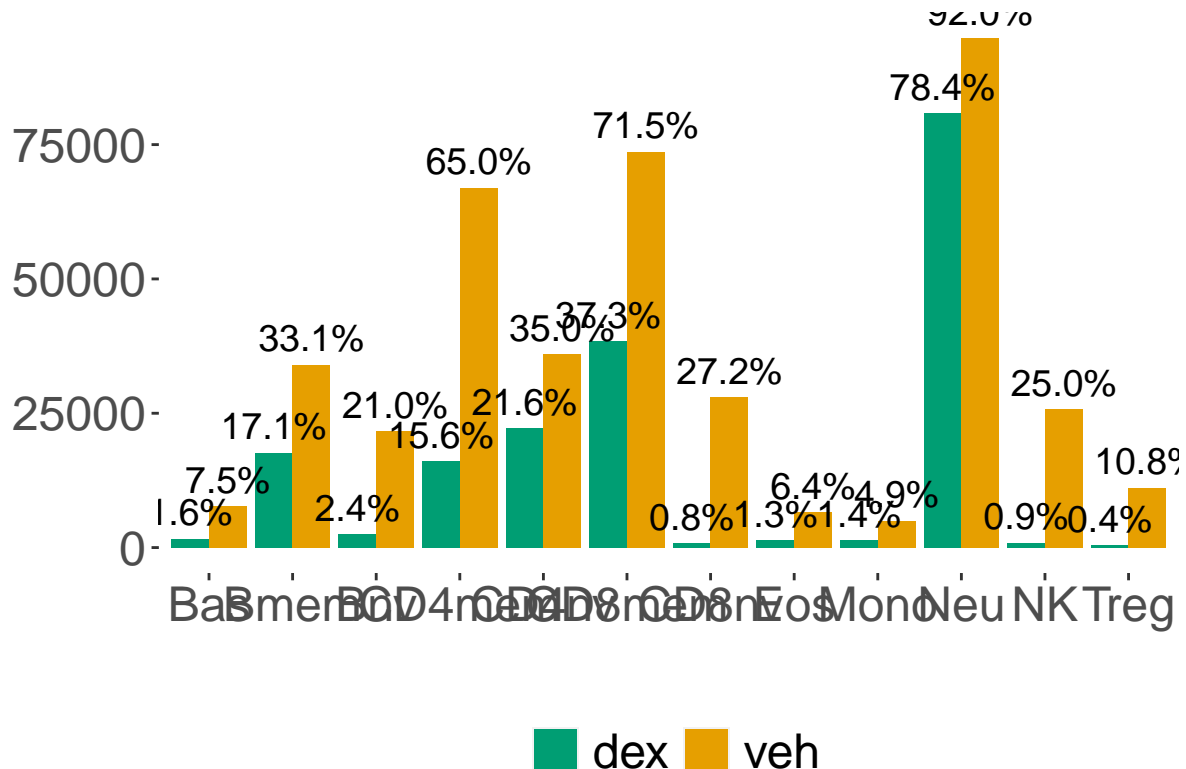
```
## [1] "Number of unique baseline CpGs: 289966"
```

```
print(paste0("Number of inteseacting CpGs: ", length(unique(intersect.cpgs))))
```

```
## [1] "Number of inteseacting CpGs: 103033"
```

```
ggplot(sign.pval.df[CpG_ID %in% intersect.cpgs], aes(x = Type, fill = Treatment)) +
  geom_bar(stat = "count", alpha = 1, position = position_dodge(width = .9)) +
  # geom_text(aes(label = scales::percent(prop.table(stat(count)))),
  # geom_text(aes(label = scales::percent(stat(count) / length(unique(sign.pval.df$CpG_ID)))),
  geom_text(aes(label = scales::percent(stat(count) / length(intersect.cpgs), accuracy = 0.1),
    stat = "count", vjust = -0.5, size = 5, position = position_dodge(width = .9)) +
  theme(legend.position = "bottom", # c(0.9, 0.9),
    legend.title = element_blank(),
    legend.text = element_text(size = 18),
    panel.grid.major = element_blank(),
    panel.background = element_blank(),
    plot.title = element_text(size = 18),
    axis.title = element_text(size = 18),
    axis.text = element_text(size = 18)) +
  labs(title = "Distribution of CpGs significant at FDR = 0.05 across 12 blood cell types",
    x = "", y = "") +
  scale_fill_manual(values = cbPalette[2:3])
```

Distribution of CpGs significant at FDR = 0.05 ac



Cell-type specificity on GR-induced (delta)-meQTLs

```
ind.meqtl.delta.df      <- fread("~/bio/code/mpip/dex-stim-human-array/output/data/integrative/matrixEQTLs/ind.meqtl.delta.df")
delta.meqtl.cpgs       <- ind.meqtl.delta.df$gene %>% unique()
sign.pval.delta.mqtl.df <- sign.pval.df[CpG_ID %in% delta.meqtl.cpgs, ]

sign.pval.cpgs.dex <- sign.pval.delta.mqtl.df[Treatment == "dex", CpG_ID]
sign.pval.cpgs.veh <- sign.pval.delta.mqtl.df[Treatment == "veh", CpG_ID]

intersect.cpgs         <- intersect(sign.pval.cpgs.dex, sign.pval.cpgs.veh)
sign.pval.delta.mqtl.df <- sign.pval.delta.mqtl.df[CpG_ID %in% intersect.cpgs,]

print(paste0("Number of unique dex CpGs: ", length(unique(sign.pval.cpgs.dex))))
```

```
## [1] "Number of unique dex CpGs: 678"
```

```
print(paste0("Number of unique baseline CpGs: ", length(unique(sign.pval.cpgs.veh))))
```

```
## [1] "Number of unique baseline CpGs: 1663"
```

```
print(paste0("Number of intersecting CpGs: ", length(unique(intersect.cpgs))))
```

```
## [1] "Number of intersecting CpGs: 653"
```

Check if changes are significant

Enrichment GR-meQTL CpGs over all CpGs

Distribution of P-values for each blood cell-type

Kolmogorov-Smirnov test: Do 2 samples follow the same distribution?

```
blood.cell.types      <- unique(sign.pval.delta.mqtl.df$Type)
# blood.cell.types    <- levels(sign.pval.delta.mqtl.df$Type)

test.res.df <- lapply(blood.cell.types, function(i){ # for each blood cell type
  dex.fdr.lst <- sign.pval.delta.mqtl.df[Type == i][Treatment == "dex", fdr]
  veh.fdr.lst <- sign.pval.delta.mqtl.df[Type == i][Treatment == "veh", fdr]
  test.rslt  <- ks.test(dex.fdr.lst, veh.fdr.lst)

  return(data.frame(Type = i, "p-value" = test.rslt$p.value))
}) %>%
  bind_rows()

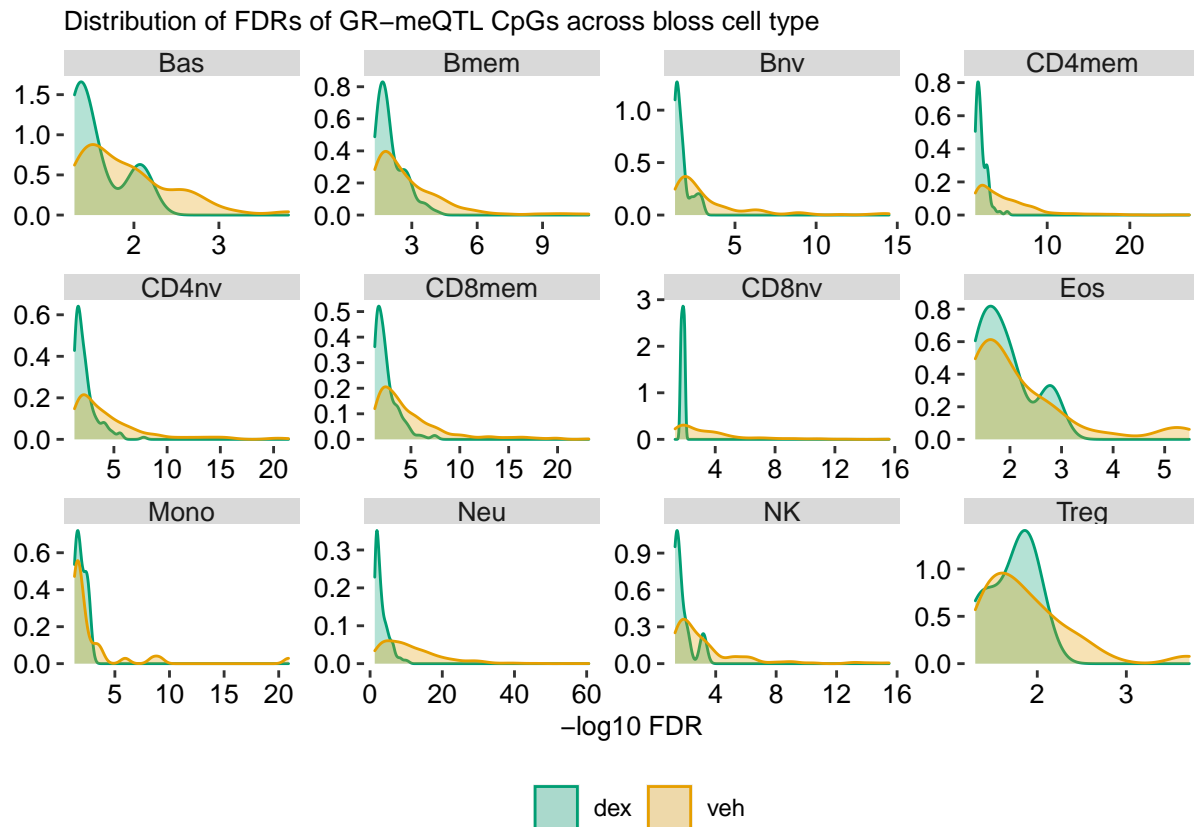
sign.pval.delta.mqtl.df <- left_join(sign.pval.delta.mqtl.df, test.res.df)
```

```
## Joining, by = "Type"
```

```

ggplot(sign.pval.delta.mqtl.df, aes(x = -log10(fdr), fill = Treatment)) +
  geom_density(alpha = 0.3, aes(color = Treatment)) +
  geom_text(data = subset(unique(sign.pval.delta.mqtl.df[, .(Treatment, Type, p.value)])),
    aes(x = -Inf, y = -Inf, label = paste0("KS test = ", signif(p.value, 3))),
    hjust = -0.5, vjust = -15, size = 5, color = "black" ) +
  facet_wrap(~ Type, scales = "free", ncol = 4) +
  theme(legend.position = "bottom", # c(.9,.9),
    legend.title = element_blank(),
    panel.grid.major = element_blank(),
    panel.background = element_blank(),
    plot.title = element_text(size = 10, color = "black"),
    axis.title = element_text(size = 10, color = "black"),
    axis.text.x = element_text(size = 10, color = "black"),
    axis.text.y = element_text(size = 10, color = "black"),
    strip.text.x = element_text(size = 10, margin = margin())) +
  labs(title = "Distribution of FDRs of GR-meQTL CpGs across blossom cell type", x = "-log10 FDR", y = "Density")
  scale_fill_manual(values = cbPalette[2:3]) +
  scale_color_manual(values = cbPalette[2:3])

```



```
plt.size <- 12
```

```

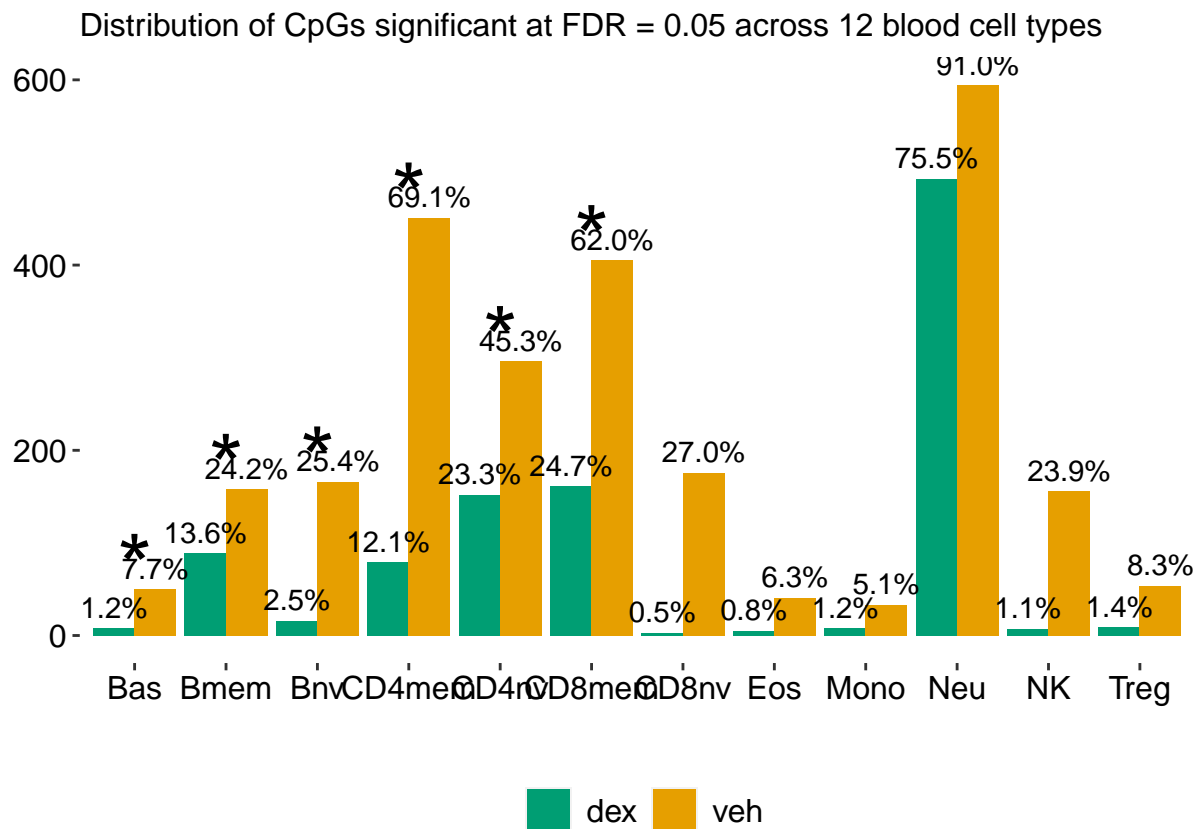
ggplot(sign.pval.delta.mqtl.df, aes(x = Type, fill = Treatment)) +
  geom_bar(stat = "count", alpha = 1, position = position_dodge(width = .9)) +
  geom_text(aes(label = scales::percent(stat(count) / length(intersect.cpgs), accuracy = 0.1)),
    stat = "count", vjust = -0.5, size = plt.size / 3, position = position_dodge(width = 0.9)) +
  geom_text(data = subset(sign.pval.delta.mqtl.df[Treatment == "veh"], p.value <= 0.05),

```

```

aes(label = "*"), stat = "count", vjust = 0.15, size = plt.size) +
theme(legend.position = "bottom", # c(0.9, 0.9),
      legend.title = element_blank(),
      legend.text = element_text(size = plt.size),
      panel.grid.major = element_blank(),
      panel.background = element_blank(),
      plot.title = element_text(size = plt.size),
      axis.title = element_text(size = plt.size),
      axis.text = element_text(size = plt.size, colour = "black")) +
labs(title = "Distribution of CpGs significant at FDR = 0.05 across 12 blood cell types",
     x = "", y = "") +
scale_fill_manual(values = cbPalette[2:3])

```



Upset plot for DEX CpGs

```

blood.types.df.lst <- split(sign.pval.delta.mqtl.df[Treatment == "dex"], by = "Type", )
blood.types.cpg.lst <- lapply(blood.types.df.lst, function(df) df$CpG_ID)

upset(fromList(blood.types.cpg.lst),
      nsets = 12,
      nintersects = 50,
      mainbar.y.label = "Number of intersections, CpGs",
      keep.order = T,
      order.by = "freq",

```

Figure 2 displays the number of intersections (C) for various cell types. The horizontal bar chart at the top shows the number of intersections for each cell type, with values ranging from 225 for CD8nv to 1 for Neu. The dot plot below shows the number of intersections for each cell type across different set sizes (500 to 0).

Cell Type	Number of Intersections (C)
CD8nv	225
Eos	72
NK	60
Bas	37
Mono	34
Treg	29
Bnv	22
CD4mem	22
Bmem	12
CD4nv	13
CD8mem	10
Neu	1

```
dmps.sign.anno.fn <- "~/bio/code/mpip/dex-stim-human-array/output/data/methylation/02_dmp/dmps_fdr01_fc
dmps.sign.anno.df <- fread(dmps.sign.anno.fn)
dmps <- dmps.sign.anno.df$PROBE_ID %>% unique()
sign.pval.dmps.df <- sign.pval.df[CpG_ID %in% dmps, ]

sign.pval.dmps.dex <- sign.pval.dmps.df[Treatment == "dex", CpG_ID]
sign.pval.dmps.veh <- sign.pval.dmps.df[Treatment == "veh", CpG_ID]

intersect.dmps <- intersect(sign.pval.dmps.dex, sign.pval.dmps.veh)
sign.pval.dmps.df <- sign.pval.dmps.df[CpG_ID %in% intersect.dmps,]

print(paste0("Number of unique dex CpGs: ", length(unique(sign.pval.dmps.dex))))

## [1] "Number of unique dex CpGs: 9095"
```

```
print(paste0("Number of unique baseline CpGs: ", length(unique(sign.pval.dmps.veh))))
```

```
## [1] "Number of unique baseline CpGs: 9848"
```

```
print(paste0("Number of inteseacting CpGs: ", length(unique(intersect.dmps))))
```

```
## [1] "Number of inteseacting CpGs: 9094"
```

Kolmogorov-Smirnov test: Do 2 samples follow the same distribution?

```
blood.cell.types      <- unique(sign.pval.delta.mqtl.df$Type)
```

```
test.res.df <- lapply(blood.cell.types, function(i){ # for each blood cell type
  dex.fdr.lst <- sign.pval.dmps.df[Type == i][Treatment == "dex", fdr]
  veh.fdr.lst <- sign.pval.dmps.df[Type == i][Treatment == "veh", fdr]
  test.rslt   <- ks.test(dex.fdr.lst, veh.fdr.lst)

  return(data.frame(Type = i, "p-value" = test.rslt$p.value))
}) %>%
bind_rows()
```

```
## Warning in ks.test(dex.fdr.lst, veh.fdr.lst): p-value will be approximate in the
## presence of ties
```

```
## Warning in ks.test(dex.fdr.lst, veh.fdr.lst): p-value will be approximate in the
## presence of ties
```

```
## Warning in ks.test(dex.fdr.lst, veh.fdr.lst): p-value will be approximate in the
## presence of ties
```

```
## Warning in ks.test(dex.fdr.lst, veh.fdr.lst): p-value will be approximate in the
## presence of ties
```

```
## Warning in ks.test(dex.fdr.lst, veh.fdr.lst): p-value will be approximate in the
## presence of ties
```

```
## Warning in ks.test(dex.fdr.lst, veh.fdr.lst): p-value will be approximate in the
## presence of ties
```

```
## Warning in ks.test(dex.fdr.lst, veh.fdr.lst): p-value will be approximate in the
## presence of ties
```

```
## Warning in ks.test(dex.fdr.lst, veh.fdr.lst): p-value will be approximate in the
## presence of ties
```

```
## Warning in ks.test(dex.fdr.lst, veh.fdr.lst): cannot compute exact p-value with
## ties
```

```
## Warning in ks.test(dex.fdr.lst, veh.fdr.lst): p-value will be approximate in the
## presence of ties
```



```
sign.pval.dmps.df <- left_join(sign.pval.dmps.df, test.res.df)
```

```
## Joining, by = "Type"
```

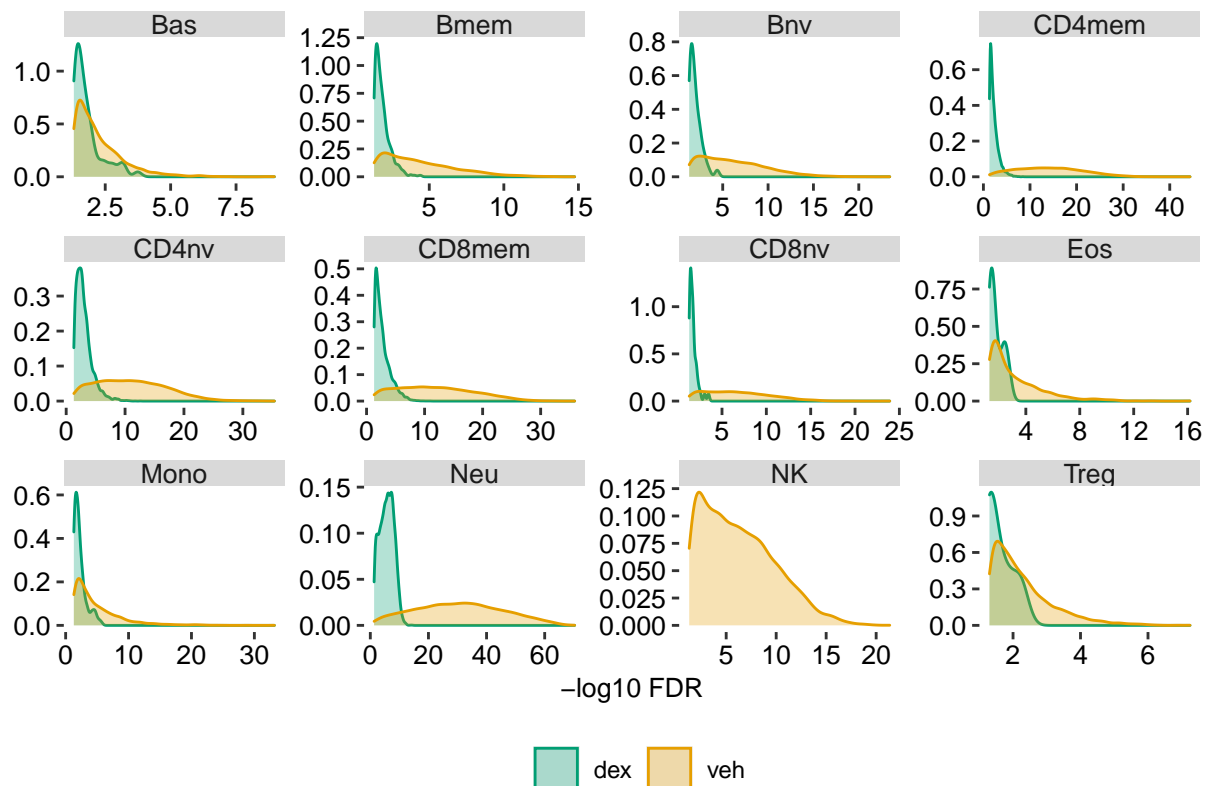
```
ggplot(sign.pval.dmps.df, aes(x = -log10(fdr), fill = Treatment)) +
  geom_density(alpha = 0.3, aes(color = Treatment)) +
  geom_text(data = subset(unique(sign.pval.dmps.df[, .(Treatment, Type, p.value)])),
    aes(x = -Inf, y = -Inf, label = paste0("KS test = ", signif(p.value, 3))),
    hjust = -0.5, vjust = -15, size = 5, color = "black" ) +
  facet_wrap(~ Type, scales = "free", ncol = 4) +
  theme(legend.position = "bottom", # c(.9,.9),
    legend.title = element_blank(),
    panel.grid.major = element_blank(),
    panel.background = element_blank(),
    plot.title = element_text(size = 10, color = "black"),
    axis.title = element_text(size = 10, color = "black"),
    axis.text.x = element_text(size = 10, color = "black"),
    axis.text.y = element_text(size = 10, color = "black"),
    strip.text.x = element_text(size = 10, margin = margin())) +
  labs(title = "Distribution of FDRs of GR-DMPs across blossom cell type", x = "-log10 FDR", y = "") +
  scale_fill_manual(values = cbPalette[2:3]) +
  scale_color_manual(values = cbPalette[2:3])
```

```
## Warning: Groups with fewer than two data points have been dropped.
```

```
## Warning in max(ids, na.rm = TRUE): no non-missing arguments to max; returning
```

```
## -Inf
```

Distribution of FDRs of GR-DMPs across blossom cell type



```
ggplot(sign.pval.dmps.df, aes(x = Type, fill = Treatment)) +
  geom_bar(stat = "count", alpha = 1, position = position_dodge(width = .9)) +
  geom_text(aes(label = scales::percent(stat(count) / length(intersect.dmps), accuracy = 0.1)),
    stat = "count", vjust = -0.1, size = 5, position = position_dodge(width = .9)) +
  geom_text(data = subset(sign.pval.dmps.df[Treatment == "veh"], p.value <= 0.05),
    aes(label = "*"), stat = "count", vjust = 0.15, size = 16) +
  theme(legend.position = "bottom", # c(0.9, 0.9),
    legend.title = element_blank(),
    legend.text = element_text(size = 16),
    panel.grid.major = element_blank(),
    panel.background = element_blank(),
    plot.title = element_text(size = 16),
    axis.title = element_text(size = 16),
    axis.text = element_text(size = 16, colour = "black")) +
  labs(title = "Distribution of GR-DMPs across 12 blood cell types",
    x = "", y = "") +
  scale_fill_manual(values = cbPalette[2:3])
```

