

Multiple Sequence Alignments

A Brief Introduction

EMBL-Australia Masterclass on Protein
Sequence Analysis

Sydney, Australia
Monday 21st October 2013

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Session Goal

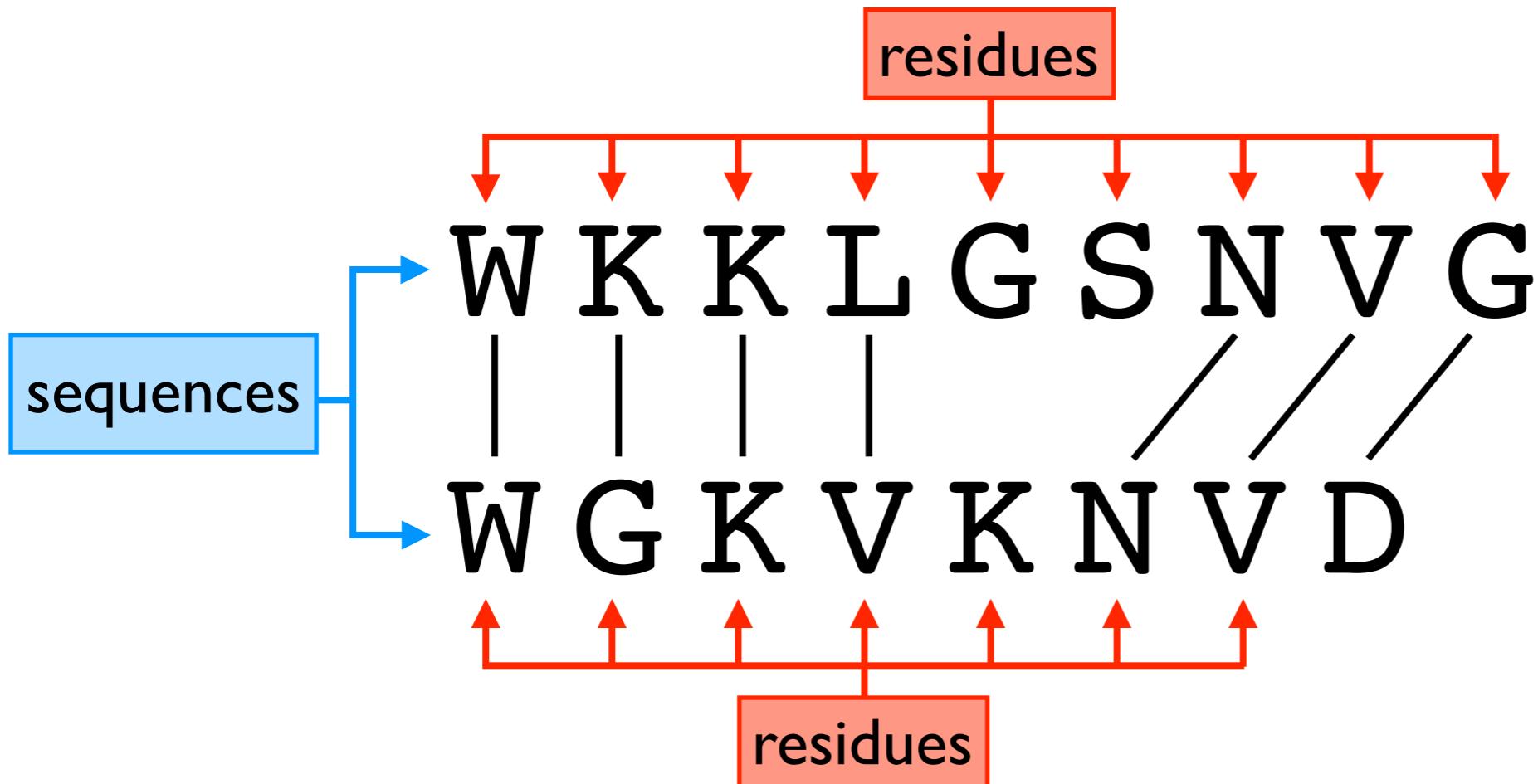
After attending today's session, we hope you will be better able to:

- build higher-quality/more appropriate MSAs for use in your own research/applications
- critically assess the quality of MSAs built by yourself and others

Why a Session on MSAs?

- Required for the development of almost all sequence analysis bioinformatics/tools
- MSAs take practice to interpret (and build) well
- Quality of downstream analysis/tools depends on quality of MSA

"Anatomy" of a Sequence Alignment



Residues:

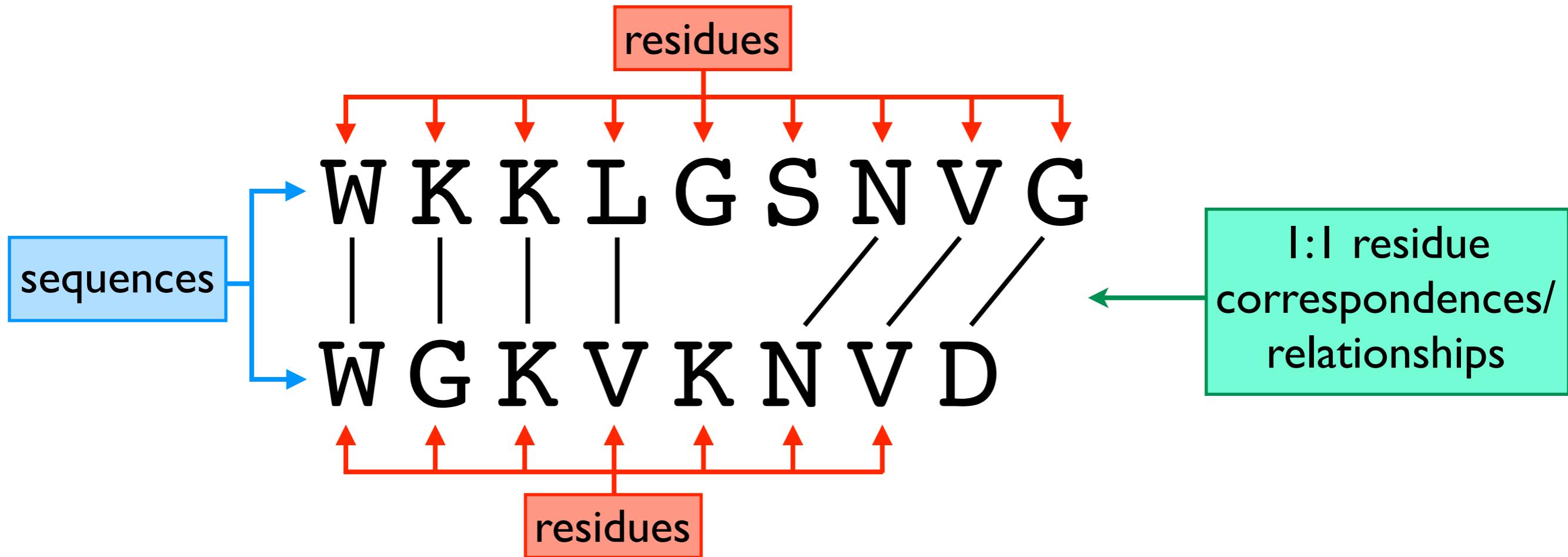
Monomers within a polymer (polypeptide or polynucleotide) chain

Sequences:

List of residues in a polymer chain...

...listed in the same order they occur within the polymer

"Anatomy" of a Sequence Alignment

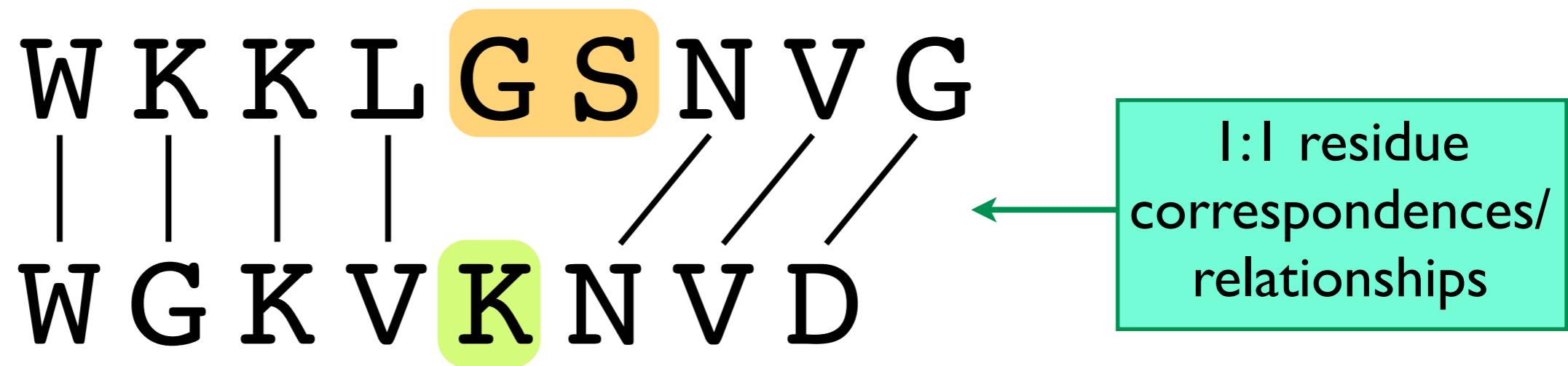


I:I residue correspondences/relationships

Correspondences between

- a single residue in one sequence and
- a single residue in another sequence

"Anatomy" of a Sequence Alignment



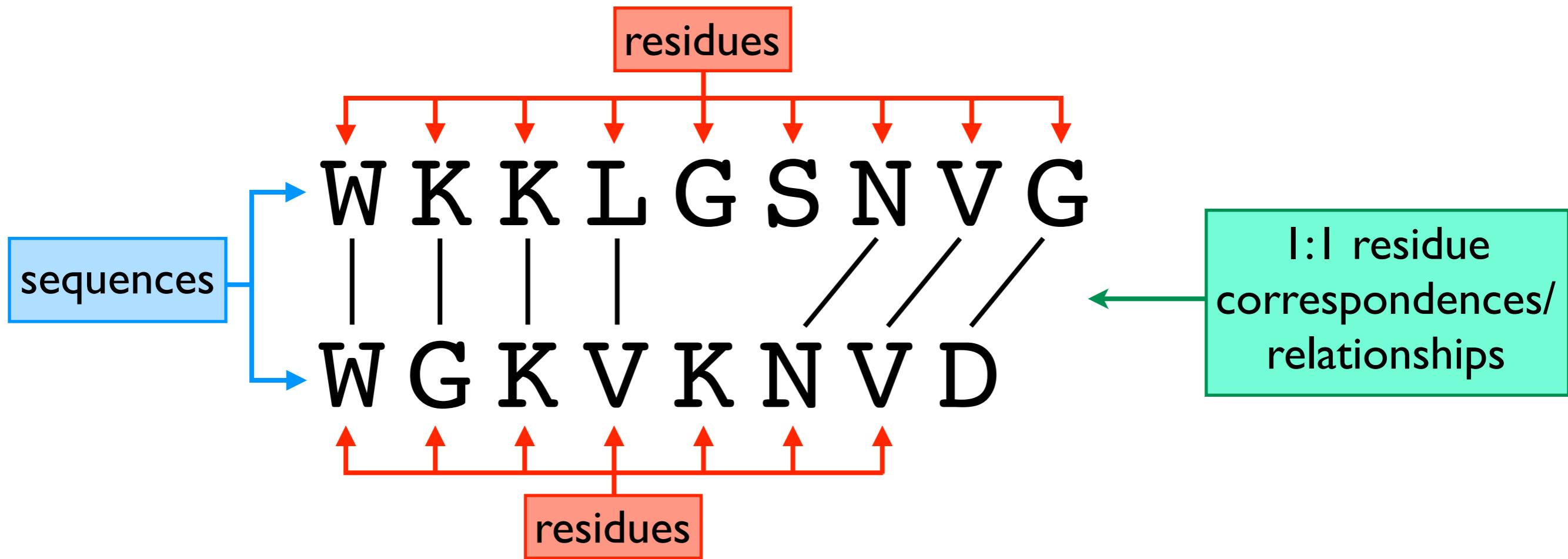
Residue has no equivalent in the top sequence

i.e. no residue in the top sequence has a I:I relationship with this residue

Could perhaps say there is a "I:2" relationship between this residue and these residues

However, alignments focus on I:I relationships

"Anatomy" of a Sequence Alignment

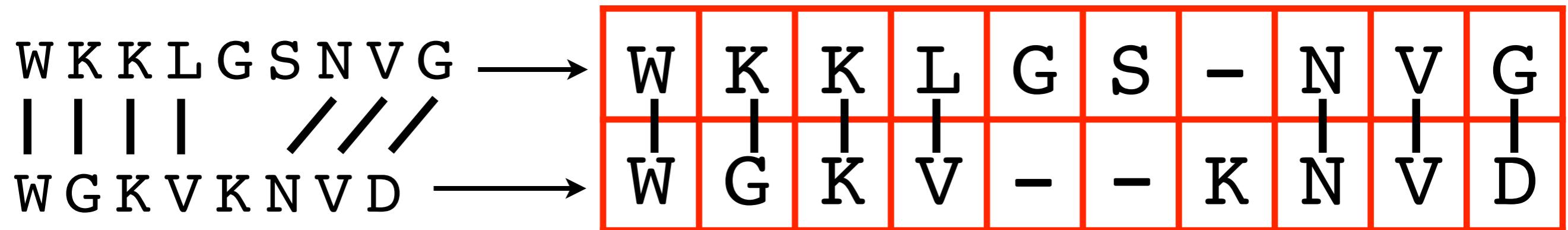


Sequence alignment

A comparison of the residues in two or more sequences...

...describing I:I correspondences/relationships/equivalences
between residues in different sequences

Sequence Alignment Within a Grid



Often represented using a **grid/matrix**:

One sequence per row

Residues in the same column are 'equivalent'

Gap characters (usually "-") indicate that the sequence contains no residues 'equivalent' to other residues in that column

Alternative Interpretations of MSAs (Evolutionary and Structural)

“Equivalence”/similarity of residues

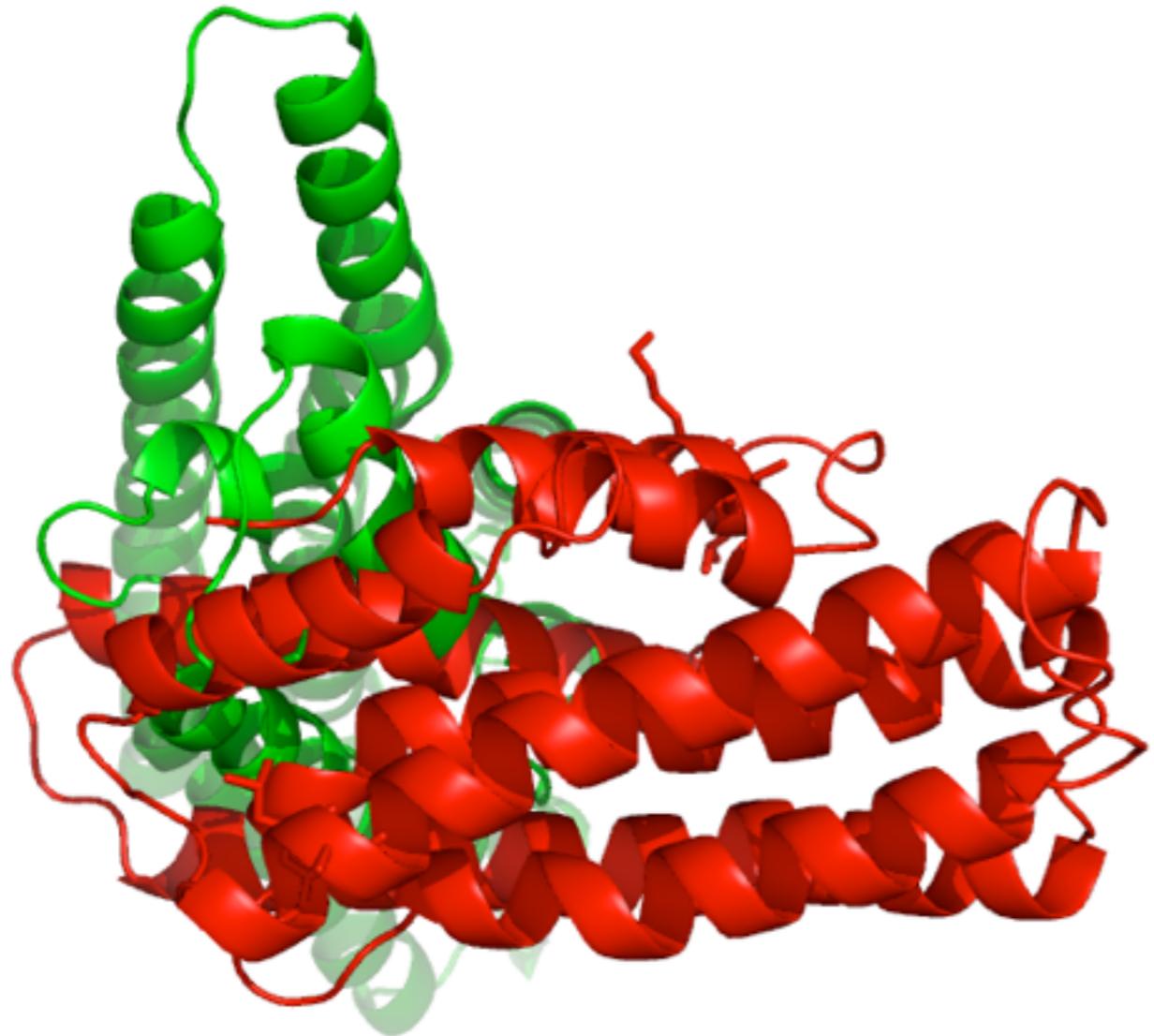
Residues in the same column either:

- Structurally equivalent/similar
- Evolutionary equivalent/related/homologous

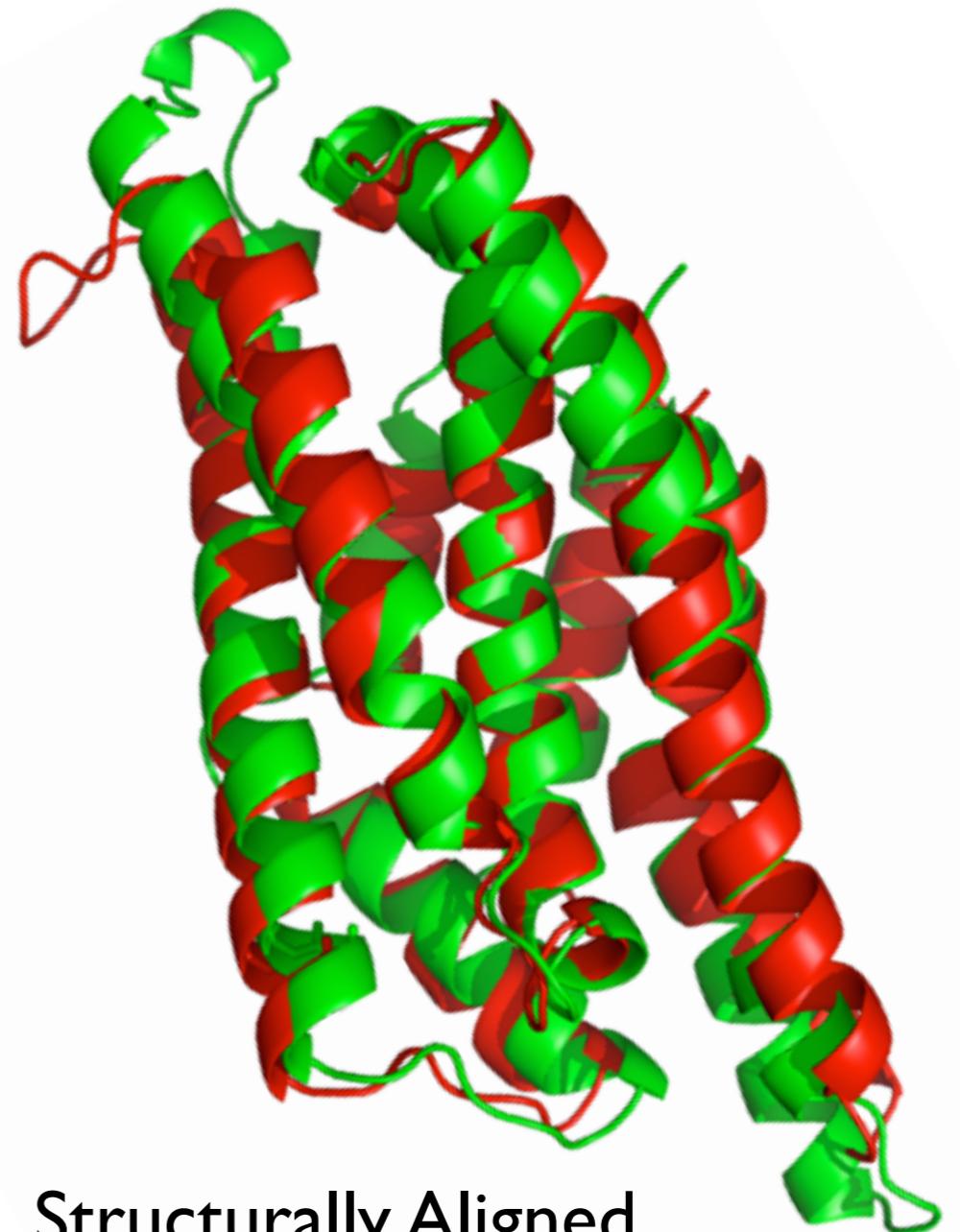
Different applications assume different types of equivalence

Different types of similarity not necessarily equivalent

Structural Similarity



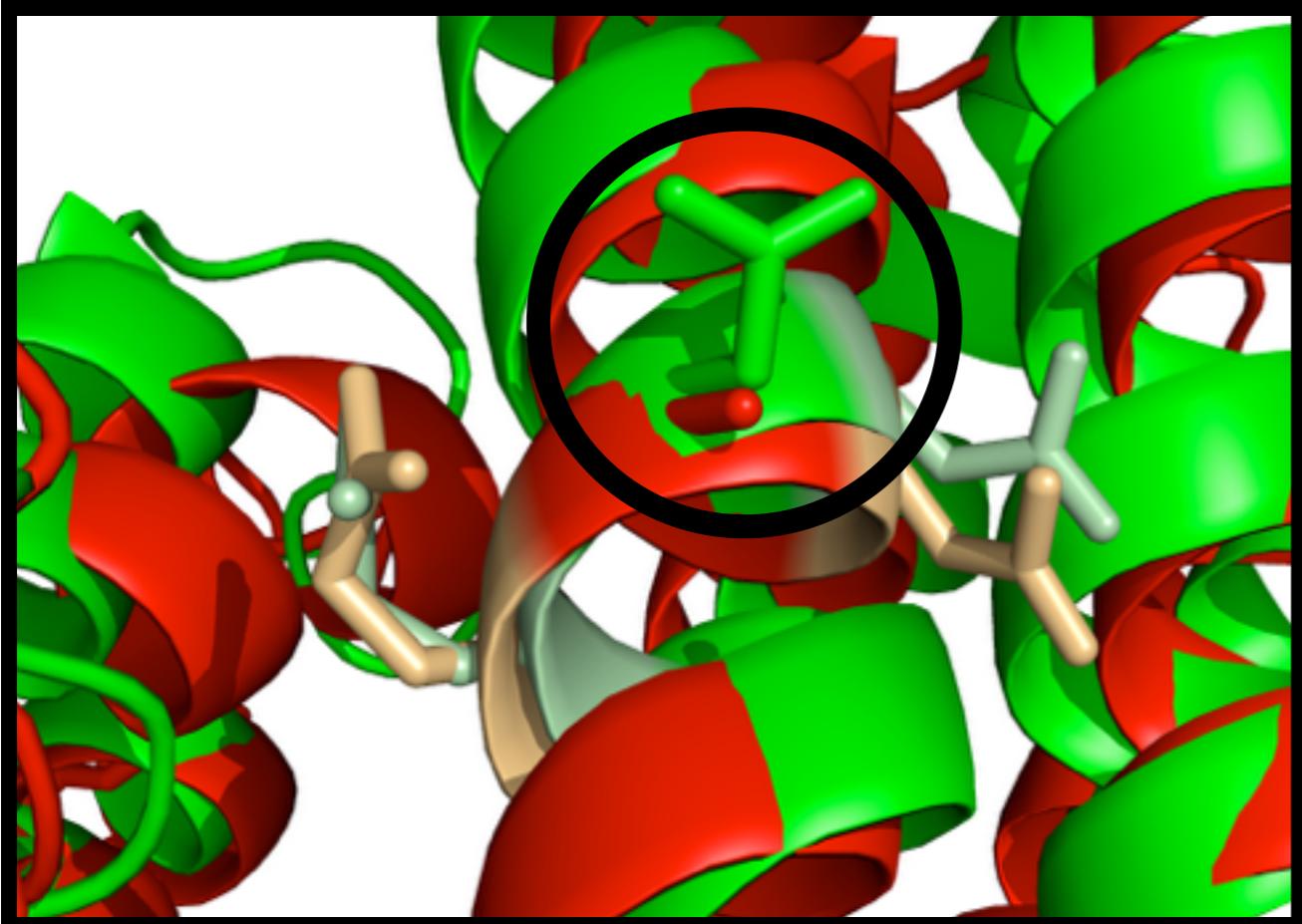
Unaligned



Structurally Aligned

Bacterial toxins **Iji6** and **Ii5p**

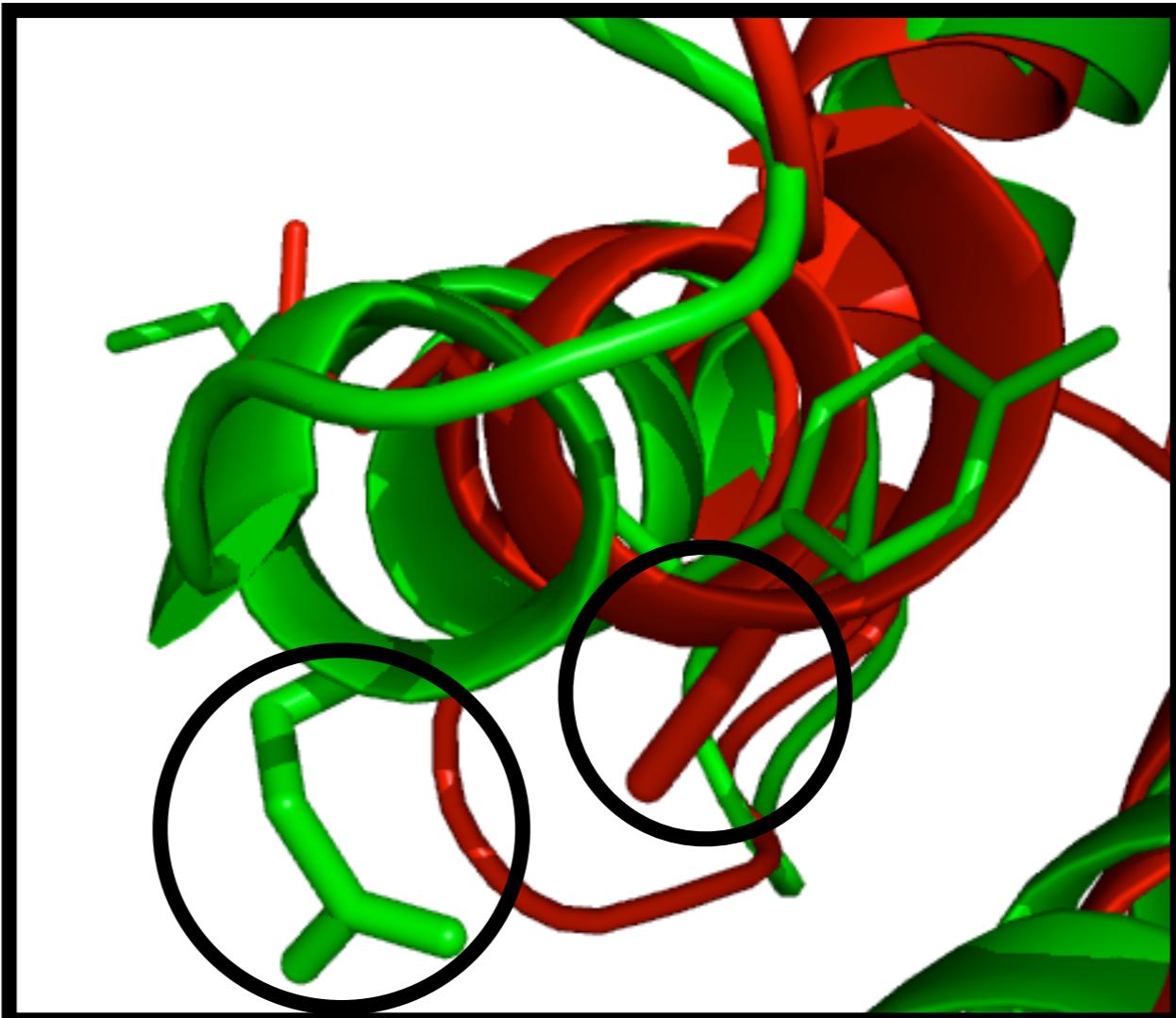
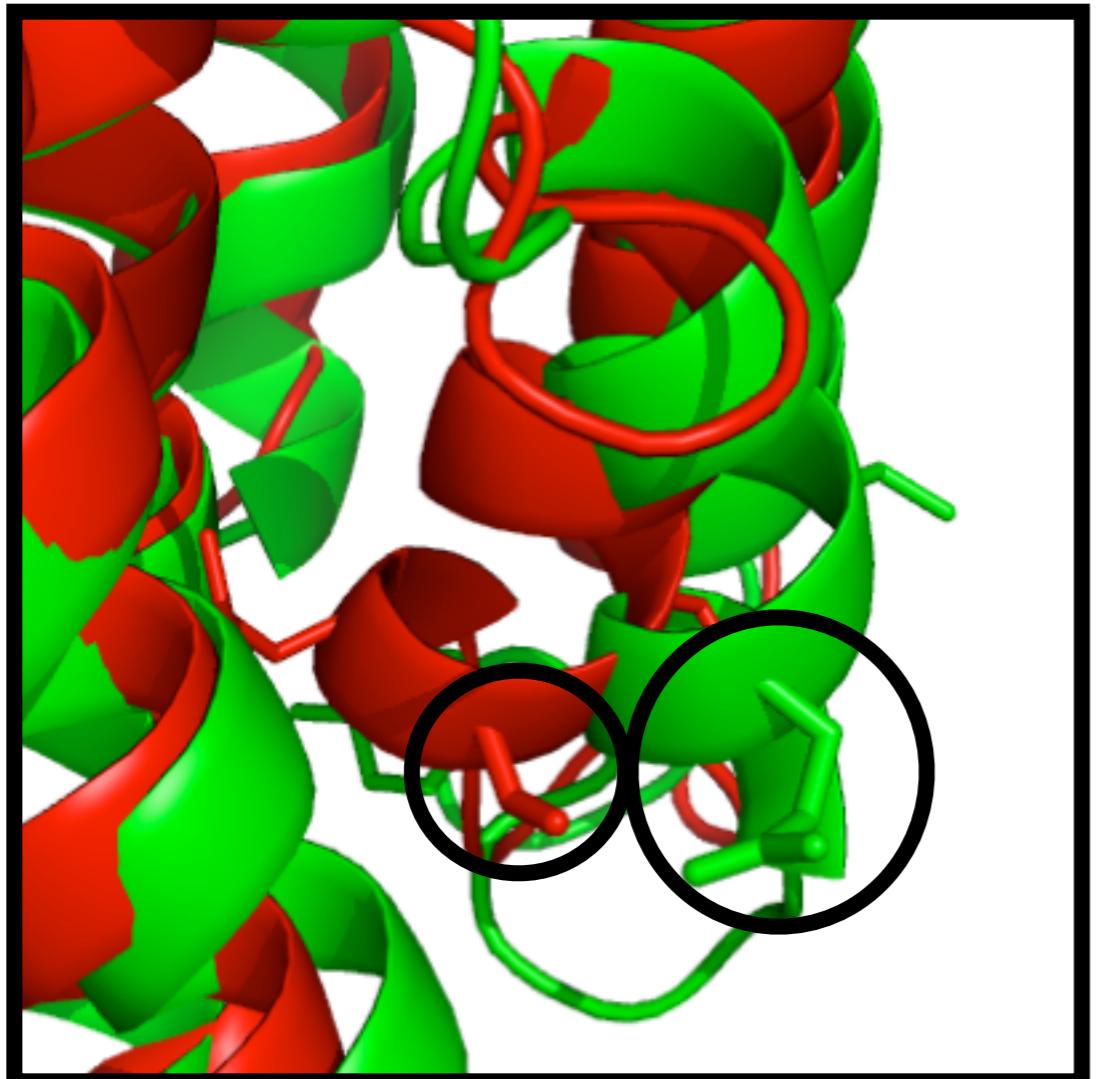
Structural Similarity



68 ELIGL**Q**A**N**IREFNQQVDNF
1111111111111111111111111111
70 ELQGL**Q**NNFEDYVNALNSW

Residues with a similar structural context may lie almost on top of each other within a structural alignment. Clearly, the dark green and red side chains have more similar structural contexts than they do with the adjacent light-coloured side chains

Structural Similarity



Chain 1: 16 KVGSLIGKR---I**LSELWGIIFPSGST**

1111111111 111111111111 111

Chain 2: 16 VVGVPFAGALTSFYQSFLNTIWP-SDA

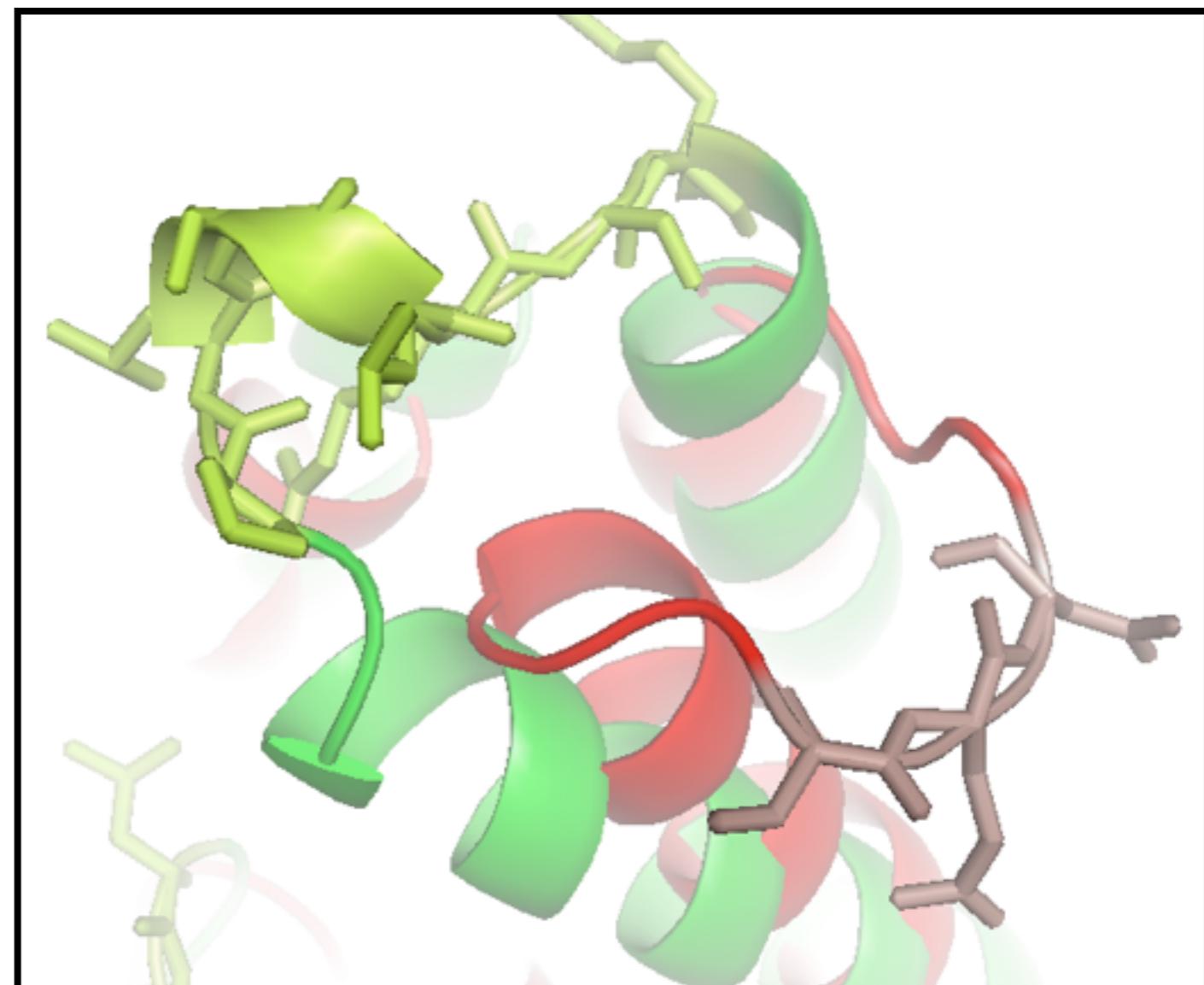
Structural equivalence

Some regions of the structures **do not have structurally equivalent residues** in the other structure

Alignment gaps are a sure sign of such residues

Placing such residues in the same column as residues from other sequences is a **misalignment** - to be avoided!

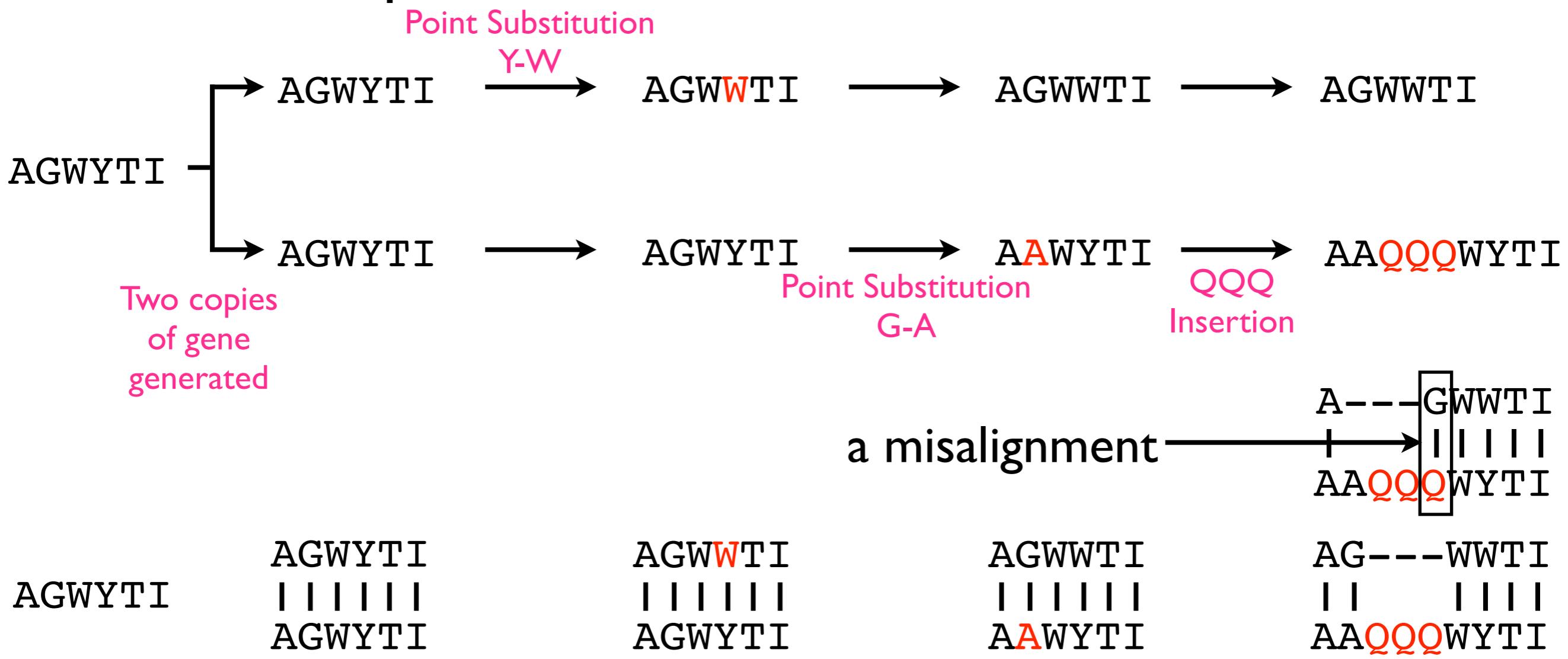
1i5p:	DNFLNPTQN	-----	PVPLSITSSVN
	111111		111111111111
1ji6:	NSWKKTPLSLRSK	RSQDRIRELFS	



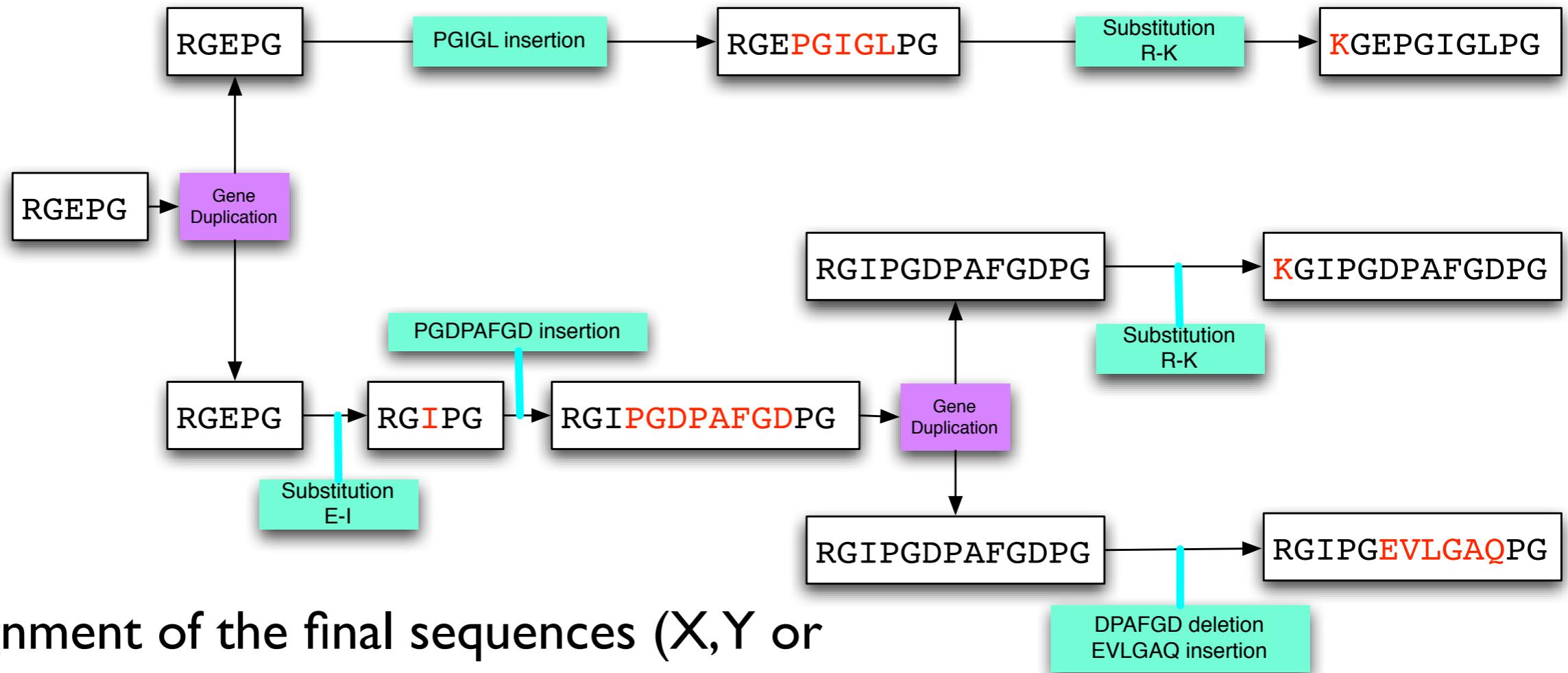
Evolutionary "Equivalence"

Residues are "evolutionarily equivalent" when:

- they are derived from the same residue in an ancestral sequence
- the only mutations experienced during divergence from this ancestral residue were **point substitutions**



Quiz - Evolutionary Interpretation of Alignments



Which alignment of the final sequences (X, Y or Z) only places residues in the same column if they are related by substitution events?

X

Y

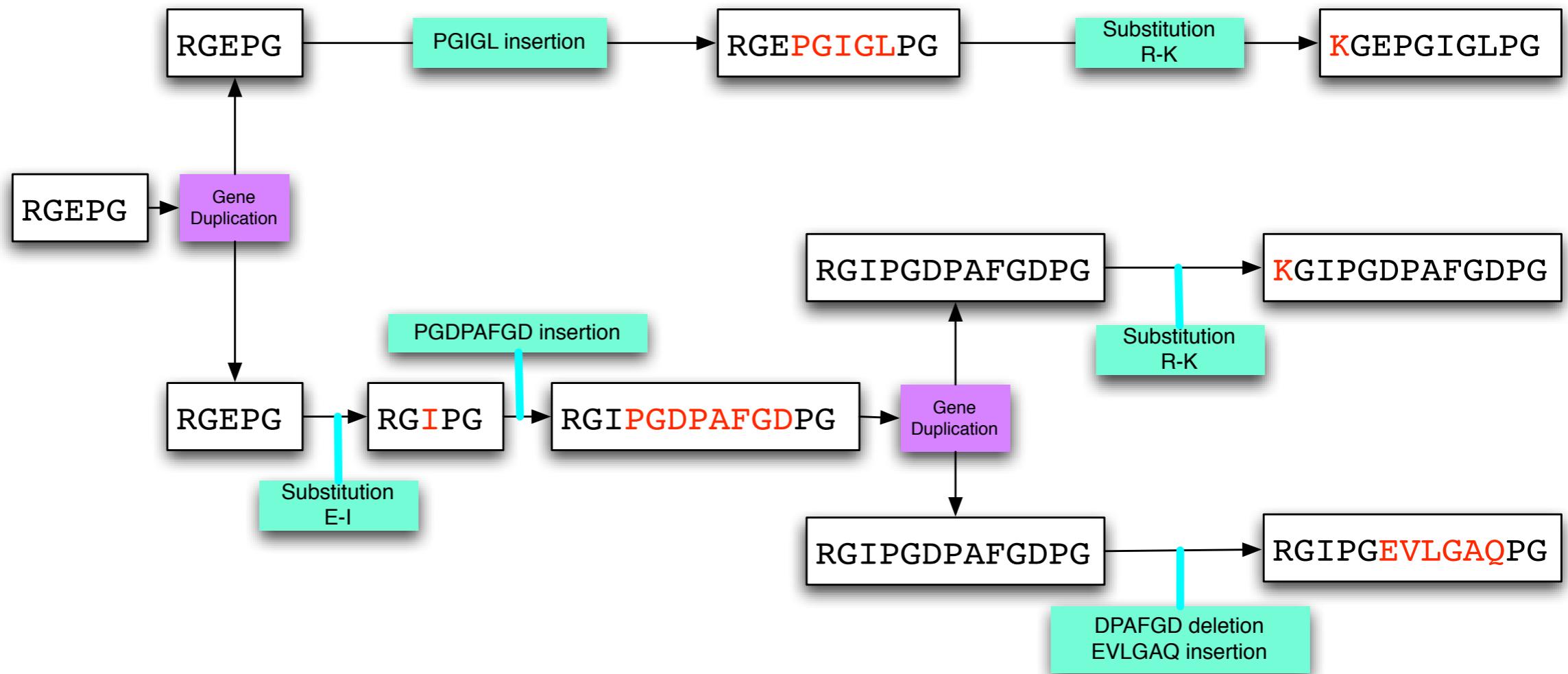
Z

KGE~~P~~G---IGLPG
KGIPGDPAFGDPG
RGIPGEVLGAQPG

KGE~~P~~G-----IGL-----PG
KGIPG-----DPAFGDPG
RGIPGEVLGAQ-----PG

KGE-----PGIGL-----PG
KGIPG-----DPAFGDPG
RGIPGEVLGAQ-----PG

Quiz - Evolutionary Interpretation of Alignments



"True" alignment given history described above

KGE-----PGIGL-----PG
 KGIPG-----DPAFGDPG
 RGIPGEVLGAQ-----PG

PRANK

RGIPGEVLGAQPG
 KGIPGDPAFGDPG
 ---KGE~~P~~GIGLPG

Quiz - Evolutionary Interpretation of Alignments

CLUSTALX

K---GEPGIGLPG
KGIPGDPAFGDPG
RGIPGEVLGAQPG

MAFFT

KGEPG---IGLPG
KGIPGDPAFGDPG
RGIPGEVLGAQPG

PRANK

RGIPGEVLGAQPG
KGIPGDPAFGDPG
---KGEPGIGLPG

Different automatic MSA software gives different results

All are different from the "true" alignment (assuming the scenario of transformation on the previous slide is true)...

... because that scenario is very unlikely under the models of evolutionary transformation incorporated within these tools

X

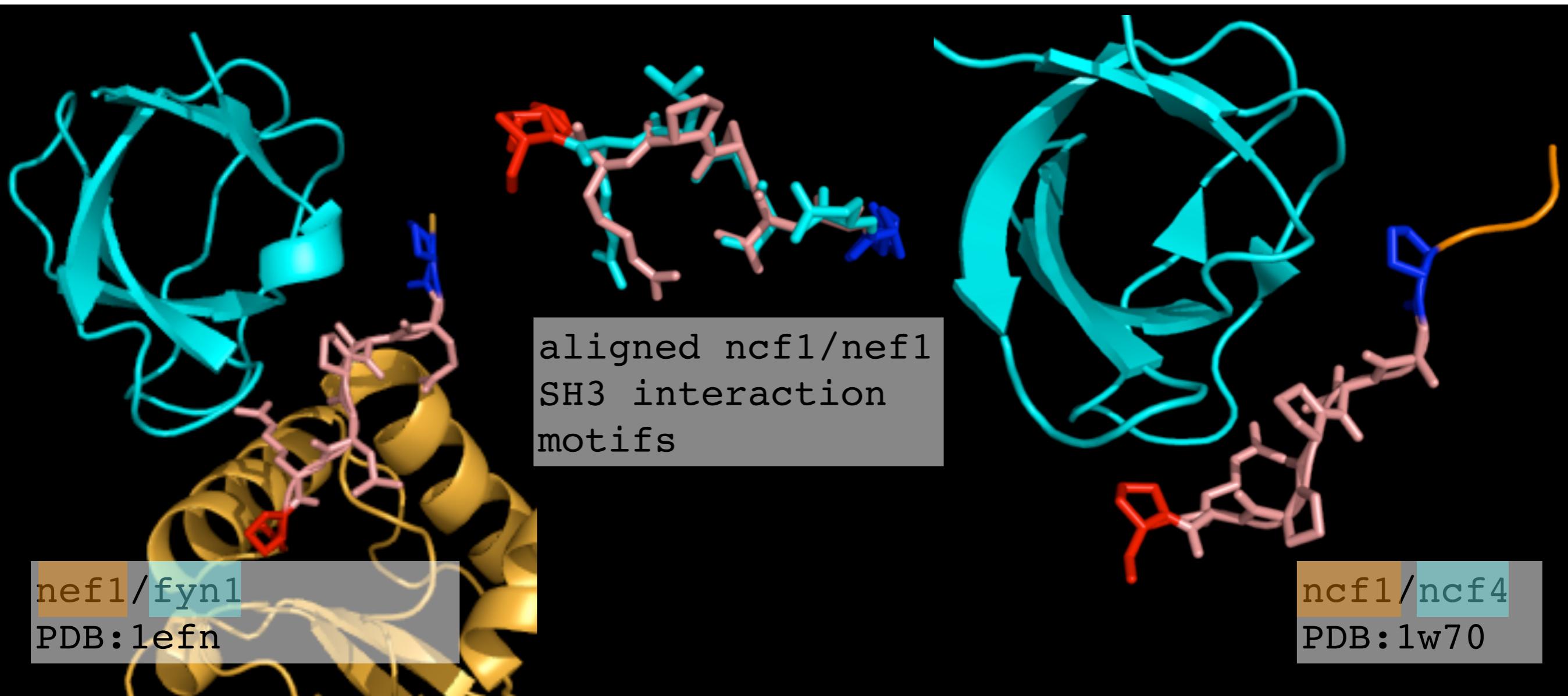
Y

Z

KGEPG---IGLPG KGEPG-----IGL-----PG KGE-----PGIGL-----PG
KGIPGDPAFGDPG KGIPG-----DPAFGDPG KGIPG-----DPAFGDPG
RGIPGEVLGAQPG RGIPGEVLGAQ-----PG RGIPGEVLGAQ-----PG

Non-Equivalence of Evolutionary and Structural Alignments

Structural equivalence without evolutionary equivalence
Structural alignment of SH3-interaction motifs from nef and ncf1



Building MSAs

Build an Automatic MSA

Load sequences into JalView, and with a few clicks you can automatically align a set of sequences

Or run an MSA tool at EBI

The screenshot shows the EBI website interface for the MUSCLE tool. At the top, there's a navigation bar with links for Databases, Tools, Research, Training, Industry, About Us, Help, and Site Index. Below the navigation is a search bar labeled "Enter Text Here" with a "Find" button. A "Help | Feedback" link is also present.

The main content area is titled "MUSCLE - Multiple Sequence Alignment". It includes a brief description: "MUSCLE stands for MULTIPLE Sequence Comparison by Log-Expectation. MUSCLE is claimed to achieve both better average accuracy and better speed than ClustalW2 or T-Coffee, depending on the chosen options." Below this is a section titled "Use this tool" with three steps:

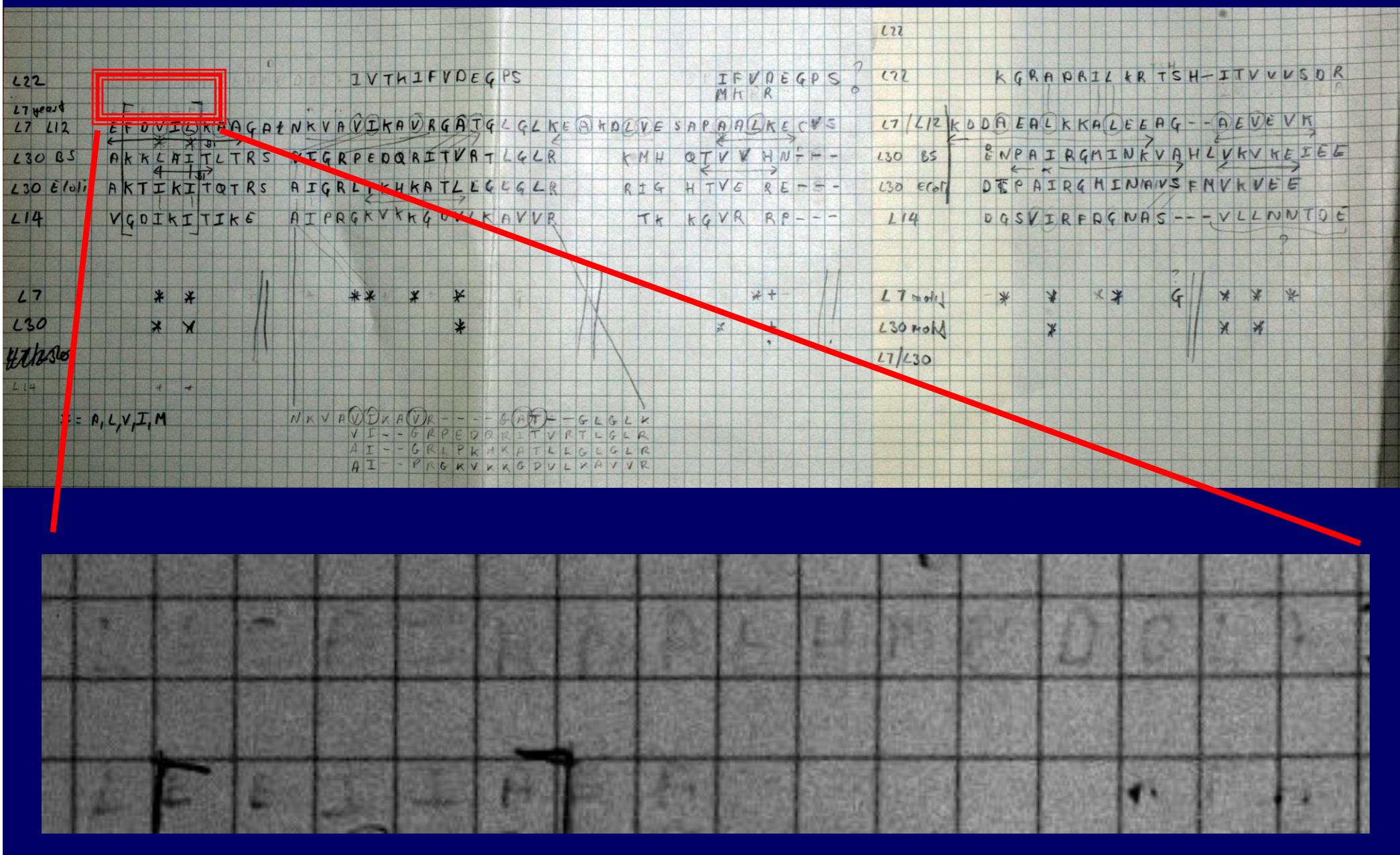
- STEP 1 - Enter your input sequences:** A text input field with placeholder text "Enter or paste a set of sequences in any supported format:" and a "Browse..." button for file upload.
- STEP 2 - Set your Parameters:** A dropdown menu for "OUTPUT FORMAT" set to "Pearson/FASTA". A note states: "The default settings will fulfill the needs of most users and, for that reason, are not visible." A "More options..." link is available.
- STEP 3 - Submit your job:** A checkbox for "Be notified by email (Tick this box if you want to be notified by email when the results are available)" and a red "Submit" button.

On the left sidebar, there's a "MUSCLE" logo with the text "multiple sequence alignment method with reduced time and space complexity". Other links include "Help", "Jalview", "Programmatic Access", "Download", "Related Applications" (with "Pairwise Sequence Alignment", "Multiple Sequence Alignment", and "Phylogeny" listed), and "MUSCLE related literature" (with a "Search for MUSCLE related literature in Medline..." link).

<http://www.ebi.ac.uk/Tools/msa/muscle/>

Build an MSA "Manually"

Multiple Sequence Alignment and Visualisation (1984/5)



Courtesy of Geoff Barton, Dundee

Aidan Budd, EMBL Heidelberg

JalView Demo and Exercises

- Loading sequences
- Changing the way the sequences are displayed
- Manual editing of alignments
- Adding/removing sequences to an alignment
- Exporting sequences/alignments from JalView for use in another application

JalView Demo and Exercises

- a process of pattern-matching/identification
- we prefer alignments where many columns contain few differences/conservative changes
- more divergent sequences are harder to align than more similar sequences
 - for divergent sequences, there are many alternative alignments are similarly good/bad
 - for rather similar sequences, there is usually one/a few alignments we feel are clearly much better than the others
- Longer sequences take longer to align than short ones

JalView Demo and Exercises

- More sequences take longer to align than fewer sequences
- Repeats cause problems
- Different positions evolve differently
- At some level, the problem is "simple"
 - we just have to choose the right place to put the gaps!

another quiz on interpreting MSAs...

Quiz - Numbers of Insertions

	A	V	S	C	L	W	G	K	V	--	N	S	D	E	V	G	G	E	A	L	G	R	L	
<i>mouseHemoglobinB1</i>																								
<i>mouseHemoglobinBZ</i>	A	I	T	S	I	W	D	K	V	--	D	L	E	K	V	G	G	E	T	L	G	R	L	
<i>mouseHemoglobinE</i>	L	I	N	G	L	W	S	K	V	--	N	V	E	E	V	G	G	E	A	L	G	R	L	
<i>humanHemoglobinAZ</i>	I	I	V	S	M	W	A	K	I	S	T	Q	A	D	T	I	G	T	E	T	L	E	R	L
<i>mouseHemoglobinAZ</i>	I	I	M	S	M	W	E	K	M	A	A	Q	A	E	P	I	G	T	E	T	L	E	R	L
<i>humanHemoglobinG2</i>	T	I	T	S	L	W	G	K	V	--	N	V	E	D	A	G	G	E	T	L	G	R	L	
<i>humanHemoglobinAT</i>	L	V	R	A	L	W	K	K	L	G	S	N	V	G	V	T	T	E	A	L	E	R	T	
<i>humanHemoglobinA</i>	N	V	K	A	A	W	G	K	V	G	A	H	A	G	E	Y	G	A	E	A	L	E	R	M
<i>humanHemoglobinB</i>	A	V	T	A	L	W	G	K	V	--	N	V	D	E	V	G	G	E	A	L	G	R	L	

The **minimum** number of insertion events required to account for the section of haemoglobin alignment shown above is?

- (a) 2
- (b) 1
- (c) 0
- (d) 3

Quiz - Numbers of Insertions

<i>mouseHemoglobinB1</i>	A	V	S	C	L	W	G	K	V	--	N	S	D	E	V	G	G	E	A	L	G	R
<i>mouseHemoglobinBZ</i>	A	I	T	S	I	W	D	K	V	--	D	L	E	K	V	G	G	E	T	L	G	R
<i>mouseHemoglobinE</i>	L	I	N	G	L	W	S	K	V	--	N	V	E	E	V	G	G	E	A	L	G	R
<i>humanHemoglobinAZ</i>	I	I	V	S	M	W	A	K	I	S	T	Q	A	D	T	I	G	T	E	T	L	E
<i>mouseHemoglobinAZ</i>	I	I	M	S	M	W	E	K	M	A	A	Q	A	E	P	I	G	T	E	T	L	E
<i>humanHemoglobinG2</i>	T	I	T	S	L	W	G	K	V	--	N	V	E	D	A	G	G	E	T	L	G	R
<i>humanHemoglobinAT</i>	L	V	R	A	L	W	K	K	L	G	S	N	V	G	V	T	T	E	A	L	E	R
<i>humanHemoglobinA</i>	N	V	K	A	A	W	G	K	V	G	A	H	A	G	E	Y	G	A	E	A	L	E
<i>humanHemoglobinB</i>	A	V	T	A	L	W	G	K	V	--	N	V	D	E	V	G	G	E	A	L	G	R

The **minimum** number of insertion events required to account for the section of haemoglobin alignment shown above is?

If all sequences are the same length, we can explain their diversity without inferring ANY insertions or deletions

If an alignment contains sequences that are all either length x or y, then we can explain their diversity by inferring just one insertion or deletion

Quiz - Numbers of Insertions

<i>mouseHemoglobinB1</i>	A V S C L W G K V -- N S D E V G G E A L G R L
<i>mouseHemoglobinBZ</i>	A I T S I W D K V -- D L E K V G G E T L G R L
<i>mouseHemoglobinE</i>	L I N G L W S K V -- N V E E V G G E A L G R L
<i>humanHemoglobinAZ</i>	I I V S M W A K I S T Q A D T I G T E T L E R L
<i>mouseHemoglobinAZ</i>	I I M S M W E K M A A Q A E P I G T E T L E R L
<i>humanHemoglobinG2</i>	T I T S L W G K V -- N V E D A G G E T L G R L
<i>humanHemoglobinAT</i>	L V R A L W K K L G S N V G V Y T T E A L E R T
<i>humanHemoglobinA</i>	N V K A A W G K V G A H A G E Y G A E A L E R M
<i>humanHemoglobinB</i>	A V T A L W G K V -- N V D E V G G E A L G R L

The **minimum** number of insertion events required to account for the section of haemoglobin alignment shown above is?

We can **ALWAYS** explain observed sequence length diversity with:

- 0 insertions (all length variation due to deletion)
- 0 deletions (all length variation due to insertion)
- a combination of insertions and deletions

Perhaps we should instead focus on inferring the **most likely** scenario?

(Although if this is not particularly relevant for our analysis, perhaps we should focus instead on something completely different!)