# Class 8 Mini-Project: Unsupervised Learning Analysis of Human Breast Cancer Cells

## Aishwarya Ramesh

#### **Exploratory data analysis**

#### **Preparing the Data**

First, getting the data:

```
# Save your input data file into your Project directory
fna.data <- "WisconsinCancer.csv"

# Complete the following code to input the data and store as wisc.df
wisc.df <- read.csv(fna.data, row.names=1)</pre>
```

Next, taking a quick look at the data:

```
head(wisc.df)
```

	diagnosis radius	s_mean	texture_mean	perimeter_mean	area_mean	
842302	М	17.99	10.38	122.80	1001.0	
842517	M	20.57	17.77	132.90	1326.0	
84300903	M	19.69	21.25	130.00	1203.0	
84348301	M	11.42	20.38	77.58	386.1	
84358402	M	20.29	14.34	135.10	1297.0	
843786	М	12.45	15.70	82.57	477.1	
	${\tt smoothness\_mean}$	compa	ctness_mean c	oncavity_mean co	oncave.points_mea	ın
842302	0.11840		0.27760	0.3001	0.1471	LO
842517	0.08474		0.07864	0.0869	0.0701	۱7
84300903	0.10960		0.15990	0.1974	0.1279	90
84348301	0.14250		0.28390	0.2414	0.1052	20
84358402	0.10030		0.13280	0.1980	0.1043	30

843786	0.1278	0	.17000	0.1578	3	0.08089
	symmetry_mean	$fractal\_dime$	nsion_mean	radius_se	texture_se	perimeter_se
842302	0.2419		0.07871	1.0950	0.9053	8.589
842517	0.1812		0.05667	0.5435	0.7339	3.398
84300903	0.2069		0.05999	0.7456	0.7869	4.585
84348301	0.2597		0.09744	0.4956	1.1560	3.445
84358402	0.1809		0.05883	0.7572	0.7813	5.438
843786	0.2087		0.07613	0.3345	0.8902	2.217
	area_se smooth	ness_se comp	actness_se	concavity	_se concave	.points_se
842302	153.40 0	.006399	0.04904	0.05	373	0.01587
842517	74.08 0	.005225	0.01308	0.018	360	0.01340
84300903	94.03 0	.006150	0.04006	0.038	332	0.02058
84348301	27.23 0	.009110	0.07458	0.056	661	0.01867
84358402	94.44 0	.011490	0.02461	0.056	688	0.01885
843786	27.19 0	.007510	0.03345	0.036	672	0.01137
	symmetry_se fr	actal_dimens	ion_se rad:	ius_worst 1	texture_wor:	st
842302	0.03003	0.	006193	25.38	17.3	33
842517	0.01389	0.	003532	24.99	23.4	<del>1</del> 1
84300903	0.02250	0.	004571	23.57	25.	53
84348301	0.05963	0.	009208	14.91	26.	50
84358402	0.01756	0.	005115	22.54	16.6	67
843786	0.02165	0.	005082	15.47	23.	75
	perimeter_wors	t area_worst	smoothness	s_worst co	mpactness_w	orst
842302	184.6	2019.0	1	0.1622	0.6	6656
842517	158.8	1956.0	1	0.1238	0.3	1866
84300903	152.5	1709.0	1	0.1444		1245
84348301	98.8	7 567.7	•	0.2098	0.8	3663
84358402	152.2	1575.0	)	0.1374	0.2	2050
843786	103.4	741.6	;	0.1791	0.!	5249
	concavity_wors	t concave.po	ints_worst	symmetry_	worst	
842302	0.711		0.2654	0	. 4601	
842517	0.241	5	0.1860		. 2750	
84300903	0.450	4	0.2430	0	.3613	
84348301	0.686		0.2575		. 6638	
84358402	0.400		0.1625	0	. 2364	
843786	0.535		0.1741	0	. 3985	
	fractal_dimens	<del>-</del>				
842302		0.11890				
842517		0.08902				
84300903		0.08758				
84348301		0.17300				
84358402		0.07678				
843786		0.12440				

Excluding the wisc.df\$diagnosis column in our analysis:

```
# We can use -1 here to remove the first column wisc.data <- wisc.df[,-1]
```

Checking whether it removed:

```
head(wisc.data)
```

	radius_mean	texture_mear	n perimet	er_mean	area_mea	an smoothi	ness_mean
842302	17.99	10.38	_	122.80	1001		0.11840
842517	20.57	17.77	7	132.90	1326	. 0	0.08474
84300903	19.69	21.25	5	130.00	1203	. 0	0.10960
84348301	11.42	20.38	3	77.58	386	. 1	0.14250
84358402	20.29	14.34	1	135.10	1297	. 0	0.10030
843786	12.45	15.70	)	82.57	477	. 1	0.12780
	compactness_	mean concavi	ity_mean	concave.	points_r	nean symme	etry_mean
842302	0.2	7760	0.3001		0.14	1710	0.2419
842517	0.0	7864	0.0869		0.0	7017	0.1812
84300903	0.1	5990	0.1974		0.12	2790	0.2069
84348301	0.2	8390	0.2414		0.10	0520	0.2597
84358402	0.1	3280	0.1980		0.10	0430	0.1809
843786	0.1	7000	0.1578		0.08	3089	0.2087
	fractal_dime	nsion_mean n	radius_se	texture	e_se per:	imeter_se	area_se
842302		0.07871	1.0950	0.9	9053	8.589	153.40
842517		0.05667	0.5435	0.7	7339	3.398	74.08
84300903		0.05999	0.7456	0.7	7869	4.585	94.03
84348301		0.09744	0.4956	1.1	1560	3.445	27.23
84358402		0.05883	0.7572	0.7	7813	5.438	94.44
843786		0.07613	0.3345	0.8	3902	2.217	27.19
	smoothness_s	e compactnes	ss_se con	cavity_s	se conca	re.points	_se
842302	0.00639		)4904	0.0537		0.01	
842517	0.00522	5 0.0	1308	0.0186	60	0.013	340
84300903	0.00615	0.0	)4006	0.0383		0.020	058
84348301	0.00911	0.0	7458	0.0566	51	0.018	367
84358402	0.01149	0.0	)2461	0.0568	38	0.018	385
843786	0.00751		3345	0.0367		0.013	
	symmetry_se	fractal_dime	_	radius_	worst to	exture_wor	rst
842302	0.03003		0.006193		25.38	17	. 33
842517	0.01389		0.003532		24.99		. 41
84300903	0.02250		0.004571		23.57		. 53
84348301	0.05963		0.009208		14.91	26	.50

84358402	0.01756	0.0	005115	22.5	54	16.67
843786	0.02165	0.0	005082	15.4	<u>1</u> 7	23.75
	perimeter_worst	area_worst	smoothness	s_worst	compactne	ess_worst
842302	184.60	2019.0		0.1622		0.6656
842517	158.80	1956.0		0.1238		0.1866
84300903	152.50	1709.0		0.1444		0.4245
84348301	98.87	567.7		0.2098		0.8663
84358402	152.20	1575.0		0.1374		0.2050
843786	103.40	741.6		0.1791		0.5249
	concavity_worst	concave.poi	ints_worst	symmetr	ry_worst	
842302	0.7119		0.2654		0.4601	
842517	0.2416		0.1860		0.2750	
84300903	0.4504		0.2430		0.3613	
84348301	0.6869		0.2575		0.6638	
84358402	0.4000		0.1625		0.2364	
843786	0.5355		0.1741		0.3985	
	fractal_dimension	on_worst				
842302		0.11890				
842517		0.08902				
84300903		0.08758				
84348301		0.17300				
84358402		0.07678				
843786		0.12440				

Creating diagnosis vector for future comparison:

```
# Create diagnosis vector for later
diagnosis <- factor(wisc.df[,1])
diagnosis</pre>
```

#### **Exploratory data analysis**

Q1. How many observations are in this dataset?

Finding dimensions of data:

```
dim(wisc.data)
[1] 569 30
```

This dataset has 569 observations.

Q2. How many of the observations have a malignant diagnosis?

```
table(diagnosis)
diagnosis
B M
357 212
```

There are 212 observations with a malignant diagnosis.

Q3. How many variables/features in the data are suffixed with \_mean?

```
wisc.columns <- colnames(wisc.data)
suffixed_list <- grep('_mean', wisc.columns)
length(suffixed_list)</pre>
```

[1] 10

## Principal component analysis

## Performing PCA

Checking whether data needs to be scaled:

# Check column means and standard deviations
colMeans(wisc.data)

radius_mean	texture_mean	perimeter_mean
1.412729e+01	1.928965e+01	9.196903e+01
area_mean	${\tt smoothness\_mean}$	compactness_mean
6.548891e+02	9.636028e-02	1.043410e-01
concavity_mean	concave.points_mean	symmetry_mean
8.879932e-02	4.891915e-02	1.811619e-01
fractal_dimension_mean	radius_se	texture_se
6.279761e-02	4.051721e-01	1.216853e+00
perimeter_se	area_se	smoothness_se
2.866059e+00	4.033708e+01	7.040979e-03
compactness_se	concavity_se	concave.points_se
2.547814e-02	3.189372e-02	1.179614e-02
symmetry_se	fractal_dimension_se	radius_worst
2.054230e-02	3.794904e-03	1.626919e+01
texture_worst	perimeter_worst	area_worst
2.567722e+01	1.072612e+02	8.805831e+02
smoothness_worst	${\tt compactness\_worst}$	concavity_worst
1.323686e-01	2.542650e-01	2.721885e-01
concave.points_worst	symmetry_worst	${\tt fractal\_dimension\_worst}$
1.146062e-01	2.900756e-01	8.394582e-02

apply(wisc.data,2,sd)

radius_mean	texture_mean	perimeter_mean
3.524049e+00	4.301036e+00	2.429898e+01
area_mean	smoothness_mean	compactness_mean
3.519141e+02	1.406413e-02	5.281276e-02
${\tt concavity\_mean}$	concave.points_mean	symmetry_mean
7.971981e-02	3.880284e-02	2.741428e-02
fractal_dimension_mean	radius_se	texture_se
7.060363e-03	2.773127e-01	5.516484e-01
perimeter_se	area_se	smoothness_se

3.002518e-03	4.549101e+01	2.021855e+00
concave.points_se	concavity_se	compactness_se
6.170285e-03	3.018606e-02	1.790818e-02
radius_worst	fractal_dimension_se	symmetry_se
4.833242e+00	2.646071e-03	8.266372e-03
area_worst	perimeter_worst	texture_worst
5.693570e+02	3.360254e+01	6.146258e+00
concavity_worst	compactness_worst	smoothness_worst
2.086243e-01	1.573365e-01	2.283243e-02
${\tt fractal\_dimension\_worst}$	symmetry_worst	concave.points_worst
1.806127e-02	6.186747e-02	6.573234e-02

There seems to be significantly different variances in the variance, so we must scale the data within PCA.

```
# Perform PCA on wisc.data by completing the following code
wisc.pr <- prcomp(wisc.data, scale=TRUE, center=TRUE)</pre>
```

To get summary of principal components, we use summary() and we can also use plot() to get visual representation of proportion of variance described by each PC.

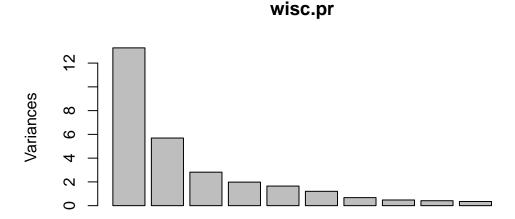
```
summary(wisc.pr)
```

#### Importance of components:

```
PC2
                                          PC3
                                                  PC4
                                                          PC5
                                                                  PC6
                                                                          PC7
                          PC1
Standard deviation
                       3.6444 2.3857 1.67867 1.40735 1.28403 1.09880 0.82172
Proportion of Variance 0.4427 0.1897 0.09393 0.06602 0.05496 0.04025 0.02251
Cumulative Proportion
                       0.4427 0.6324 0.72636 0.79239 0.84734 0.88759 0.91010
                           PC8
                                  PC9
                                          PC10
                                                 PC11
                                                         PC12
                                                                 PC13
                                                                         PC14
Standard deviation
                       0.69037 0.6457 0.59219 0.5421 0.51104 0.49128 0.39624
Proportion of Variance 0.01589 0.0139 0.01169 0.0098 0.00871 0.00805 0.00523
                       0.92598 0.9399 0.95157 0.9614 0.97007 0.97812 0.98335
Cumulative Proportion
                          PC15
                                  PC16
                                          PC17
                                                   PC18
                                                           PC19
                                                                   PC20
                                                                          PC21
                       0.30681 0.28260 0.24372 0.22939 0.22244 0.17652 0.1731
Standard deviation
Proportion of Variance 0.00314 0.00266 0.00198 0.00175 0.00165 0.00104 0.0010
Cumulative Proportion
                       0.98649 0.98915 0.99113 0.99288 0.99453 0.99557 0.9966
                          PC22
                                  PC23
                                          PC24
                                                  PC25
                                                          PC26
                                                                  PC27
Standard deviation
                       0.16565 0.15602 0.1344 0.12442 0.09043 0.08307 0.03987
Proportion of Variance 0.00091 0.00081 0.0006 0.00052 0.00027 0.00023 0.00005
Cumulative Proportion 0.99749 0.99830 0.9989 0.99942 0.99969 0.99992 0.99997
                          PC29
                                  PC30
Standard deviation
                       0.02736 0.01153
```

Proportion of Variance 0.00002 0.00000 Cumulative Proportion 1.00000 1.00000

plot(wisc.pr)



Q4. From your results, what proportion of the original variance is captured by the first principal components (PC1)?

From the data above, we see that PC1 captures 44.27% of the original variance in the dataset.

Q5. How many principal components (PCs) are required to describe at least 70% of the original variance in the data?

Three principal components (PC1, PC2 and PC3) are required to describe 72.64% of the variance in the original data.

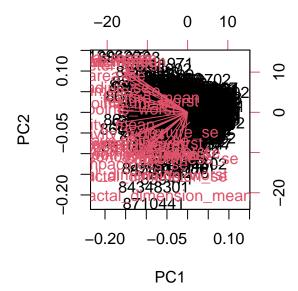
Q6. How many principal components (PCs) are required to describe at least 90% of the original variance in the data?

Seven principal components (PC1 to PC7) are required to capture 91.0% of the variance in the original data.

### Interpreting PCA Results

First, we create a biplot of our PC data.

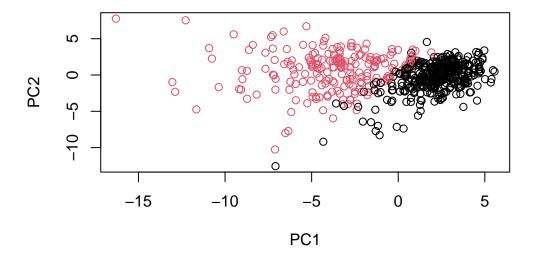
```
biplot(wisc.pr)
```



Q7. What stands out to you about this plot? Is it easy or difficult to understand? Why?

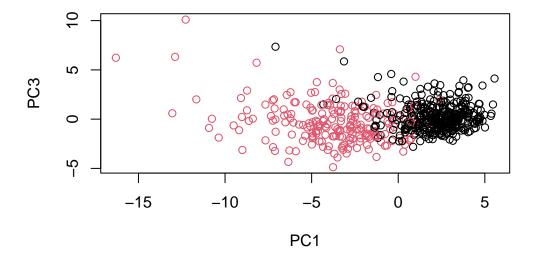
This plot is very hard to understand, due to the large number of dimensions in the original data and the fact that axes are labeled with rownames. They make this plot basically unreadable.

Let's try creating scatterplot of PC1 and PC2  $\,$ 



The above plot has PC1 on the x-axis, PC2 on the y-axis and the points are colored by diagnosis.

Q8. Generate a similar plot for principal components 1 and 3. What do you notice about these plots?



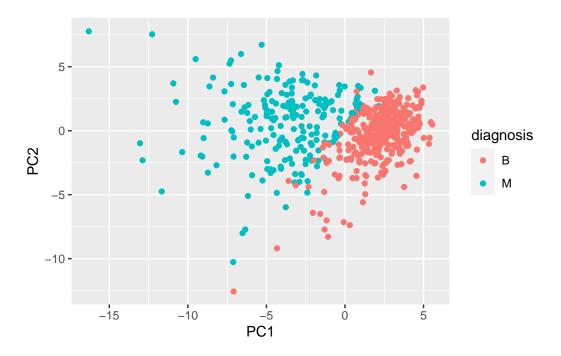
In both the above plots, the points clustered further along PC1 tend to be benign while the points less further along tend to be malignant. Moreover, benign cell points are clustered together so they tend to be more similar to each other than malignant cells are to each other. In the above figures, red color indicates malignant cells.

Using ggplot to make a fancier version of this plot:

```
# Create a data.frame for ggplot
df <- as.data.frame(wisc.pr$x)
df$diagnosis <- diagnosis

# Load the ggplot2 package
library(ggplot2)

# Make a scatter plot colored by diagnosis
ggplot(df) +
   aes(PC1, PC2, col=diagnosis) +
   geom_point()</pre>
```



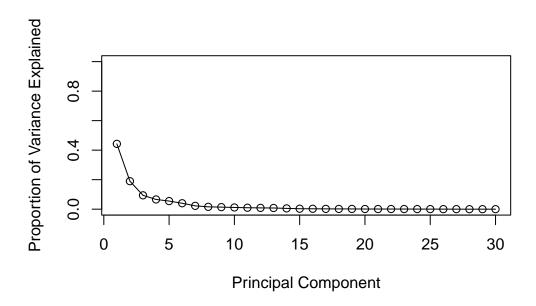
#### Variance explained

Calculating variance for each principal component

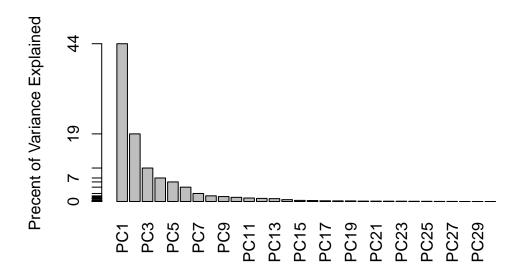
```
# Calculate variance of each component
pr.var <- wisc.pr$sdev^2
head(pr.var)</pre>
```

[1] 13.281608 5.691355 2.817949 1.980640 1.648731 1.207357

Next, calculating the proportion of variance explained by each principal component:



#### Creating alternative plot of same data:

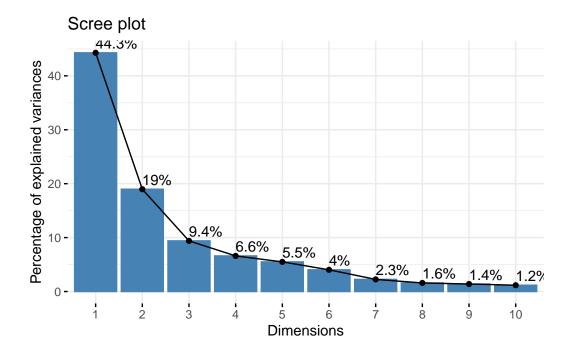


#### Creating ggplot graph with factoextra

```
## ggplot based graph
#install.packages("factoextra")
library(factoextra)
```

Welcome! Want to learn more? See two factoextra-related books at https://goo.gl/ve3WBa

```
fviz_eig(wisc.pr, addlabels = TRUE)
```



#### **Communicating PCA results**

Q9. For the first principal component, what is the component of the loading vector (i.e. wisc.pr\$rotation[,1]) for the feature concave.points\_mean?

```
wisc.pr$rotation["concave.points_mean",1]
```

#### [1] -0.2608538

The PC1 value for concave.points\_mean is -0.2608538.

Q10. What is the minimum number of principal components required to explain 80% of the variance of the data?

Below we get the attributes of the summary of the data set.

```
y <- summary(wisc.pr)
attributes(y)</pre>
```

Then, finding number of required components.

```
num_comp = sum(y$importance[3,] <= 0.8)
num_comp</pre>
```

[1] 4

The number of components required to describe roughly 80% of variation in the data is 4.

#### Hierarchical clustering

First, scaling wisc data.

```
# Scale the wisc.data data using the "scale()" function
data.scaled <- scale(wisc.data)</pre>
```

Then, calculating Euclidean distances between all pairs of observations in scaled data.

```
data.dist <- dist(data.scaled)</pre>
```

Next, creating HC model using complete linkage.

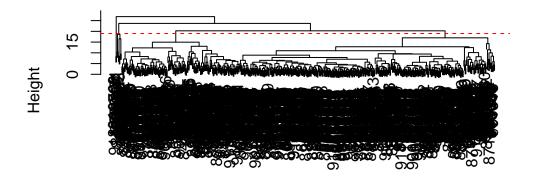
```
wisc.hclust <- hclust(data.dist, method='complete')</pre>
```

Q11. Using the plot() and abline() functions, what is the height at which the clustering model has 4 clusters?

The height at which the clustering model has 4 clusters is 19.

```
plot(wisc.hclust)
abline(h=19, col="red", lty=2)
```

## **Cluster Dendrogram**



data.dist hclust (\*, "complete")

#### Selecting number of clusters

Cutting the tree so that it has 4 clusters;

```
wisc.hclust.clusters <- cutree(wisc.hclust, 4)</pre>
```

Then, comparing cluster membership to actual diagnoses:

```
table(wisc.hclust.clusters, diagnosis)
```

```
diagnosis
wisc.hclust.clusters B M
1 12 165
2 2 5
3 343 40
4 0 2
```

Q12. Can you find a better cluster vs diagnoses match by cutting into a different number of clusters between 2 and 10?

Four clusters seems to be the minimum number of clusters required for creating groups stratified by diagnosis. False positives and false negatives are not reduced by increasing the number

of groups, and reducing the number of groups leads to groups that are not stratified by diagnosis. Therefore, 4 is the optimal number of clusters.

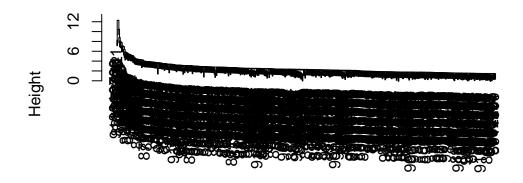
#### Using different methods

Q13. Which method gives your favorite results for the same data.dist dataset? Explain your reasoning.

Trying 'single' method

```
wisc.hclust.single <- hclust(data.dist, method='single')
plot(wisc.hclust.single)</pre>
```

## **Cluster Dendrogram**

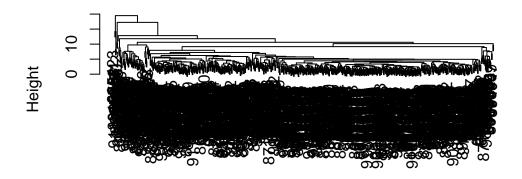


data.dist hclust (\*, "single")

Trying 'average' method

```
wisc.hclust.avg <- hclust(data.dist, method='average')
plot(wisc.hclust.avg)</pre>
```

# **Cluster Dendrogram**

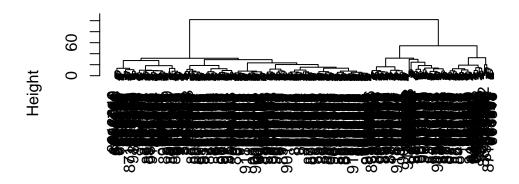


data.dist hclust (\*, "average")

Trying 'ward.D2' method

```
wisc.hclust.ward <- hclust(data.dist, method='ward.D2')
plot(wisc.hclust.ward)</pre>
```

## **Cluster Dendrogram**



data.dist hclust (\*, "ward.D2")

My favorite method is the ward.D2 method because it is easiest to see the clusters. The clusters are not so easily visible in the other methods.

#### K-means clustering

```
Creating k-means model
```

```
wisc.km <- kmeans(data.scaled, centers=2, nstart=20)
Using table to compare cluster membership

table(wisc.km$cluster, diagnosis)

diagnosis
    B M
1 14 175
2 343 37

table(wisc.hclust.clusters, diagnosis)</pre>
```

```
diagnosis
wisc.hclust.clusters B M
1 12 165
2 2 5
3 343 40
4 0 2
```

Q14. How well does k-means separate the two diagnoses? How does it compare to your hclust results?

K-means does a good job of separating the two diagnoses. It does a better job than the hclust method as it is able to get all the points into just two groups.

#### **Combining methods**

#### Clustering on PCA results

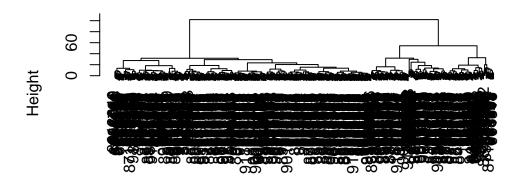
First, creating distance matrix from scaled 30 PCA variables.

```
scaled_pca_dist = scale(wisc.pr$x)
pca_dist = dist(wisc.pr$x)
```

Then, performing hierarchical clustering.

```
wisc.pr.hclust <- hclust(pca_dist, method='ward.D2')
plot(wisc.pr.hclust)</pre>
```

# **Cluster Dendrogram**



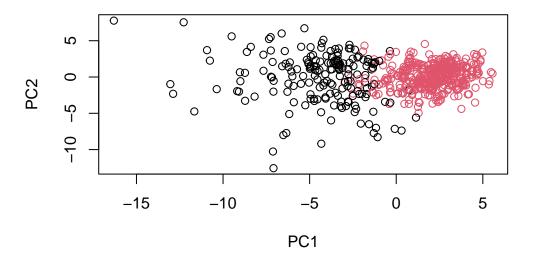
pca\_dist hclust (\*, "ward.D2")

Finding cluster membership vector with cutree() function.

Plotting the data colored by group:

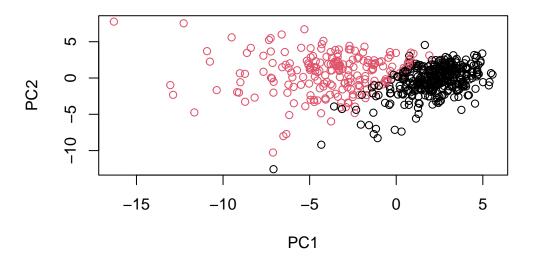
2 337 48

```
plot(wisc.pr$x[,1:2], col=grps)
```



Plotting the data colored by diagnosis:

```
plot(wisc.pr$x[,1:2], col=diagnosis)
```



## Recoloring:

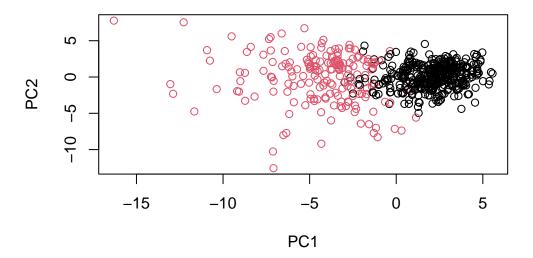
```
g <- as.factor(grps)
levels(g)

[1] "1" "2"

g <- relevel(g,2)
levels(g)

[1] "2" "1"

# Plot using our re-ordered factor
plot(wisc.pr$x[,1:2], col=g)</pre>
```



Plotting with rgl

```
library(rgl)
plot3d(wisc.pr$x[,1:3], xlab="PC 1", ylab="PC 2", zlab="PC 3", cex=1.5, size=1, type="s",
```

Next, using only first 7 PC's we perform hierarchical clustering

```
## Use the distance along the first 7 PCs for clustering i.e. wisc.pr$x[, 1:7]
seven_pc = dist(wisc.pr$x[ , 1:7])
wisc.pr.hclust <- hclust(seven_pc, method="ward.D2")</pre>
```

Then, cutting into 2 clusters

```
wisc.pr.hclust.clusters <- cutree(wisc.pr.hclust, k=2)</pre>
```

Now, comparing results from new model with actual diagnoses

Q15. How well does the newly created model with two clusters separate out the two diagnoses?

```
# Compare to actual diagnoses
table(wisc.pr.hclust.clusters, diagnosis)
```

```
diagnosis
wisc.pr.hclust.clusters B M
1 28 188
2 329 24
```

Q16. How well do the k-means and hierarchical clustering models you created in previous sections (i.e. before PCA) do in terms of separating the diagnoses? Again, use the table() function to compare the output of each model (wisc.km\$cluster and wisc.hclust.clusters) with the vector containing the actual diagnoses.

Comparing to hierarchical clustering before PCA

```
diagnosis

diagnosis

wisc.hclust.clusters B M

1 12 165

2 2 5

3 343 40

4 0 2
```

The PCA model identifies less people who are actually malignant as benign, but identifies more people who are actually benign as malignant than the other models.

### Sensitivity/Specificity

Q17. Which of your analysis procedures resulted in a clustering model with the best specificity? How about sensitivity?

Sensitivity for regular helust = 0.8 Specificity for regular helust = 0.96

Sensitivity for pca hclust = 0.886 Specificty for pca hclust = 0.922

The regular helust model was more specific as there were less false positives. The PCA helust model was more sensitive as it had less false negatives.

#### Prediction

Getting new data and projecting into PCA space.

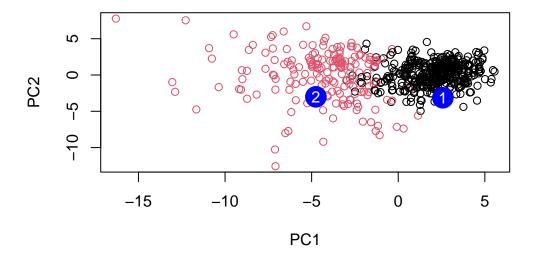
```
#url <- "new_samples.csv"
url <- "https://tinyurl.com/new-samples-CSV"</pre>
```

```
new <- read.csv(url)
npc <- predict(wisc.pr, newdata=new)
npc</pre>
```

```
PC1
                   PC2
                             PC3
                                       PC4
                                                PC5
                                                          PC6
                                                                     PC7
[1,] 2.576616 -3.135913 1.3990492 -0.7631950 2.781648 -0.8150185 -0.3959098
[2,] -4.754928 -3.009033 -0.1660946 -0.6052952 -1.140698 -1.2189945 0.8193031
          PC8
                    PC9
                             PC10
                                      PC11
                                               PC12
                                                        PC13
                                                                 PC14
[1,] -0.2307350 0.1029569 -0.9272861 0.3411457 0.375921 0.1610764 1.187882
[2,] -0.3307423 0.5281896 -0.4855301 0.7173233 -1.185917 0.5893856 0.303029
         PC15
                   PC16
                              PC17
                                         PC18
                                                    PC19
                                                              PC20
[1,] 0.3216974 -0.1743616 -0.07875393 -0.11207028 -0.08802955 -0.2495216
PC21
                    PC22
                              PC23
                                        PC24
                                                   PC25
                                                               PC26
[1,] 0.1228233 0.09358453 0.08347651 0.1223396 0.02124121 0.078884581
[2,] -0.1224776 0.01732146 0.06316631 -0.2338618 -0.20755948 -0.009833238
                      PC28
                                  PC29
                                              PC30
           PC27
[1,] 0.220199544 -0.02946023 -0.015620933 0.005269029
[2,] -0.001134152  0.09638361  0.002795349 -0.019015820
```

#### Plotting

```
plot(wisc.pr$x[,1:2], col = g)
points(npc[,1], npc[,2], col="blue", pch=16, cex=3)
text(npc[,1], npc[,2], c(1,2), col="white")
```



Q18. Which of these new patients should we prioritize for follow up based on your results?

We should prioritize patient 2 as their scores are in the region of the graph that has many malignant points.