Class 10

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Introduction to the RCSB Protein Data Bank (PDB).

First let's see what's in the database, the main repository of protein structures.

Downloaded composition stats from: https://www.rcsb.orgs/stats/summary

For context: Release 2023_04 of 13-Sep-2023 of UnitProt/TrEMBL contains 251600,768 sequence entries. The PDB only contains 183,201.

```
stats <- read.csv("PDBstats.csv", row.names = 1)
stats
```

	X.ray	EM	NMR	Multiple.methods	Neutron	Other
Protein (only)	158,844	11,759	12,296	197	73	32
Protein/Oligosaccharide	9,260	2,054	34	8	1	0
Protein/NA	8,307	3,667	284	7	0	0
Nucleic acid (only)	2,730	113	1,467	13	3	1
Other	164	9	32	0	0	0
Oligosaccharide (only)	11	0	6	1	0	4

	Total
Protein (only)	183,201
Protein/Oligosaccharide	11,357
Protein/NA	12,265
Nucleic acid (only)	4,327
Other	205
Oligosaccharide (only)	22

```
dim(stats)
```

```
[1] 6 7
```

There is a problem here due to commas in the numbers. This causes them to be treated as characters.

```
x <- stats$X.ray
x
```

```
[1] "158,844" "9,260"   "8,307"   "2,730"   "164"     "11"
```

```
as.numeric( gsub(",", "", x) )
```

```
[1] 158844   9260   8307   2730    164     11
```

Making a function of how to do this.

```
rm.comma <- function(x) {
  as.numeric( gsub(",", "", x))
}
```

```
rm.comma(stats$EM)
```

```
[1] 11759  2054  3667   113     9     0
```

I can use the `apply()` function to fix this whole table.

```
apply(stats, 2, rm.comma)
```

```
     X.ray    EM   NMR Multiple.methods Neutron Other  Total
[1,] 158844 11759 12296              197      73    32 183201
[2,]   9260  2054    34                8       1     0  11357
[3,]   8307  3667   284                7       0     0  12265
[4,]   2730   113  1467               13       3     1   4327
[5,]    164     9    32                0       0     0    205
[6,]     11     0     6                1       0     4     22
```

```
pdbstats <- apply(stats, 2, rm.comma)
rownames(pdbstats) <- rownames(stats)
head(pdbstats)
```

```
                        X.ray    EM   NMR Multiple.methods Neutron Other
Protein (only)         158844 11759 12296              197      73    32
Protein/Oligosaccharide  9260  2054    34                8       1     0
Protein/NA               8307  3667   284                7       0     0
Nucleic acid (only)      2730   113  1467               13       3     1
Other                     164     9    32                0       0     0
Oligosaccharide (only)     11     0     6                1       0     4
                        Total
Protein (only)         183201
Protein/Oligosaccharide  11357
Protein/NA               12265
Nucleic acid (only)       4327
Other                      205
Oligosaccharide (only)      22
```

```
totals <- apply(pdbstats, 2, sum)
totals
```

```
        X.ray              EM             NMR Multiple.methods
       179316           17602           14119             226
     Neutron           Other           Total
           77              37          211377
```

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
round(totals/totals["Total"] * 100, 2 )
```

```
        X.ray              EM             NMR Multiple.methods
        84.83            8.33            6.68            0.11
     Neutron           Other           Total
         0.04            0.02          100.00
```

Q2: What proportion of structures in the PDB are protein?

```
pdbstats[1, "Total"]
```

```
[1] 183201
```

```
pdbstats[1, "Total"] / sum(pdbstats[, "Total"] * 100, 2)
```

```
[1] 0.008667025
```

```
round((pdbstats[1, "Total"] / 251600768) *100, 2)
```

```
[1] 0.07
```

> Q4: Water molecules normally have 3 atoms. Why do we see just oneatom per water molecule inthis structure?

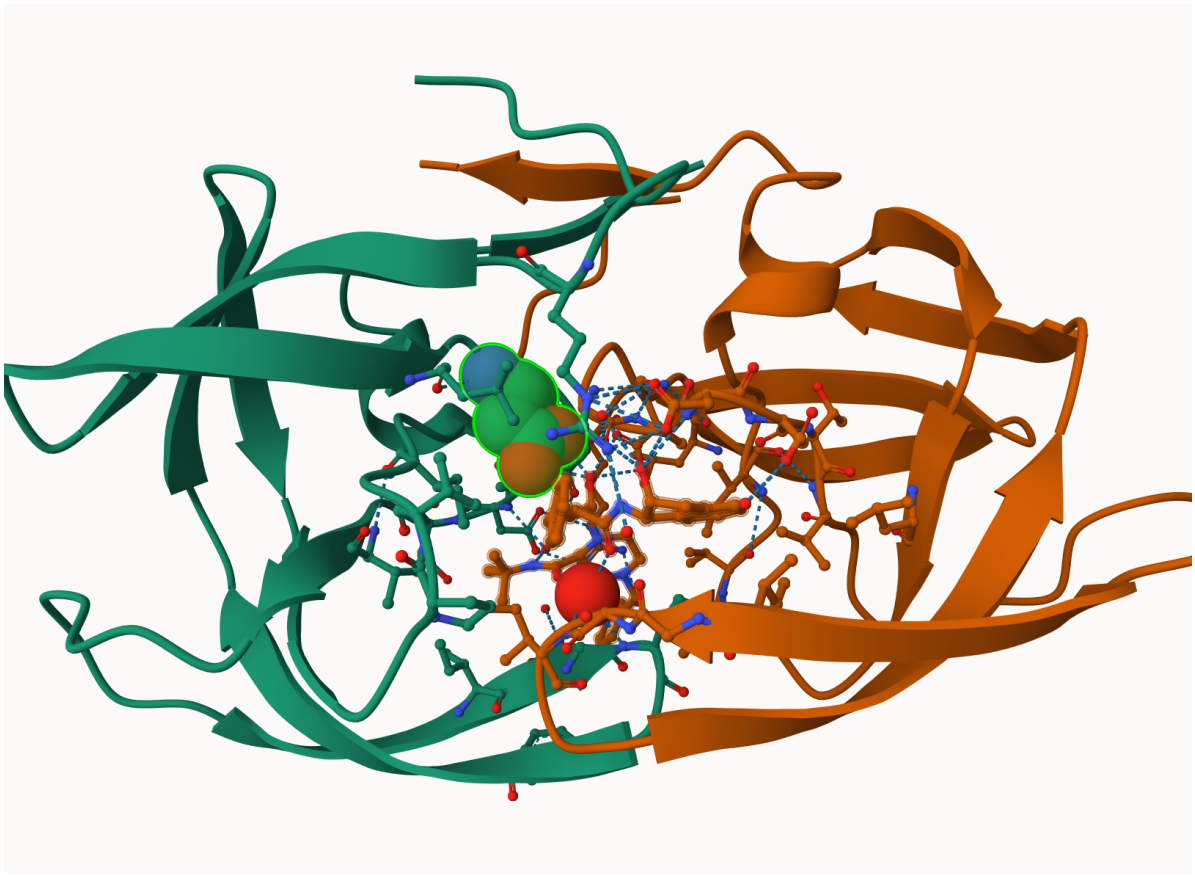
This is a 2 Angstrom structure and hydrogen is not visible at this resolution. 1 Angrstrom or better is needed.

> Q5: There is a critical "conserved" water molecule in the binding site. Can you identify this water molecule?

Water 308 is the conserved water molecule in the binding site.

> Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend "Ball & Stick" for these side-chains). Add thisfigure to your Quarto document.

Here is a figure of HIP-Pr with the catalytic ASP residues, the MK1 compound and all the important water 308.



The bio3d package for structural bioinformatics.

```
library(bio3d)

pdb <- read.pdb("1hsg")
```

```
 Note: Accessing on-line PDB file
```

```
pdb
```

```
Call:  read.pdb(file = "1hsg")

  Total Models#: 1
    Total Atoms#: 1686,  XYZs#: 5058  Chains#: 2  (values: A B)
```

```
     Protein Atoms#: 1514  (residues/Calpha atoms#: 198)
     Nucleic acid Atoms#: 0  (residues/phosphate atoms#: 0)

     Non-protein/nucleic Atoms#: 172  (residues: 128)
     Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]

   Protein sequence:
      PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
      QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
      ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
      VNIIGRNLLTQIGCTLNF

+ attr: atom, xyz, seqres, helix, sheet,
        calpha, remark, call
```

```
head(pdb$atom)
```

```
  type eleno elety  alt resid chain resno insert      x      y     z o     b
1 ATOM     1     N <NA>   PRO     A     1   <NA> 29.361 39.686 5.862 1 38.10
2 ATOM     2    CA <NA>   PRO     A     1   <NA> 30.307 38.663 5.319 1 40.62
3 ATOM     3     C <NA>   PRO     A     1   <NA> 29.760 38.071 4.022 1 42.64
4 ATOM     4     O <NA>   PRO     A     1   <NA> 28.600 38.302 3.676 1 43.40
5 ATOM     5    CB <NA>   PRO     A     1   <NA> 30.508 37.541 6.342 1 37.87
6 ATOM     6    CG <NA>   PRO     A     1   <NA> 29.296 37.591 7.162 1 38.40
  segid elesy charge
1  <NA>     N   <NA>
2  <NA>     C   <NA>
3  <NA>     C   <NA>
4  <NA>     O   <NA>
5  <NA>     C   <NA>
6  <NA>     C   <NA>
```

Predicting functional motions of a single structure

Let's finish today with a bioinformatics calculation to predict the functional motions of a PDB structure.

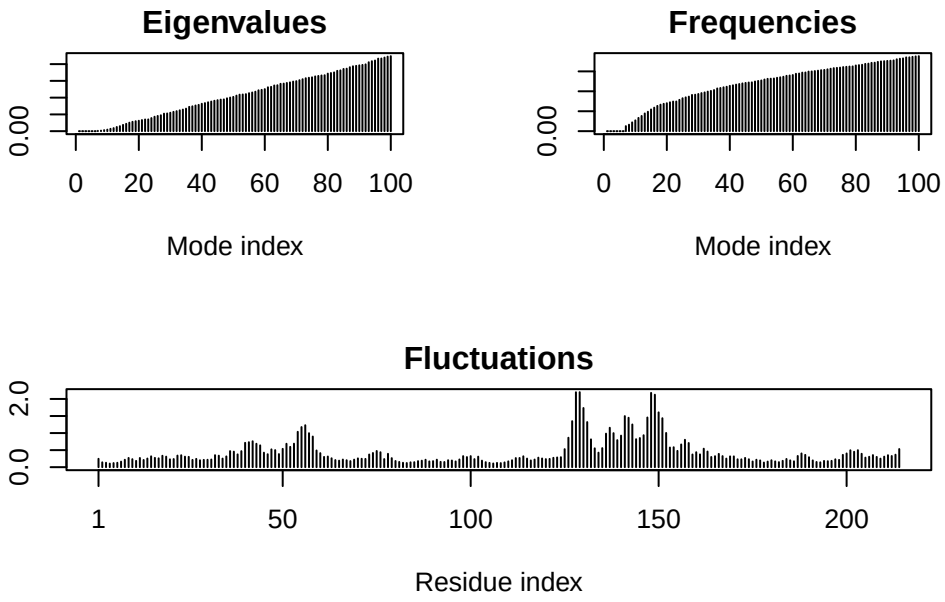
```
adk <- read.pdb("6s36")
```

```
 Note: Accessing on-line PDB file
  PDB has ALT records, taking A only, rm.alt=TRUE
```

```
m <- nma(adk)
```

```
Building Hessian...        Done in 0.039 seconds.
Diagonalizing Hessian...   Done in 0.396 seconds.
```

```
plot(m)
```




```
mktrj(m, file="adk_m7.pdb")
```