

DRAFT

**Identified and Unidentified Diatom Genera
Found on *Zostera marina*,
Montague Harbour, Galiano Island, BC.
Morphological and Molecular Data**

IMERSS, UBC and UVIC-AMF

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Abstract

A diverse range of microscopic biota are found on eelgrasses. Diatoms play a significant role in this microbiota and often cover eelgrass leaves in thick formations. The role of diatoms and their interactions with bacteria and eukaryotes is under-studied. To initiate an understanding of the ecological spatial distribution and dynamics, including the total microbiota found on the eelgrass *Zostera marina* at Montague Harbour Marine Provincial Park (MHMPP), Galiano Island, British Columbia, molecular and physical sampling for diatom identification was coordinated on March 7, 2021. DNA barcoding of 16S, 18S, and RBCL target regions from extracted DNA of swabs was taken on proximal, medial and distal sections. Physical sampling, from the same plants were acquired, across a stretch of the MHMPP beach. Each sample was physically divided into three 8-10 mm sections and preserved for diatom identification; a) by imaging intact sections with scanning electron microscopy (SEM) and b) cleaned frustules for SEM and light microscopy. Additionally, five samples were taken on different dates over two years. We have identified 60 genera of diatoms through morphological methods over the five sampling dates. Confident molecular sequence matches are 45 (85%) of the 53 genera identified morphologically on the co-sampling date of March 7, 2021. Ten additional genera have been identified through molecular sequences but not yet identified morphologically, though we presume these diatoms will be rare occurrences. Early results of counting on sections indicates that less than 10 genera will make up the vast majority of diatoms. Currently, adding together the morphological identifications and confident sequence data, results in a total of 70 genera over five samplings in a two-year period. With a high percentage of the diatom genera identified, we have begun the next phase of the diatom portion of this project; the counting of genera in numerous randomly selected 200 μm squares of in situ leaf sections.

Summary of Results

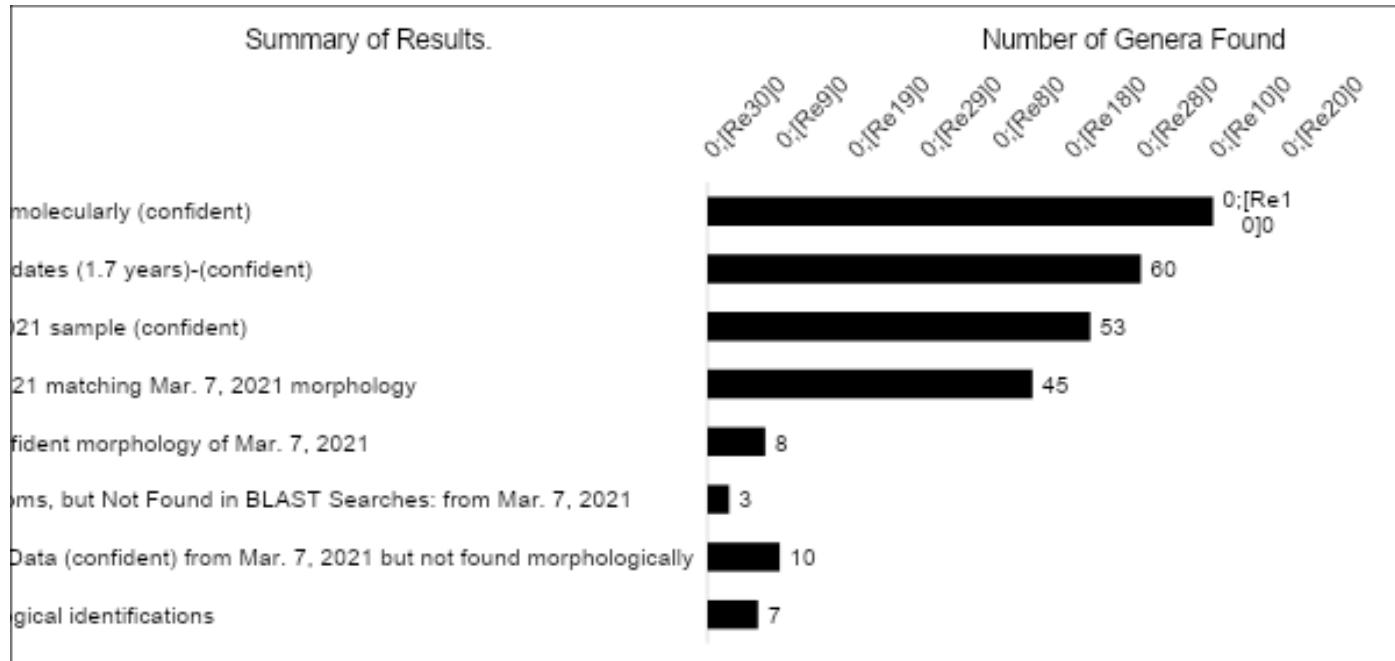


Table 2: Diatoms Found on *Zostera marina*, Montague Harbour Marine Provincial Park, Galiano Island, 2020-2022

		1	2	3	4	5	6	7	8
	All Genera Reported: morphologically and molecularly (confident)	Identified Morphologically over 5 sampling dates (1.7 years)- (confident)	Identified Morphologically from March 7, 2021 sample (confident)	Confident Molecular Sequences Mar. 7, 2021 matching Mar. 7, 2021 morphology	Non-confident Sequence Matches, but confident morphology of Mar. 7, 2021	Confident morphology of Mar. 7, 2021 Diatoms, but Not Found in BLAST Searches from Mar. 7, 2021	Additional Genera Found in the Molecular Data (confident) from Mar. 7, 2021 but not found morphologically	Additional genera: Non-confident morphological identifications	
1	ACHN	<i>Achnanthes</i> Bory 1822	<i>Achnanthes</i> Bory 1822	<i>Achnanthes</i> Bory 1822					<i>Dominika</i> Haas, '861
2	ACHNA	<i>Achnanthidium</i> Kützing, F.T. (1844)						<i>Achnanthidium</i> Kützing, F.T. (1844)	<i>Gomphosphaerula</i> Medi, n., '866
3	ACTI	<i>Actinophrys</i> Ehrenberg 1813	<i>Actinophrys</i> Ehrenberg 1813						<i>Hanzelia</i> Grunow, A. 1877
4	AMPHI	<i>Amphora</i> Ehrenberg ex Kützing, '847	<i>Amphora</i> Ehrenberg ex Kützing, '847	<i>Amphora</i> Ehrenberg ex Kützing, '847	<i>Amphora</i> Ehrenberg ex Kützing, '847				<i>Podosira</i> Ehrenberg, 1813
5	ATTI	<i>Attheya</i> I. West, 1850	<i>Attheya</i> I. West, 1850	<i>Attheya</i> I. West, 1850	<i>Attheya</i> I. West, 1850				<i>Rhopalodia</i> G. Müller, 1895 nom.
6	BACI	<i>Bacillaria</i> Grün, J.F. (1791)	<i>Bacillaria</i> Grün, J.F. (1791)	<i>Bacillaria</i> Grün, J.F. (1791)	<i>Bacillaria</i> Grün, J.F. (1791)				<i>Undatella</i> Piddock & M. S. Sims, 1999
7	CAMP	<i>Campylodiscus</i> Kützing 1844	<i>Campylodiscus</i> Kützing 1844	<i>Campylodiscus</i> Kützing 1844	<i>Campylodiscus</i> Kützing 1844				
8	CHAE	<i>Chlaetoceras</i> Ehrenberg 1842	<i>Chlaetoceras</i> Ehrenberg 1842	<i>Chlaetoceras</i> Ehrenberg 1842	<i>Chlaetoceras</i> Ehrenberg 1842				
9	COCC	<i>Cocconeis</i> Ehrenberg 1846	<i>Cocconeis</i> Ehrenberg 1846	<i>Cocconeis</i> Ehrenberg 1846	<i>Cocconeis</i> Ehrenberg 1846				
10	COSC	<i>Coscinodiscus</i> Ehrenberg, 1849	<i>Coscinodiscus</i> Ehrenberg, 1849	<i>Coscinodiscus</i> Ehrenberg, 1849	<i>Coscinodiscus</i> Ehrenberg, 1849				
11	CYCL	<i>Cyclotella</i> Kütz? St. 1860, nom. et syn. cons.	<i>Cyclotella</i> Kütz? St. 1860, nom. et syn. cons.						
12	CYLI	<i>Cylindrotheca</i> Habermann, 1840 nom. corr.	<i>Cylindrotheca</i> Habermann, 1840 nom. corr.	<i>Cylindrotheca</i> Habermann, 1840 nom. corr.	<i>Cylindrotheca</i> Habermann, 1840 nom. corr.				
13	DMME	<i>Dimerogramma</i> Taaffe 1861	<i>Dimerogramma</i> Taaffe 1861	<i>Dimerogramma</i> Taaffe 1861	<i>Dimerogramma</i> Taaffe 1861				
14	DIPL	<i>Diplonema</i> Ehrenberg ex Cleve, 1844	<i>Diplonema</i> Ehrenberg ex Cleve, 1844	<i>Diplonema</i> Ehrenberg ex Cleve, 1844	<i>Diplonema</i> Ehrenberg ex Cleve, 1844				
15	DITY	<i>Ditylum</i> (W.Bailey ex L.W.Bailey) 1861	<i>Ditylum</i> (W.Bailey ex L.W.Bailey) 1861	<i>Ditylum</i> (W.Bailey ex L.W.Bailey) 1861	<i>Ditylum</i> (W.Bailey ex L.W.Bailey) 1861				
16	ELLE	<i>Ellerbeckia</i> Crawford, R. M. 1938	<i>Ellerbeckia</i> Crawford, R. M. 1938	<i>Ellerbeckia</i> Crawford, R. M. 1938	<i>Ellerbeckia</i> Crawford, R. M. 1938				
17	ENTO	<i>Entomoneus</i> Ehrenberg, 1846	<i>Entomoneus</i> Ehrenberg, 1846	<i>Entomoneus</i> Ehrenberg, 1846	<i>Entomoneus</i> Ehrenberg, 1846				
18	EPIT	<i>Epithemia</i> Kütz? 1811	<i>Epithemia</i> Kütz? 1811	<i>Epithemia</i> Kützing, 1847	<i>Epithemia</i> Kützing, 1847				
19	EUCA	<i>Eucypris</i> Ehrenberg 1889	<i>Eucypris</i> Ehrenberg 1889						
20	ENCY	<i>Encyonema</i> Kützing, F.T. (1834)	<i>Encyonema</i> Kützing, F.T. (1834)	<i>Encyonema</i> Kützing, F.T. (1834)	<i>Encyonema</i> Kützing, F.T. (1834)				
21	EXTU	<i>Extubocellulus</i> Hasle, var. Stoebeli 1883						<i>Extubocellulus</i> Hasle, von Stoebeli & Syverstsen 1933	
22	FALL	<i>Fallertia</i> Simonart, Marin & Nicoud et al. 1960	<i>Fallertia</i> Simonart, Marin & Nicoud et al. 1960	<i>Fallertia</i> Simonart, Marin & Nicoud et al. 1960	<i>Fallertia</i> Simonart, Marin & Nicoud et al. 1960				
23	FOCE	<i>Fogedia</i> Witkowski, Linge-Durkot & Vetsch 1997	<i>Fogedia</i> Witkowski, Linge-Durkot & Vetsch 1997	<i>Fogedia</i> Witkowski, Linge-Durkot & Vetsch 1997	<i>Fogedia</i> Witkowski, Linge-Durkot & Vetsch 1997				
24	FRAG	<i>Fragilaria</i> Lyngbya 1819						<i>Fragilaria</i> Lyngbya 1819	
25	FRAGI	<i>Fragilariopsis</i> Hustedt 1913	<i>Fragilariopsis</i> Hustedt 1913	<i>Fragilariopsis</i> Hustedt 1913	<i>Fragilariopsis</i> Hustedt 1913				
26	GEDA	<i>Gedanieffia</i> Chudík, A. Witkowski & M.P. Ashworth 2018						<i>Gedanieffia</i> Chudík, A. Witkowski & M.P. Ashworth 2018	
27	GOMP	<i>Gomphonemopsis</i> Medlin 1988	<i>Gomphonemopsis</i> Medlin 1988	<i>Gomphonemopsis</i> Medlin 1988	<i>Gomphonemopsis</i> Medlin 1988				
28	GRAM	<i>Grammatophora</i> Ehrenberg 1840	<i>Grammatophora</i> Ehrenberg 1840	<i>Grammatophora</i> Ehrenberg 1840	<i>Grammatophora</i> Ehrenberg 1840				

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29 GYRO	<i>Gyrosigma</i> Hassall, 1845, nom. cons.	<i>Gyrosigma</i> Hassall, 1845, nom. cons.							
30 HALA	<i>Halimphora</i> (C.van Vredenbrouck, 1923)								
31 HASL	<i>Haslea</i> Simonsen, R. (1974)								
32 HOBA	<i>Hobartella</i> P.A.Sims & J.M.Williams, 2018; synonym of <i>Ostracocystis</i>	<i>Hobartella</i> P.A.Sims & J.M.Williams, 2018; synonym of <i>Ostracocystis</i>	<i>Hobartella</i> P.A.Sims & J.M.Williams, 2018; synonym of <i>Ostracocystis</i>	<i>Hobartella</i> P.A.Sims & J.M.Williams, 2018; synonym of <i>Ostracocystis</i>					
33 HYAL	<i>Hydrodictyon</i> Thümeling (1849)	<i>Hydrodictyon</i> Thümeling (1849)	<i>Hydrodictyon</i> Thümeling (1849)	<i>Hydrodictyon</i> Thümeling (1849)					
34 ISTH	<i>Isthmia</i> Agardh 1852							<i>Isthmia</i> Agardh - 832	
35 LEPT	<i>Leptocylindrus</i> Cleve, 1895	<i>Leptocylindrus</i> Clev. 1895	<i>Leptocylindrus</i> Clev. 1895	<i>Leptocylindrus</i> Cleve, 1895					
36 LIOM	<i>Lionophora</i> C.Agardh, 1827	<i>Lionophora</i> C.Agardh, 1827	<i>Lionophora</i> C.Agardh, 1827	<i>Lionophora</i> C.Agardh, 1827					
37 LYRE	<i>Lyrella</i> Ehrenberg Karayeva 1978								
38 MELO	<i>Melosira</i> C. Agardh, 1821, nom. cons.								
39 MINI	<i>Minidiscus</i> Hahn, 1975	<i>Minidiscus</i> Hahn, 1975	<i>Minidiscus</i> Hahn, 1975	<i>Minidiscus</i> Hahn, 1975					
40 MINU	<i>Minutocellus</i> Hahn, var. <i>stoeberi</i> and <i>syverdii</i> 1903							<i>Minutocellus</i> Hahn, var. <i>stoeberi</i> and <i>syverdii</i> 1903	
41 NAVI	<i>Nauvola</i> Bory 1822	<i>Nauvola</i> Bory 1822	<i>Nauvola</i> Bory 1822	<i>Nauvola</i> Bory 1822					
42 NITZ	<i>Nitzschia</i> Hassall, 1818, no n. cons.								
43 ODON	<i>Odontella</i> (Lyngbya) C.Agardh 1	<i>Odontella</i> (Lyngbya) C.Agardh 1	<i>Odontella</i> (Lyngbya) C.Agardh	<i>Odontella</i> (Lyngbya) C.Agardh					
44 OREP	<i>Opephora</i> F. Petit, 1899								
45 PARA	<i>Paralia</i> siberg., 1853	<i>Paralia</i> siberg., 1853	<i>Paralia</i> siberg., 1853	<i>Paralia</i> siberg., 1853					
46 PARI	<i>Panellulus</i> Cux 1568	<i>Panellulus</i> Cux 1568							
47 PERI	<i>Peridiphyle</i> , <i>Peridiphyle</i> nom. nov.								
48 PETR	<i>Petromenia</i> A.J. Strickland & D.G. Mann, 1990								
49 PLAG	<i>Plagiomgramma</i> Graville, 1859	<i>Plagiomgramma</i> Graville, 1859	<i>Plagiomgramma</i> Graville, 1859	<i>Plagiomgramma</i> Graville, 1859					
50 PLAGI	<i>Plagiotropis</i> Pfitzer, 1871	<i>Plagiotropis</i> Pfitzer, 1871	<i>Plagiotropis</i> Pfitzer, 1871	<i>Plagiotropis</i> Pfitzer, 1871					
51 PLAN	<i>Planothidium</i> Hustedt 1924	<i>Planothidium</i> Hustedt 1924	<i>Planothidium</i> Hustedt 1924	<i>Planothidium</i> Hustedt 1924					
52 PLEU	<i>Pleurosigma</i> W. Smith, 1852								
53 PODO	<i>Pododesmus</i> Ehrenberg, 1810	<i>Pododesmus</i> Ehrenberg, 1810	<i>Pododesmus</i> Ehrenberg, 1810	<i>Pododesmus</i> Ehrenberg, 1810					
54 PROS	<i>Proschkinia</i> Karayeva 1978							<i>Proschkinia</i> Karayeva, 1978	
55 PSAM	<i>Psammodictyon</i> D. G.Mann, 1990								
56 PSEU	<i>Pseudogymnophore</i> nra. Mex. - 1941								
57 PSEUN	<i>Pseudo-nitzschia</i> 4. Pringsheim, 1922	<i>Pseudo-nitzschia</i> 4. Pringsheim, 1922		<i>Pseudo-nitzschia</i> H. Pringsheim, 1941					

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18 RIAB	<i>Rhabdonema</i> Küzing, 1844, nom. cons.	<i>Rhabdonema</i> Küzing, 1844, nom. cons.	<i>Rhabdonema</i> Küzing, 1844, nom. cons.	<i>Rhabdonema</i> Küzing, 1844, nom. cons.					
55 RHIZ	<i>Rhizosolenia</i> <td><i>Rhizosolenia</i><td></td><td></td><td><i>Rhizosolenia</i><td></td><td></td><td></td><td></td></td></td>	<i>Rhizosolenia</i> <td></td> <td></td> <td><i>Rhizosolenia</i><td></td><td></td><td></td><td></td></td>			<i>Rhizosolenia</i> <td></td> <td></td> <td></td> <td></td>				
62 RMH	<i>Rhoicosphenia</i> <td><i>Rhoicosphenia</i><td><i>Rhoicosphenia</i><td><i>Rhoicosphenia</i><td></td><td></td><td></td><td></td><td></td></td></td></td>	<i>Rhoicosphenia</i> <td><i>Rhoicosphenia</i><td><i>Rhoicosphenia</i><td></td><td></td><td></td><td></td><td></td></td></td>	<i>Rhoicosphenia</i> <td><i>Rhoicosphenia</i><td></td><td></td><td></td><td></td><td></td></td>	<i>Rhoicosphenia</i> <td></td> <td></td> <td></td> <td></td> <td></td>					
8 SHIO	<i>Shionodiscus</i> <td></td> <td></td> <td></td> <td></td> <td></td> <td><i>Shionodiscus</i><td></td><td></td></td>						<i>Shionodiscus</i> <td></td> <td></td>		
22 SEMI	<i>Sentusavis</i> <td></td> <td></td> <td></td> <td></td> <td></td> <td><i>Sentusavis</i><td></td><td></td></td>						<i>Sentusavis</i> <td></td> <td></td>		
22 SERR	<i>Serratella</i> <td></td> <td></td> <td></td> <td></td> <td></td> <td><i>Serratella</i><td></td><td></td></td>						<i>Serratella</i> <td></td> <td></td>		
64 SKEL	<i>Skeletonema</i> <td><i>Skeletonema</i><td><i>Skeletonema</i><td><i>Skeletonema</i><td></td><td></td><td></td><td></td><td></td></td></td></td>	<i>Skeletonema</i> <td><i>Skeletonema</i><td><i>Skeletonema</i><td></td><td></td><td></td><td></td><td></td></td></td>	<i>Skeletonema</i> <td><i>Skeletonema</i><td></td><td></td><td></td><td></td><td></td></td>	<i>Skeletonema</i> <td></td> <td></td> <td></td> <td></td> <td></td>					
45 STEP	<i>Stephanopyxis</i> <td><i>Stephanopyxis</i><td><i>Stephanopyxis</i><td><i>Stephanopyxis</i><td></td><td></td><td></td><td></td><td></td></td></td></td>	<i>Stephanopyxis</i> <td><i>Stephanopyxis</i><td><i>Stephanopyxis</i><td></td><td></td><td></td><td></td><td></td></td></td>	<i>Stephanopyxis</i> <td><i>Stephanopyxis</i><td></td><td></td><td></td><td></td><td></td></td>	<i>Stephanopyxis</i> <td></td> <td></td> <td></td> <td></td> <td></td>					
36 TABU	<i>Tabularia</i> (Küzing) D.M.Williams & Lind, 1955	<i>Tabularia</i> (Küzing) D.M.Williams & Lind, 1955	<i>Tabularia</i> (Küzing) D.M.Williams & Lind, 1955	<i>Tabularia</i> (Küzing) D.M.Williams & Lind, 1955					
47 THAL	<i>Thalassiosira</i> <td><i>Thalassiosira</i><td><i>Thalassiosira</i><td><i>Thalassiosira</i><td></td><td></td><td></td><td></td><td></td></td></td></td>	<i>Thalassiosira</i> <td><i>Thalassiosira</i><td><i>Thalassiosira</i><td></td><td></td><td></td><td></td><td></td></td></td>	<i>Thalassiosira</i> <td><i>Thalassiosira</i><td></td><td></td><td></td><td></td><td></td></td>	<i>Thalassiosira</i> <td></td> <td></td> <td></td> <td></td> <td></td>					
48 TRAC	<i>Trachyneis</i> <td><i>Trachyneis</i><td><i>Trachyneis</i><td><i>Trachyneis</i><td><i>Trachyneis</i><td></td><td></td><td></td><td></td></td></td></td></td>	<i>Trachyneis</i> <td><i>Trachyneis</i><td><i>Trachyneis</i><td><i>Trachyneis</i><td></td><td></td><td></td><td></td></td></td></td>	<i>Trachyneis</i> <td><i>Trachyneis</i><td><i>Trachyneis</i><td></td><td></td><td></td><td></td></td></td>	<i>Trachyneis</i> <td><i>Trachyneis</i><td></td><td></td><td></td><td></td></td>	<i>Trachyneis</i> <td></td> <td></td> <td></td> <td></td>				
22 TRIG	<i>Trigonium</i> <td><i>Trigonium</i><td><i>Trigonium</i><td><i>Trigonium</i><td><i>Trigonium</i><td><i>Trigonium</i><td></td><td></td><td></td></td></td></td></td></td>	<i>Trigonium</i> <td><i>Trigonium</i><td><i>Trigonium</i><td><i>Trigonium</i><td><i>Trigonium</i><td></td><td></td><td></td></td></td></td></td>	<i>Trigonium</i> <td><i>Trigonium</i><td><i>Trigonium</i><td><i>Trigonium</i><td></td><td></td><td></td></td></td></td>	<i>Trigonium</i> <td><i>Trigonium</i><td><i>Trigonium</i><td></td><td></td><td></td></td></td>	<i>Trigonium</i> <td><i>Trigonium</i><td></td><td></td><td></td></td>	<i>Trigonium</i> <td></td> <td></td> <td></td>			
75 TRYB	<i>Tryblionella</i> W. Smith 1852	<i>Tryblionella</i> W. Smith 1852	<i>Tryblionella</i> W. Smith 1852	<i>Tryblionella</i> W. Smith 1852					
		70	60	55	45	3	3	10	7

1. Sixty genera are reported with certainty (Chart 1, col. 2).¹ from all five samplings over two years, based on morphological methods.
2. Five additional genera (Chart 1, col. 7) are of uncertain (uc) taxonomic placement and require more images, taxonomic investigations and or molecular confirmation: 1) *Donkinia* Ralfs, 1861; 2) *Gomphoseptatum* Medlin, 1986; 3) *Hanzschia* Grunow, A. 1877); 4) *Rhopalodia* O. Müller, 1895, nom. cons. and 5) *Undatella* Paddock & P.A.Sims, 1980.
3. Fifty-three genera are reported with certainty (Chart 1, col. 3) from the March 7, 2021 sampling based on morphological methods; 88% of the 60 genera identified over five sampling dates in two years.
4. Forty-five genera are confidently supported by the March 7, 2021 molecular data (MOL) (Chart 1, col. 4). The molecular data supports 85% of all genera confidently reported using morphology methods from the March 7, 2021 sample.
5. Eleven additional genera are non-confident in reasonable alignment stats or without sequence matches (Chart 1, col. 5, 6). Nine genera identified morphologically are lacking in high alignment stats: 1) *Campylodiscus* Kützing 1844²; 2) *Ditylum* J.W.Bailey ex L.W.Bailey, 1861³; 3) *Epithemia* Kützing, 1844⁴; 4) *Fogedia* Witkowski, Lange-Beralot et Metzeltin 1997⁵; 5) *Plagiotropis*⁶; 6) *Rhizosolenia* Brightwell, 1858, nom. et typ. Cons⁷; 7) *Stephanopyxis* Ehrenberg, C.G. (1845)⁸; 8) *Trachyneis* Cleve, P.T. (1894)⁹ (Chart 1, col. 5). Three genera morphologically identified are without sequence matches: 1) *Fogedia* Witkowski, Lange-Beralot et Metzeltin 1997; 3) *Grammatophora* Ehrenberg 1840 and 2) *Trigonium* Cleve, P.T. 1867 (Chart 1, col. 6).
6. The molecular data from March 7, 2023 matches 76% of the diatoms identified over a 1.7-year period on five sampling dates.
7. Eight genera identified morphologically are ‘probable’ matches to the molecular data (Chart 1, col. 5): they are found repeatedly but they lack very high percentage sequence identity matches¹⁰ to sequences found with BLAST (NIH 2023) and the distance tree results are only moderate to weak. Three more genera identified by morphological methods have sequences with matches in the BLAST databases, however the alignment data and distance tree results are not dependable.
8. An additional 10 genera have been identified with confidence based on BLAST results from two molecular sequence datasets but have not yet been identified by morphological methods (Chart 1, col. 6). Results listed in these two sequence datasets were examined with the databases accessed by BLAST set to *Sequences producing only significant alignments* and double checked in the BLAST Distance Tree of results program and only with repeated entries that have excellent or high scoring alignments.¹¹ The following genera have repeated results of very low e values, high query cover, sequences producing significant alignments (very high Per. Ident) and excellent distance tree results (here listed as: confid.): 1) *Achnanthidium* Kützing, F.T. (1844) (confid¹²); 2) *Extubocellulus* Hasle, von Stosch & Syvertsen 1983 (confid.); 3) *Fragilaria* Lyngbye 1819¹³ (confid.); 4) *Gedaniella* Chunlian Li, A. Witkowski & M.P.

¹ ‘Pariraphis like’ is included in the certain list of genera since it is found regularly and has a consistent taxonomy.

² *Campylodiscus*: Sequences found in BLAST, but low stat confidence in the alignment and distance tree results

³ *Ditylum*: Sequences found in BLAST, but low stat confidence in the alignment and distance tree results.. Part and entire valves of *Ditylum sp.* are commonly observed in slides from March 7, 2021 sections.

⁴ *Epithemia*: Sequences found in BLAST, but low stat confidence in the alignment and distance tree results.

⁵ *Fogedia*: No sequences found.

⁶ *Plagiotropis*: Sequences found in BLAST, but low stat confidence in the alignment and poor distance tree results.

⁷ *Rhizosolenia*: Sequences found in BLAST, but low stat confidence in the alignment and poor distance tree results.

⁸ *Stephanopyxis*: Numerous hits in BLAST, but low stat confidence in the alignment and moderate distance tree results.

⁹ *Trachyneis*: Sequences found in BLAST, but low stat confidence in the alignment and distance tree results.

¹⁰ BLAST, Per. Ident.

¹¹ At this time we have not developed a precise set of numeric guidelines for determining how confident we are of good matches to BLAST entries; except that we consider confident matches to have: 1) very good base alignment stats (high query cover and percent ident.(usually no less than 90-93% percent ident.), 2) in the top 10 genera listed in the BLAST search table and top query genera are separated by percent ident. from the next matches and 3) excellent distance tree results; horizontally very close to the query sequence and linked to the same or a very close node.

¹² *Arcocellulus*: though many sequences are listed, only one sequence, SLIVA 2586 has an excellent alignment match and very excellent distance tree results. Also, sequences occur in the top list with *Minutocellus* and in excellent distance tree results; maybe both genera are present or one.

¹³ *Fragilaria*: Two good query sequences so far (March 26, 2023). Their alignment stats and distance tree results are excellent. Marine species.

- Ashworth, 2018 (confid.); 5) *Isthmia* Agardh 1832 (confid.); 6) *Minutocellus* Hasle, von Stosch and Syvertsen 1983 (confid.); 7) *Podosira* Ehrenberg, 1840 (confid.); 8) *Proschkinia* Karayeva 1978 (confid.); 9) *Shionodiscus* Alverson, A.J., Kang, S.-H. & Theriot, E.C. (2006); 10) *Seminavis* D.G. Mann 1990 (confid.); 11) *Serratifera* M.P.Ashworth, C.Li & A.Witkowski, 2016 (confid.).
9. *Arcocellulus* Hasle, von Stosch & Syvertsen (1983) is occasionally found in BLAST searches with strong alignment and distance results, with only one excellent sequence match in BLAST, and often indistinguishable stats relative to *Minutocellus*, is herein is listed as probable (probab.).
 10. Five more genera are showing moderate indications of being present in the March 7, 2021 molecular samples, but not yet observed by morphology. The lower confidence is due to weaknesses in alignment stats, distance tree results or a low number of occurrences in the datasets (here listed as: possible): 1) *Bacterosira Gran 1900*¹⁴ (possible); 2) *Berkeleya Graville 1827* (possible); 5) *Cymbellonitzschia* Hustedt 1924;¹⁵ 6) *Lemincola* Round & Basson, 1997.¹⁶; 7) *Leyanella* and *Pseudoleyanella* (possible); 3) *Pinnularia* Ehrenberg, 1843, nom. et. typ. cons (possible to uncertain); 4) *Phaeodactylum* Bohlin, K. (1898) (possible); 5) *Ralfsiella* (Ralfs) P.A.Sims, D.M.Williams & Ashworth 2018 (possible¹⁷). We are continuing to search the sequence data for good BLAST results occasionally found in the Family, Order or Genera columns of the sequence data.
 11. Seventy genera are confirmed by adding together the morphological identifications (60) and the genera confidently identified through molecular sequences (10) (Chart 1, col. 1) but are not yet identified using morphology, found on *Z. marina* over a 1.7-year period from 5 samples.
 12. One fairly common genus is currently undergoing taxonomic placement (paper in progress) and is tentatively identified as *Pariraphis* like (Witkowski, pers. comm.) is included in the total genera recorded.
 13. *Rhoicosphenia*, a genus regularly seen in all the *Z. marina* samples and straightforward to identify, to date, has only appeared in one record (SILVA 519, Bacillariophyceae_fa) by a search using BLAST.¹⁸ The excellent alignment and distance tree stats from BLAST makes the molecular identification certain.
 14. *Achnanthes cf. elongata*, is common on some proximal and medial leaf sections and is straightforward to identify. It is the only *Achnanthes* species observed on March 7, 2021 *Z. marina* sections. However, though it has appeared in BLAST searches using all the sequence data, all three matches in BLAST gave moderate alignment stats and poor distance tree results. It is possible that the species we observe is a different or undescribed species compared to the morphologically very similar *Achnanthes cf. elongata*, reported as epizoic by Majewska et al. 2017.
 15. *Cocconeis*, possibly the most abundant diatom on *Z. marina* shows few hits within the molecular data. It is possible that swabbing a flat, well silicified and strongly adhering diatom missed collecting sufficient quantities of the diverse taxon range of *Cocconeis* observed on *Z. marina* leaves during all samplings.
 16. A considerable number of SEM and LM images have been taken. Not all prepared SEM stubs and LM slides of cleaned material have been examined or re-examined. Out of 200+ folders of SEM and LM images, there are approximately 20 unprocessed folders yet to be examined in detail since the first sampling in August 2020. In the March 7, 2021 sampling: approximately 85% of a possible 70 folders have been examined and processed. Based on the folders left to examine, it is expected that more genera will be identified, especially those only identified by molecular methods. Re-examining SEM and LM material has continued to yield new confirmed genera (i.e. *Podosira* and *Campylodiscus*), better images and taxonomic features of previously identified images.

¹⁴ *Bacterosira*: Only one query sequence so far (March 26, 2023). However, it's alignment stats and distance tree results are very good. It was previously placed as a species under *Thalassiosira*.

¹⁵ *Cymbellonitzschia*: Only one query sequence so far (April 3, 2023). However, it's alignment stats and distance tree results are good.

¹⁶ *Lemincola*: Only two query sequences so far (March 27, 2023) and it is considered a non-marine diatom. However, it's alignment stats and distance tree results are very good.

¹⁷ *Ralfsiella*: only to sequences in SILVA records. However, the alignment and distance tree results are excellent.

¹⁸ See the notes: Distinguishing Rhoicosphenia, Gomphonemoplia, Pseudogomphonema, Gomphoseptatum & Licmophora-(MW-Dec 23-2021-Aug 28-2021 AvA Edits Nov14-2021)-AC.docx

2. Introduction

3. Methods and Materials

2.1 Study area and sampling collection



Fig. 1. Map of the Salish Sea, Gulf Islands and Galiano Island.



Fig. 2. Map of Galiano Island and study area. The inset shows the location of the study area, located at the Montague Harbour Marine Provincial Park.

At low tides, 10 *Zosteria marina* eelgrass leaf samples were harvested evenly across the entire 400 m long shallow gravel beach at Montague Harbour marine Provincial Park (MHMPP) (Lat: 48.896732, Long: -123.403318), Galiano Island, British Columbia, Canada (Fig. 1-2.). Galiano Island is a narrow, 26 km long island in the Gulf Islands within the Salish Sea. Authorization to harvest eelgrass leaves in the provincial park was granted to the research team by BC Parks. Although five separate samplings have been taken at MHMPP over a three-year period, the March 7, 2021 samples were taken together for the purposes of both molecular and morphological analysis. Leaves were cut in-situ 10-20 mm above the rhizome base providing 10 secondary and 10 tertiary leaves.

For morphological research, each leaf was placed in a freshly opened pre-labeled Ziplock plastic bag containing 300 mL of 0.2 µm filtered MHMPP water that was collected 2 days before the harvesting of the leaves. On the shore, with sterile gloves, a polypropylene cutting board, ruler tweezer and razor blades (all items washed between samples with 100% ethanol) each leaf was sliced into three sections--proximal, media and dorsal--and each of those three sections cut into three 8 mm sections for a total of 90 individual samples. Each 8mm section was placed in labeled 15 mL sterile poly Falcon tubes with 4% final concentration of Formalin and inserted upright into test tube racks and transported within 1 hour undisturbed to a 2-3°C refrigerator. [Need the protocol for the sampling of leaves for molecular analysis: Similarly, at the same time and location, X sections of secondary and tertiary leaves of proximal, medial and distal sections were swabbed with sterile Q-tips and stored in liquid nitrogen for molecular analysis. The use of secondary and tertiary leaves was alternated between the molecular analysis and the microscopy teams.

2.2. DNA extraction, PCR amplification and high-throughput sequencing (HTS) library preparation

[Need the full protocol] Molecular data (MOL) obtained by performing DNA barcoding of 16S, 18S, and RBCL target regions from extracted DNA of swabs. Sequence data processing, including taxonomic assignment of the Amplicon Sequence Variants (ASVs), was performed in the DADA2 pipeline (Callahan et al. 2016). 16S and 18S taxonomy was assigned from the SILVA 138 database and RBCL sequences assigned with DiatBarcode V10 database. The taxonomy database from Siobhan Schenk (Parfrey lab, UBC) sent on February 25, 2022 is the output (3221 sequence records) of the denoised output of DADA2 pipeline. A second database of 3,965 sequences was generated from the March 7, 2021 data with SILVA by Siobhan Schenk on Feb. 10, 2023 (SILVA #X,).

2.3 Microscopical analysis

Samples for morphological analysis were prepared in two ways: 1) sections of *Z. marina* were scraped with a sterile razor or toothbrush in a few mL of 0.2 µm MHMPP water to produce a concentrated pool of diatoms of which 5 mL was pipetted and placed in a 50 mL borosilicate centrifuge tube. Diatoms were cleaned with concentrated reagent grade concentrated sulfuric acid in a ratio by volume of 1:4 sample to acid at between 95-100°C for 2-3 hours or until the mixture was faint straw coloured or clear. The same procedure was used for whole 8 mm leaf sections to capture all of the diatoms. Concentrated reagent grade Nitric acid (47%) to reagent grade concentrated sulphuric acid (97%) in a 1:3 ratio was also used for bulk cleaning of diatom and leaf samples in the same manner as with the sulfuric acid methods used previously. In some cases where samples showed some residual organics, concentrated (37%) hydrogen peroxide was further added and held at 95-100°C for 1 hour. Thereafter samples were washed 8-9 times, each time removing the supernatant to 5 mL and washing with 35 mL of distilled and deionized water at 12 h intervals to a neutral pH. Cleaned diatom suspensions were pipetted onto cover slips, covered and left at room temperature overnight to dry. Permanent slides were mounted with Naphrax (Brunel Microscopes: <http://www.brunelmicroscopes.co.uk/>), which has a high refractive index. Diatom analysis was performed using a Nikon E800 with a Nikon 1.4 n.a. condenser and either Nikon 60x PlanApo n.a. 1.4 DIC or 100× PlanApo (n.a. 1.4) DIC objectives.

For SEM examination, 2-3 drops of the cleaned sample was filtered onto 13 mm Whatman Nuclepore polycarbonate membranes (Fisher Scientific, Toronto, Canada) using Swinnex, polypropylene, 13mm Filter Holders (Sterlitech Corp. <https://www.sterlitech.com>) and washed with 60 mL of distilled and deionized water. Filters were air-dried overnight or in a 60°C oven for 1 hr., mounted onto aluminum stubs and sputtered coated with gold. Besides acid cleaned samples, a second method was used to take images of intact diatoms in-situ on the eelgrass: a) The formalin solution used to store the *Z. marina* 8 mm sections in 15 mL polytubes was removed by a sterile glass Pasteur pipette, being careful to not touch or disturb the specimen, b) Specimen solution was diluted with 15 mL of distilled and deionized water carefully pipetted into the tube. Let sit for 2 hrs. to dissolve marine salts. c) after removing the water to 1-2 mL a sequential ethanol series of 50%, 60%, 70%, 90% and 100% 3x (ethanol dried with zeolites) of 10 mL at 30 minutes was carefully pipetted and removed by pipette to not disturb or touch the specimen. d) Then, 1x 5 mL application of 1:1 Ethanol to HMDS (Hexamethyldisilizane, Ted Pella Inc. <https://www.tedpella.com/>) followed by 3 mL of 100% HMDS at 30 min. each was pipetted, and the supernatant removed. e) After the last application of HMDS the section of eelgrass was slid out of the tube onto a cleaned and sterile 100 mm glass petri dish and allowed to air dry in a fume hood. Specimens were transferred to a 60°C oven to fully dry. f) Specimens were mounted on black carbon stickies (Ted Pella Inc.): To attach it to the carbon sticky, the tip of a sterile fine tweezer was used to press the section down in 6 equidistant spots around the edge of the section. 3) Another method was used to image semi-cleaned eelgrass sections. Before using the in-situ method, after a soak in deionized and distilled water, 30% hydrogen peroxide was added to the eelgrass sections for 24 hrs. at 20°C to remove some of the organics to make genus identifications easier and examine girdle bands of intact cells, without resorting to full acid cleaning. After gold sputter coating, samples were imaged with a Hitachi s4800 FESM or a Hitachi

TM4000 SEM instruments at the Advanced Microscope Facility, University of Victoria, British Columbia, Canada.

Identified Diatom Genera

Abbreviations used:

1. Observed approximate frequency (not by numerical counts): **1** = rare, **2** = common, **3** = very common, **4** = abundant¹⁹
 2. a) **X** = reported/imaged, b) **blank space** (either not found yet or due to the non-examination of samples), c) **uc**= uncertain or unconfirmed, though they fit some genus characteristics but require further investigation.
 3. **SEM** = Scanning Electron Microscopy, **LM** = light microscopy (cleaned frustules, prepared slides), **Live** = live cells in light microscopy, **MOL** = molecular data: confid. (confid.), prob. (prob.), indic. (indications).
- All diatom genera and species names are from AlgaeBase (Guiry and Guiry, 2021) recently cited papers or diatom taxonomic references.
 - Further information is found on the detailed a) **Checklist of Diatoms of the Southern Gulf Isl.(in process)**, b) **Updates on Genera Identified** (with images) and c) **Taxonomic Notes**.

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
1.	ACHN	<i>Achnanthes</i> Bory 1822. The only <i>Achnanthes</i> observed for March 7, 2021 leaf sections is an elongate-linear <i>Achnanthes</i> , fairly abundant and with unusual characteristics: they are highly elongate, lack apical pore fields, are mono-raphid, noticeably heteropolar (bent valves) and have other characteristics typical of <i>Achnanthes</i> . ²⁰ A variety of more common <i>Achnanthes</i> species are present in other eelgrass samplings. <i>Achnanthes bongrainii</i> (which should be spelt <i>A. bongrainii</i>) (M.Peragallo) Mann 1937: 16, shows up in the molecular sequences. (#319, SILVA-BLAST: <i>Achnanthes bongrainii</i> 100%, 2e-179, 95.53%; <i>Achnanthes</i> sp. 100%, 1e-161, 92.02%); <i>Achnanthalles</i> (MOL-52-V60, MOL-262-V306); <i>Achnanthalles</i> (MOL 104-V121, <i>Planothidium lanceolatum</i> isolate 100% BLAST). Comparing <i>Achnanthalles</i> sequences in the March 7, 2021 sampling	2-3	x	x	x SEM, (MOL-confid.)	x		SEM, LM, Live, (MOL -confid.)

¹⁹ Names for relative abundance used by Jacob and Noten (1980).

²⁰ Detailed taxonomic notes are forthcoming. This diatom is placed in the *Achnanthes* taxon since the cells are bent along the transapical axis and broad in the centre girdle view and are monoraphid (Round et al. 1990, Plinski and Witkowski 2020: 62). The closest fit to the observed diatoms are a recently described species, *Achnanthes elongata* Majewska & Van de Vijver 2017 found on the carapaces of marine turtles in Costa Rica (Majewska et al. (2017), a variety of *A. groenlandica* (Cleve) Grunow 1880 or possibly an undescribed *Achnanthes* species. A. Witkowski confirms it is similar to

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		<p>against data in BLAST has so far three matches for <i>A. elongata</i> (SILVA 546 (BLAST: <i>Achnanthes elongata</i>, 688, 100, 0.0, 97.519, MT441502,-could not find in distance tree); SILVA 1632 (96.91% ident.); SILVA 2150 (97.27% ident.), all with poor distance tree results.</p> <p><i>Achnanthes spp. show significant close alignments to Achnanthes in BLAST.</i></p> <p>#35-SILVA Feb 10-2023: (BLAST: Achnanthidium sp. 98% 2e; Achnanthidium saprophilum 98%, 2e); SILVA # 1453 A. bongranii, (BLAST: Achnanthes bongranii, 608, 608, 100%, 2e-169, 94.04% - poor distance tree alignment). BLAST searches: 33, 37 (95.44% ident), 38, 249, 284, 319 (good), 326, 453, 454, 461, 466, 467 (99.27% ident), 477 97.46% ident.), 479 (98.99% ident.- Halamphora), 481 (98.99% ident. Halamphora), 497, 499, 587 (Achnanthes chlidanos isolate, 483, 74, 1.33e-131, 98.535, KJ658412 (Achnanthes chlidanos has lots of poor quality hits); SILVA 319, Bacillariophyceae, BLAST: Achnanthes bongranii (M.Peragallo) Mann 1937: 16, 100%, 2e-179, 95.53%; Achnanthes sp. 100%, 1e, 92.02%, good distance tree result); SILVA 1453, Coscinodiscophytina_cl (BLAST: <i>Achnanthes bongranii</i>, 608, 608, 100%, 2e-169, 94.04% - 1st in alignments, however, poor distance tree alignment)</p>							
2.	ACTI	<i>Actinoptychus</i> Ehrenberg 1843 No BLAST hits from the SILVA library of sequences (see Appendix D)	1				x		LM
3.	AMPH	<i>Amphora</i> Ehrenberg ex Kützing, 1844 SILVA #477 (BLAST: <i>Halamphora bicapitata</i> (Hohn & Hellerman) Stepanek & Kociolek, 2018 #477, 673, 673, 100%, 0.0e, 97.46%; <i>Amphora coffeiformis</i> , 673, 673, 100%, 0.0e, 97.46%); SILVA #2123, under the taxon Fragilariales --BLAST: <i>Amphora</i> sp. Kützing, F.T. (1844, 634, 634, 100%, 4e-177, 95.24% - good on distance tree results; <i>Amphora_sulcata</i> (MOL-43-V51); <i>Amphora_lineolata</i> (MOL-57-V67); <i>Amphora_abludens</i> (MOL-82-V97) <i>Amphora_helenensis</i> (MOL-221-V256); <i>Amphora_beaufortiana</i> (MOL-331-V386); <i>Amphora_vixvisibilis</i> (MOL-380-V437)	2	x	x	x (MOL-c onfid.)	x	SEM, LM Live (MOL-c onfid.)	

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
4.	ATTH	Attheya T.West, 1860 (MOL-431-V493) SILVA #1108 (BLAST: Attheya longicornis, 427, 427, 98%, 5e-115, 93.68%--Attheya spp. Sequences show significant close alignments to Attheya sequences in BLAST); Attheya_decora (MOL-349-V405); MOL 189-V222, 289-V336 Mediophyceae (<i>Attheya longicornis</i> , max score, 98%, 93.33; <i>A. septentrionalis</i> same score and percentages, but lower on the list-BLAST); Mediophyceae SILVA #2572 (BLAST: <i>Attheya longicornis</i> or <i>Attheya septentrionalis</i> : 422, 422, 98%, 2e-113, 93.33%— good on distance tree); SILVA #2573 BLAST: Attheya longicornis & <i>A. Septentrionalis</i> , 416, 416, 98%, 1e-111, 92.98%- excellent distance tree results)	1	x SEM		X (MOL-confid.)			SEM (MOL-confid)
5.	BACI – double check sequences	Bacillaria Gmelin, J.F. (1791) <i>Bacillariales</i> (MOL-53-V61) Under Bacillariophyceae - BLAST: Bacillaria paxillifer, 449, 449, 98%, 1e-121, 95.09%); MOL: <i>Bacillaria paxillifer</i> max score & 97% BLAST. Bacillariales (BLAST: 2e-188, 1e-116, 1e-111: Tryblionella, Nitzschia and Bacillaria paxillifer in that order—not very specific in the distance tree) Lots of entries for Cylindrotheca_closterium: 1e to 2e (BLAST; SILVA, Cylindrotheca closterium, 98%, 1e, 96.14%); Bacillaria paxillifer (BLAST: <i>B. paxillifer</i> 427, 427, 98%, 5e-115, 93.68%—distance tree results inconclusive) Note: Bacillariales refers to a family that has many genera, including Bacillaria, listed in Appendix A. Others in the family include Cylindrotheca, Nitzschia, Pseudo-nitzschia, Tryblionella, amongst others found in the identified genera.	2	x	x	X (MOL-confid.)	x	x	SEM, LM Live (MOL-confid.)
6.	CAMP	Campylocardia Kützing 1844 Found on: Zm_MHMPP-Mar 7-2021_slide 1-T42_4Db+c nitric acid (imaged May 2, 2023) (MW). DADA2 sequences: V55 (poor), V386 (moderate-poor distance tree), V452,V515, V2240 SILVA: 242, 434, 438 (too far away on nodes), 588, 2492, 3660, 3716, 2983.	1			X-LM (MOL-Possible)			LM
7.	CHAE	Chaetoceros Ehrenberg 1844 (MOL-1843-V200) Chaetoceros_diadema (MOL-401-V460); SILVA #2580, Chaetoceros cf.wighamii	1			X SEM (MOL-confid.)			SEM (MOL-confid.)

No.	Counting Codes	General	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		BLAST: Chaetoceros cf. wighamii and Chaetoceros sp. 739, 739, 100%, 0.0, 100.00% - excellent distance tree alignment; SILVA #2581, <i>Chaetoceros convolutus</i> , (BLAST: C. convolutus Castracane 1886 Castracane, 1886, 749, 749, 100%, 0.0, 100.00%- good distance tree alignment); SILVA #2582, Chaetoceros diadema, (BLAST: C. diadema, 745, 745, 100%, 0.0, 100.00%)							
8.	COCC	Cocconeis Ehrenberg 1836 Cocconeis (MOL-20-V24, 100-V117) Bacillariophyceae 243-V282 97%, 93.26%BLAST; #30 SILVA Feb 10-2023 Cocconeis (Cocconeis cf. cupulifera SZCZCH662) 98% 1e-121, 95.01%) (BLAST: <i>Cocconeis mascarenica</i> Riaux-Gobin & Compere, 2008, 477, 477, 98%, 5e-130, 96.84%)-good distance tree alignment, better than C. cf. cupulifera); 682-V780-Cocconeis, BLAST: likely C. Cocconeis stauroneiformis, 689, 689, 100%, 0.0, 98.22%; SILVA # 323, BLAST: C. stauroneiformis,— 711, 711, 100%, 0.0, 100.00%—perfect alignment score— and matching the query sequence on the distance tree results.	4	x	x	X (MOL-c onfid.)	x	x	SEM, LM Live (MOL-c onfid.)
9.	COSC	Coscinodiscus Ehrenberg, 1839 Coscinodiscus sp on Zm MHMPP-Nov 15-2020-environ-stub 4_m004.tif (Mar 7-2021) MQ-Feb 3, 2022 (not processed); SILVA #1452, Ok alignments for Coscinodiscus granii, 355, 355, 100%, 2e-93, 88.89%; SILVA #1345, Coscinodiscus granii 457, 914, 100%, 5e-124, 98.83%—which is a perfect match horizontally and node in the distance tree results; -(Mar 12-21_m002_2_edited-1.jpg; Box 8 stub 72b	2		x	X (MOL- confid.)			SEM (MOL-c onfid.)
10.	CYCL	Cyclotella (Kützing) Brébisson, 1838, nom. et typ. cons. BLAST searches: Weak match with SILVA sequences 2596, 2983, 3731.	1	x					LM
11.	CYLI	Cylindrotheca Rabenhorst, 1859, nom. cons. (MOL-58-V68) Cylindrotheca_closterium (MOL-5-V6); #231 SILVA, Feb 10-2023, Cylindrotheca_closterium (BLAST; C. closterium, 98%, 1e-131, 96.14%; 494, 494, 98%, 5e-135, 97.89%); SILVA #328,	2		x	x (MOL-c onfid.)	x		SEM Live (MOL-c onfid.)

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		BLAST: Cylindrotheca closterium, 726, 726, 100%, 0.0, 100.00%— excellent alignment score — and excellent result in distance tree results; found under a record “Bacillariophyceae”);							
12.	DIME	Dimeregramma Ralfs 1861: SILVA #2123, Found in the taxon Fragilariales—BLAST <i>Dimeregramma minor</i> var. <i>nanum</i> (W.Gregory) Van Heurck 1896, 571, 571, 100%, 3e-158, 92.36%—in the distance tree results; Note: For <i>Dimeregramma</i> the class is Bacillariophyceae, the order is <u>Plagiogrammatales</u> and the family is <u>Plagiogrammaceae</u> . So, something is not quite right here (same with finding <i>Amphora</i> sp. in the same alignments); #2591, BLAST: <i>Dimeregramma</i> sp. SZCZCH437, 747, 747, 100%, 0.0, 100.00%—excellent alignment stats—very good distance tree results, <i>Plagiogramma</i> sp., 747, 747, 100%, 0.0, 100.00%—) <i>Plagiogramma</i> is very slightly closer to the query than <i>Dimeregramma</i> on the distance tree results)	1			X (MOL – confid.)			LM (MOL-c onfid.)
13.	DIPL	Diploneis Ehrenberg ex Cleve, 1894 <i>Diploneis</i> (MOL-379-V436); SILVA #2755, BLAST: <i>Diploneis vacillans</i> (A.W.F.Schmidt) Cleve 1894 444, 444, 100%, 5e-120, 94.44%—distance tree results good); SILVA 2755 is very good alignment stats and distance tree results; SILVA #2850, <i>Diploneis vacillans</i> , 438, 438, 100%, 2e-118, 94.10%—good distance tree alignment); 1065-BLAST: <i>Diploneis</i> cf. <i>smithii</i> 701, 701, 100%, 0.0, 98.73%—very good distance tree results and alignment score)	1	x		X (MOL-c onfid.)			LM (MOL-c onfid.)
14.	DITY	Ditylum J.W.Bailey ex L.W.Bailey, 1861 Note: Ditylum external processes and valves are commonly found in the SEM images and slides of cleaned frustules. There are no entries in the molecular data for the Order Lithodesmiales, however 116 entries for the class Mediophyceae. BLAST: Weak matches with SILVA sequences 588, 1627, 1631.	2			X	x	x	SEM, LM
15.	DONK	Donkinia Ralfs, 1861	1			(MOL -uc)	x-uc		Live

No.	Counting Codes	General	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		Uncertain identification: only one live sample and it looks similar to <i>Pleurosigma directum</i> in valve view. Requires more samples and cleaned. Donkinia possibly showed up in the MOL under Naviculales MOL- 123-V143 BLAST , 100%, 93.42%, 3e-115; BLAST: Donkinia sp. 427, 427, 100%, 5e-115, 93.43%-good result in the distance tree spans. Also, good result for <i>Lithodesmium</i> sp.)							(MOL uc)
16.	ELLE	<i>Ellerbeckia</i> Crawford, R. M. 1988.: (MOL-328-V383). <i>Ellerbeckia</i> _sp. Silva #2983; <i>Ellerbeckia</i> _sp., BLAST #2983; 399, 399, 98%, 1e-106, 91.93%; good on distance tree results; (SILVA #2982-BLAST 98% 2e-108, 92.28%, 98% 1e-101, 90.88%)—best match on a distance tree is a Rhodophyta; <i>Gayliella</i> sp.; DADA2 V452, <i>Ellerbeckia</i> _sp., BLAST: 399, 399, 98%, 1e-106, 91.93%, good on distance tree, 2 nd node. <i>Ellerbeckia</i> Stb 1-Zm NA cleaned-MHMPP-Nov 15-20(x500)-TM4000-Dec 29-2020-2.tif; Box 9 16P 23 Feb 2022_m006.tif (Mar 7-2022) RR (unprocessed)-valve view; <i>Ellerbeckia</i> _Zm MHMPP_Mar 7-2021_Box 9 16P_RR (Feb 23 2022)_m006_3.tif	1		x	X-SEM (MOL-confd.)	x	SEM (MOL- -confd..)	
17.	ENTO- Double check sequences	<i>Entomoneis</i> Ehrenberg, 1845 (MOL-452-V515), #3660 SILVA Entomoneis cf. alta (BLAST: 98% 1e-126, 96.14%); found under Surirellales SILVA #3659 (BLAST: <i>Entomoneis</i> sp. strain TA350, 444, 444, 98%, 5e-120, 94.74%)-good distance tree & with lots of <i>Entomoneis</i> sp. listed first on alignments; SILVA 515, top of list 466, 466, 98%, 1e-126, 96.14%, good distance tree and lots of hits.	2	x		X SEM LM (MOL-confid.)	x		SEM (MOL-confid.)
18.	EPIT	<i>Epithemia</i> Kützing, 1844: Certain morphological. SILVA #3742 (BLAST: <i>Bacillaria</i> sp. 433, 433, 98%, 1e-116, 94.04%; <i>Epithemia sorex</i> Kuetzing 1844, 405, 405, 98%, 2e-108, 92.28%-distance tree shows close to <i>Epithema sorex</i> , but closer to <i>Bacillaria</i> ; (BLAST: unknown diatoms, <i>Epithemia pelagica</i> Schvarcz, Stancheva & Steward, 2022 , 468, 936, 100%, 3e-127, 99.61%-very good	1		x	X MOL (UC)			LM MOL (UC)

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		alignment stats, tree distance ok.—uncertain MOL.							
19.	EUCA	<i>Eucampia</i> Ehrenberg 1839	1				X		SEM
20.	ENCY/CYMB	<i>Encyonema</i> Kützing, F.T. (1834) (freshw only?.) (MOL-548-V620; & #29 98% e2 SILVA-BLAST-Feb 102-23); SILVA #29, BLAST: (<i>Encyonema prostratum</i> 392, 392, 97%, 2e-104, 91.67%—1st in alignments-and on distance tree results, & Or <i>Cymbella</i> (freshw.)(Mol 224-V260, 222-V258): So far, only a live cell has been imaged and possibly one SEM image. Until cleaned frustules are found, it's generic placement is uncertain. It is unlikely to be <i>Cymbella</i> due to its plastid shape and it lacks a large central pyrenoid. ²¹ Both <i>Encyonema</i> and <i>Cymbella</i> are reported by Plientz et al. (2003) and <i>Cymbella</i> by Riznyk (1973), Shim (1976) and Tynni (1986) in Salish Sea marine habitats. [<i>Encyonema</i> present in March 7 samples, confirmed by molecular data-genus only]. The order Cymbellales is also confirmed in molecular data (Mol 224-V260, 222-V258, 280-V326, mostly <i>Cymbella lanceolata</i> & <i>Cymbella cistula</i>); (BLAST: <i>Encyonema silesiacum</i> , 590 590, 100%, 8e-164, 93.70%- very good distance tree results	1		X-(MOL-confid.)	x-uc	SEM Live (MOL- -confid.)		
21.	FALL	<i>Fallacia</i> Stickle et Mann in Round et al. 1990. Fig. 9. <i>Fallacia</i> (MOL-194-V22); <i>Fallacia laevis</i> (MOL 79-V94,100%, 93.75% BLAST); <i>Fallacia hodgeana</i> , #509 SILVA (BLAST:, 100% 0e, 96.28%); 256-V298 Bacillariophyceae (<i>Fallacia litoricola</i> isolate, max score 100%, 98.61% BLAST); <i>Fallacia litoricola</i> isolate, max score 100%, 98.26%, same for <i>F. hodgeana</i> and <i>F. bosoensis</i> at 97.92 and 97.57% respectively found under Bacillariophyceae – BLAST; MOL-297-V347-Bacillariophyceae (<i>Fallacia litoricola</i> isolate, max score 100%, 97.92%, same <i>F. bosoensis</i> and <i>F. hodgeana</i> at 97.22 and 96.88% respectively_ BLAST); <i>Fallacia</i> (BLAST: <i>Fallacia laevis</i> Li & Suzuki, 2022, 433, 433,			X-(MOL-confid.)		SEM (MOL-confid.)		

²¹ Round et al. (1990).

No.	Counting Codes	General	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		100%, 1e-116, 93.75%—distance tree results for <i>Fallacia</i> sp. is a good alignment).							
22.	FOGE	<i>Fogedia</i> Witkowski, Lange-Beralot et Metzeltin 1997. Fig. 7-8. Genus not found in any of the SILVA sequences				X LM			LM
23.	FRAG	<i>Fragilariopsis</i> Hustedt 1913 Fragilariopsis?-Zm MHMPP-stb-T45 distal-cleaned Mar 7-2021-TM4000-MW-(Sept 4-2021)_013(x4.0k)_2.tif1913 (MOL-444-V507). <i>Fragilariopsis_kerguelensis</i> (O'Meara) Hustedt (MOL-446-V509); BLAST: F. kerguelensis, 460 460, 98%, 5e-125, 95.79%—excellent distance tree results. <i>Fragilariopsis</i> or <i>Nitzschia</i> on Zm stub 20 En-7-medial-Mar 7-2021 (MW) (Aug 24-2021)_035(x3.0k)_2.tif				X- (MOL-confid.)			SEM (MOL-confid.)
24.	GOMP	<i>Gomphonemopsis</i> Medlin, 1986 <i>Gomphonemopsis_pseudoexigua</i> (MOL-449-V512) <i>Gomphonemopsis_pseudoexigua</i> (BLAST: 499, 499, 100%, 1e-136, 97.92%- excellent distance tree result. SILVA #1524, BLAST: <i>Gomphonemopsis pseudoexigua</i> 499, 499, 100%, 1e-136, 97.92%--excellent distance tree result.	4		X	X SEM LM (MOL-confid.)			SEM LM (MOL-confid.)
25.	GOMS	<i>Gomphoseptatum</i> Medlin, 1986 (uc-only one SEM image)	1			X-uc			SEM (uc)
26.	GRAM	<i>Grammatophora</i> Ehrenberg 1840. Note: This genus is commonly found in samples, however not yet found in the molecular data, yet there 1,507 entries for the Bacillariophyceae in which <i>Grammatophora</i> is a sub-taxon.	2		X	X-			SEM LM Live
27.	GYRO-Scan database for any good results	<i>Gyrosigma</i> Hassall, 1845, nom. cons: 96-V91, BLAST: <i>Trachyneis</i> sp., 3rd on list of alignments, 449, 449, 100%, 1e-121, 94.79%—good score, ok distance tree result); (BLAST: <i>Gyrosigma limosum</i> Sterrenburg & Underwood 1997 & <i>Gyrosigma acuminatum</i> (Kuetzing) Rabenhorst, 699, 699, 100%, 0.0, 97.78%-some gaps, good alignment stats, distance tree is OK.; found in SILVA #279, 349, 407, 449, 500 (good), 576, 597, 671, 672, 673, 674, 676, 1621, 2149, 2692. SILVA 2706,	1		X (MOL-prob.)				SEM (MOL-prob.)

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		Naviculales (BLAST: <i>Gyrosigma acuminatum</i> (Kuetzing) Rabenhorst, 466, 466, 98%, 1e-126, 96.14%, poor distance tree result, though Gyrosigma was near the top of the matches. Pleurosigma is a better match.							
28.	HALA	<i>Halamphora</i> (Cleve) Mereschkowsky, 1903: SILVA #425, BLAST: <i>Halamphora nagumoi</i> Stepanek et al. in Stepanek & Kocolek, 2018, 449, 449, 98%, 1e-121, 95.09%—good distance tree results); SILVA #477 (BLAST: <i>Halamphora bicapitata</i> (Hohn & Hellerman) Stepanek & Kocolek, 2018, 673, 673, 100%, 0.0, 97.46%; <i>Amphora coffeiformis</i> , 673, 673, 100%, 0.0, 97.46%—very good distance tree results) <i>Halamphora</i> (MOL-174-V205, 218 V252); <i>Halamphora_montana</i> (MOL-354-V410) As we learn to distinguish between <i>Amphora</i> and <i>Halamphora</i> , more <i>Amphora</i> identifications will be moved to <i>Halamphora</i> . However, <i>Halamphora coffeiformis</i> (formerly <i>Amphora coffeiformis</i>) appears to be common on all <i>Z. marina</i> samplings.	2	x	x	X (MOL - confid.)			SEM (MOL - confid.)
29.	HANZ	<i>Hanzschia</i> Grunow, A. 1877 (<i>H. vergata</i> , only one LM image, not enough images). No sequences matching Hanzschia in all SILVA data.	?	X-uc					LM-uc
30.	HASL	<i>Haslea</i> Simonsen, R. (1974): Haslea on Zm-Mar 7-2021-leaf 4 T42 4Db_c Nit A-E800-MU2003-Mar 25-2022-slide 1-0079a_2.tif; <i>Haslea crucigera</i> _Box 11-Zm Mar 7-2021-stb2 T45 SA1-H2O2-TM4000 MW-Oct 24-2022-24_4(x1).tif <i>Haslea</i> (MOL-25-V31); <i>Haslea_howeana</i> or <i>H. avium</i> (MOL-60-V70 BLAST 3e-135) <i>Haslea_howeana</i> (MOL-84-V99); #2734 SILVA <i>Haslea_howeana</i> ; <i>Haslea_pseudostrearia</i> (MOL-313-V367); <i>Haslea</i> cf. <i>howeana</i> -BLAST: <i>Haslea</i> cf. <i>howeana</i> , 510, 510, 100%, 5e-140, 98.61% - marginal on distance tree; SILVA 2799, BLAST: <i>Haslea</i> cf. <i>howeana</i> 510, 510, 100%, 5e-140, 98.61% excellent alignment score- good on distance tree, however Navicular is slightly closer; 76-V91 <i>Haslea nipkowii</i> , 455, 455, 100%, 2e-123, 95.14%—ok distance tree, <i>Navicular</i> sp. a				X LM SEM (MOL-confid.)		LM SEM (MOL -confid.)	

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		bit closer, but Haslea has a better alignment match; 84-V99 (BLAST: <i>H howeana</i> , 510, 510, 100%, 5e-140, 98.61%—excellent alignment scores, top of list, but slightly distant on distance tree results; SILVA 501, SILVA 2710, Haslea, BLAST: 505, 505, 100%, 2e-138, 98.26%—good distance tree results)							
31.	HOBA Search MOL 2022 records (SILVA has been searched)	<i>Hobaniella</i> P.A.Sims & D.M.Williams, 2018: synonym is <i>Odontella longicrucis</i> -monogenic, (rare) July 22, 2021 (Aug. 4, 2020), slide 1a, 0194-0240, slide 1b, 0126-0160. Molecular data confirms <i>Odontella</i> sp. MOL 464-V529, (BLAST: <i>Odontella</i> sp. 743, 743, 100%, 0.0, 99.75%; <i>Odontella longicurvis</i> 722, 722, 100%, 0.0, 99.51%, also V529 , BLAST: <i>Ralfsiella smithii</i> (Ralfs) Sims, Williams & Ashworth, 2018, 743, 743, 100%, 0.0, 99.75%; <i>Odontella</i> sp. 743, 743, 100%, 0.0, 99.75%; <i>Odontella longicurvis</i> 722, 722, 100%, 0.0, 99.51% — <i>O. longicurvis</i> found in distance results better than <i>O. aurita</i> .) which could be <i>Odontella longicrucis</i> = <i>Hobaniella</i> : <i>Hobaniella</i> found in SILVA 2587, <i>Hobaniella longicurvis</i> with a 0-e, 98.27% alignment, however distance tree results are stronger for <i>O. aurita</i> . Conclusion:confident on the genus Hobaniella as it is a synonym for Odontella which is a confident match of sequences.	1	x		X-LM (MOL-confid. On genus)		LM (MOL-confid. On genus)	
32.	HYAL	<i>Hyalodiscus</i> Ehrenberg (1845) <i>Hyalodiscus_scoticus</i> (MOL-70-V82); #2603 SILVA, <i>Hyalodiscus_scoticus</i> ; <i>Hyalodiscus_scoticus</i> (BLAST: <i>Podosira stelligera</i> 527, 527, 100%, 5e-145, 99.65%; <i>Hyalodiscus_scoticus</i> 527, 527, 100%, 5e-145, 99.65% — alignment stats and distance tree results are same for <i>P. stelligera</i> and <i>H. scoticus</i> ; SILVA #1459 (BLAST: <i>H. scoticus</i> , 779, 779, 100%, 0.0, 100.00%, same stats for <i>Podosira stelligera</i> synonym = <i>H. stelligera</i> — perfect match and distance in distance tree results for both genera/species.)	2		x	X-SEM (MOL-confid.)	x	x?	SEM LM Live (MOL-confid.)
33.	LEPT	<i>Leptocylindrus</i> Cleve, 1889 <i>Leptocylindrus_danicus</i> (MOL-384-V441); # 2492 SILVA, <i>Leptocylindrus danicus</i> , <i>Leptocylindrus_danicus</i> BLAST: 472, 472, 100%, 2e-128, 96.18%-excellent distance	1			X-SEM confid.			SEM (MOL-confid.)

No.	Counting Codes	General	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		tree results ; BLAST: L. danicus 472, 472, 100%, 2e-128, 96.18%—good to excellent distance tree results)							
34.	LICM	Licmophora C.Agardh, 1827, nom. et typ. cons. (MOL-1745-VV189); (BLAST: Licmophora paradoxa and Licmophora abbreviata, 745, 745, 100%, 0.0, 100.00%) Licmophora_paradoxa (MOL-59-V69, 213-V247 100% BLAST) SILVA data has many entries of Limporphora including 2 entries for Licmophora_paradoxa #2494 and 2496; SILVA #2497 BLAST: L. abbreviata 466, 466, 100%, 1e-126, 95.85%—distance tree results good); SILVA # 2127, Licmophora juergensii Agardh 1831 BLAST: L. juergensii Agardh 1831, 47, 747, 100%, 0.0, 100.00%—excellent distance tree results); SILVA 2126, BLAST: Licmophora paradoxa and Licmophora abbreviata, 745, 745, 100%, 0.0, 100.00%-very excellent distance tree results)	2-3			X- SEM (MOL-confid.)	x	x	SEM LM Live (MOL-confid.)
35.	LYRE	Lyrella (Ehrenberg) Karayeva 1978: Lyrella_hennedyi (MOL-216-V250); SILVA 30 is marginal result; SILVA #40, Feb 10-2023 (BLAST, possibly Lyrella_hennedyi 100%, 1e, 93.77%); #2517 SILVA, BLAST: Lyrella_hennedyi 438, 438, 100%, 2e-118, 94.10%, but 1st entry is Petroneis humerosa, 395, 395, 100%, 2e-105, 94.44% with a good alignment score. Next scores are Lyrella, owerer, distance tree result for Lyrella is very good, same node as query sequence, much better than Petroneis) ; Lyrella sp_Box 11-Zm Mar 7-2021-stb2 T45 SA1-H2O2-TM4000 MW-Oct 24-2022-39_2(x3.0k).tif	1	x	?	X- LM SEM (MOL-confid.)			SEM LM (MOL-confid..)
36.	MELO	Melosira C. Agardh, 1824, nom. cons. (MOL-916-V101, 11 hits): SILVA #1623, #1622, (BLAST: Melosira nummuloides 656, 656, 100%, 0.0, 96.26% - good to excellent distance tree result); 916-BLAST: Melosira sp.446, 446, 100%, 2e-120, 86.85%—good alignment stats, maybe <i>Melosira capsularum</i> Yang & Wang, 2022—but is freshwater); 1263 BLAST: Melosira sp. 446, 446, 100%, 2e-120, 86.85%—top alignment stats—could not find in distance tree, <i>Melosira moniliformis</i> 100% 3e109,	2-3	x	x	X- (MOL-confid.)	x	x	SEM LM Live (MOL-confid.)

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		85.11%—good alignment stats); SILVA 1454, Melosira (BLAST: 736, 736, 100%, 0.0, 99.75%, excellent distance tree results);							
37.	MINI	<i>Minidiscus</i> Hasle, 1973 <i>Minidiscus_triloculatus</i> (MOL-17-V21, 225-V261); SLIVA #3732, <i>Minidiscus_triloculatus</i> (BLAST: T. <i>triloculatus</i> , 483, 483, 100%, 1e-131, 96.88%—good distance tree result); SILVA # 2584 (BLAST: <i>Minidiscus triloculatus</i> (F.J.R.Taylor) Hasle 1973749, 749, 100%, 0.0, 100.00% - excellent distance tree results); SILVA #2585, (BLAST: <i>Minidiscus variabilis</i> Kaczmarśka in Kaczmarśka et al. 2009, 749, 749, 100%, 0.0, 100.00%—distance tree reflects close distance to <i>Minidiscus triloculatus</i> .	1			X- SEM (MOL-confid.)			SEM (MOL - confid.)
38.	NAVI	<i>Navicula</i> Bory 1822: individual, chain and tube dwelling species: <i>Navicula_rhynchocephala_var._hankensis</i> (MOL-2-V3_1e-139); <i>Navicula_kongfjordensis</i> (MOL-3-V4); <i>Navicula_perminuta</i> (MOL-6-V9); <i>Navicula_avium</i> (MOL-8-V11); <i>Navicula_pseudacceptata</i> (MOL-44-V52); <i>Navicula_glaciei</i> (MOL-108-V125); <i>Navicula_ramosissima</i> (MOL-141-V166); <i>Navicula_slesvicensis</i> (MOL-167-V197); <i>Navicula_reinhardtii</i> (MOL-415-V475) <i>Navicula_hippodontafallax</i> (MOL-266-V310); <i>Navicula_rhynchotella</i> (MOL-301 V351); <i>Navicula_cari</i> (MOL-302-v352); SILVA # 2147 (BLAST: various species of <i>Navicula</i> . <i>Navicula phyllepta</i> Kützing 1844, 520, 520, 100%, 1e-142, 89.90%—not in the distance tree results. Best tree result is close to <i>Navicula trivialis</i> and <i>Navicula salinarum</i> Kolbe 1927; <i>Navicula perminuta</i> Grunow, BLAST: N._perminuta, 532, 532, 100%, 1e-146, 100.00%—excellent distance tree result); SILVA 2694, <i>Navicula_rhynchocephala</i> var. <i>hankensis</i> (BLAST: <i>Navicula</i> sp. 532, 532, 100%, 1e-146 ,100.00%, very excellent distance tree results;	4	x	x	X- LM SEM (MOL-confid.)	x	x	SEM LM Live (MOL-confid.)
39.	NITZ	<i>Nitzschia</i> Hassall, 1845, nom. cons.	4	x	x	X- SEM LM	x		SEM LM Live

No.	Counting Codes	General	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		Nitzschia dissipata, #232, #270, SILVA Feb. 10-2023: (BLAST: Nitzschia dissipata, 100%, 1e, 93.77%); Nitzschia_dradeillensis (#308, SILVA, (BLAST: 98% 1e); Nitzschia_dubiiformis (MOL-18-V22) & #233 SILVA Feb. 10-2023; Nitzschia frustulum (#252, SILVA, Feb 10-2023: BLAST: 100% 1e; Nitzschia_inconspicua (#252, SILVA, Feb 10-2023: BLAST 100% 1e; Nitzschia_laevigata (MOL-85-V101) & SILVA #257, #268 (98%, 2e); Nitzschia_microcephala (MOL-350-V406); Nitzschia_placea (#261, SILVA) Nitzschia_promare (MOL-99-V116); #259 SILVA Nitzschia_pumila (MOL-197-V230); Nitzschia_recta (MOL-36-V43); #232 SILVA Nitzschia cf. recta 100%, 1e, 98%; Nitzschia_reskoi (#296, SILVA, 98% 1e, 96.14%); Nitzschia_sigmoidea (MOL-38-V45); #235 SILVA, Feb 10-2023 (BLAST: Nitzschia sigmoidea 100%, 1e); Nitzschia_spathulata (MOL-86-V102, 308-V359) & morphology, #269, SILVA (100% 2e); Nitzschia_volvendirostrata (MOL-97-V144); #249, #258 & #265 (2e) SILVA Feb 10-2023; SILVA 232, Nitzschia_dissipata (BLAST: Nitzschia cf. recta 466, 466, 100%, 1e-126, 95.83%, very excellent distance tree results for the genus)				(MOL--confid.)			(MOL-confid.)
40.	ODON	Odontella (Lyngbye) C Agardh Odontella_aurita (BLAST: O. aurita, 516, 516, 98%, 1e-141, 99.30%—excellent alignment and excellent in distance tree result) (MOL-464-V529) Odontella_aurita (MOL-48-V56, MOL 260-V304); SILVA # 2587 BLAST: O. aurita, perfect match in sequence alignment to databases and distance tree)	2	x	x	X- (MOL-confid.)			SEM LM Live (MOL-confid.)
41.	OPEP	Opephora P. Petit, 1889 Opephora pacifica_Box 11-Zm Mar 7-2021-stb2 T45 SA1-H2O2-TM4000 MW-Oct 24-2022-10_3-(x2.0k).tif. Opephora sp. #431, SILVA, Bacillariophyceae (BLAST: Opephora sp., 494, 494, 100%, 5e-135, 97.57%-excellent	2-3	x	x	X- SEM LM, (MOL-confid.)			SEM, LM, (MOL-confid.)

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		distance tree match); Bacillariophyceae (BLAST: Opephora sp. 532, 532, 100%, 1e-146, 100.00%, very excellent distance tree result); SILVA 2130, 499, 100, 1.01e-136, 97.917, KR048204, Opephora sp. SZCZCH153 good alignment stats; SILVA 2132, 516, 100, 1.01e-141, 98.958, KR048204, Opephora sp. SZCZCH153; Note: It is possible that some of the <i>Opephora</i> observed are similar looking species of <i>Serratifera</i> and <i>Gedenella</i> found in the molecular data, but not yet confirmed by morphology.							
42.	PARA	Paralia Heiberg, 1863 Note: This genus is commonly found in samples, can even be seen at 50x SEM overview images, however it has not been found in the Feb. 25, 2022 molecular data. Yet there are 8 entries for the class Coscinodiscophyceae in which <i>Paralia</i> is a sub-taxon. #1345 SILVA = Paralia sulcate. Sequence data using BLAST on Coscinodiscophyceae (98-V115) gave only readings for the genus Paralia (taxid:216907 & 216927): query cover of 100% for Paralia sulcate and 85% for Paralia fenestrata.; 1405-V153, BLAST: Paralia longspina or Paralia sulcata, 100%, e0, 99.75% ident; Paralia sulcata, 473, 947, 100%, 5e-129, 100.00%; SILVA #1458, (BLAST: Paralia longspina. 741, 741, 100%, 0.0, 99.75%—excellent alignment and excellent distance tree results.	2	x		X (MOL-confid.)		SEM (MOL--confid.)	
43.	PARI	Paribellus Cox 1988. Fig. 10.: Paribellus_delognei_f._ellipticus (MOL-49_V57); SILVA 2693; Paribellus_protracta (BLAST: Prestauroneis integra, 468, 468, 96%, 3e-127, 96.79%—uncertain tree placement for the genus) Paribellus_protracta (MOL-145-V170); SILVA # 2732 Paribellus_delognei_f._ellipticus (BLAST: <i>Paribellus delognei f. ellipticus</i> (Lobban) E.J.Cox, 499, 499, 98%, 1e-136, 98.25%—distance tree results are excellent)		x		(MOL-confid.)		LM (MOL-confid.);	
44.	PERI	Periraphis like. (<i>Periraphis nom. nud.</i>). Previously listed as <i>Catenula-Catenulopsis</i> , but it's taxonomic placement is still	2-3			X-uncer-tain taxon-omi-c		SEM	

No.	Counting Codes	General	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		uncertain but moving closer to a new genus called <i>Pairaphis</i> . ²² SILVA 320, BLAST: Catenulaceae sp., 725, 725, 100%, 0.0, 100.00%—Pinnularia sp. has same excellent alignment stats, then less exact fits are Halamphora, Amphora, Diploneis, and Brachysira. On distance tree results, this sequence is close to Biremis sp. and Diploneis sp.)				place-me nt			
45.	PETR	Petroneis A. J. Stickle & D. G. Mann, 1990 Showed up SILVA #2517, BLAST: under Lyrella_hennedyi, but 1 st significant alignment is Petroneis humerosa P. humerosa, 395, 395, 100%, 2e-105, 94.44% with a very good alignment score and an excellent horizontal distance tree result by eliminating other diatoms from the search); SILVA #37, P. humerosa, Max score 444, Total Score 100%, 3e-120 & Per. Ident. 94.44%, very good distance tree result.; SILVA #287, BLAST Petroneis humerosa, 462, 462, 100%, 2e-125, 94.44%)	1-2	x		X (MOL- confid.)			LM (MOL-c onfid.))
46.	PLAGG	Plagiogramma Greville, 1859 (MOL-2059-V223, e value 0.0) <i>Plagiogramma on Zm MHMP-Mar 7-2021-Box 1b T32_6_1Mb_c MQ-Mar 29-2022_m003_2.tif</i> Plagiogramma tsawwassen 100%, 92.01% BLAST); SILVA #2591 BLAST: Plagiogramma sp., 747, 747, 100%, 0.0, 100.00%—for sure #2591, BLAST: <i>Dimeregramma</i> sp. SZCZCH437, 747, 747, 100%, 0.0, 100.00%—excellent alignment stats—very good distance tree results, Plagiogramma sp., 747, 747, 100%, 0.0, 100.00%—) Plagiogramma is very slightly closer to the query than Dimeregramma on the distance tree results)	1			X- (MOL-c onfid.)	x		LM SEM (MOL-c onfid.)
47.	PLAGT	Plagiotropis Pfitzer, 1871, nom. illeg. <i>Plagiotropis</i> sp. MOL 220-V255, BLAST: Plagiotropis sp. strain 450, 450, 100%, 6e-122, 93.75% - alignment score is OK, match for query sequence. SILVA 379, Bacillariophyceae (BLAST: Plagiotropis sp., 433, 433, 100%, 1e-116, 93.75%, 1 st in alignment score, uncertain, distance tree result is not confid.;	2	x	x	X (MOL- prob.)			SEM LM Live (MOL-pr ob.)

²² A. Witkowski, 2022, pers. commun.

No.	Counting Codes	General	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
48.	PLAN	<p><i>Planothidium</i> Round and Bukhtiyarova 1996. Fig. 1-3, 5-6 (MOL-584-VV657); MOL4-V5 <i>Planothidium lanceolatum</i> strain D16_002 (98% BLAST_2e-108); #33, SILVA, Feb 1—2023: <i>Planothidium</i> sp. BLAST: 98% 1e, <i>Pauliella_taeiata</i> 98% 1e); #39, SILVA, Feb 10-2023 (BLAST-<i>Planothidium lanceolatum</i> 100%, 2e); SILVA # 582, BLAST: <i>Planothidium</i> sp. SZCZCH26, 717, 717, 100%, 0.0, 99.00%—excellent distance tree results); SILVA # 335, BLAST: <i>Planothidium lanceolatum</i> (Brebisson ex Kuetzing) Lange-Bertalot 1999, homotypic synonym: <i>Achnanthidium lanceolatum</i>, 434, 434, 100%, 6e-117, 93.40%—perfect alignment score, excellent alignment result and distance tree results have excellent horizontal results for many <i>Achnanthidium</i> sp.); SILVA 467, Bacillariophyceae. BLAST: <i>Planothidium</i> confid. and excellent distance tree results)</p>	1-2			X (MOL-confid.)			SEM (MOL-confid.)
49.	PLEU	<p><i>Pleurosigma</i> W. Smith, 1852, nom. et typ. cons. (MOL-55-V64-BLAST: (BLAST: <i>Pleurosigma</i> sp. 477, 477, 100%, 5e-130, 96.53%)—for sure. Distance tree good. Maybe <i>P. stuxbergerii</i> isolate; SILVA #671, <i>Pleurosigma intermedium</i> var. <i>mauritianum</i> (Grunow ex Cleve) M. Peragallo 1903 (BLAST: 743, 743, 100%, 0.0, 99.75%, distance tree results excellent and close to query sequence).</p>	2		x	X (MOL-confid.)			SEM LM Live (MOL-confid.)
50.	PODO	<p><i>Podosira</i> Ehrenberg, 1840: (<i>morphologically confirmed, with one exterior and one interior valve views; Podosira</i> sp. <i>Zm</i> MHMPP(Mar 7-2021-Box 1b Stub T32-7 March 31 2022_m021_2.tif, <i>Podosira</i> sp-<i>Zm</i>, MHMPP-Mar 7-21-T57 9D C H2O2 Hot-TM4000-before(x2.0k) July 23-2021-ECH_2.tif SEM images from March 7, 2021 and an LM live image from Oct 2020): There is confident molecular evidence for it being in the Mar. 7, 2021 eelgrass sample. MOL-98-V115, BLAST: 405, 405, 100%, 2e-108, 92.01%, 1st alignment line, good distance tree result, under Coscinodiscophyceae = <i>Podosira baldjickiana</i> Sequence data under Coscinodiscophyceae : 98-V115 = <i>Podosira baldjickiana</i> 405, 405, 100%, 2e-108,</p>	1		x-uc	SEM-uc (MOL-confid.)			SEM, Live, (MOL-confid.)

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		92.01%—Abe,K., Lam,D.W., Ashworth,M.P., Harwood,D.M. and Jordan,R.W. (2022). Observations on Praecorethron crawfordii gen. et sp. nov. (Corethrales, Bacillariophyceae) from Upper Cretaceous marine sediments, southwest Pacific. Nova Hedwig Beih 151 (2022) In press; SILVA #1452, <i>Podosira baldjickiana</i> (BLAST: <i>P. baldjickiana</i> Grunow, 1888, 405, 405, 100%, 2e-108, 92.01%-good distance tree alignment); SILVA #2603 under <i>Hyalodiscus_scoticus</i> -BLAST: <i>Podosira stelligera</i> 527, 527, 100%, 5e-145, 99.65% & <i>Hyalodiscus_scoticus</i> 527, 527, 100%, 5e-145, 99.65% —alignment stats and distance tree results are same for <i>P. stelligera</i> and <i>H. scoticus</i>); SILVA 705, Bacillariophyta (BLAST: <i>Podosira baldjickiana</i> Grunow, 1888, 399, 399, 100%, 1e-106, 91.67%, very good distance tree result)							
51.	PSAM	<i>Psammodictyon</i> D. G. Mann, 1990 <i>Psammodictyon</i> (MOL-403-V462); <i>Psammodictyon</i> sp. (#310, SILVA-Feb 10, 2023 <i>Psammodictyon</i> _sp. (BLAST: <i>Psammodictyon panduriforme</i> var. <i>continuum</i> , 516, 516, 98%, 1e-141, 99.30%—excellent top alignments scores with multiple entries and very excellent distance tree results to query seq.) Note: only one entry record to date with 15 hits.	1-2			X SEM (MOL-confid.)			SEM (MOL-confid.)
52.	PSEUG	<i>Pseudogomphonema</i> Medlin 1986 <i>Pseudogomphonema_kamschaticum</i> (MOL-12-V15. 20 entries in the SILVA molecular data, e.g. #2701. Entries in the orginal Mol data have generated a species name with an 2e value, i.e.: <i>Pseudogomphonema_kamschaticum</i> (BLAST: 521, 521, 98%, 2e-143, 99.65%); SILVA #2701, BLAST: <i>P. kamschaticum</i> , 521, 521, 98%, 2e-143, 99.65%-top 3 alignments and excellent score and excellent distance tree result); <i>Navicula</i> (BLAST: <i>Pseudogomphonema</i> sp. LM-2002, 726, 726, 100%, 0.0, 99.25%, excellent distance tree result);	4		x	X (MOL-confid.)			SEM LM (MOL-confid.)
53.	PSEUN	<i>Pseudo-nitzschia</i> H. Peragallo, 1900 Note: found in the molecular data, SILVA 1333, BLAST: <i>Pseudo-nitzschia</i>	1-2	x		(MOL-confid.)			SEM (MOL-confid.)

No.	Counting Codes	General	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		<p>hainanensis Chen & Li, 2021, 427, 855, 99%, 4e-115, 96.86%—top of list, OK to good alignment, fair distance tree results; SILVA 487, Bacillariophyceae_fa (BLAST: Bacillaria sp. and B. Bacillaria paxillifer, 737, 737, 100%, 0.0, 99.50%—very excellent on distance tree results); There are 158 entries for the genus <i>Bacillaria</i> in which <i>Pseudo-nitzschia</i> is closely related. <i>Pseudo-nitzschia</i> was found under a search of a sequence for SILVA #302, <i>Nitzschia_microcephala</i>, BLAST- found <i>Pseudo-nitzschia</i> way down the list: 411, 411, 98%, 5e-110, 92.63%—but good alignment score and shows up somewhat distant and many nodes away on the distance tree-PDF file saved of the tree. <i>Pseudo-nitzschia</i> was only seen on Aug. 4, 2020 Z. marina leaves. Also shows up in SILVA #275, but far down the alignment list.and could not find in distance tree results; SILVA #484, (BLAST: <i>Pseudo-nitzschia</i> sp. 702, 702, 100%, 0.0, 98.26%—found far down the alignment list, but stats are good; SILVA #485 BLAST: <i>Bacillaria</i> sp. and B. <i>Bacillaria paxillifer</i>, 737, 737, 100%, 0.0, 99.75%—very good on distance tree, <i>Pseudo-nitzschia</i> sp. 686, 686, 100%, 0.0, 97.51%; SILVA # 487, BLAST: <i>Bacillaria</i> sp. and B. <i>Bacillaria paxillifer</i>, 737, 737, 100%, 0.0, 99.50%—very good on distance tree, <i>Pseudo-nitzschia</i> sp. 691 691 100% 0.0, 97.76%- far down in the alignment runs.</p>							
54.	RHAB	<p><i>Rhabdonema</i> Kützing, 1844, nom. cons.: Aug 4, 2021 slide 1b E800 cleaned 0076-0101, valve view. MOL-259-V302 Found under Fragilarophyceae (BLAST: <i>Rhabdonema arcuatum</i>, 521, 521, 98%, 2e-143, 99.65%—good alignment at the top of the list, excellent distance tree results) Not found yet in the SILVA molecular data; Fragilarophyceae (BLAST: <i>Rhabdonema arcuatum</i> (Lyngbye) Kuetzing 1844, 521, 521, 98%, 2e-143, 99.65%, very excellent distance tree results)</p>	2	X	X	X-(MOL-confid.)	X		Live LM (MOL-confid.)
55.	RHIZ	<p><i>Rhizosolenia</i> Brightwell, 1858, nom. et typ. Cons; <i>Rhizosolenia</i> was not listed in the MH table of sequences from Feb 25, 2022. <i>Rhizosolenia</i> is found in the SILVA</p>	2			(MOL-u c)	X		SEM LM (MOL-u c)

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		molecular data in three sequences: 350, 369 and 371; all three sequences give poor alignment and distance tree results. Found distantly in SILVA 2755.							
56.	RHOI	Rhoicosphenia Grunow, 1860: Bacillariophyceae_fa SILVA 519, BLAST: Rhoicosphenia cf. abbreviata, 715, 715, 100%, 0.0, 99.24%—3 base pair difference to query seq., top of alignment list and top 12, excellent distance tree result. Note: Rhoicosphenia is one of the most abundant diatoms on <i>Z. marina</i> medial and distal sections, however to date, only one molecular sequence hit with excellent and confid. stats has been found and all the diatom sequences in SILVA have been reviewed. Another sequence, SILVA 2755 is all low quality matches.	4	x	x	x (MOL-confid.)	x	SEM, LM, Live, (MOL-confid.)	
57.	RHOP	Rhopalodia O. Müller, 1895, nom. cons. (unconfirmed-only one image). No sequence matches found in all of the SILVA data.	1		X-UC				LM-uc
58.	SHIO	Shionodiscus Alverson, A.J., Kang, S.-H. & Theriot, E.C. (2006): SILVA #2583 Thalassiosira (BLAST: Minidiscus sp. & Thalassiosira bioculata (Shionodiscus is the strongest match and in distance tree results, 749, 749, 100%, 0.0, 100.00%—more hits for T. bioculata and excellent results for T. bioculata, however T. oestrupii (Shionodiscus oestrupii) and Shionodiscus biocultus are equal for distance result and very excellent in distance tree result. Shionodiscus species are common in the Salish Sea.				(MOL-confid.)		(MOL-confid.)	
59.	SKEL	Skeletonema Greville, 1865, nom. et typ. cons.(MOL-2149-V232) Skeletonema_costatum (MOL-39-V47, 219-V254); SILVA #2592 (BLAST: S. costatum or S. marinii; 745, 745, 100%, 0.0, 100.00%); SILVA 2593, 2594, 3727 are also excellent matches; 39-V47 (BLAST: S. costatum, 527, 527, 100%, 5e-145, 99.65%—excellent alignment score and excellent distance tree results)	2	x		x (MOL-confid.)	x	SEM (MOL-confid.)	
60.	STEP Double check	Stephanopyxis Ehrenberg, C.G. (1845) (synonym Eupyxis Blanco, S. & Wetzel, C.E. (2016))	1			x (MOL-prob.)		SEM (MOL-prob.)	

No.	Counting Codes	General	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
	sequences	(MOL-1405-V153, Stephanopyxis, 608, 608, 100%, 2e-169, 93.84%; DADA2-V1536, V2169, V2120, V3190, V1039 (all mediocre quality matches). (<i>S. turris</i> and <i>S. nipponica</i> are commonly observed diatoms in Trincomali Channel, Galiano Island waters) MOL (1 st sheet) SEM- March 7, 2022 T32_7 (RR-Mar. 16-2022); SILVA #1458, BLAST: Stephanopyxis, 608, 608, 100%, 2e-169, 93.84%—good for distance tree results.							
61.	TABU	<i>Tabularia</i> (Kützing) D.M.Williams & Round, 1986 (MOL-1298-V141) Tabularia_tabulata (MOL-45-V53, and a close match to 287 V334); 3 species that have been identified by morphology are also in the MOL data as Fragilaria (MOL: MOL-748-V831 Fragilaria (BLAST: Tabularia sp. 100% 0e, 99.75; Tabularia laevis 100% 0e, 99.75; Tabularia tabulata 100%, 0e, 99.75; T. fasiculata 100%, e0, 98.77%; Also for SIVA searches, e.g. #2160, Tabularia,: (BLAST: Tabularia sp. 741, 741, 100%, 0.0, 99.75% excellent alignment scores in top 4 and excellent distance tree results. 100% 0e, 99.75; Tabularia laevis 100% 0e, 99.75; Tabularia tabulata 100%, 0e, 99.75; T. fasiculata 100%, e0, 98.77%)	4	x	x	x SEM LM (MOL -confid.)	x	x	SEM, LM, Live, (MOL -confid.)
62.	THAL	<i>Thalassiosira</i> Cleve, 1873 SILVA, #3724, BLAST: Thalassiosira sp. 527, 527, 100%, 5e-145, 99.65%-good alignment on distance tree); Thalassiosira (MOL 252-V293) Thalassiosira_antarctica SILVA #3739- BLAST: Thalassiosira angulata Max Score 483, Total Score 483, 100%, 1e-131, 96.88%; Thalassiosira antarctica Max Score 483, Total Score 483, 100%, 1e-131, 96.88%-slightly far away in the distance tree); SILVA # 3730- Thalassiosira_minima (BLAST: doubtful it is T. minima, closer to T. antartica, or better Thalassiosira) Thalassiosira_nordenskioeldii (MOL-19-V23); SILVA #3741, BLAST: T. nordenskioeldii, 527, 527, 100%, 5e-145, 99.65%-excellent distance tree alignment); Thalassiosira_angulata (MOL-41-V49); Thalassiosira_eccentrica (MOL-115-V135); SILVA #2602, Thalassiosira (BLAST: T. eccentrica, 749, 749, 100%, 0.0, 100.00%	2	x	x	x (MOL -confid.)			SEM, LM, (MOL -confid.)

No.	Counting Codes	General	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		& <i>T. minima</i> 749, 749, 100%, 0.0, 100.00%—same alignment stats and same distance tree results-excellent); <i>Thalassiosira_pacifica</i> (MOL-191-V224); <i>Thalassiosira_oceanica</i> (MOL-381-V438); <i>Thalassiosira_minima</i> (MOL-429-V491); <i>Thalassiosira_spinulifera</i> (MOL-430-V492); (BLAST: <i>Thalassiosira hendeyi</i> , 527, 527, 100%, 5e-145, 99.65%-good alignment on distance tree, but sits very close to <i>Thalassiosira anguste-lineata</i> ; BLAST: <i>T. tenera</i> , 505, 505, 100%, 2e-138, 98.26%—good alignment in distance tree); (BLAST: <i>Thalassiosira</i> sp. 494, 494, 100%, 5e-135, 97.57%; <i>Thalassiosira decipiens</i> (Grunow) E.G.Jrgensen 1905, 488, 488, 100%, 2e-133, 97.22%; SILVA #3735. <i>Thalassiosira antarctica</i> Comber 1896: 491, pl. 11, 488, 488, 98%, 2e-133, 97.54%—closest alignment in the distance tree); SILVA # 3728, <i>Thalassiosira_angulata</i> (BLAST: <i>T. baltica</i> . 521, 521, 98%, 2e-143, 99.65%, <i>T. angulata</i> 499, 499, 100%, 1e-136, 97.92%).							
63.	TRAC	<i>Trachyneis</i> Cleve, P.T. (1894): SILVA 2786, BLAST: <i>Trachyneis</i> sp., 472, 472, 100%, 2e-128, 96.18%, good alignment stats, but <i>Navicula</i> and <i>Gyrosigma</i> have better results on the distance tree; SILV 2832, BLAST: <i>Navicular</i> sp, best matches; <i>Trachyneis</i> sp., 445, 445, 100%, 2e-123, 95.14%, good alignment stats, but <i>Navicula</i> and <i>Gyrosigma</i> have better results on the distance tree results. MOL-76-V91 (possible): <i>Naviculales</i> (<i>Haslea nipkowii</i> , max score, 100% & 95.15; <i>Trachyneis</i> sp., 100%, 94.79%, but high e value of 7e-122 BLAST; SILVA 373, BLAST: <i>Trachyneis</i> sp., 422, 100, 2.24e-113, 93.056, KX981824; Also found in SILVA 702, 2706, 2750, 2751, 2772, 2777, 2778, 2786, 2788, 2790, 2791, 2797, 2801, 2803, 2810, 2833 (good one); SILVA 2731, BLAST: <i>Trachyneis</i> sp 449, 100, 1.04e-121, 94.792, KX981824, close match to sequences, however distance tree is unreliable for <i>Trachyneis</i> . Distinctive morphology. Not yet conclusively found in the molecular data,	2	X	X	X-(MOL-prob.)			LM Live, (MOL--prob.)

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		however, note there are only 2 entries for Trachyneis in BLAST.							
64.	TRIG	Trigonium Cleve, P.T. (1867) Not found in a search of all the SILVA molecular sequences.	2			X			Live
65.	TRYB	Tryblionella W. Smith, 1853 ²³ SILVA #299, BLAST: Tryblionella sp. and T. apiculata, 499, 499, 98%, 1e-136, 98.25%, 1st, 10x alignments are Tryblionella species all excellent results, excellent distance tree results; Tryblionella_gaoana (MOL-81-V96; BLAST: 494, 494, 98%, 5e-135, 97.89% or 399, 399, 98%, 1e-106, 91.93%-- Stats and distance tree indicates it could be <i>Nitzschia ligowskii</i> . ; SILVA #251, Feb 10-2023 (BLAST: Tryblionella gaoana, 494, 494, 98%, 5e-135, 97.89% (<i>Tryblionella gaoana</i> Witkowski & Chunlian Li 2016: 191, figs 3 i, 15) or equally BLAST = <i>Nitzschia ligowskii</i> Witkowski, Lange-Bertalot, Kociolek & Brzezinska 2004: 584-586, figs 26-81—both are equal for excellent quality on the distance tree results; Tryblionella_apiculata (MOL-443-V506, 309-V360), #299, SILVA, 98%, 1e); Tryblionella queries match many sequences in BLAST, however the best one is #251.	2			X (MOL -confid.)	X	SEM LM, (MOL -confid.)	
66.	UNDA	Undatella Paddock & P.A.Sims, 1980 (occasionally observed, however it's taxonomic placement is uncertain): Only one hit for the entire SILVA, #239, BLAST: 355, 100, 2.30e-93, 88.966, KX981831, Undatella quadrata isolate, --poor on distance tree and poor stats for alignment.	2	X-UC	X-UC		X-UC		SEM (uc) LM (uc)
		Recently Identified Diatoms from Known Genera							
	NAVI	<i>Navicula cf. cancellata</i> Donkin 1872 (girdle view) (Unknown diatom on Zm stub	1			X			SEM

²³ A guide to distinguishing these two genera is forthcoming.

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		20_Ma En March 7-2021 (MW)_003(x1.5)_3.tif). Fig. 13.							
	NAVI	<i>Navicula directa</i> (Unknown diatom-on ZM MHMPP-T56 9M_B H2O2-TM4000-(MQ-July 29-22 (x1.2k)_2.tif) Fig. 14.	1			x			SEM
	NAVI	<i>Navicula cf. perminuta</i> : Unknown diatom Zm-MHMPP Mar 7-2021 stb 9M RT-H2O2-TM4000_MW Sept 6-2021_011_3.tif Appears to be common on March 7, 2021 leaf sections Fig. 15.	1-2			x			SEM
		<i>Unidentified Genera</i>							
	UNKN	Unknown raphid diatom: a) unknown raphid on <i>Z marina</i> -Mar 7-2020-T45 leaf#5 Da+c slide 3 SA1b-E800-MU2003-Mar 29-2021-0052_3.tif & unknown raphid on <i>Z marina</i> -Mar 7-2020-T45 leaf#5 Da+c slide 3 SA1b-E800-MU2003-Mar 29-2021-0001(1)_3.tif Figs. 11-12.				x			LM
		<i>Genera identified from Molecular Data (ITS and RCLB, Jan. 2022 from</i>							

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		Parfrey Lab) not yet reported by microscopy. Must have good BLAST e values, alignment stats and strong distance tree results (confident, probable and and weak)							
1	ACHNA	Achnanthidium: has 8 hits in the molecular data: SILVA #35, (BLAST: Achnanthidium sp. 98% 2e-118 (possible); Achnanthidium saprophilum 98%, 2e, <i>Achnanthidium minutissimum</i> Kuetzing) Czarnecki—good alignment stats and best on distance tree results). (MOL-35-V42, (BLAST_2e-113); SILVA 490 (BLAST: <i>Achnanthidium spp.</i> -A. cavitatum 689, 689, 100%, 0.0, 98.22%—good on distance tree result, but <i>Cocconeis stauroneiformis</i> is #1 on alignment stats, followed by 6 <i>Achnanthidium</i> x 4 different species); SILVA #564 BLAST: <i>Achnanthidium ovatum</i> Watanabe & Tuji, 689, 689, 100%, 0.0, 98.22%—distance tree results oK); SILVA # 584, BLAST: <i>Achnanthidium daonense</i> (Lange-Bertalot) Lange-Bertalot, O.Monnier & L.Ector in Monnier et al. 2007, 684, 684, 100%, 0.0, 97.97%-poor distance tree result, though on the tree; SILVA # 335, BLAST: <i>Planothidium lanceolatum</i> (Brebisson ex Kuetzing) Lange-Bertalot 1999, homotypic synonym: <i>Achnanthidium lanceolatum</i> , 434, 434, 100%, 6e-117, 93.40%—perfect alignment score, excellent alignment result and distance tree results have excellent horizontal results for many <i>Achnanthidium</i> sp—can no longer find these strong scores and excellent distance tree results-possible); SILVA 490, BLAST: <i>Achnanthidium spp.</i> -A. cavitatum 689, 689, 100%, 0.0, 98.22%—good on distance tree result, but <i>Cocconeis stauroneiformis</i> is #1 on alignment stats, followed by 6 <i>Achnanthidium</i> x 4 different species) — very possible, but not certain in distance tree results; Bacillariophyceae (BLAST: <i>Achnanthidium daonense</i> (Lange-Bertalot) Lange-Bertalot, O.Monnier & L.Ector in Monnier et al. 2007, 678, 678, 100%, 0.0, 97.72% (1st hit. <i>Achanthes</i> is 2nd hit, the many <i>Achnanthidium</i> sp. - distance tree is good for <i>Achnanthidium</i> sp.)			MOL-confid.			(MOL confid.	

No.	Counting Codes	General	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
2.	ARCO	<p><i>Arcocellulus</i>: SILVA 2856; (BLAST: 749, 749, 100%, 0.0, 100.00%); MOL-1158-V127 (BLAST e value 0.0, but Minutocellus has an equal e-value and score);</p> <p><i>Arcocellulus_mammifer</i> (Mol-440-V503_BLAST 3e-135, but best hit is an unidentified uncultured microorganism with an e value of 2e-138); SILVA #1521-BLAST: Arcocellulus sp., 494, 494, 100%, 5e-135, 97.57%-not in distance tree' SILVA #2586, (BLAST: Best distance tree result is <i>Minutocellus</i> sp. 749, 749, 100%, 0.0, 100.00%, alignment is <i>Minutocellus cf. polymorphus</i> (Hargraves & Guillard) Hasle, Stosch, & Syvertsen; less possible is <i>Arcocellulus cornucervis</i> G.R.Hasle, H.A.von Stosch & E.E.Syvertsen 1983, 749, 749, 100%, 0.0, 100.00%-good distance tree results and so is <i>Minutocellus polymorphus</i> (Hargraves & Guillard) Hasle, Stosch, & Syvertsen with the same alignment stats); SILVA 2586, BLAST: <i>Arcocellulus cornucervis</i> G.R.Hasle, H.A.von Stosch & E.E.Syvertsen 1983, 749, 749, 100%, 0.0, 100.00%-excellent alignment stats and very excellent distance tree results)</p>				MOL-very strong-probable			MOL- - very strong-probable
3.		<p><i>Asterionella</i>:</p> <p>has decent % identity scores with sequences 1290 (weak), 1292 (weak), 1317 (weak), 1328 (weak) although it appears lower down the list for all. Weak match with sequences 2164, 2166, 2167, 2495, 2575.</p>				MOL-weak			MOL- uc
4.		<p><i>Bacterosira</i> Gran 1900 (Basonym; <i>Thalassiosira constricta</i>: SILVA #3745</p> <p>BLAST: <i>Bacterosira constricta</i> (Gaarder) J.S.Park & J.H.Lee 2016: 11, 521, 521, 98%, 2e-143, 99.65%-good alignment in distanced tree)</p>				MOL-possible			MOL-possible
5.	done	<p><i>Berkeleya</i>: Berkeleya sp. MOL 109-V126-100%, 100% BLAST_3e-120 to 7e-147) found in Bacillariophyceae -uncertain: Achnanthales (Planothidium 98%, 93.33%or Berkeleya sp. 100%, 93.06% BLAST). #29-Berkelya sp. 98% e2 (SILVA & BLAST-Feb 10-2023-close in distance tree, but not certain); #31 SILVA (BLAST-Berkeleya sp. 100% 1e-116);</p>				MOL-strong			MOL-strong

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		(BLAST: Berkeleya fennica, 516, 516, 98%, 1e-141, 99.30%)							
6.		Biddulphiopsis: Stosch, H.A. von & Simonsen, R. (1984) Biddulphia alternans, 217-V251, 2e-116 Very uncertain, though occasionally found in Trincomali Channel marine plankton samples. Family: Biddulphiaceae. Round et al. 1990: 248-249.				MOL			MOL uc
7.		Cymbellales (Order): is showing up in the molecular data (Mol 224-V260) However, Staurophora sp. Max score, 98%, 2e-133, 97.54% per. Ident.- marine and brackish waters. 222-V258 is the closest fit under Cymbellales. Bacillariophyceae (<i>Cymbella lanceolata</i> , <i>Cymbella cistula</i> 98%, 95.45% 2e-123; <i>Gomphonella olivacea</i> 98%, 95.09% high e value of 7e-122 BLAST).				MOL			MOL uc
8.		Cymbellonitzschia: SILVA 284, BLAST: Cymbellonitzschia banzuensis isolate, 433, 99 1.03e-116, 94.035—good distance tree result; 98%, 94%, low e value of 7e-117 BLAST, Tokyo Bay, Japan)-unlikely				MOL-uc			MOL-uc
9.		Cymboleura sp.: (Freshwater only?) max score 98%, 94.39%, 2e-118 found under Bacillariophyceae – BLAST. In this search, both Planothidium and Achnanthidium have higher e-values at 1e-116.				MOL-uc			MOL-uc
10		Eunotia sp.: (BLAST: likely Eunotia sp. 488, 488, 100%, 4e-133, 87.13%-lots of sequences matches for Eunotia. However far away on the distance tree results, but good indication.)				Mol -uc			MOL -uc
11	EXTU	Extubocellulus: Extubocellulus_spinifer (MOL-77-V92); Extubocellulus_cribriger (MOL-251-V292); #1519, SILVA, Extubocellulus_spinifer, (BLAST: Extubocellus sp. 505, 505, 100%, 2e-138, 98.26%-very good distance tree results);; #1520, SILVA Extubocellulus_cribriger - BLAST: E. cribriger, 527, 527, 100%, 5e-145, 99.65-very good distance tree results); 251 V292 (BLAST: E._cribriger 548,				MOL -confid.			MOL-confid. - - certa in

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		548, 100%, 2e-151, 99.65%—very good distance tree result)							
12	FRAG	<p>Fragilaria: there are 6 entries in the molecular data for this genus:</p> <p>SILVA #2124, Fragilaria (BLAST: Fragilaria striatula Lyngbye 1819 (<i>Grammonema striatula</i>-not accepted by AlgaeBase-https://www.diatombase.org/aphia.php?p=axdetails&id=968692), 745, 745, 100%, 0.0, 100.00%—excellent distance tree results;</p> <p>SILVA #2125 Fragilaria (BLAST: <i>Fragilaria striatula</i> entered as <i>Grammonema striatula</i>, 739, 739, 100%, 0.0, 99.75%—excellent distance tree results); SILVA #2133, BLAST: Fragilaria sp. 510, 510, 100%, 5e-140, 98.61%); SILVA #2151 BLAST: Fragilaria striatula entered as <i>Grammonema striatula</i>, 739, 739, 100%, 0.0, 99.75%—good distance tree results); (i.e. MOL-748-V831 Fragilaria (BLAST: Tabularia sp. 100% 0e, 99.75; Tabularia laevis 100% 0e, 99.75; Tabularia tabulata 100%, 0e, 99.75; T. fasiculata 100%, e0, 98.77%; MOL- 1087-V119, MOL 287-V-334: 748, 1087, 1317, 1741, 1943 and 3214) (are these the <i>Tabularia</i> that have clear central areas? The raw molecular data showed <i>Tabularia</i> spp. As well as <i>Tabularia tabulata</i>)</p>			MOL-confid.			MOL-confid.	
13	GEDA	<p>Gedaniella Chunlian Li, A. Witkowski & M.P. Ashworth, 2018:</p> <p>Gedaniella_panicellus, BLAST: Gedaniella panicellus SZCZCH1350 Chunlian Li, Yu & Witkowski 2018, 532, 532, 100%, 1e-146, 100.00%, excellent distance tree result for genus and species; Gedaniella SILVA #2131; BLAST: Gedaniella paucistriata 518, 518, 100%, 3e-142, 98.96%; Gedaniella_mutabilis 516, 516, 100%, 1e-141, 98.96%); Gedaniella (MOL-102-V119); Gedaniella_mutabilis (MOL-117-V137); Gedaniella_panicellus (MOL-250-V291); #2137, SILVA (BLAST: Gedaniella 100% 1e, 93.77, Nanofrustulum 100%, 2e, 94.10%; SILVA # 2138, BLAST: G. panicellus, 516, 516, 100%, 1e-141, 98.96%—good distance tree results); SILVA #2131BLAST: Gedaniella sp. 444, 444, 100%, 5e-120, 94.44%-good distance tree</p>			MOL-confid.			MOL-confid.	

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		results); 28-V35, possibly Gedaniella boltinii SZCZP, 416, 416, 100%, 1e-111, 92.71%, though 12 down the list of alignments, but ok for distance tree results)							
14		Gomphonella: Gomphonella olivacea:, 98%, 5e and 2e = Gomphonema olivaceum (Hornem.) Bréb. 1838 (Hofmann et al. 2013, Levkov et al. 2016); #438, SILVA, Gomphonella olivacea, 98%, 2e, 1e)				MOL-uc			MOL-uc
15		Grammonema (needs investigation of number of hits and what it looks like-see Fragilaria striatula) (Agardh, C.A. (1832). Conspectus Criticus Diatomacearum. Part 4. Lundae. Litteris Berlingianis. pp. 48-66.) Grammonema striatula SILVA #2124, Fragilaria (BLAST: Grammonema striatula (old name = Fragilaria striatula Lyngbye 1819- https://www.diatombase.org/aphia.php?p=taxdetails&id=968692), 745, 745, 100%, 0.0, 100.00%—good distance tree results;				MOL-uc			MOL-uc
16	ISTH	Isthmia: Isthmia enervis Ehrenberg 1838, (BLAST: Isthmia enervis Ehrenberg, 632, 632, 100%, 1e-176, 94.81%) - good distance tree data. Found in abundance on Elaine Humphrey's eelgrass sample from Sidney Island, Gulf Island, B.C.				MOL-confid.			MOL-confid.
17	Look for more hits	Lemnicola Round & Basson, 1997: Bacillariophyceae (BLAST: Lemnicola sp. 499, 499, 98%, 1e-136, 98.25%—good distance tree alignment). NOTE: Regarded as non-marine genera; SILVA # 584, BLAST: Lemnicola sp., 712, 712, 100%, 0.0, 99.24% and good distance tree result.				MOL-possible			MOL-possible
18	Look for more hits	Leyanella Hasle, Stosch von & Syvertsen 1983 :SILVA #2578, Leyanella probus strain, 721, 721, 100%, 0.0, 98.77%) Hasle, G.R., Stosch, H.A. von & Syvertsen, E.E. (1983). Cymatosiraceae, a new diatom family. Bacillaria 6: 9-156. Distance tree data looks good but not a firm phylogeny match; SILVA #2577, Leyanella probus strain, 726, 726, 100%, 0.0, 99.01%); SILVA 2577, (BLAST: Leyanella probus strain, 726, 726, 100%, 0.0, 99.01%) Hasle, G.R., Stosch, H.A. von & Syvertsen, E.E. (1983);				MOL-strong			MOL - strong

No.	Counting Codes	General	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		best for distance tree result is Pseudoleyanella lunata isolate NG0002 18S ribosomal RNA gene, partial sequence, then Leyanella sp.; Cymatosiraceae, a new diatom family. <i>Bacillaria</i> 6: 9-156.							
19		<i>Medlinella amphoroidea:</i> (MOL #263, SILVA Feb 10-2023, 98%, 2e, 97.54%)				MOL-uc			MOL-uc
20	MINU	<i>Minutocellus:</i> <i>Minutocellus</i> has an equal e-value and score as <i>Arcocellulus</i> MOL-1158-V127 (BLAST e value 0.0); SILVA #1521, <i>Minutocellus polymorphus</i> , 483, 483, 100%, 1e-131, 96.88%—good distance tree results). (Hargraves & Guillard) Hasle, Stosch, & Syvertsen 1983; SILVA # 2586, BLAST: and so is <i>Minutocellus polymorphus</i> (Hargraves & Guillard) Hasle, Stosch, & Syvertsen 749, 749, 100%, 0.0, 100.00% (shows a perfect match)-good to excellent distance tree results with the same alignment stats)				MOL-confid.			MOL-confid.
21		<i>Nanofrustulum wachnickianum</i> Chunlian Li, Witkowski & Ashworth 2018: MOL-248-V288 under Fragilariales (<i>Nanofrustulum wachnickianum</i> SZCZCH194, max score 100%, 94.10% 2e-118- BLAST)				MOL			MOL
22		<i>Pauliella</i> Round, F.E & Basson, P.W. (1997): <i>Pauliella_taeniata</i> (MOL-187-V219, #32-2e-118 & SILVA_DiatBarcde_PR2_summarized-Feb 10-2023; SILVA #320, 327, <i>Pauliella toeniata</i> shows up as a distant node and in the right horizontal distance to the query sequence, but is 17 sequences down from the top score, but is still a good score at 680, 680, 100%, 0.0, 98.22%); SILVA 29, BLAST <i>Pauliella taeniata</i> <i>Pauliella taeniata</i> (381, 381, 96%, 4e-101, 91.23%) is very good in the distance tree results, alignment				MOL-prob.		MOL-prob.	
23	PHEO	<i>Phaeodactylum</i> Bohlin, K. (1898): SILVA #521: (BLAST: <i>Phaeodactylum tricornutum</i> 654, 654, 100%, 0.0, 96.03%) good distance tree result MOL-513-V58; (BLAST: <i>Phaeodactylum tricornutum</i> , 302, 450, 100%, 4e-77, 100.00%, can't find <i>Phaeodactylum tricornutum</i> in the distance				MOL-Strong			MOL-Strong -

No.	Counting Codes	General	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		tree; (BLAST: Phaeodactylum tricornutum, 656, 656, 100%, 0.0, 96.03%, good distance tree results, excellent alignment and lots of tops hits. However, Gomphonemopsis cf. pseudoexigua is a little better for distance tree result, but only one entry, where as Phaeodactylum is close but many more entries.); (BLAST: Phaeodactylum tricornutum, 656, 656, 100%, 0.0, 96.03%, good on distance tree result but entry 2, Gomphonemopsis is a better tree result, however there are many top results for Phaeodactylum tricornutum; (have checked all sequences in SILVA-only strong, not certain)							
24	Look for more hits	Pinnularia: (MOL-976-V107, e value = 0) Or Caloneis amphisbaena:SILVA 2815; (BLAST: Caloneis amphisbaena 449, 449, 100%, 1e-121, 94.79%; Pinnularia subanglica K.Krammer 449, 449, 100%, 1e-121, 94.79%); SILVA 320, BLAST: Pinnularia sp. 725, 725, 100%, 0.0, 100.00% has same excellent alignment stats and good distance tree result.; SILVA 2185, Pinnularia (BLAST: Caloneis amphisbaena 449, 449, 100%, 1e-121, 94.79%; Pinnularia subanglica K.Krammer 449, 449, 100%, 1e-121, 94.79%)-good distance results for Pinnularia; SILVA 2826, BLAST: Fallacia monoculata (Hustedt) D.G.Mann & 427, 427, 100%, 5e-115, 93.40%, Pinnularia sp. 427, 427, 100%, 5e-115, 93.40% — Ok distance tree results (hundreds of hits with the sequences, but distance tree results moderate-prob. to uncertain)				MOL-UC		MOL-uc-	
25	PROS	Proschkinia (highly likely to be found in the microscopic images): SILVA 334, BLAST: Proschkinia cf. complanatula, 732, 732, 100%, 0.0, 99.75%; distance tree results are very strong for Proschkinia and good for Proschkinia cf. complanatula); Proschkinia_complanatula # 2761, SILVA (BLAST: Proschkinia 98%, 1e, 99.30); Proschkinia cf. complanatula #450, SILVA (100%, 0e) or Proschkinia staurospectiosa (100%, 0e); Proschkinia cf. complanatula				MOL-confid.			MOL-confid.

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		(BLAST: 510, 510, 98%, 5e-140, 98.95%, with distance tree, for sure Proschkinia sp. but slight uncertainty regarding the species P. complanatula; SILVA 369, Bacillariophyceae (BLAST: Proschkinia sp. 1 RoG-2019444, 444, 98%, 5e-120, 94.74%, excellent distanced tree results)							
26	Look for more hits	<p><i>Ralfsiella smithii</i> (=Cerataulus smithii Ralfs in Pritchard) marine species.</p> <p>V529, BLAST: <i>Ralfsiella smithii</i> (Ralfs) Sims, Williams & Ashworth, 2018, 743, 743, 100%, 0.0, 99.75%; SILVA 2590, BLAST: <i>Ralfsiella smithii</i> (Ralfs) Sims, Williams & Ashworth, 2018 synonym = Cerataulus smithii var. smithii Ralfs in Pritchard, 1861, 727, 727, 100%, 0.0, 99.75%—excellent match to query in distance tree results, as is Odontella sp. and O. aurita; O. longicruis is 11th on the list of alignments, 722, 722, 100%, 0.0, 99.51%—not in the distance tree results); <i>Ralfsiella smithii</i> (Ralfs) Sims, Williams & Ashworth, 2018, 743, 743, 100%, 0.0, 99.75%- excellent distance result to query (Odontella sp.);</p> <p>2x excellent query matches in the molecular data, found under Odontella which will show almost the exact same alignment stats and distance tree results. However, have never seen this taxon in any reports or LME or SEM on the West Coast. <i>Ralfsiella smithii</i>:</p> <p>https://www.marinespecies.org/aphia.php?p=taxdetails&id=974475 and AlgaeBase: https://www.algaebase.org/search/species/detail/?species_id=171157</p>				MOL?? ? 2 hits-pro b.		UC MOL--prob.	
27	SEMI	<p>Seminavis D.G. Mann, 1990 (marine): SILVA # (BLAST: Seminavis robusta, 510, 510, 98%, 5e-140, 98.95%- distance tree alignment excellent); (MOL-65-V77) Seminavis_robusta (MOL-131-V156, 299-V349 e values ranging from 7e-137 to 2e-113); SILVA 2798, BLAST: Seminavis</p>					MOL-confid.		MOL—c onfid.

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		robusta, 499, 499, 98%, 1e-136, 98.25%-distance tree alignment excellent); (BLAST: Seminavis sp., 499, 499, 98%, 1e-136, 98.25%—excellent distance tree alignment); 122-V142 BLAST: Seminavis robusta 427, 427, 98%, 5e-115, 93.68%-good distance tree results); Seminavis, 699, 699, 100%, 0.0, 98.01%-good distance tree; (BLAST: Seminavis sp. 4a LKM-2015 & Seminavis cf. robusta, 743, 743, 100%, 0.0, 100.00%, very excellent distance tree result)							
28	SERR	<p><i>Serratifera:</i> <i>Serratifera_andersonii</i> SZCZP695 (BLAST: 510, 510, 98%, 5e-140, 98.95%-high confidence-very excellent distance tree results); MOL-348-V404, 3e-140 to 2e-138-high confidence) <u>Considered a synonym of <i>Pseudostaurosira andersonii</i> (Chunlian Li, Dabek & Wachnicka) E.A.Morales, C.E.Wetzel & Ector²⁴ (Guiry and Guiry 2022)</u> <u>Note: some identified <i>Opephora</i> may be Serratifera. Images need re-examination.</u></p>				MOL-confid.			MOL—confid. LM?
		<p><i>Stauroforma:</i> <i>Stauroforma_rinceana</i> SZCZCH, max score, 100%, 92.01-BLAST in Bacillariophyta MOL 290-V337, 2e-108</p>				MOL-uc			MOL-uc
		Possible Genera with less likely hits from BLAST molecular data sequences (ITS and RCLB, Jan. 2022 from Parfrey Lab) not yet reported by microscopy.							

²⁴ According to Morales et al. (2019) *Serratifera* and *Gedaniella* were both erected by molecular data (reference) but may not be valid based on morphological evaluations.

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		High E values or iffy alignment percentage and poor or not distance tree results							
1.		<i>Astartiella</i> Witkowski, Lange-Bertalot & Metzeltin, 1998: SILVA #1524, BLAST 405, 405, 98%, 2e-108, 92.28%—better, but OK distance tree result)				X MOL			MOL?
2.	Look for more hits	<i>Ctenophora pulchella</i> (a brackish water to freshwater species), SILVA #2133, C. pulchella 394, 394, 98%, 5e-105, 91.61% is the closest match in the distance tree results but is far down the list of the best sequence alignment. <i>C. pulchella</i> is a brackish water diatom that was previously <i>Synedra pulchella</i> or <i>Fragilaria pulchella</i> . This species has these distinctive characteristics similar to a Tabularia sp. we see on occasion on the eelgrass: Valves are elongate. The central area has a thickened, dome-like silicic acid fascia and ghost striae may be present. Each pole has a rimoportula positioned near the axial area, positioned diagonally to the apical axis (https://diatoms.org/species/ctenophora_pulchella_a/guide)				X MOL?			MOL?
3.	Look for more hits	<i>Cymbellonitzschia banzuensis</i> SILVA 284, BLAST: <i>Cymbellonitzschia banzuensis</i> isolate, 433, 99.1.03e-116, 94.035—good distance tree result; 98%, 94%, low e value of 7e-117 BLAST, Tokyo Bay, Japan)-unlikley				X?			MOL?
4.	Look for more hits	<i>Gomphoneis</i> : found under <i>Cymbellales</i> , 100% match in GeneBank MOL224-V260, 3e-130 : <i>Gomphoneis okunoii</i> Ak513 chloroplast <i>rbcL</i> gene for ribulose-1,5-bisphosphate carboxylase/oxygenase large subunit, partial cds Sequence ID: LC530728.1 Length: 1485 Number of Matches: 1 Syn: <i>Gomphonella</i> and <i>Gomphonema</i> This genera has a higher e value than <i>Stauropora</i> sp. at 2e-133				X?			MOL?
5.		<i>Gomphonema parvulum</i> MOL 68-V80 <i>Gomphonema pseudoaugur</i> _3e-115: BLAST, MOL-241-V280, 98%, 92.28%				X?			MOL?
6.		<i>Scoliopleura peisonis</i> strain (freshwater species), max score. 100%, 93.75%, high e- value = 7e-117 BLAST				X MOL			MOL

No.	Counting Codes	Genera	Frequency	July-Aug. 2020	Nov. 15, 2020	Mar. 7, 2021	July 22, 2021	Nov. 7, 2021	SEM LM Live MOL
		then Halaphora and Panothidium lower percentages-BLAST found under Bacillariophyceae							
7.		Staurophora: 98%, 98.60; Gomphoneis olivacea 98%, 96.49%—Staurophora is a closer match in BLAST for MOL 260 V304; SILVA # 1523 (BLAST: Staurophora sp. 505, 505, 98%, 2e-138, 98.60%—reasonable distance tree results —found under Cymbelles				x-MOL?			MOL
8.		Sternimirus shandongensis Witkowski & Chunlian Li 2016 (MOL-352-V408, not found in BLAST, however the best alignment is with 2e-133 is Bacillariophyta sp. SZCZCH968 ribulose-1,5-bisphosphate carboxylase/oxygenase large subunit (rbcL) gene, partial cds; chloroplast)				x-MOL			MOL

Acknowledgements

- Molecular data (MOL) obtained by performing DNA barcoding of 16S, 18S, and RBCL target regions from extracted DNA of swabs. Sequence data processing, including taxonomy taxonomic assignment of the Amplicon Sequence Variants (ASVs), was performed in the DADA2 pipeline (Callahan et al. 2016). 16S and 18S taxonomy was assigned from the SILVA 138 database and RBCL sequences assigned with DiatBarcode V10 database. The taxonomy table from Siobhan Schenk (Parfrey lab, UBC) sent on February 25, 2022 is the output (3221 records) of the denoised output of DADA2 pipeline. A second data set was generated from the March 7, 2021 data with SILVA by Siobhan Schenk on Feb. 10, 2023 (#X, SILVA). Kirsty Smith, Manager of Molecular Algae Ecology at the Cawthonn Institute in Nelson, New Zealand for helpful discussions regarding examination and assessment of BLAST searches against sequence data.
- SEM imaging, support, research and assistance by Elaine Humphrey, Manager of the Advanced Microscope Facility at UVIC & IMERSS. Sample organization, collection, environmental stub preparation and editing assisted by Arjan van Asselt (IMERSS). SEM imaging contributions and taxonomic discussions from the projects' volunteers; Melanie Quenneville and Ron Read. Henry McGee developed an effective program for downloading all the BLAST searches on the eelgrass sequence records.
- Andrzej Witkowski (University of Szczecin, Institute of Marine and Environmental Sciences, Szczecin, Poland) generously made edits to the unidentified images, Feb. 10, 2022: 1) Unknown but suspected *Achnanthidium* images were identified as *Planothidium*, 2) a possible LM image of *Aneumastus* was identified as *Fogedia*, 3) an SEM image of a questionable *Planothidium* has been identified as *Fallacia*, 4) unknown, possibly *Petroneis*, identified as *Paribellus*, 5) Fig. 13, unknown was identified as *Navicula cf.*

cancellata, 6) Fig. 14 has been identified as *Navicula directa*, and 7) Fig. 15. Unknown, identified as *Navicula cf. perminuta*. Also, Catenula is unconfirmed. It may be *Catenula* or *Catenulopsis*, but Andrzej Witkowski is suggesting it may fit closer to a newly proposed (nov. nud.: no valid nomenclatural status) genera called *Pariraphis* Sh. Mayama (Mayama et al. 2018: p. 42, Plate 9, fig. 10, Kryk et. al. 2020) So for now it is listed as *Periraphis* like.

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Images of Previously Unidentified Diatoms



Fig. 1. Nine white arrows highlight a group of *Planothidium* frustules on a March 7, 2021 distal section of *Z. marina*.

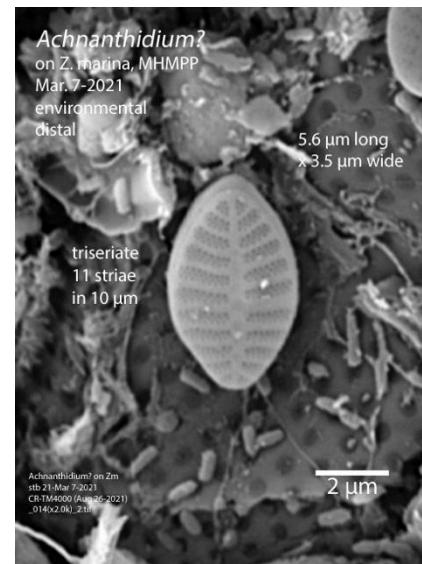


Fig. 2. A *Planothidium* frustule on a March 7, 2021 distal section of *Z. marina*. (same frustule in Fig. 1 with the image rotated 90 degrees)

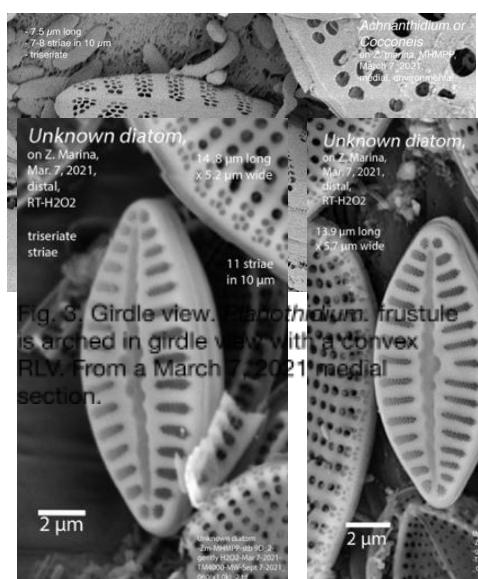
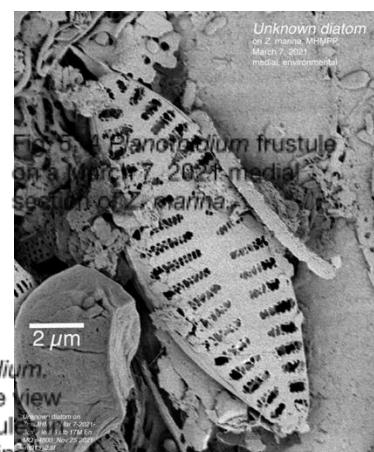


Fig. 3. Girdle view. *Leptothrixum*. fruit is arched in girdle view with a convex RLV. From a March 7, 2021 medial section.



Frustularis cretacea in guide with a convex RV or RLV of frustul Fig. 1-3.? Has a distinctive opening along the apical axis where a raphe would normally be located. From a March 7, 2021 distal section, RT-H2O2 cleaned.



Fogedia,
on *Z. marina*, MHMPP,
March 7, 2021, leaf 9 T57
cleaned, distal section

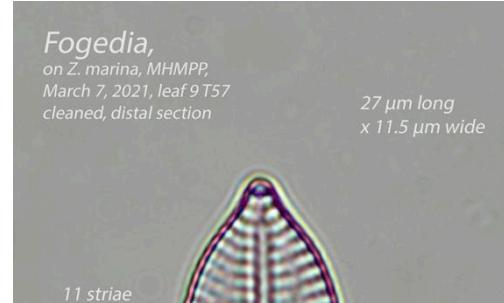


Fig. 7. *Fogedia*. Stack of seven LM images. (Plinski and Witkowski, 2020: p. 109-110, Fig. 536-537, pp. 320-321)

Fig. 8. *Fogedia*. Single LM image. (Plinski and Witkowski, 2020: p. 109-110, Fig. 536-537, pp. 320-321)

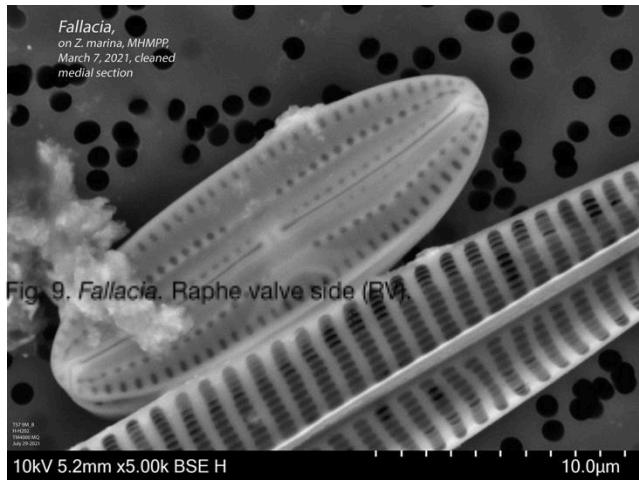
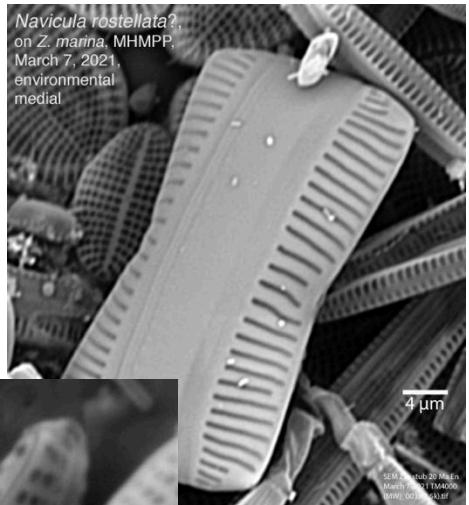
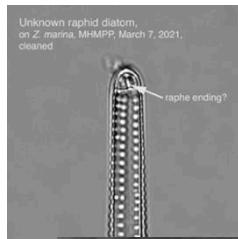
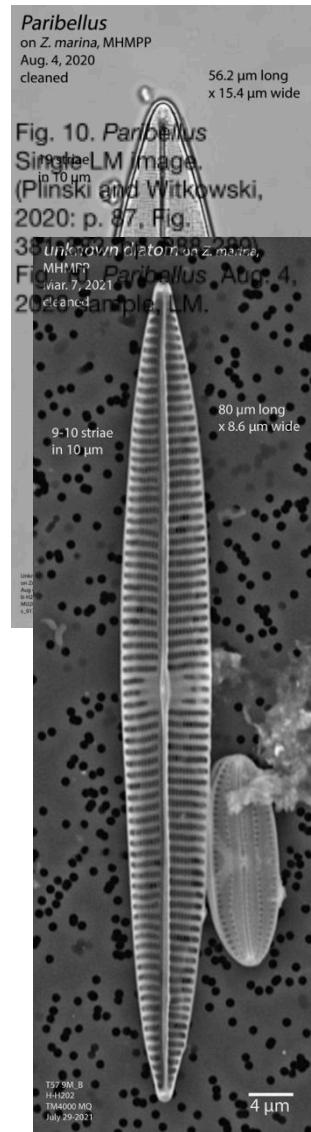


Fig. 9. *Fallacia*. Raphe valve side (RV).

Fig. 9. *Fallacia*. March 7, 2022
sample, with *Navicula directa*. SEM



Unknown diatom
on Zm, MHMPP
Mar. 7, 2021,
RT-H2O2
Achnanthes media; single
LM image.

wide striae
raphe ending
in center

16 μm long
x 4.3 μm wide

Slide 3_00033
MHMPP Mar 7-2021

Fig. 11. *RT-H2O2*
Achnanthes? single
LM image.

Fig. 12. Unknown,
Achnanthes? Single
LM image.

Fig. 13. *Navicula cf. cancellata* SEM
image. (Plinski and Witkowski, 2020: p.
104, Fig. 491, pp. 312-313)

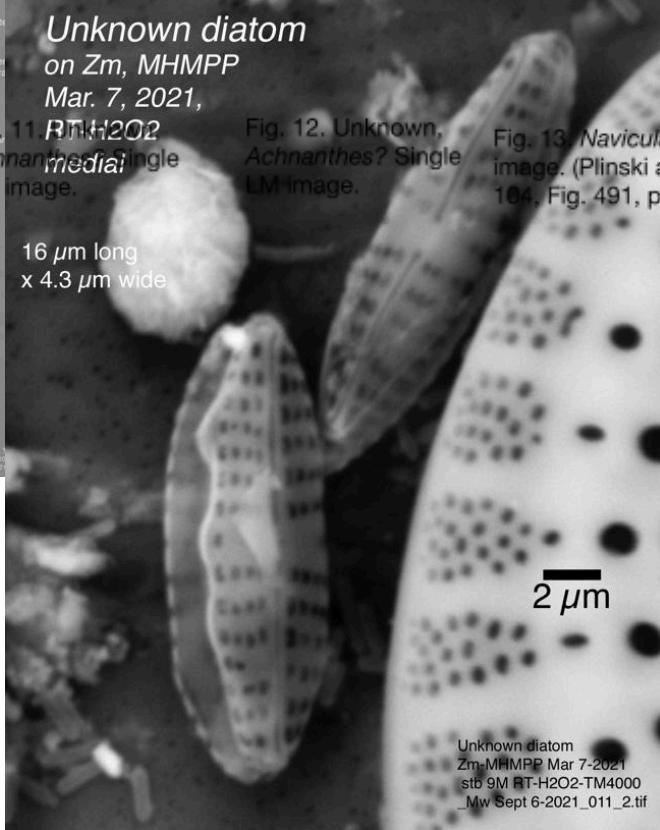


Fig. 14. *Navicula directa*.
Hendey 1964: 202. SEM
image.

Fig. 15. *Navicula cf. perminuta*. (Plinski and Witkowski, 2020: p. 106, Fig. 502, pp. 312-313)
Appears to be common on March 7, 2021 leaf sections. SEM image.

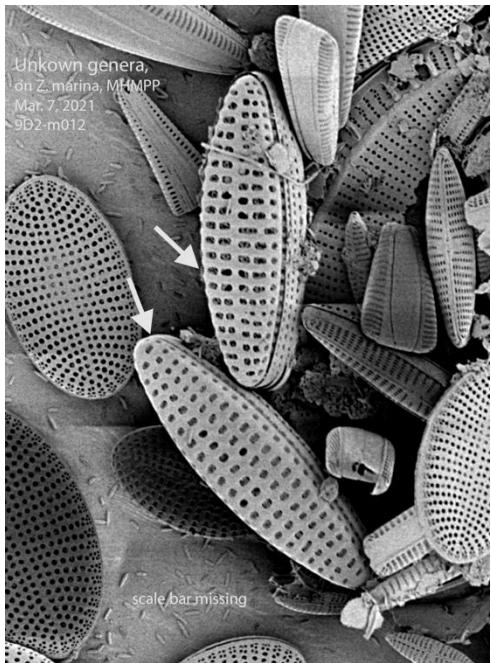
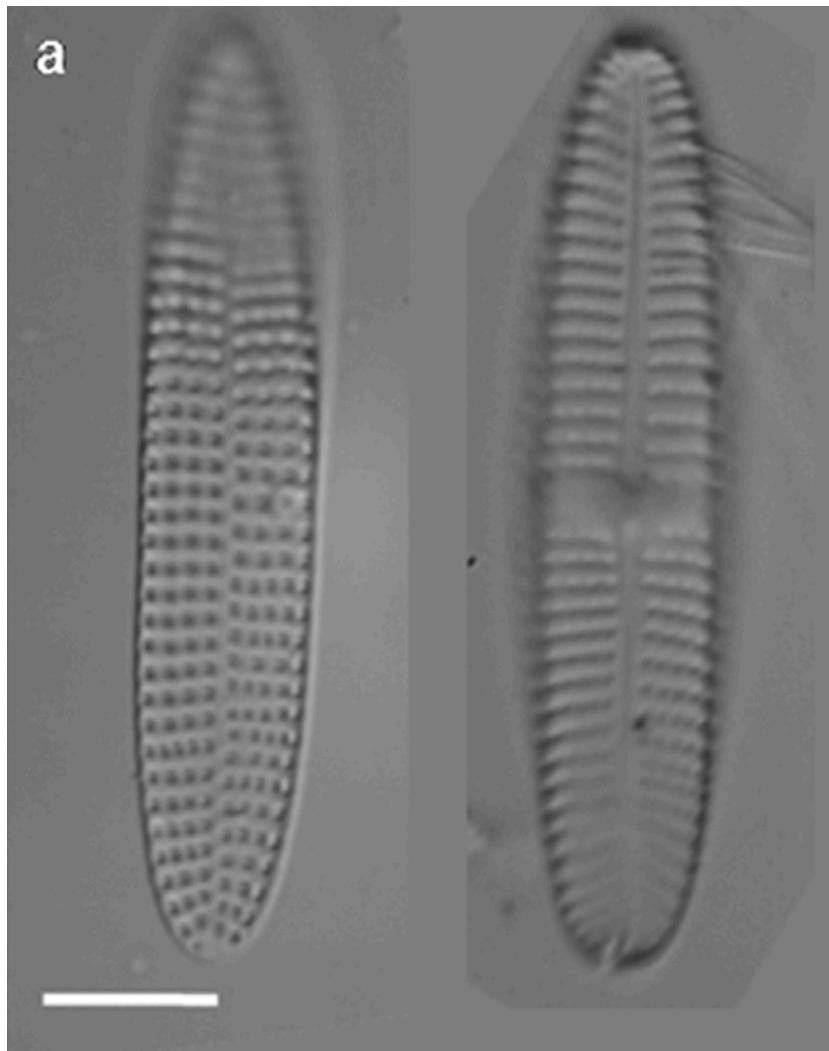
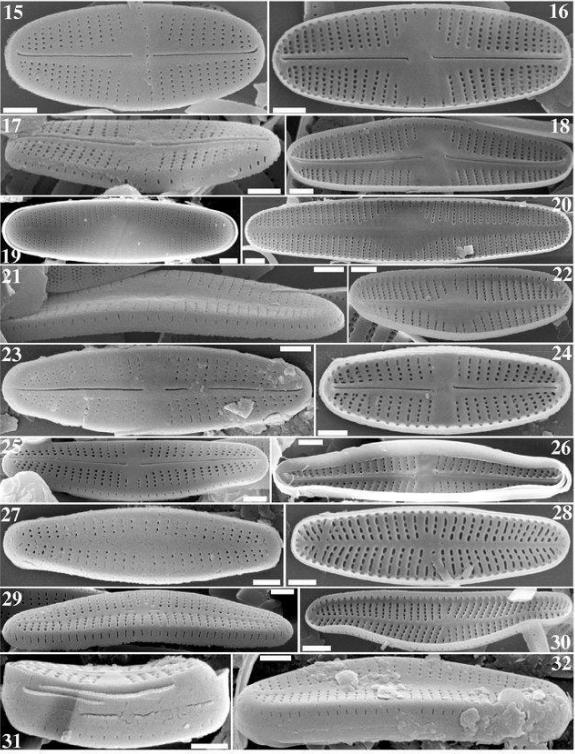
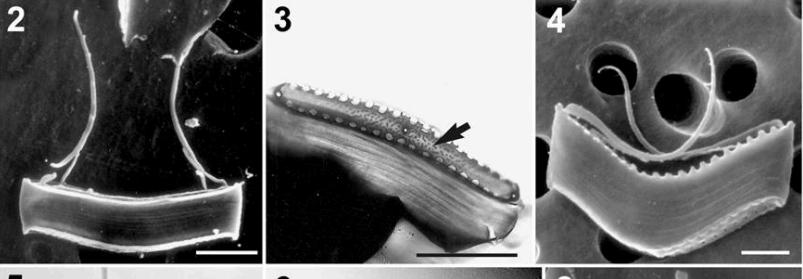
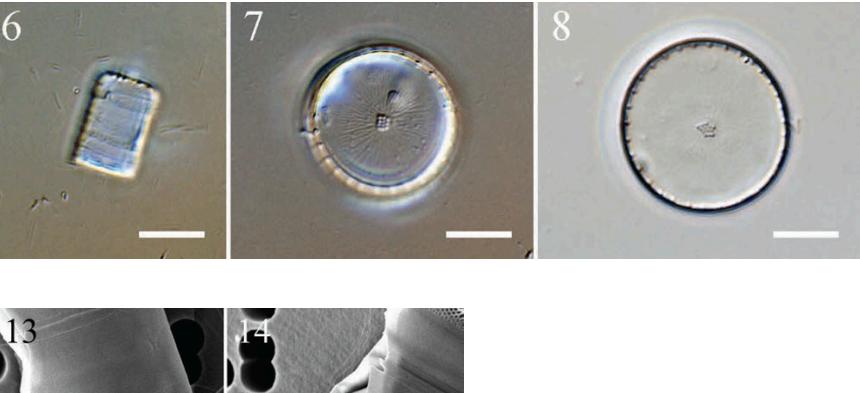


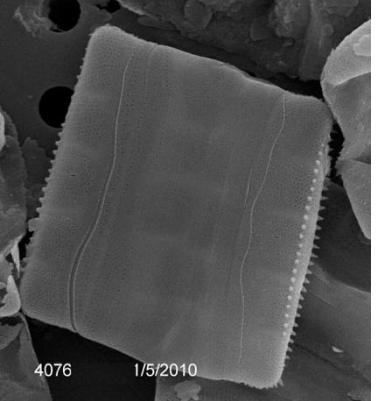
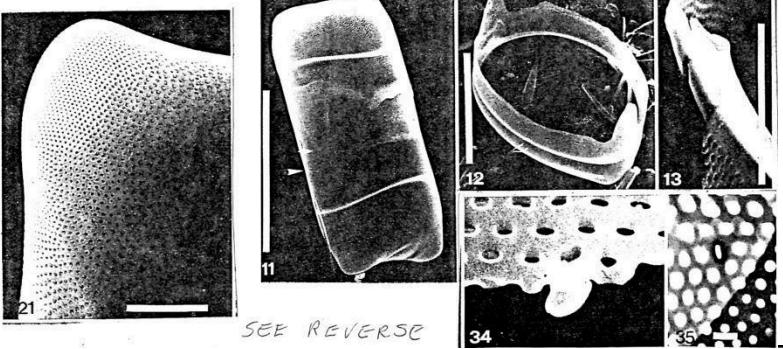
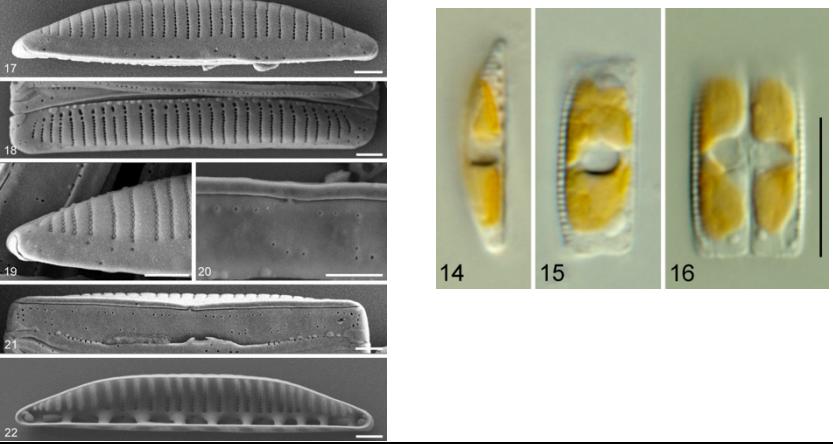
Fig. 16. Unknown genera. March 7, 2021.
SEMs-Zm-MHMPP-Mar
7-2021-9D2-9M2-9P2-MQ-Sept
18-2021-m012.tif



Achnanthes bongrainii is frequently found in the molecular data from March 7, 2021. This series of images is from: Daglio et al. (2018). Benthic Diatoms from Potter Cove Antarctica-(2018)- Fig. 3. Marine benthic diatoms from 25 de Mayo (Potter Cove) Antarctica on colonized ceramic tiles, viewed in LM.
a *Achnanthes bongrainii*, b *Entopyla ocellata*, c *Parlibellus delognei*, d *Licmophora antarctica*, e *Odontella litigiosa*, f *Amphora* sp. G *Actinocyclus actinochilus*, h *Cocconeis pinnata*, I *Cocconeis costata* var. *antarctica*, j *Cocconeis fasciolata*. Scale bar=10 μ m.

**Images (not from this project) of Genera found in the Molecular Data,
but not yet observed by our Microscopy**

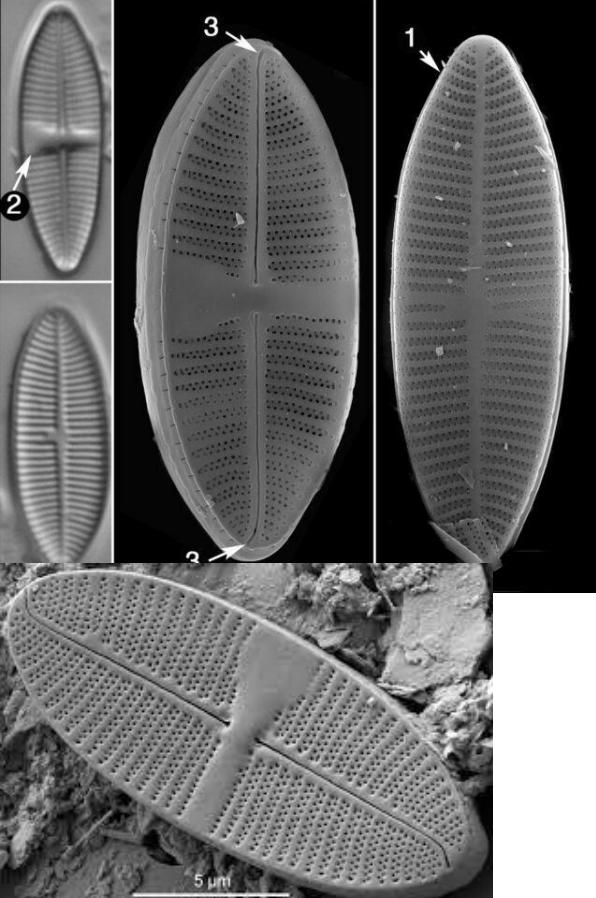
Genera identified by molecular sequences		Source of images
Achnanthidium		https://www.researchgate.net/publication/298988399_The_genera_Achnanthidium_Kutzing_and_Psammothidium_Bukhtiyarova_et_Round_in_the_family_Achnanthidiaceae_Bacillariophyceae_A_reappraisal_of_the_differential_criteria/figures?lo=1
Arcocellulus		Percopo et al. 2011 https://fce-lter.fi.u.edu/data/database/diatom/?species=3416
Bacterosira Gran 1900 (Basonym; <i>Thalassiosira constricta</i>		<i>A phylogenetic re-definition of the diatom genus Bacterosira (Thalassiosirales, Park et al. (2016). A phylogenetic re-definition of the diatom genus Bacterosira</i>

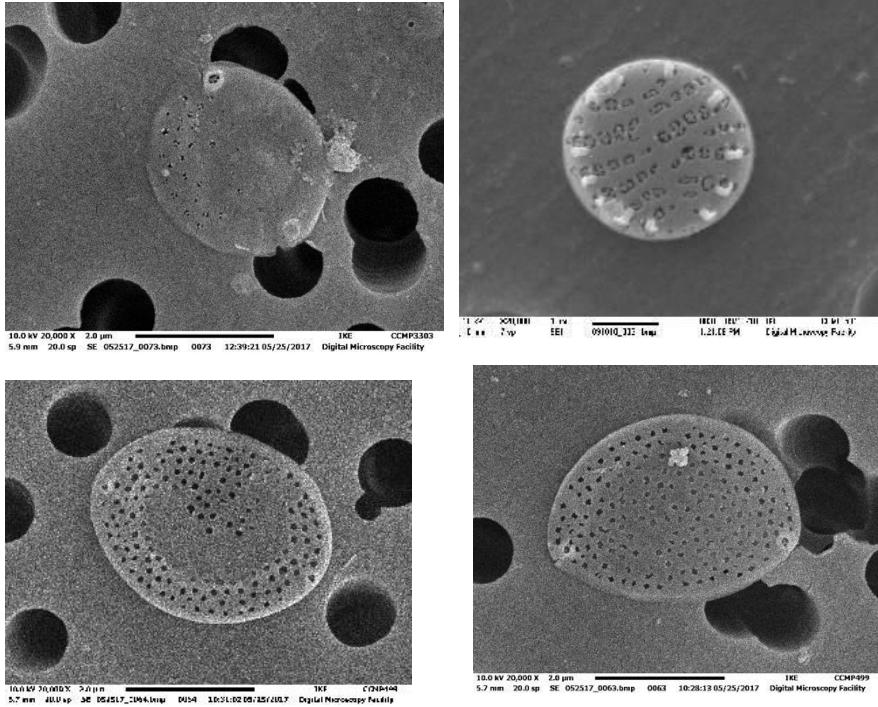
		(<i>Thalassiosirales, Bacillariophyta</i>), with the transfer of <i>Thalassiosira constricta</i> based on morphological and molecular characters. <i>Phytotaxa</i> 245 (1): 001–016
<i>Biddulphiopsis</i>	 <p>4076 1/5/2010</p> <p><i>Biddulphiopsis titiana</i> (Grunow) von Stosch & Simonsen n. comb. 1984 Basionym: <i>Cerataulus titianus</i> Grunow 1863, p. 159, pl. 4, figs. 25a,b Type locality: Adriatic Sea (Biddulphiopsis, a new genus of the Biddulphiaceae.) <i>Bacillaria</i>, 7: 12–15, figs. 1–35. CWR Copy International code: Valid</p>  <p>SEE REVERSE</p>	http://symbiont.anp.org/dntf/details.php?id=007846 http://symbiont.anp.org/dntf/details.php?id=007846
<i>Cymbellonitzchia banzuensis</i> 98%, 94% BLAST, Tokyo Bay, Japan)		Stepanek, Joshua G.; Hamsher, Sarah E.; Mayama, Shigeki; Jewson, David H.; Kociolek, J. Patrick (2016). Observations of two marine members of the genus <i>Cymbellonitzchia</i>

		(Bacillariophyta) from Tokyo Bay, Japan, with the description of the new species <i>Cymbellonitzchia banzuensis</i>. Phycological Research, 64(1), 26–34. doi:10.1111/pre.12110
<i>Extubocellulus</i>		http://v3.boldsystems.org/index.php/Taxbrowser_Taxonpage?taxid=187471
<i>Fragilaria</i>		FRAGILARIA STRIATULA LYNGBYE: A LITTLE KNOWN MARINE DIATOM FROM CHILE. Patricio Rivera1 & Fabiola Cruces (2002)
<i>Fragilaropsis</i>		https://en.wikipedia.org/wiki/Fragilaropsis_kerguelensis https://www.authorea.com/users/359729/articles/481556-ongoing-specialization-within-the-diatom-fragilaropsis-kerguelensis-in-the

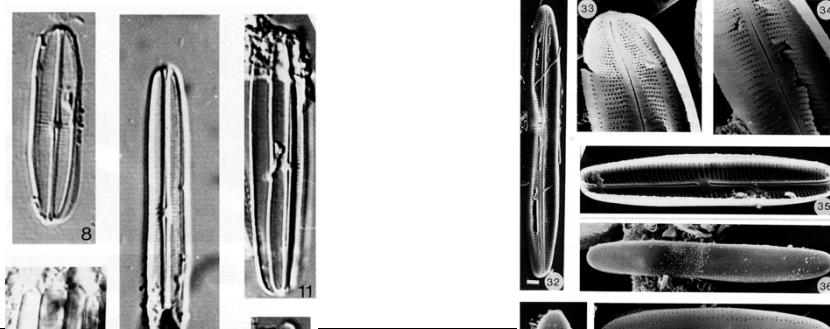
		-southern-ocean (Grattepanche 2020)
Gedaniella		https://twitter.com/krawczyk_diana/status/102464955390622515 2 Li et al. (2018)
Gomphoneis okunoi- MOL224-V260 (GeneBank-100 % match). See images from Tuji (2005: Plate 12-13.) considered a freshwater species.		Tuji (2005: Plate 12-13.) (left) https://diatoms.org/species/gomphonella_olivacea/guide

	sourounding brackish waters	
	<p>Haslea</p> <p>http://nordicmicroalgae.org/taxon/Haslea%20ostraria</p>	<p>Haslea</p>

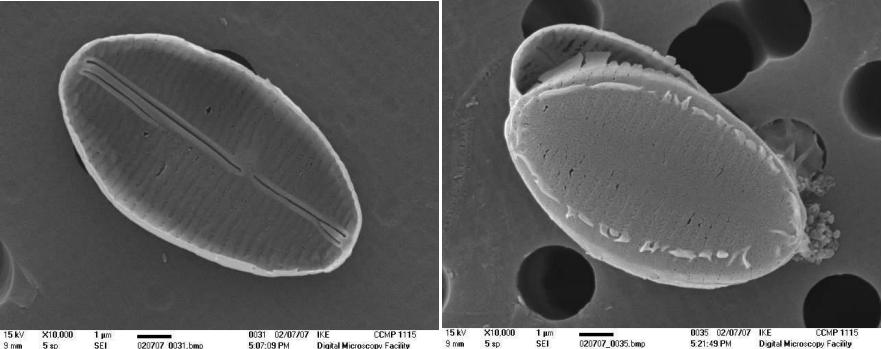
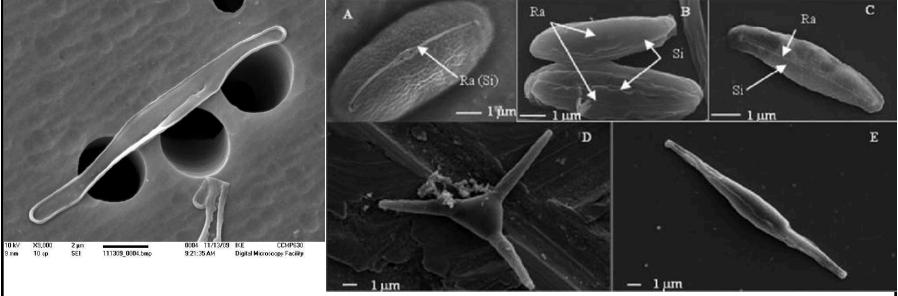
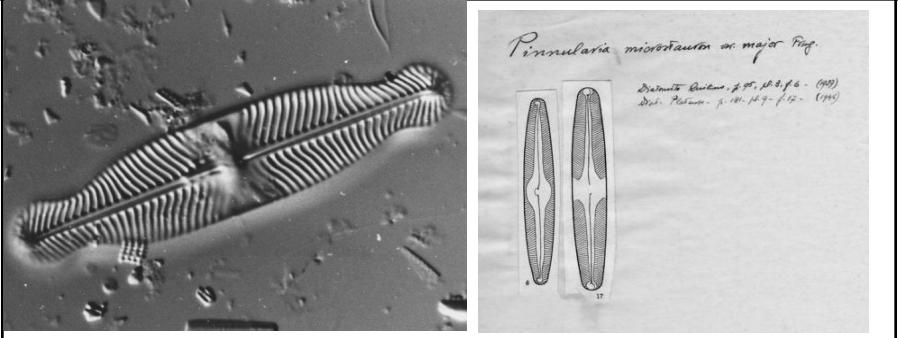
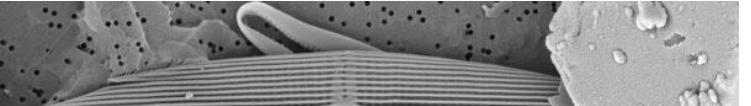
			https://diatoms.org/genera/haslea/guide
<i>Lemnicola</i> Synonyms: <i>Achnanthidium hungaricum</i> Grunow <i>Achnanthes hungarica</i> (Grunow) Grunow in Cleve and Grunow <i>Microneis hungarica</i> (Grunow) Cleve	<p>1. Valves linear to linear-elliptic, with subrostrate apices 2. Raphe valve with asymmetric fascia 3. Terminal raphe fissures turned to opposite sides</p> <p>Valves are linear to linear-elliptic with subrostrate apices. The raphe valve has a fascia. The fascia is typically asymmetric. Terminal raphe fissures are turned toward opposite sides. Striae are biseriate, 18-24 in 10 µm.</p> 	https://diatoms.org/species/lemncola_hungarica/guide	Regarded as non-marine genera.
Minutocellus			



Pauliella

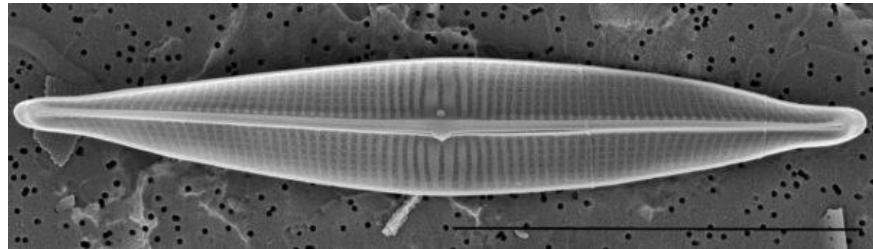


NEW
MONORAPHID
DIATOM GENUS
(POGONEZ)
FROM BAHRAIN
AND THE
TRANSFER OF
PREVIOUSLY
DESCRIBED
SPECIES A.
HUNGARZCA
AND A.
TAENZATA TO
NEW GENERA
F. E. Round
(1997)

		http://v3.boldsystems.org/index.php/Taxbrowser_Taxonpage?taxid=87690
<i>Phaeodactylum</i>		http://v3.boldsystems.org/index.php/Taxbrowser_Taxonpage?taxon=Phaeodactylum&searchTax=95_Phaeodactylum_tricornutum_polymorphism_an_overview/figures?lo=1 https://www.researchgate.net/publication/235993095_Phaeodactylum_tricornutum_polymorphism_an_overview/figures?lo=1
<i>Pinnularia</i>		http://nordicmicroalgae.org/taxon/Pinnularia%20lundii%20(Pauli%20Snoeijs)-marine https://www.marinespecies.org/photogallery.php?album=4394&pic=149512 (Drawing by Frenguelli -marine Pinnularia)
<i>Proschkinia</i>		https://diatoms.org/species/proschkinia-browderia

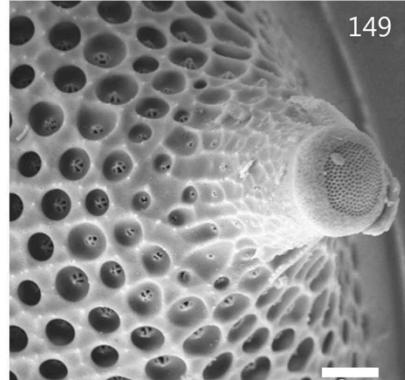
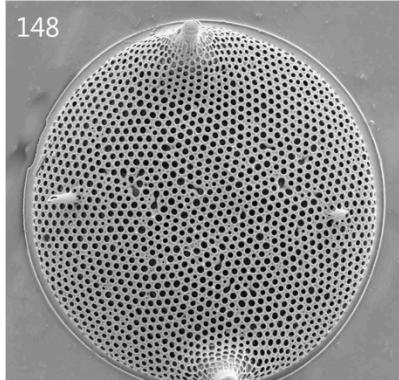
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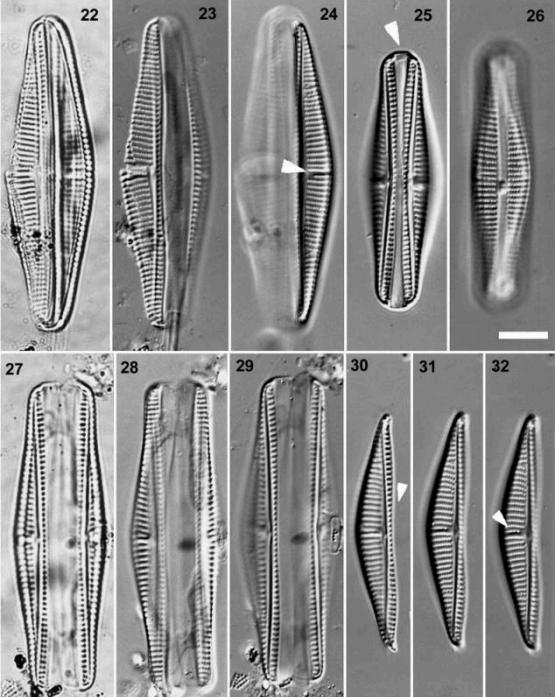
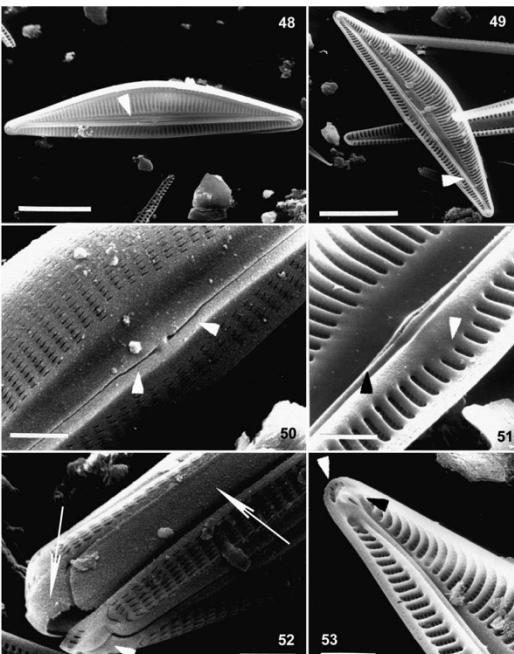
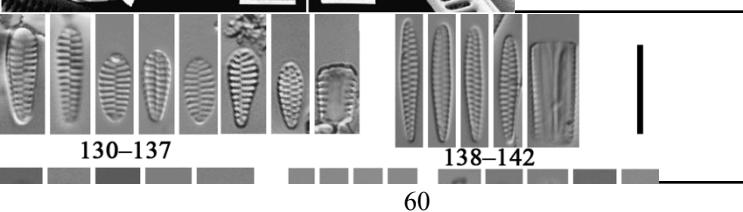
Proschkinia
Proschkinia continued.



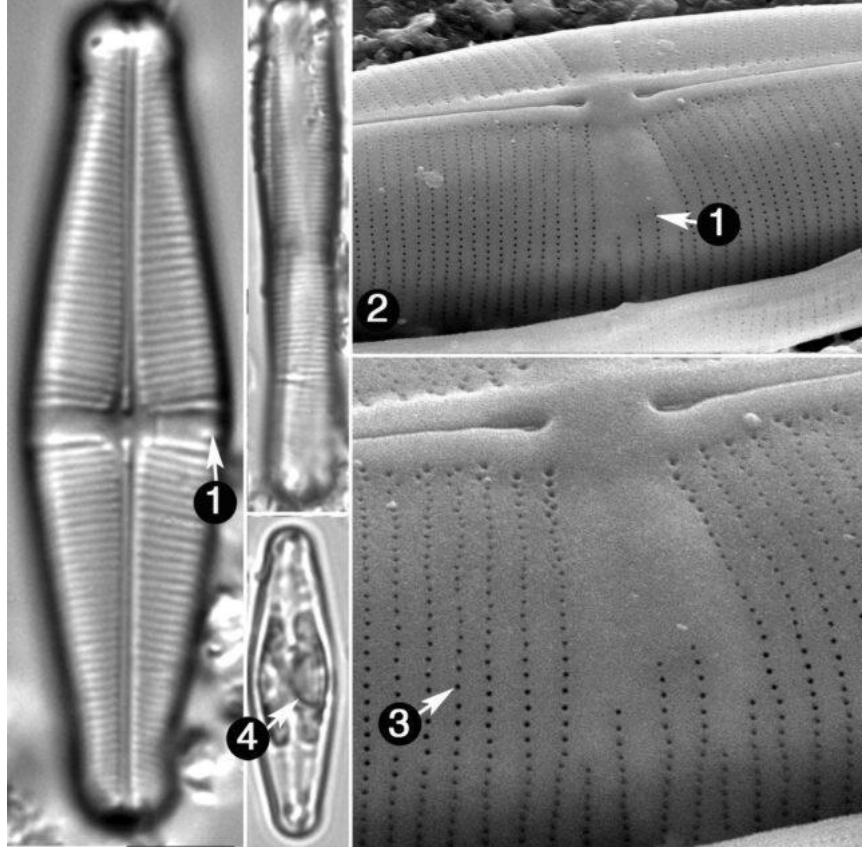
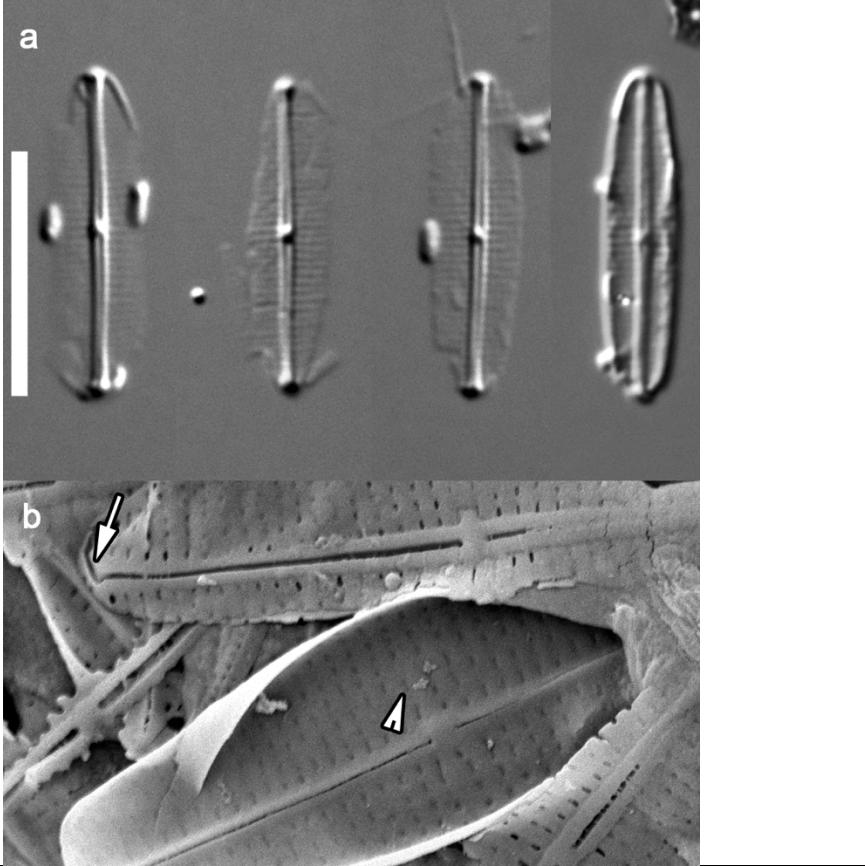
Ralfsiella smithii
(=Cerataulus smithii Ralfs in Pritchard)

2 excellent query matches in the molecular data, found under *Odontella* which will show almost the exact



same alignment stats and distance tree results.		
<i>Seminavis</i>	 	https://www.researchgate.net/publication/231980487_The_systematics_of_Seminavis_Bacillariophyta_The_lost_identities_of_Amphora_angusta_A_venericosa_and_A_malicilenta/figures?lo=1 Danielidis and Mann (2002)
<i>Serratifera andersonii</i>		Li et al. (2018)

DRAFT

<p>Staurophora: epipelagic in marine and brackish waters</p>		https://diatoms.org/genera/staurophora/guide
<p>Sternimirus <i>shandongensis</i></p>		Witkowski et al. (2016)

Possible Genera identified by molecular sequences		
<p>Berkeleya: #31 <i>SILVA</i>, <i>(BLAST-Berkelya sp. 100% 1e-116)</i> <i>Bacillariophyceae: Berkeleya sp. 100%, 100% BLAST;</i> <i>Achnanthales (Planothidium 98%, 93.33% or</i> <i>Berkeleya sp. 100%, 93.06% BLAST)</i></p> 