

Lab 9-10: Structural Bioinformatics

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PDB Statistics:

PDB is the main database for structural information on proteins.

```
db <- read.csv("PDB.csv")
head(db)
```

	Molecular.Type	X.ray	EM	NMR	Multiple.methods	Neutron	Other	
## 1	Protein (only)	154,904	10,218	12,189		191	72	32
## 2	Protein/Oligosaccharide	9,089	1,805	32		7	1	0
## 3	Protein/NA	8,129	3,184	283		6	0	0
## 4	Nucleic acid (only)	2,675	94	1,450		12	2	1
## 5	Other	163	9	32		0	0	0
## 6	Oligosaccharide (only)	11	0	6		1	0	4
##	Total							
## 1		177,606						
## 2		10,934						
## 3		11,602						
## 4		4,234						
## 5		204						
## 6		22						

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
sum(as.numeric(gsub(",", "", db$X.ray)))
```

```
## [1] 174971
```

```
em.total <- sum(as.numeric(gsub(",", "", db$EM)))
em.total
```

```
## [1] 15310
```

```
# Writing function
sum_comma <- function(x) {
  # substitute comma and convert to numeric
  sum(as.numeric(gsub(",", "", x)))
}

sum_comma(db$X.ray) / sum_comma(db$Total)
```

```
## [1] 0.8551774
```

For EM:

```
round(sum_comma(db$EM) / sum_comma(db$Total), 3)
```

```
## [1] 0.075
```

Q2: What proportion of structures in the PDB are protein?

```
round(sum_comma(db$Total[1]) / sum_comma(db$Total), 2)
```

```
## [1] 0.87
```

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

N/A

Q4. Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

The resolution of the structure is too low to be able to see H atoms.

Q5. There is a critical “conserved” water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have?

HOH308

Working with structures in R

We can use the bio3d package to read and perform bioinformatics calculations on PDB structures.

```
library(bio3d)
```

```
## Warning: package 'bio3d' was built under R version 4.3.0
```

```
pdb <- read.pdb("1hsg")
```

```
## Note: Accessing on-line PDB file
```

```
pdb
```

```
##
## Call: read.pdb(file = "1hsg")
##
## Total Models#: 1
## Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
##
## Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
## Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
##
## Non-protein/nucleic Atoms#: 172 (residues: 128)
## Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
##
## Protein sequence:
## PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
## QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
## ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
## VNIIGRNLLTQIGCTLNF
##
## + attr: atom, xyz, seqres, helix, sheet,
## calpha, remark, call
```

```
attributes(pdb)
```

```
## $names
## [1] "atom" "xyz" "seqres" "helix" "sheet" "calpha" "remark" "call"
##
## $class
## [1] "pdb" "sse"
```

```
head(pdb$atom)
```

```
## type eleno elety alt resid chain resno insert x y z o b
## 1 ATOM 1 N <NA> PRO A 1 <NA> 29.361 39.686 5.862 1 38.10
## 2 ATOM 2 CA <NA> PRO A 1 <NA> 30.307 38.663 5.319 1 40.62
## 3 ATOM 3 C <NA> PRO A 1 <NA> 29.760 38.071 4.022 1 42.64
## 4 ATOM 4 O <NA> PRO A 1 <NA> 28.600 38.302 3.676 1 43.40
## 5 ATOM 5 CB <NA> PRO A 1 <NA> 30.508 37.541 6.342 1 37.87
## 6 ATOM 6 CG <NA> PRO A 1 <NA> 29.296 37.591 7.162 1 38.40
## segid elesy charge
## 1 <NA> N <NA>
## 2 <NA> C <NA>
## 3 <NA> C <NA>
## 4 <NA> O <NA>
## 5 <NA> C <NA>
## 6 <NA> C <NA>
```

Q7. How many amino acid residues are there in this pdb object?

198

Q8. Name one of the two non-protein residues?

Mk1 and water

Q9. How many protein chains are in this structure?

2

Predicting function motions

```
adk <- read.pdb("6s36")
```

```
## Note: Accessing on-line PDB file
## PDB has ALT records, taking A only, rm.alt=TRUE
```

```
adk
```

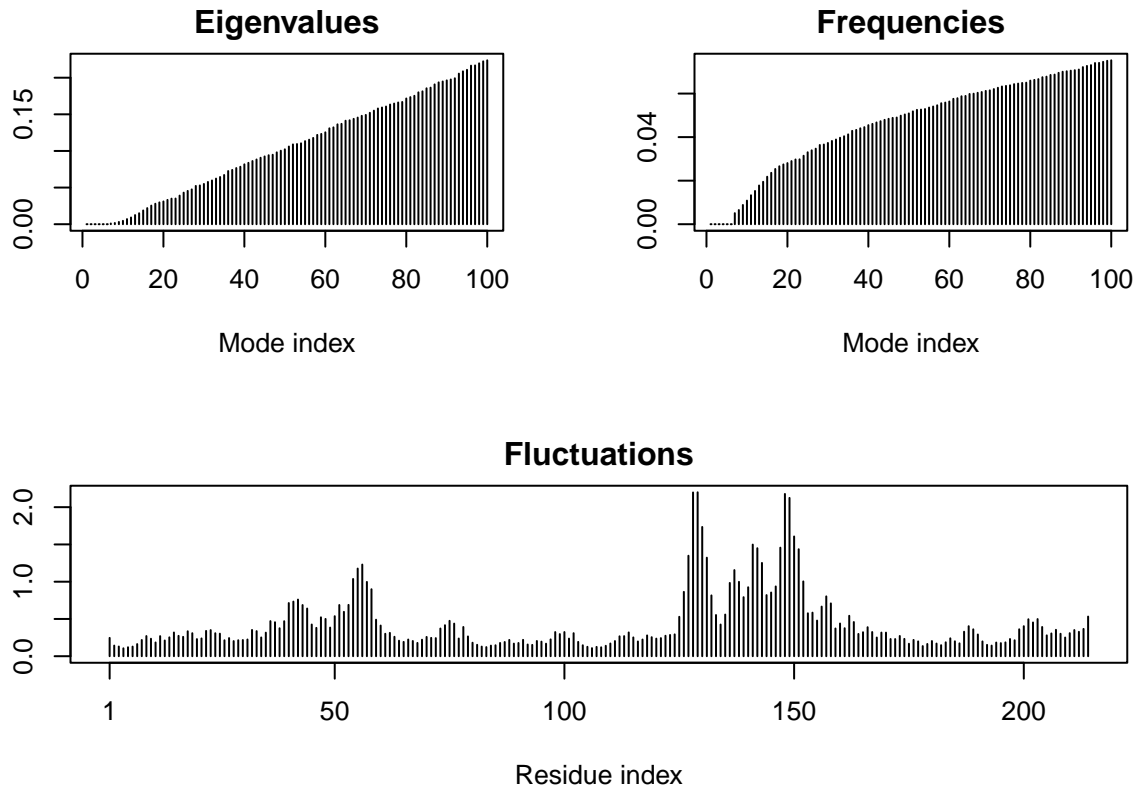
```
##
## Call: read.pdb(file = "6s36")
##
## Total Models#: 1
## Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)
##
## Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
## Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
##
## Non-protein/nucleic Atoms#: 244 (residues: 244)
## Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]
##
## Protein sequence:
## MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAAVKSGSELGKQAKDIMDAGKLV
## DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVDKI
## VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTRKDDQEETVRKRLVEYHQM TAPLIG
## YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
##
## + attr: atom, xyz, seqres, helix, sheet,
## calpha, remark, call
```

Perform a prediction of flexibility with a technique called NMA (normal mode analysis).

```
m <- nma(adk)
```

```
## Building Hessian... Done in 0.06 seconds.
## Diagonalizing Hessian... Done in 0.61 seconds.
```

```
plot(m)
```



Write out a 'movie' (trajectory) of the motion for viewing in Molstar.

```
mktrj(m, file="adk_m7.pdb")
```

```
# Install packages in the R console NOT your Rmd/Quarto file

#install.packages("bio3d")
#install.packages("devtools")
#install.packages("BiocManager")

#BiocManager::install("msa")
#devtools::install_bitbucket("Grantlab/bio3d-view")
```

Q10. Which of the packages above is found only on BioConductor and not CRAN?

msa

Q11. Which of the above packages is not found on BioConductor on CRAN?

bio3d-view

Q12. True or False? Functions from the devtools package can be used to install packages from GitHub and BitBucket?

True

```
library(bio3d)
```

```
aa <- get.seq("lake_A")
```

```
## Warning in get.seq("lake_A"): Removing existing file: seqs.fasta
```

```
## Fetching... Please wait. Done.
```

```
aa
```

```
##           1           .           .           .           .           .           60
## pdb|1AKE|A  MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAAVKSGSELGKQAKDIMDAGKLV
##           1           .           .           .           .           .           60
##
##           61           .           .           .           .           .           120
## pdb|1AKE|A  DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVDRI
##           61           .           .           .           .           .           120
##
##          121           .           .           .           .           .           180
## pdb|1AKE|A  VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQM
##          121           .           .           .           .           .           180
##
##          181           .           .           .           .           .           214
## pdb|1AKE|A  YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
##          181           .           .           .           .           .           214
##
## Call:
##   read.fasta(file = outfile)
##
## Class:
##   fasta
##
## Alignment dimensions:
##   1 sequence rows; 214 position columns (214 non-gap, 0 gap)
##
## + attr: id, ali, call
```

Q13. How many amino acids are in this sequence, i.e. how long is this sequence?

214 amino acids long

```
# Blast search
# b <- blast.pdb(aa)
```

```
# hits <- plot(b)
```

```
# Checking what's in the hits object
# hits$pdb.id
```

```

hits <- NULL
hits$pdb.id <- c('1AKE_A','6S36_A','6RZE_A','3HPR_A','1E4V_A','5EJE_A','1E4Y_A','3X2S_A','6HAP_A','6HAM_A')

# Download related PDB files
files <- get.pdb(hits$pdb.id, path="pdbs", split=TRUE, gzip=TRUE)

## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
## pdbs/1AKE.pdb exists. Skipping download

## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
## pdbs/6S36.pdb exists. Skipping download

## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
## pdbs/6RZE.pdb exists. Skipping download

## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
## pdbs/3HPR.pdb exists. Skipping download

## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
## pdbs/1E4V.pdb exists. Skipping download

## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
## pdbs/5EJE.pdb exists. Skipping download

## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
## pdbs/1E4Y.pdb exists. Skipping download

## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
## pdbs/3X2S.pdb exists. Skipping download

## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
## pdbs/6HAP.pdb exists. Skipping download

## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
## pdbs/6HAM.pdb exists. Skipping download

## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
## pdbs/4K46.pdb exists. Skipping download

## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
## pdbs/3GMT.pdb exists. Skipping download

## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
## pdbs/4PZL.pdb exists. Skipping download

##      |

```

```
# Align related PDBs
```

```
pdbs <- pdbaln(files, fit = TRUE, exefile="msa")
```

```
## Reading PDB files:
```

```
## pdbs/split_chain/1AKE_A.pdb
```

```
## pdbs/split_chain/6S36_A.pdb
```

```
## pdbs/split_chain/6RZE_A.pdb
```

```
## pdbs/split_chain/3HPR_A.pdb
```

```
## pdbs/split_chain/1E4V_A.pdb
```

```
## pdbs/split_chain/5EJE_A.pdb
```

```
## pdbs/split_chain/1E4Y_A.pdb
```

```
## pdbs/split_chain/3X2S_A.pdb
```

```
## pdbs/split_chain/6HAP_A.pdb
```

```
## pdbs/split_chain/6HAM_A.pdb
```

```
## pdbs/split_chain/4K46_A.pdb
```

```
## pdbs/split_chain/3GMT_A.pdb
```

```
## pdbs/split_chain/4PZL_A.pdb
```

```
## PDB has ALT records, taking A only, rm.alt=TRUE
```

```
## . PDB has ALT records, taking A only, rm.alt=TRUE
```

```
## . PDB has ALT records, taking A only, rm.alt=TRUE
```

```
## . PDB has ALT records, taking A only, rm.alt=TRUE
```

```
## .. PDB has ALT records, taking A only, rm.alt=TRUE
```

```
## .... PDB has ALT records, taking A only, rm.alt=TRUE
```

```
## . PDB has ALT records, taking A only, rm.alt=TRUE
```

```
## ...
```

```
##
```

```
## Extracting sequences
```

```
##
```

```
## pdb/seq: 1 name: pdbs/split_chain/1AKE_A.pdb
```

```
## PDB has ALT records, taking A only, rm.alt=TRUE
```

```
## pdb/seq: 2 name: pdbs/split_chain/6S36_A.pdb
```

```
## PDB has ALT records, taking A only, rm.alt=TRUE
```

```
## pdb/seq: 3 name: pdbs/split_chain/6RZE_A.pdb
```

```
## PDB has ALT records, taking A only, rm.alt=TRUE
```

```
## pdb/seq: 4 name: pdbs/split_chain/3HPR_A.pdb
```

```
## PDB has ALT records, taking A only, rm.alt=TRUE
```

```
## pdb/seq: 5 name: pdbs/split_chain/1E4V_A.pdb
```

```
## pdb/seq: 6 name: pdbs/split_chain/5EJE_A.pdb
```

```
## PDB has ALT records, taking A only, rm.alt=TRUE
```

```
## pdb/seq: 7 name: pdbs/split_chain/1E4Y_A.pdb
```

```
## pdb/seq: 8 name: pdbs/split_chain/3X2S_A.pdb
```

```
## pdb/seq: 9 name: pdbs/split_chain/6HAP_A.pdb
```

```
## pdb/seq: 10 name: pdbs/split_chain/6HAM_A.pdb
```

```
## PDB has ALT records, taking A only, rm.alt=TRUE
```

```
## pdb/seq: 11 name: pdbs/split_chain/4K46_A.pdb
```

```
## PDB has ALT records, taking A only, rm.alt=TRUE
```

```
## pdb/seq: 12 name: pdbs/split_chain/3GMT_A.pdb
```

```
## pdb/seq: 13 name: pdbs/split_chain/4PZL_A.pdb
```

```
# Looking at the 'pdbs' object
```

```
pdbs
```

```
##
```

```
1
```

```
.
```

```
.
```

```
.
```

```
40
```



```

## [Truncated_Name:1] 1AKE_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
## [Truncated_Name:2] 6S36_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
## [Truncated_Name:3] 6RZE_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
## [Truncated_Name:4] 3HPR_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
## [Truncated_Name:5] 1E4V_A.pdb -----MRIILLGAPVAGKGTQAQFIMEKYGIPQIS
## [Truncated_Name:6] 5EJE_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
## [Truncated_Name:7] 1E4Y_A.pdb -----MRIILLGALVAGKGTQAQFIMEKYGIPQIS
## [Truncated_Name:8] 3X2S_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
## [Truncated_Name:9] 6HAP_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
## [Truncated_Name:10] 6HAM_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
## [Truncated_Name:11] 4K46_A.pdb -----MRIILLGAPGAGKGTQAQFIMAKFGIPQIS
## [Truncated_Name:12] 3GMT_A.pdb -----MRLILLGAPGAGKGTQANFIKEKFGIPQIS
## [Truncated_Name:13] 4PZL_A.pdb TENLYFQSNAMRIILLGAPGAGKGTQAKIIEQKYNIAHIS
##
##
##
##
## [Truncated_Name:1] 1AKE_A.pdb
## [Truncated_Name:2] 6S36_A.pdb
## [Truncated_Name:3] 6RZE_A.pdb
## [Truncated_Name:4] 3HPR_A.pdb
## [Truncated_Name:5] 1E4V_A.pdb
## [Truncated_Name:6] 5EJE_A.pdb
## [Truncated_Name:7] 1E4Y_A.pdb
## [Truncated_Name:8] 3X2S_A.pdb
## [Truncated_Name:9] 6HAP_A.pdb
## [Truncated_Name:10] 6HAM_A.pdb
## [Truncated_Name:11] 4K46_A.pdb
## [Truncated_Name:12] 3GMT_A.pdb
## [Truncated_Name:13] 4PZL_A.pdb
##
##
##
##
## [Truncated_Name:1] 1AKE_A.pdb
## [Truncated_Name:2] 6S36_A.pdb
## [Truncated_Name:3] 6RZE_A.pdb
## [Truncated_Name:4] 3HPR_A.pdb
## [Truncated_Name:5] 1E4V_A.pdb
## [Truncated_Name:6] 5EJE_A.pdb
## [Truncated_Name:7] 1E4Y_A.pdb
## [Truncated_Name:8] 3X2S_A.pdb
## [Truncated_Name:9] 6HAP_A.pdb
## [Truncated_Name:10] 6HAM_A.pdb
## [Truncated_Name:11] 4K46_A.pdb
## [Truncated_Name:12] 3GMT_A.pdb
## [Truncated_Name:13] 4PZL_A.pdb
##
##
##
##
## [Truncated_Name:1] 1AKE_A.pdb
## [Truncated_Name:2] 6S36_A.pdb
## [Truncated_Name:3] 6RZE_A.pdb
## [Truncated_Name:4] 3HPR_A.pdb
## [Truncated_Name:5] 1E4V_A.pdb
## [Truncated_Name:6] 5EJE_A.pdb
## [Truncated_Name:7] 1E4Y_A.pdb
## [Truncated_Name:8] 3X2S_A.pdb
## [Truncated_Name:9] 6HAP_A.pdb
## [Truncated_Name:10] 6HAM_A.pdb
## [Truncated_Name:11] 4K46_A.pdb
## [Truncated_Name:12] 3GMT_A.pdb
## [Truncated_Name:13] 4PZL_A.pdb
##
##
##
##
## [Truncated_Name:1] 1AKE_A.pdb
## [Truncated_Name:2] 6S36_A.pdb
## [Truncated_Name:3] 6RZE_A.pdb

```

```

## [Truncated_Name:4] 3HPR_A.pdb      VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDGTG
## [Truncated_Name:5] 1E4V_A.pdb      VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG
## [Truncated_Name:6] 5EJE_A.pdb      VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG
## [Truncated_Name:7] 1E4Y_A.pdb      VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG
## [Truncated_Name:8] 3X2S_A.pdb      VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG
## [Truncated_Name:9] 6HAP_A.pdb      VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG
## [Truncated_Name:10] 6HAM_A.pdb     VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG
## [Truncated_Name:11] 4K46_A.pdb     VADSVIVERMAGRAHLASGRTYHNVYNPPKVEGKDDVTG
## [Truncated_Name:12] 3GMT_A.pdb     VPFSEIIERMSGRRTHPASGRTYHVKNPPKVEGKDDVTG
## [Truncated_Name:13] 4PZL_A.pdb     VADNLLIERITGRIHPASGRTYHTKFNPPKVADKDDVTG
##                                     *   ^^^ ^ *** * *** ** ^***** ** **
##                                     121       .       .       .       160
##
##                                     161       .       .       .       200
## [Truncated_Name:1] 1AKE_A.pdb      EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN
## [Truncated_Name:2] 6S36_A.pdb      EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN
## [Truncated_Name:3] 6RZE_A.pdb      EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN
## [Truncated_Name:4] 3HPR_A.pdb      EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN
## [Truncated_Name:5] 1E4V_A.pdb      EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN
## [Truncated_Name:6] 5EJE_A.pdb      EELTTRKDDQEECVRKRLVEYHQM TAPLIGYYSKEAEAGN
## [Truncated_Name:7] 1E4Y_A.pdb      EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN
## [Truncated_Name:8] 3X2S_A.pdb      EELTTRKDDQEETVRKRLCEYHQM TAPLIGYYSKEAEAGN
## [Truncated_Name:9] 6HAP_A.pdb      EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN
## [Truncated_Name:10] 6HAM_A.pdb     EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN
## [Truncated_Name:11] 4K46_A.pdb     EDLVIREDDKEETVLARLGVYHNQ TAPLIAYYGKEAEAGN
## [Truncated_Name:12] 3GMT_A.pdb     EPLVQRDDDKKEETVKKRLDVYEAQTKPLITYYGDWARRGA
## [Truncated_Name:13] 4PZL_A.pdb     EPLITRTDDNEDTVKQRLSVYHAQTAKLIDFYRNFSSNT
##                                     * * * ** * ^ * ** * * ** ^*
##                                     161       .       .       .       200
##
##                                     201       .       .       227
## [Truncated_Name:1] 1AKE_A.pdb      T--KYAKVDGTPVAEVRADLEKILG-
## [Truncated_Name:2] 6S36_A.pdb      T--KYAKVDGTPVAEVRADLEKILG-
## [Truncated_Name:3] 6RZE_A.pdb      T--KYAKVDGTPVAEVRADLEKILG-
## [Truncated_Name:4] 3HPR_A.pdb      T--KYAKVDGTPVAEVRADLEKILG-
## [Truncated_Name:5] 1E4V_A.pdb      T--KYAKVDGTPVAEVRADLEKILG-
## [Truncated_Name:6] 5EJE_A.pdb      T--KYAKVDGTPVAEVRADLEKILG-
## [Truncated_Name:7] 1E4Y_A.pdb      T--KYAKVDGTPVAEVRADLEKILG-
## [Truncated_Name:8] 3X2S_A.pdb      T--KYAKVDGTPVAEVRADLEKILG-
## [Truncated_Name:9] 6HAP_A.pdb      T--KYAKVDGTPVCEVRADLEKILG-
## [Truncated_Name:10] 6HAM_A.pdb     T--KYAKVDGTPVCEVRADLEKILG-
## [Truncated_Name:11] 4K46_A.pdb     T--QYLKFDGTPKAVAEVSAELEKALA-
## [Truncated_Name:12] 3GMT_A.pdb     E-----NGLKAPA-----YRKISG-
## [Truncated_Name:13] 4PZL_A.pdb     KIPKYIKINGDQAVEKVSQDIFDQLNK
##                                     *
##                                     201       .       .       227
##
## Call:
##   pdbaln(files = files, fit = TRUE, exefile = "msa")
##
## Class:
##   pdb, fasta
##
## Alignment dimensions:

```

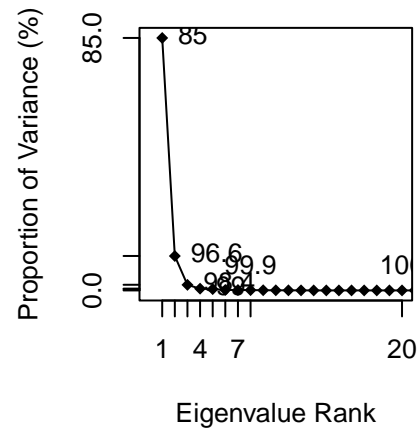
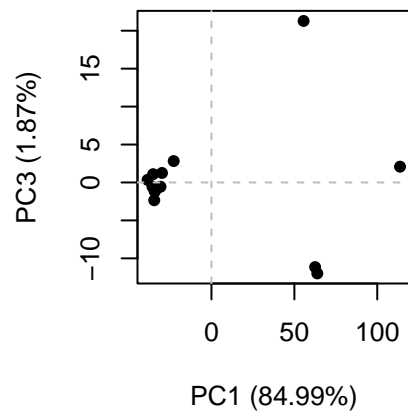
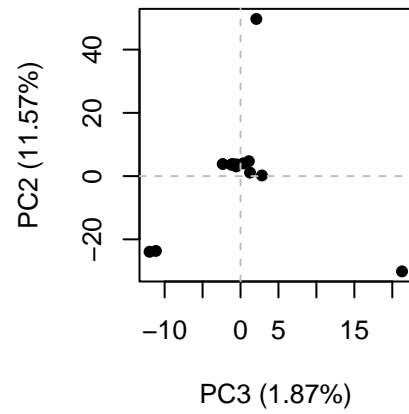
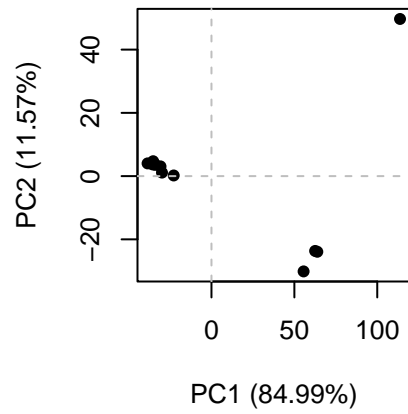
```
## 13 sequence rows; 227 position columns (204 non-gap, 23 gap)
##
## + attr: xyz, resno, b, chain, id, ali, resid, sse, call
```

```
# Now that we have our aligned and superposed structures, we can perform all sorts of analyses on them
# Let's do PCA:
```

```
pc.xray <- pca(pdbbs)
pc.xray
```

```
##
## Call:
##  pca.pdbbs(pdbbs = pdbbs)
##
## Class:
##  pca
##
## Number of eigenvalues:
##  612
##
##      Eigenvalue Variance Cumulative
## PC 1    2824.299   84.993    84.993
## PC 2     384.613   11.574    96.568
## PC 3      62.077    1.868    98.436
## PC 4      19.614    0.590    99.026
## PC 5      14.644    0.441    99.467
## PC 6       5.228    0.157    99.624
##
##      (Obtained from 13 conformers with 612 xyz input values).
##
## + attr: L, U, z, au, sdev, mean, call
```

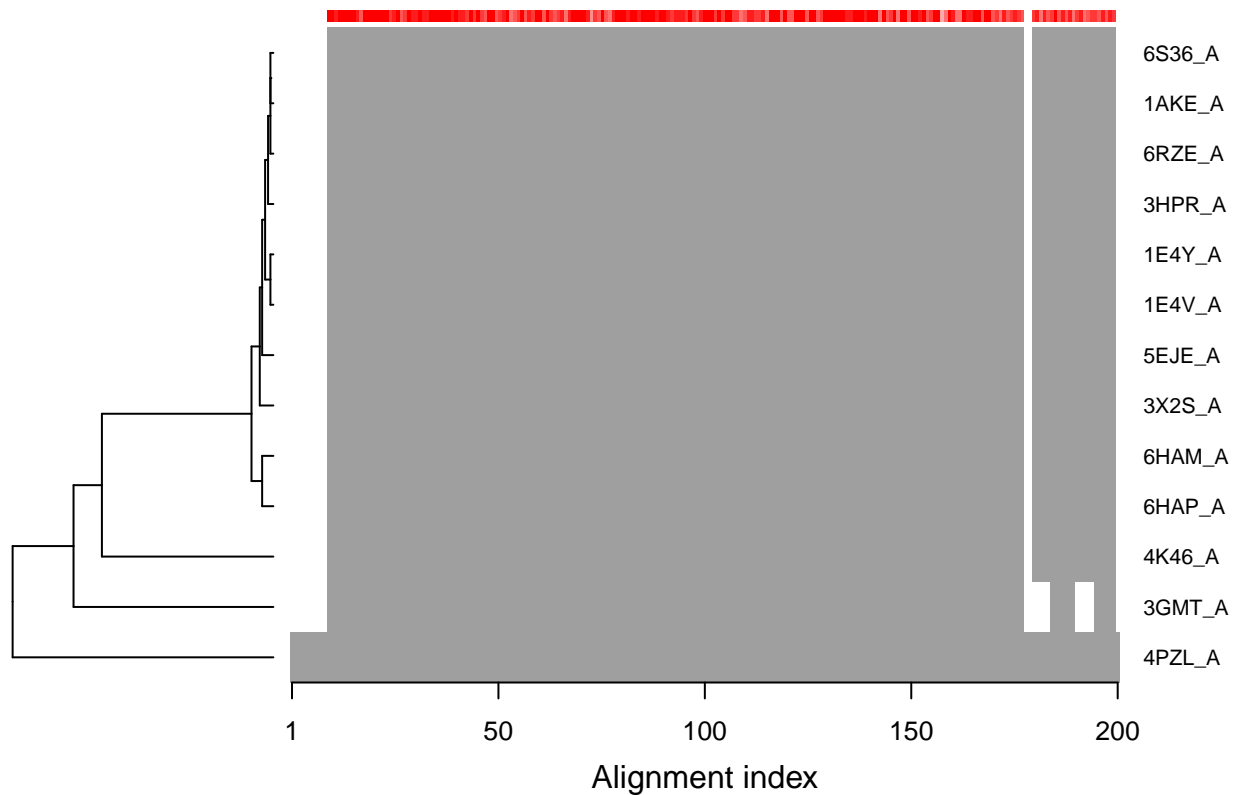
```
plot(pc.xray)
```



```
# Vector containing PDB codes for figure axis
ids <- basename.pdb(pdb$ids)

# Draw schematic alignment
plot(pdb, labels=ids)
```

Sequence Alignment Overview



```
# Results of PCA on Adenylate kinase X-ray structures:
```

```
# Calculate RMSD
```

```
rd <- rmsd(pdb)
```

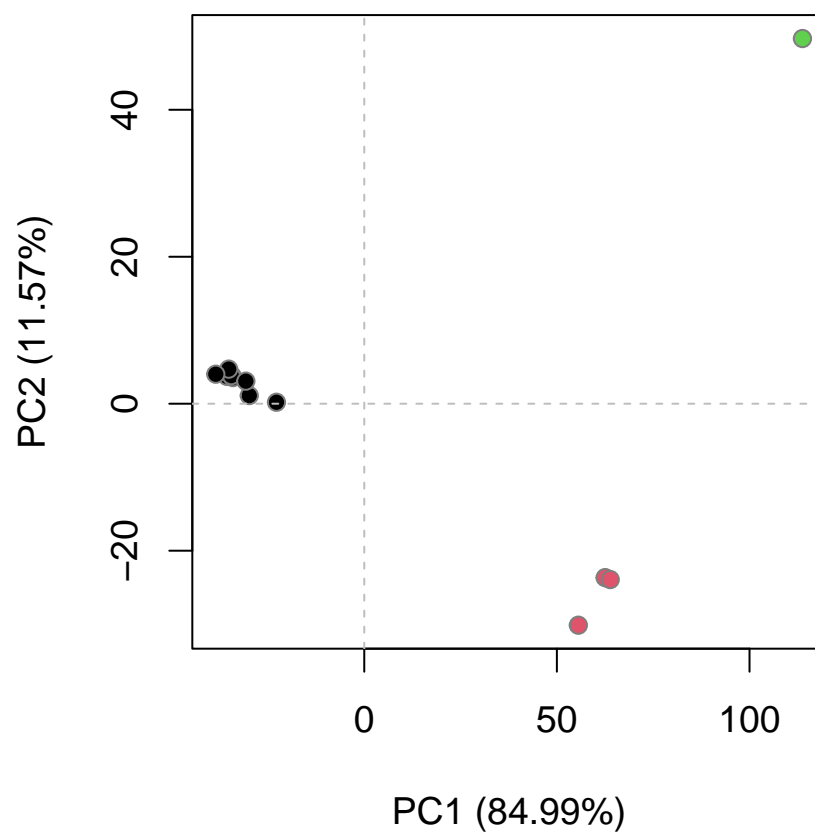
```
## Warning in rmsd(pdb): No indices provided, using the 204 non NA positions
```

```
# Structure-based clustering
```

```
hc.rd <- hclust(dist(rd))
```

```
grps.rd <- cutree(hc.rd, k=3)
```

```
plot(pc.xray, 1:2, col="grey50", bg=grps.rd, pch=21, cex=1)
```



```
#plot(pc.xray, 1:2, col=grps.rd)
```

```
# Visualize first principal component
pc1 <- mktrj(pc.xray, pc=1, file="pc_1.pdb")
pc1
```

```
##
##   Total Frames#: 34
##   Total XYZs#:   612, (Atoms#:  204)
##
##   [1] 26.787 52.261 40.414 <...> 15.653 53.622 42.018 [20808]
##
## + attr: Matrix DIM = 34 x 612
```