



Authentic Check

 Preventive Health Check-up | Pathology | Digital X-Ray | Sonography | Colour Doppler | Mammography | BMD (DXA Scan) | OPG | ECG | 2D Echo
 Stress Test/TMT | Spirometry | Eye Examination | Dental Examination | Diet Consultation | Audiometry | OT Sterility | Water Sterility | Clinical Research

CID	: 2103145500	SID	: 177802813825	R E P O R T
Name	: MR.GYANENDRA PRATAP CHANDRAPATI SINGH	Registered	: 20-Jun-2021 / 16:31	
Age / Gender	: 34 Years / Male	Collected	: 20-Jun-2021 / 16:00	
Dr.	: -	Reported	: 22-Jun-2021 / 12:10	
Reg. Location	: Lanka, Varanasi	Printed	: 22-Jun-2021 / 13:25	

Real time Qualitative RT-PCR detection of 2019-nCoV RNA / COVID-19 RNA

<u>PARAMETER</u>	<u>RESULT</u>
Result	SARS-CoV-2: Not Detected (Negative)
Kit Specification	Genestore SARS-CoV-2, Target gene (N1 & N2), Cutoff <37

ICMR Registration No: CHSLLUP

Specimen: Nasopharyngeal & Oropharyngeal swab in VTM

Method: Real time RT-PCR

Note:

- ICMR recommended kits are used for reporting. All the positive cases will be notified to ICMR for further surveillance.
- Clinical correlation with patient history, radiology findings and co-infection with other virus infection is necessary to be determined especially in cases with Borderline positive Ct values.
- Borderline positive cases (Ct Value >30) may give variable results on repeat testing. The possible reasons could be the variations in kits and instruments used.

Limitations:

- Optimum specimen types and timing of peak viral levels during infections caused by 2019-nCoV have not been determined. Collection of multiple specimens (Types & Time points) may be necessary in view of suspected clinical history. The repeat specimen may be considered after a gap of 2-4 days after the collection of first specimen for additional testing if required. (other respiratory pathogens)
- Negative results do not preclude SARS - CoV - 2 infection and should not be used as the sole basis for patient management decisions.
- This test is a qualitative assay and does not quantify viral load. Various host factors, viral factors, variability in sample collection / site and techniques used by the laboratories can affect Ct values. Therefore, Ct values are not an absolute indication of viral load and should be interpreted with caution.

Factors leading to false negative RT-PCR report:

- Inadequate specimen collection, Poor quality of sample and non-representative sample.
- Sample collected too early or too late in the infection, Improper sample handling and shipment.
- Technical reasons- PCR Inhibitor, analytical sensitivity of kit used.
- Active recombination &/ mutations in target genes used for detection of SARS-CoV-2 virus.

References:

- Diagnostic detection of 2019-nCoV by real-time RT-PCR, Berlin Jan 17th, 2020.
- Labcorp COVID-19 RT-PCR test EUA Summary / COVID-19 RT-PCR test (laboratory corporation of America).

*** End Of Report ***



MC-2111

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