

# Physical Mapping of DNA

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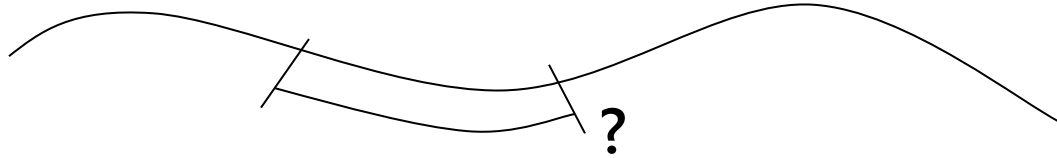
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# Background

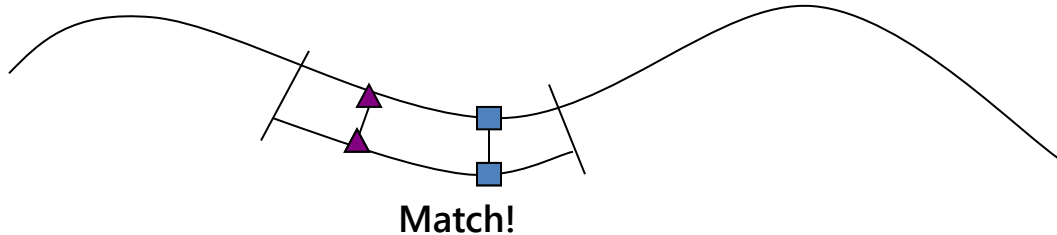
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Given a sequence of DNA, how do we figure out where on some larger chromosome the sequence lies?



Look for markers that match in both the chromosome and the shorter sequence.

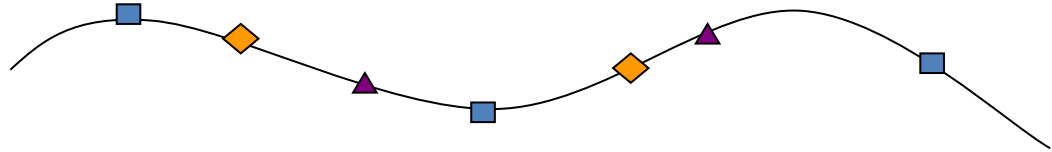
- Markers: Usually short, precisely defined sequences



# Creating the Physical Map

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How do we create the original map?



Generate fingerprints (markers) with:

- Restriction site mapping
- Hybridization

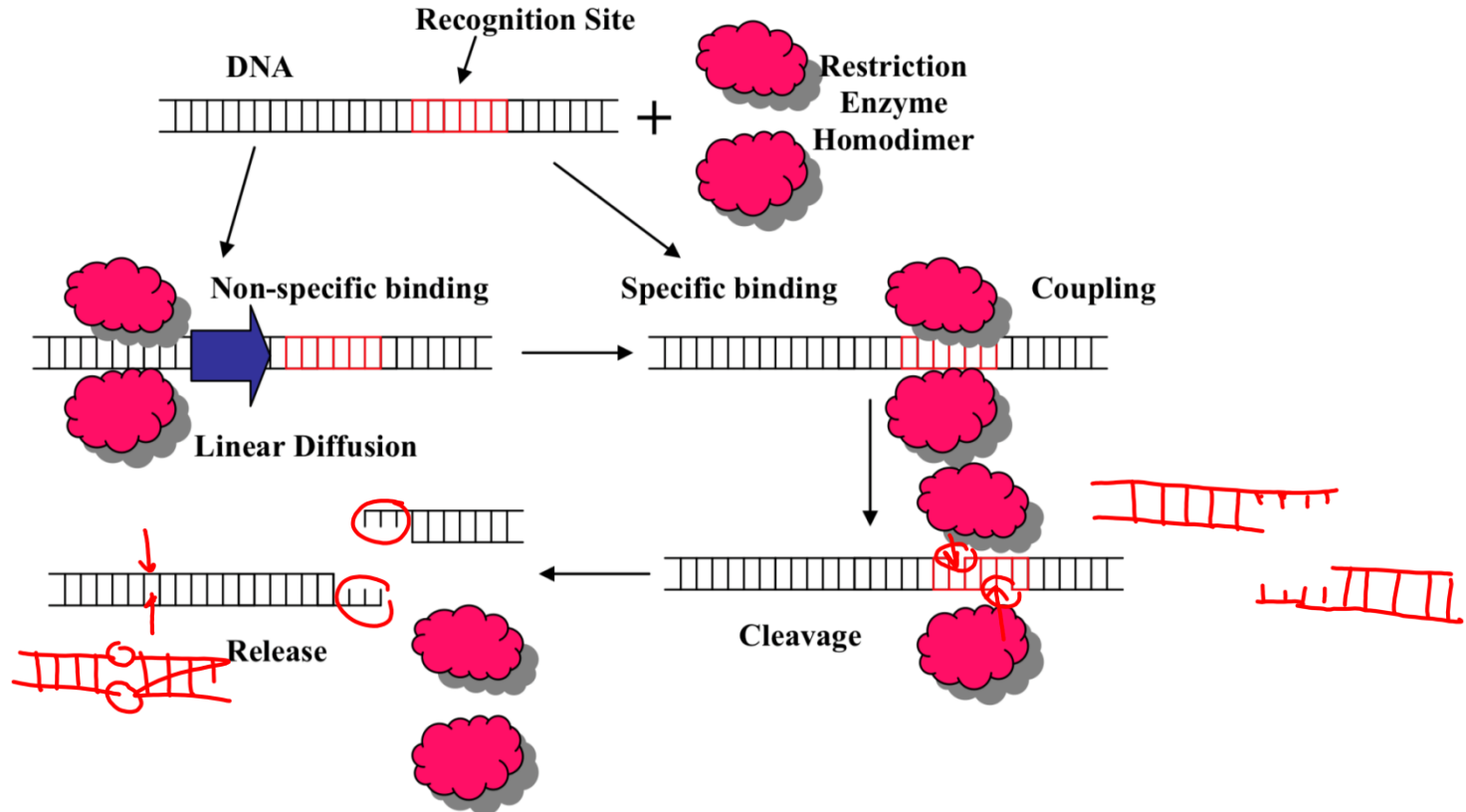
Can't we just expand the sequence assembly techniques we've already learned?  
No!

Why not?

- A chromosome isn't just a few kilo bps long.
- Human chromosomes range in length from 51 million to 245 million base pairs.

# Restriction enzymes

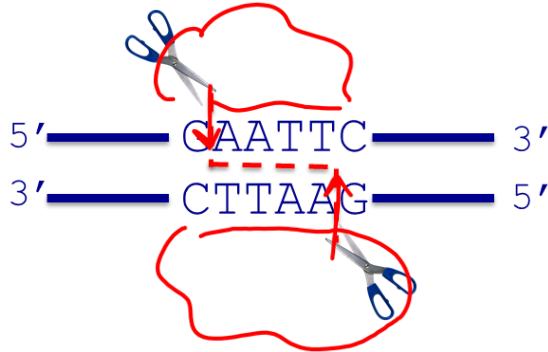
Molecular scissors that cut double stranded DNA molecules at specific points



# Restriction enzymes



EcoRI recognition site is a palindrome with an axis of symmetry



EcoRI dimer binds sequence and catalyzes double-strand cleavage



Products have “sticky ends” or overhanging bases.

# Restriction enzymes

## Examples of Restriction Enzymes

<u>Enzyme</u>	<u>Microorganism</u>	<u>Recognition Sequence</u>	<u>Isoschizomers</u>
Alu I	<i>Arthrobacter luteus</i>	AG CT	
Apa I	<i>Acetobacter pasteurianus</i>	GGGCC C	Bsp120 I, PspOM I
Bam HI	<i>Bacillus amiloliquifaciens</i>	G GATCC	
Bgl II	<i>Bacillus globigii</i>	A GATCT	
Cla I	<i>Caryophanon latum</i> L	AT CGAT	Bsp DI, Bsc I, BspX I
Dde I	<i>Desulfovibrio desulfuricans</i>	C TNAG	BstDE I
Dra I	<i>Deinococcus radiophilus</i>	TTT AAA	
Eco RI	<i>Escherichia coli</i> RY13	G AATTC	
Eco RV	<i>Escherichia coli</i> J62	GAT ATC	Eco32 I
Fnu4H I	<i>Fusobacterium nucleatum</i> 4H	GC NGC	Fsp4H I, Ita I
Hae III	<i>Haemophilus aegyptius</i>	<u>GG CC</u>	Bsh I, <u>BsuR I</u> , Pal I
Hind II	<i>Haemophilus influenzae</i> Rd	A AGCTT	
Hinf I	<i>Haemophilus influenzae</i> Rf	<u>G ANTC</u>	
Kpn I	<i>Klebsiella pneumoniae</i> OK8	GGTAC C	Acc65 I, Asp718 I
Mbo I	<i>Moraxella bovis</i>	GATC	Dpn II, Nde II, Sau3A I
Msp I	<i>Moraxella</i> sp.	C CGG	BsiS I, Hap II, Hpa II
Nde I	<i>Neisseria dentrificans</i>	CA TATG	FauND I
Not I	<i>Nocardia otitidis-caviarum</i>	GC GGCCGC	CciN I
Pst I	<i>Providencia stuartii</i> 164	CTGCA G	
Pvu II	<i>Proteus vulgaris</i>	CAG CTG	
Rsa I	<i>Rhodopseudomonas sphaeroides</i>	GT AC	
Sma I	<i>Serratia marcescens</i> S	CCC GGG	Cfr9 I, Psp A I, Xma I
Taq I	<i>Thermus aquaticus</i> YT1	T CGA	TtaHB8 I
Xba I	<i>Xanthomonas badrii</i>	T CTAGA	
Xho I	<i>Xanthomonas holcicola</i>	C TCGAG	PaeR7 I, Sfr274 I, Tli I

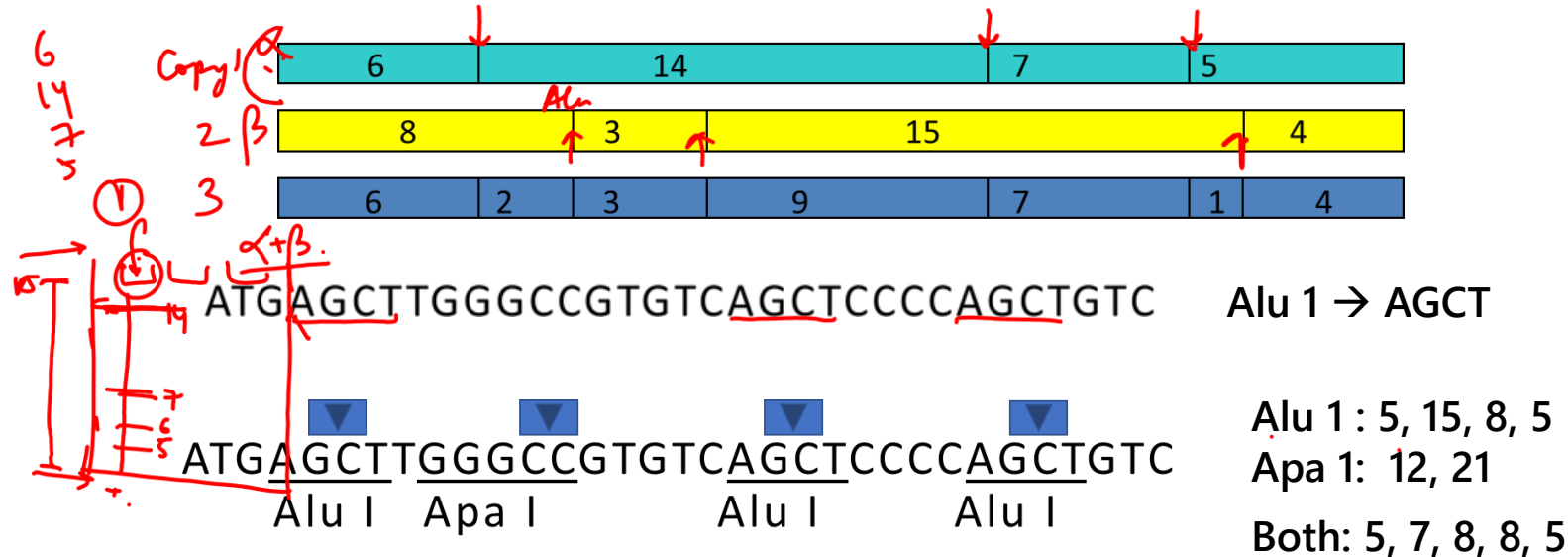
# Restriction Site Mapping

In this situation, the fingerprint is the length between restriction sites of given enzymes.

Make three copies of target DNA: strings A, B, C.

Apply one enzyme ( $\alpha$ ) to string A, another ( $\beta$ ) to string B, and both ( $\alpha$  and  $\beta$ ) to string C.

Line up the fragments in A and B so they match C: This is the double digestion problem.



# Restriction Site Mapping

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A variant is the *partial digestion approach*:

Use only one enzyme, but allow it to act for different time periods.

Different restriction sites will be recognized.

6	14	7	5
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Fragment sites: 6, 20, 27, 32;

14, 21, 26;

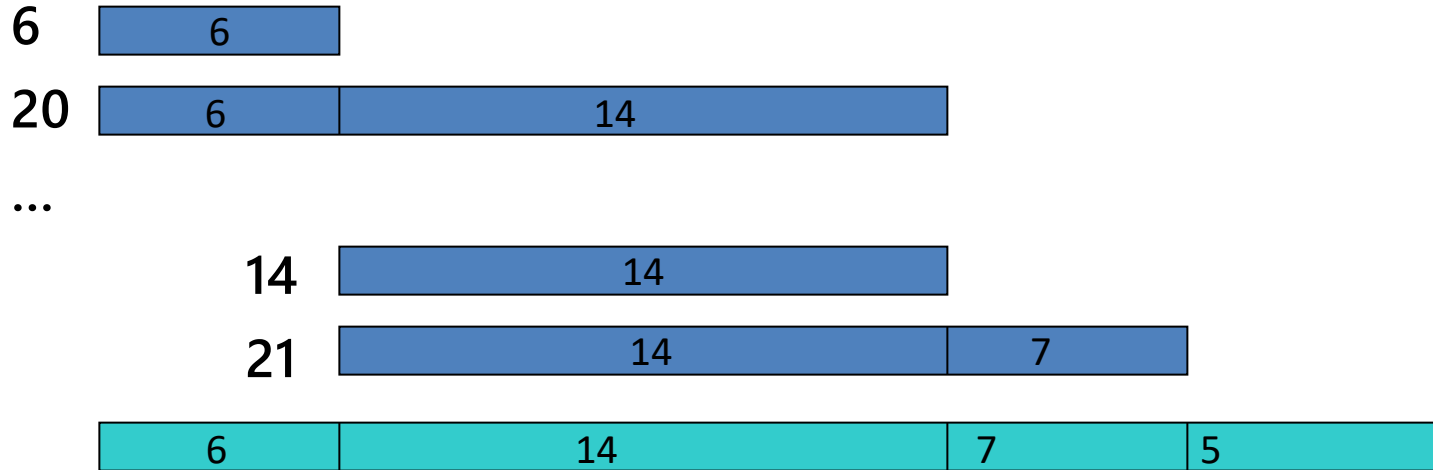
7, 12;

and 5



# Restriction Site Mapping

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etc ...

Fragment sites: 6, 20, 27, 32; 14, 21, 26; 7, 12; and 5

# Restriction site model

Back to the double digest problem:

Enzyme A: 1000, 2100, 1400, 500

Enzyme B: 1200, 2500, 1300

Enzyme A+B: 1000, 200, 1900, 600, 800, 500

Find permutations of A and B such that there is a one-to-one correspondence between all the subintervals and C.



A+B: 1000    200    1900    600    800    500  
          a        b        c        d        e        f

A: 1000 = a, 2100 = b+c, 1400 = d+e, 500 = f

B: 1200 = a+b, 2500 = c+d, 1300 = e+f

1000 = a	2100 = b+c	1400 = d+e	500 = f
1200 = a+b	2500 = c+d	1300 = e+f	

# Restriction site models

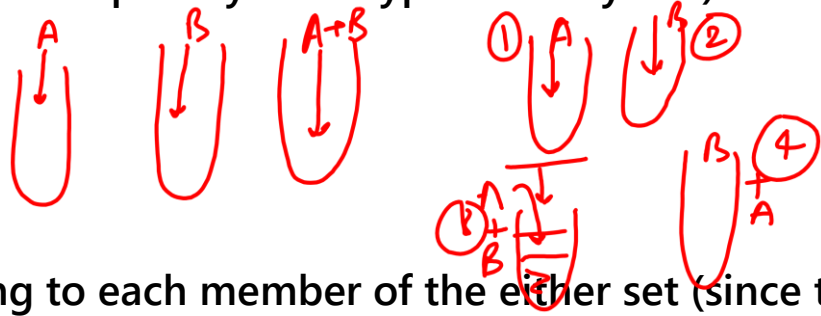
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## Limitations:

- This double digestion problem is NP-complete.
- Between 2 sites cut by A, there are three sites – b1, b2, b3 cut by enzyme B. It will be difficult to tell the order of the fragments [b1,b2] and [b2,b3].
- The number of solutions is  $k!$  for  $k$  = number of restriction sites.

# Enhanced Double Digestion Problem

- The Enhanced Double Digest (EDD) problem is NP-hard in the general case, but if the lengths of fragments in C (the string acted upon by both types of enzymes) are distinct, it can be solved in linear time!
- We have the multisets A and B.  
 $A = \{6, 14, 7, 5\}$   
 $B = \{8, 3, 15, 4\}$
- Take the actual fragments corresponding to each member of the either set (since the sets are only lengths). Apply the other enzyme to the fragment (i.e. apply enzyme  $\beta$  to fragments from A, and vice versa) to create subfragments.
- $AB_i$  is the multiset of subfragments created by applying enzyme  $\beta$  to fragments from A;
- $BA_j$  from applying enzyme  $\alpha$  to fragments of B.



# Enhanced Double Digestion Problem

Example:

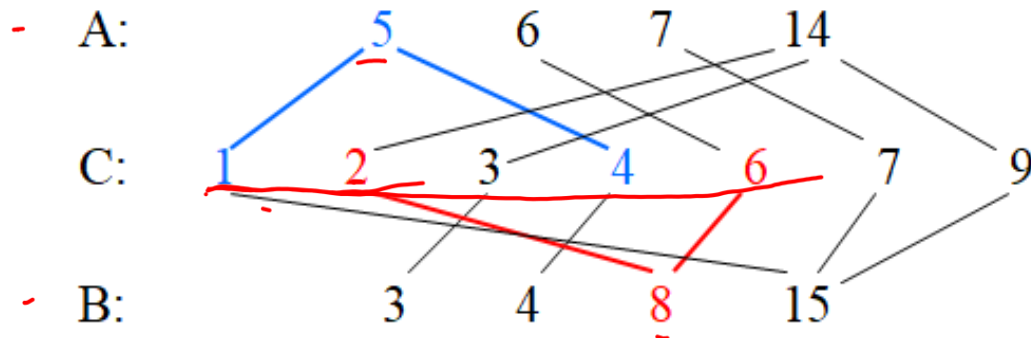
$A = \{5, 6, 7, 14\}$

$B = \{3, 4, 8, 15\}$

$AB_1 = \{1, 4\}$ ,  $AB_2 = \{6\}$ ,  $AB_3 = \{7\}$ ,  $AB_4 = \{2, 3, 9\}$

$BA_1 = \{3\}$ ,  $BA_2 = \{4\}$ ,  $BA_3 = \{2, 6\}$ ,  $BA_4 = \{1, 7, 9\}$

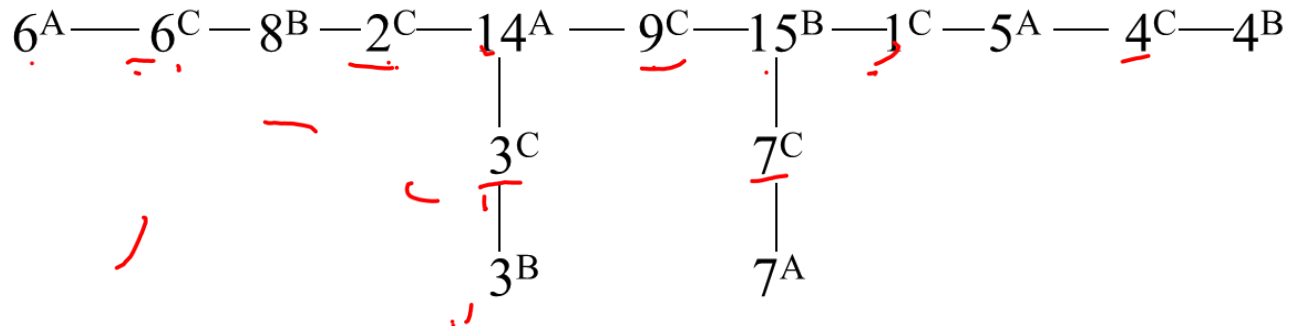
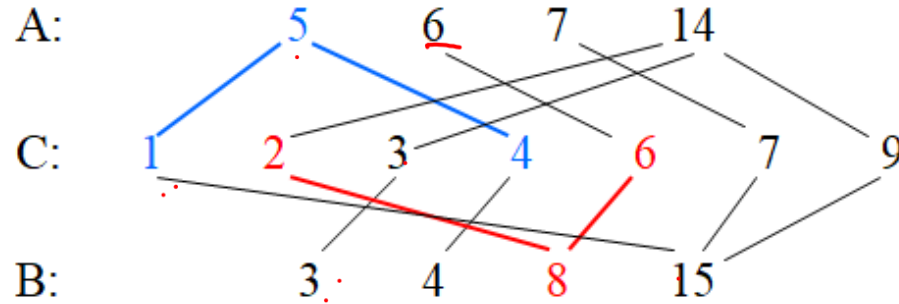
Given  $A$ ,  $B$ ,  $AB_i$  and  $BA_j$  for all  $i, j$ , construct an undirected graph that connects each element of  $A$  and  $B$  to its corresponding  $AB/BA$ . Note that all elements in  $C$  will be covered



# Enhanced Double Digestion Problem

Create a spanning tree:

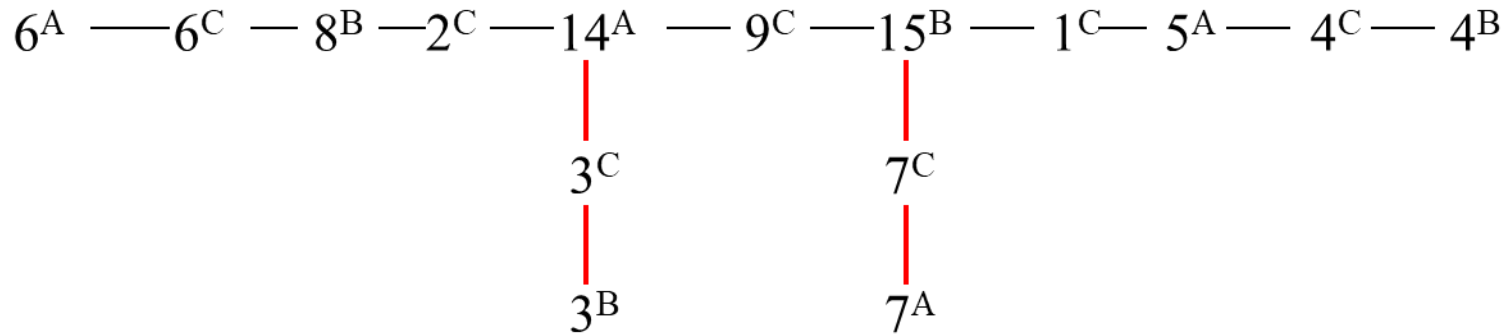
Starting at random node, follow all paths from the node without repeating the edges.



# Enhanced Double Digestion Problem

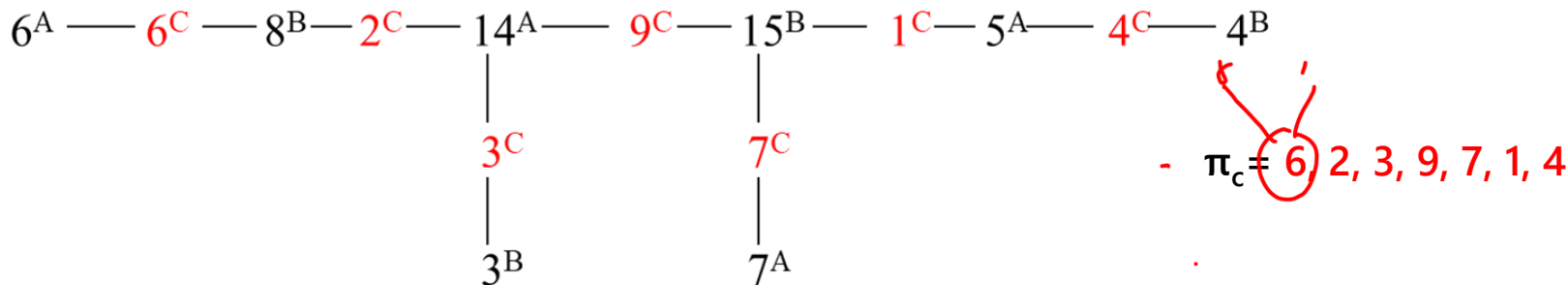
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- The graph (G) will always be connected, and every node in A and B will only be adjacent to nodes from C. Each node from C connects to only one node each from A and B.
- If the problem can be solved: G will be a spanning tree, and any subtree that “hangs” on the longest path will be a 2-node length path (dangler).



# Enhanced Double Digestion Problem

- If the graph  $G$  is not a spanning tree, and not all subtrees hanged off the longest path are dangles, then there is no valid permutation.
- Perform Dangler-first search on the graph  $G$ ...
- Traverse  $G$  starting at one end of a path  $S$  with the largest number of edges, reading only the nodes from  $C$ . Whenever reaching a node with degree greater than 2 (must have a dangle), read the nodes in  $C$  from the hanging dangles first, then continue to traverse  $S$ . This sequence is  $\pi_C$ .

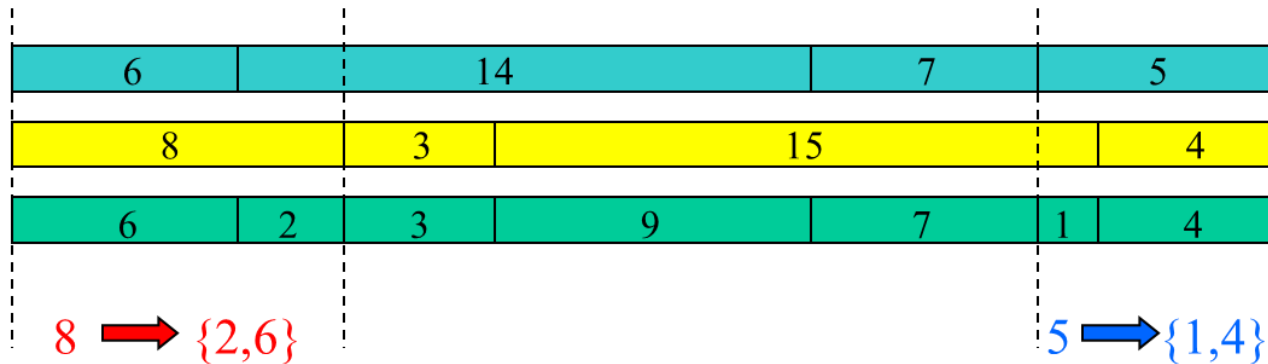




# Enhanced Double Digestion Problem

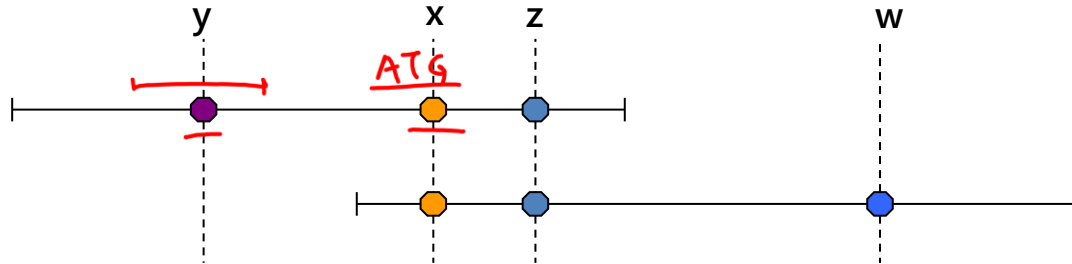
The elements in each  $AB_i$  form a consecutive subsequence in  $\pi_c$ . Likewise, the elements in each  $BA_j$  also form a consecutive subsequence in  $\pi_c$ .

This permutation is a valid permutation... meaning: we have the answer!



# Hybridization Mapping

- Check whether specific small sequences (called probes) bind (hybridize) to fragments (clones)
- The fingerprint is the subset of probes that successfully hybridize to the clone.
- If some portion of one clone's fingerprint matches another, they are likely to be from overlapping regions of the target.
- Probes x, y, z, bound to clone A; x, w and z bound to clone B... overlap in x and z.



# Hybridization mapping model

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- Consecutive Ones Property Model (C1P)
- This can be solved in linear time!
- Assumptions:
  - The probes are unique.
  - There are no errors.
  - All of the correspondences of clones and probes have been found.

# Hybridization mapping model

- Build a matrix ( $n \times m$ ),  $n$  = number of clones,  $m$  = number of probes.
- Entry  $i,j$  is a binary code for whether probe  $j$  hybridized to clone  $i$ .

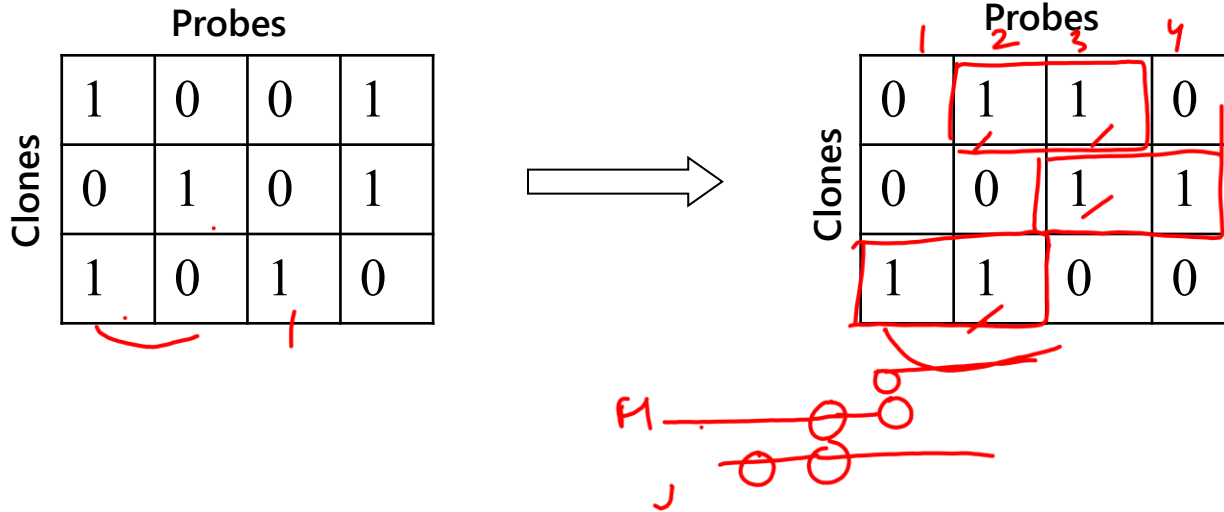
	Probes →			
	1	2	3	4
F1	1	1	0	1
F2	0	1	0	1
F3	1	0	1	1

4 markers

Here, probe 1 hybridized to clone 1, probe 2 hybridized to clone 1,  
probe 1 hybridized to clone 3, probe 4 hybridized to clone 3.

# Hybridization mapping model

- Find a permutation of the columns (probes) such that all the 1s in each row (clone) are consecutive.



# Hybridization mapping model

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- This algorithm can be run in linear time!
- Unfortunately, the assumption that there are no errors isn't useful because biology isn't a mathematical model. Probes may not bind; DNA may be replicated incorrectly.