

Alaska rex sole assessment

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1 Draft conclusions and recommendations

- The assessment of Alaska rex sole was conducted in March 2025 using the Stock Synthesis (SS) model (Methot & Wetzel 2013). This document is a comparison between the proposed “BaseCase” by the Assessment Team and alternative model configurations. A model that includes time varying selectivity and natural mortality might constitute a candidate alternative configuration to reduce misfit of length compositions and survey indices present in the current proposed baseline
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2 BaseCase Model Development

- The assessment model is a two-area yearly model where the population is comprised of 20+ age-classes with two sexes (males and females are considered as separated). The model is a length-based model where the numbers at length in the fisheries and survey data are converted into ages using the von Bertalanffy growth function. The model started in 1982 and it includes 1 commercial fisheries and 2 surveys.

- This document presents a comparison between the model configurations tested by the assessment Team and additional configurations built to improve the fitting of the LFDs and survey indices.
- SSB is above the target reference point and fishing pressure was estimated to be below the target reference point for all model configurations tested.

2.1 Data revisions

- Revision of survey data
- Revision of composition data

2.2 Model setting

- Maximum age in the population set to 20 years (population plus group)
- Growth parameters separated for females and males using the von Bertalanffy function
- CV young and old individuals is estimated
- Some of the growth parameters by areas and sex are estimated
- Estimating selectivity for the commercial fleet and surveys
- Early recruitment era start in 1965
- Recruitment apportioning between areas is constant over time

2.3 Benchmark alternative model configuration

Model Directory

- Reference run (BaseCase; run7_sampler_survbio_edits_new_data)
- BaseCase_TVsel_fleet: As Reference run but assuming time varying selectivity for the fishery
- BaseCase_TVM : As Reference run but assuming time varying natural mortality
- BaseCase_TVsel_fleet_TVM : As Reference run, but assuming time varying natural mortality and time varying selectivity for the fishery
- run9_early_maturity : As Reference run, but assuming earlier maturity
- run10_late_maturity : As Reference run, but assuming later maturity
- run11_est_male_M : As Reference run, but estimating natural mortality for males
- BaseCase_TVsel_fleet_TVM_h072 : As Reference run, but assuming time varying natural mortality and time varying selectivity for the fishery and steepness 0.72 (from Fishlife)

2.4 Work flow

- Run first the file Ensemble_grid.Rmd (as an R file, the Rmd is too large to compile), which creates input files for this files (Basecase model Alaska rex sole.Rmd). The Ensemble_grid.Rmd is used as guidance for selecting the BestCase using diagnostic as in Carvahlo et al., 2021 (BaseCase_TVsel_fleet_TVM in this case), and prepare objects for this file, where the BaseCase and the BestCase are compared.

Load R packages

```
library(r4ss)
library(ss3diags)
library(ggplot2)
library(ggpubr)
```

```

library(FLCore)
library(ggplotFL)
library(FLSRTMB)
library(FLRef)
library(ss3om)
library(png)
library(parallel)
library(doParallel)
cl <- makeCluster(10, type = "PSOCK")
registerDoParallel(cl)

```

Some system specific setups (Linux vs Windows)

```

if (Sys.info()["sysname"] == "Windows") {

  ss.exe <- "~/Max/Stock_synthesis/ss3_3.23.1/ss3.exe"
  main.dir <- "~/Max/Commitees/CIE/Alaska rex sole/Ensemble"
  MC.CORES <- 3

} else {

  SS_EXE <- "~/Max/Executives_SS/ss_linux"
  main.dir <- "~/Max/WKBENCH 2023/Central Baltic herring/Ensemble"
}

```

Load BestCase (produced by Ensemble_grid_example.R)

```

# 2025 BestCase
load("rdata_runs/Alaska rex sole_basecase_TVsel_fleet_TVM.rdata",
     verbose = F)
ref = ss3rep

load("rdata_runs/Alaska rex sole_basecase.rdata", verbose = F)
base = ss3rep

# load retrospective runs
load("rdata_retros/RetroModels_basecase_TVsel_fleet_TVM.rdata",
     verbose = F)

# load steepness profile (to be run only for the BestCase
# outside here; see last part of the script)
load("BaseCase_TVsel_fleet_TVM/profile/profile_h.ref.rdata",
     verbose = F)

```

2.5 Fishery Data

- Catch data were available for the commercial fleets.
-

```

mod = ref
df.catch = mod$catch
df.catch$Season = factor(df.catch$Seas)

pc1 = ggplot(df.catch, aes(Yr, Obs, fill = Fleet_Name)) + theme_bw() +
  geom_bar(position = "stack", stat = "identity") + ylab("Catch") +

```

```

xlab("Year") + scale_x_continuous(expand = c(0.01, 0)) +
scale_y_continuous(expand = c(0.01, 0)) + scale_fill_manual(values = sscol(length(unique(df.catch$F

```

pc1

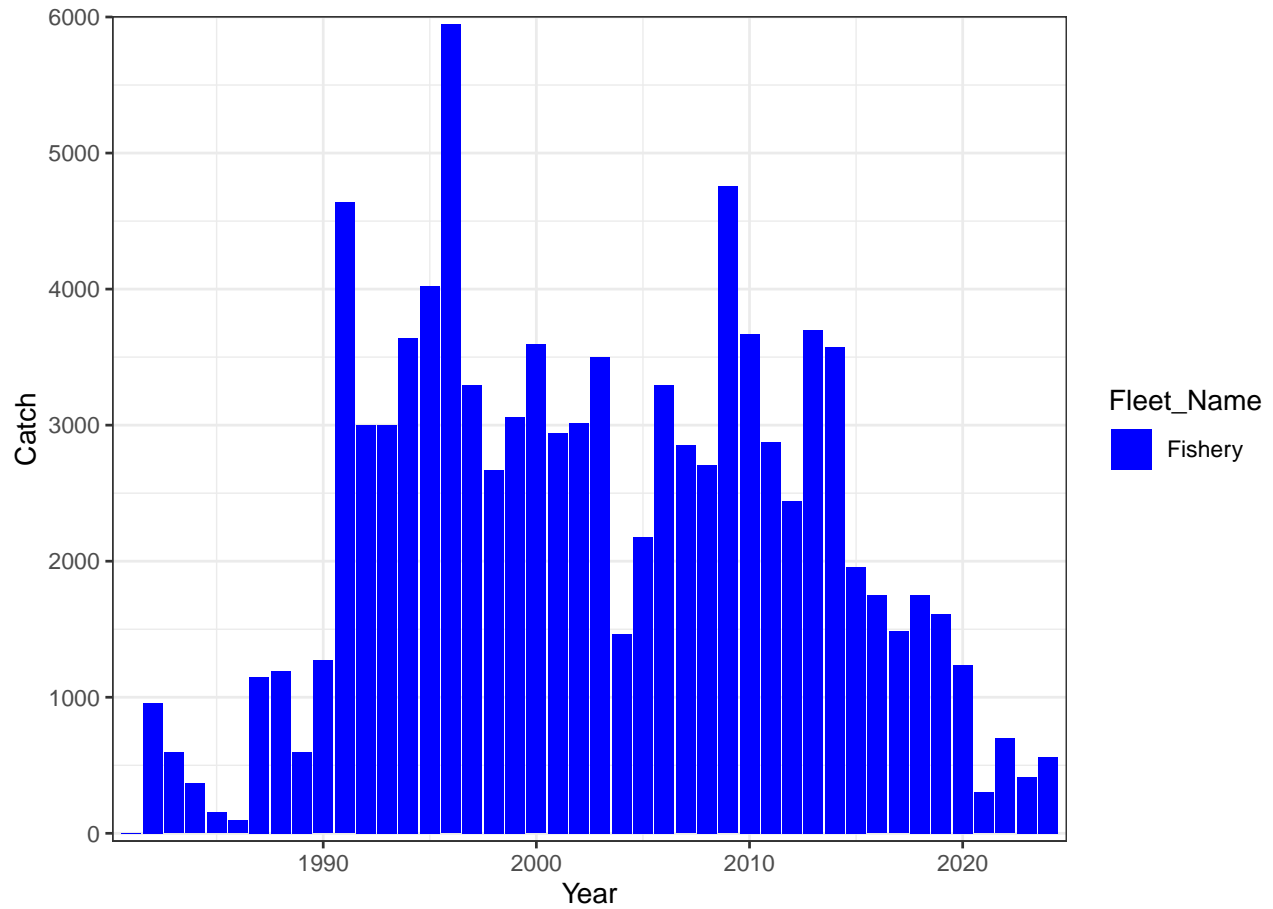


Figure 1: Time series of annual total catches for Alaska rex sole

•
•

```
SSplotData(ref, subplots = 2)
```

2.6 Survey

- Two survey indices were include in the model.

•

```
knitr::include_graphics("basecase_TVsel_fleet_TVM/plots/index2_cpuefit_EasternSurvey.png")
```

```
knitr::include_graphics("basecase_TVsel_fleet_TVM/plots/index2_cpuefit_NonEasternSurvey.png")
```

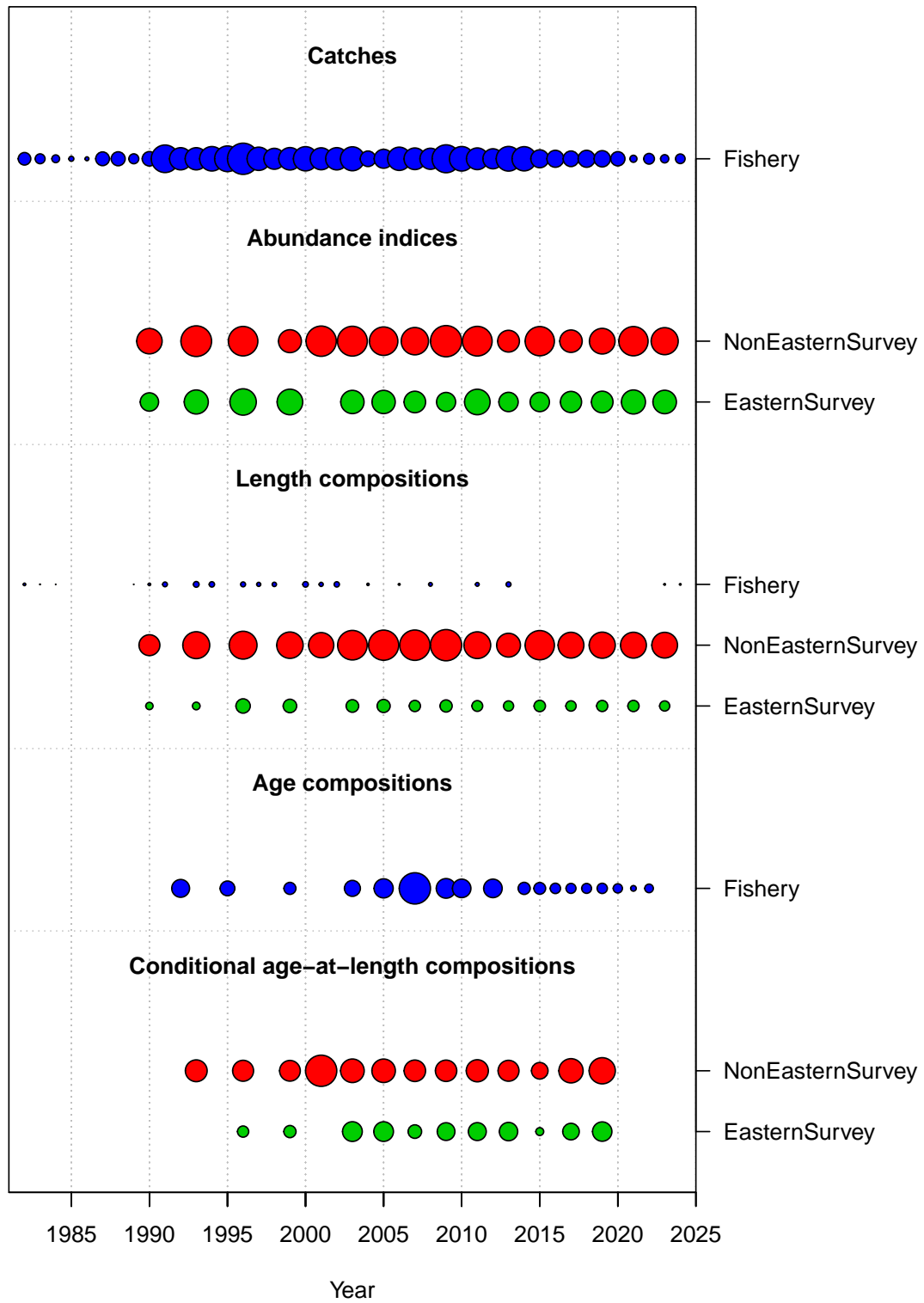


Figure 2: Illustration of time series data for catches, LFDs age compositions and the survey indeces that were adopted as input for the Stock Synthesis model

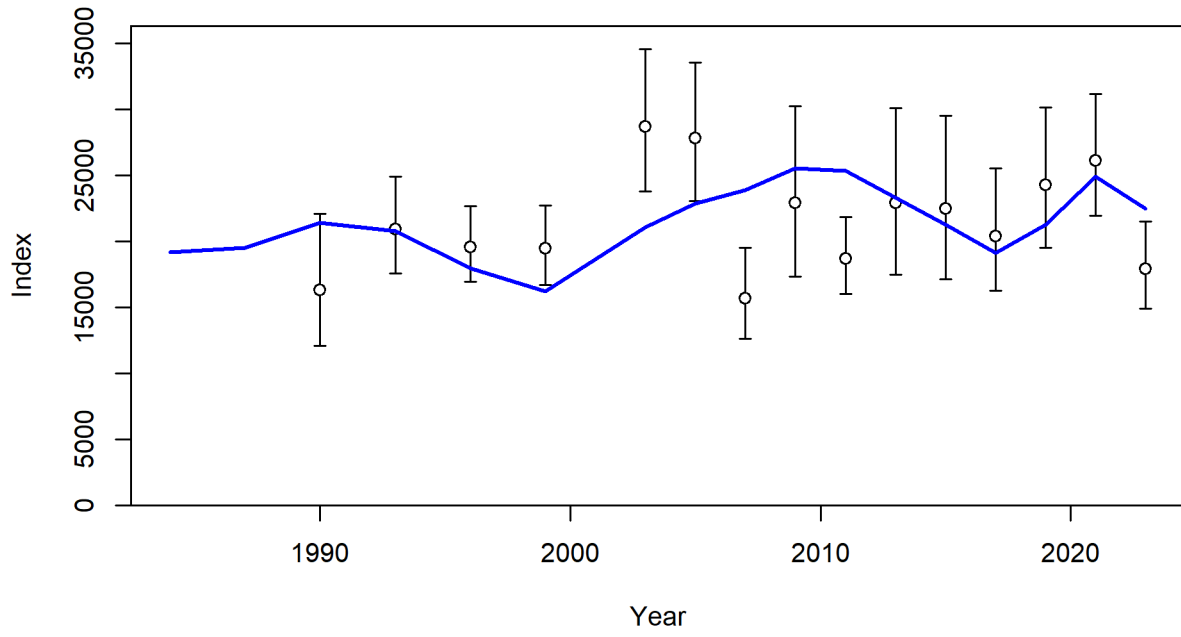


Figure 3: EasternSurvey indices fit

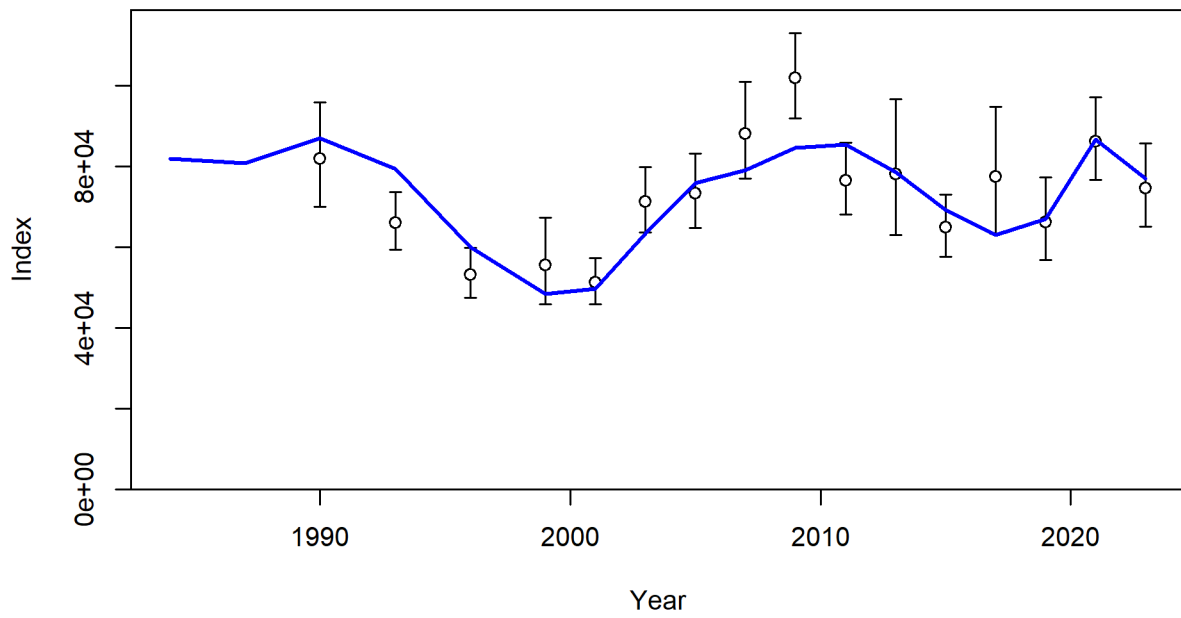


Figure 4: NonEasternSurvey indices fit

2.7 Model specifications

- The reference model for Alaska rex sole is an annual, sex-structured length-age based Stock Synthesis model. The underlying age-structured dynamics is set up to comprise ages 0-20, where age 20 was treated as a plus group. The population was modeled as sex-structured with sex-specific parametrisation for somatic growth. Stock fecundity was assumed to be proportional to female spawning stock biomass.
- For the reference model, the sex specific growth is modeled for both sexes using Von Bertalanffy model. L_{atAmin} and L_{atAmax} were estimated. The CVs for L_{atAmin} and L_{atAmax} were estimated.

```
sspar(mfrow = c(1, 1), plot.cex = 0.7)
SSplotBiology(mod, subplots = 1, main = F)
```

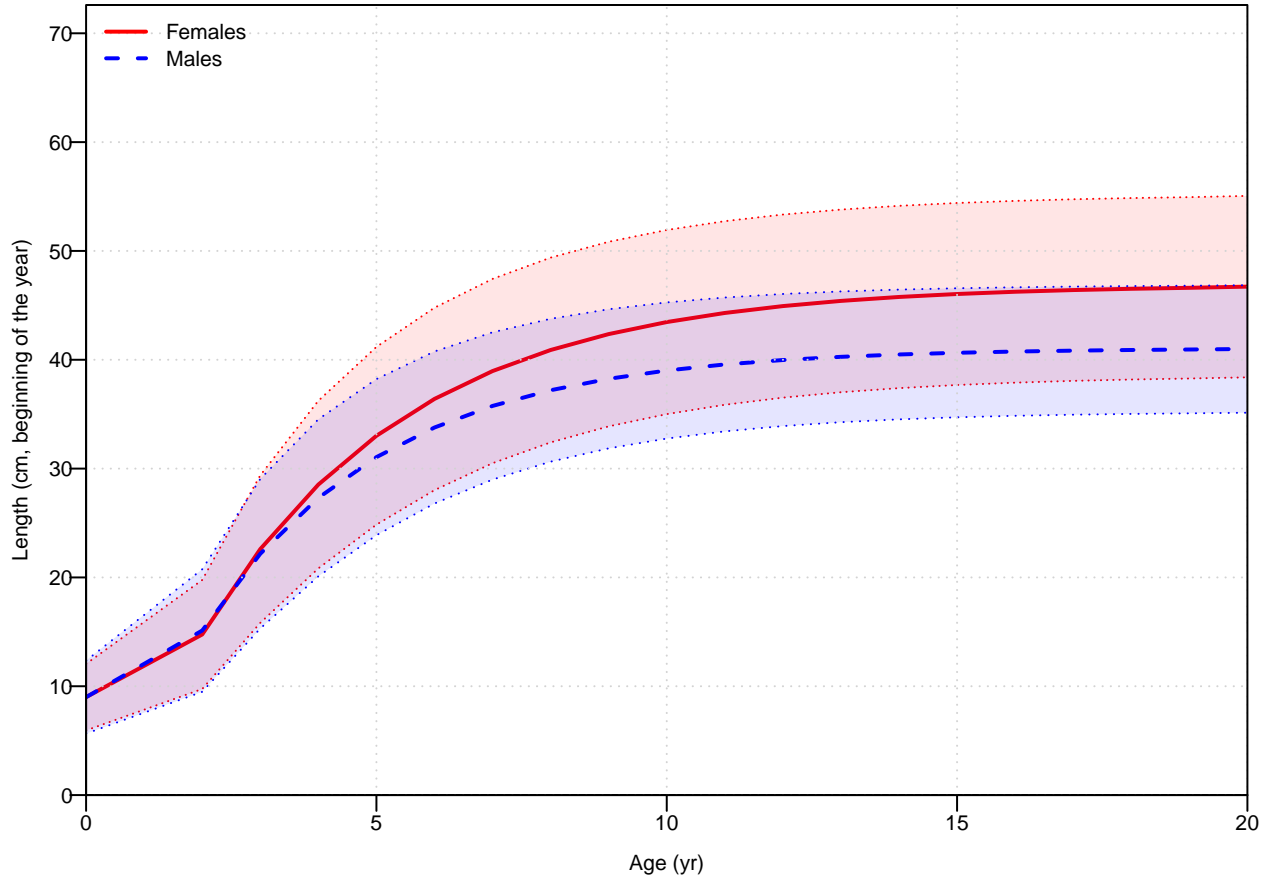


Figure 5: Growth functions for Female and Males.

- Female maturity was assumed to have the form of a logistic ogive with an age at 50% maturity (A_{m50}) being attained at 5.7 years and a slope of -1.11 1/cm (Figure 7).

```
sspar(mfrow = c(1, 1), plot.cex = 0.7)
SSplotBiology(mod, subplots = 6, main = F)
```

- Sex-specific natural mortality at age (M_a) were inputted as a single value (0.17)

```
knitr::include_graphics("basecase_TVsel_fleet_TVM/plots/bio24_time-varying_NatM_uniform_Fem_GP_1.png")
```

- Nominal spawning and settling time were set to January 1st. The expected mean recruitment was assumed to follow a Beverton and Holt stock recruitment relationship. For the BaseCase a steepness of $h = 1.0$ was assumed. Recruitment deviations were estimated for 1982-2022 as main recruitment

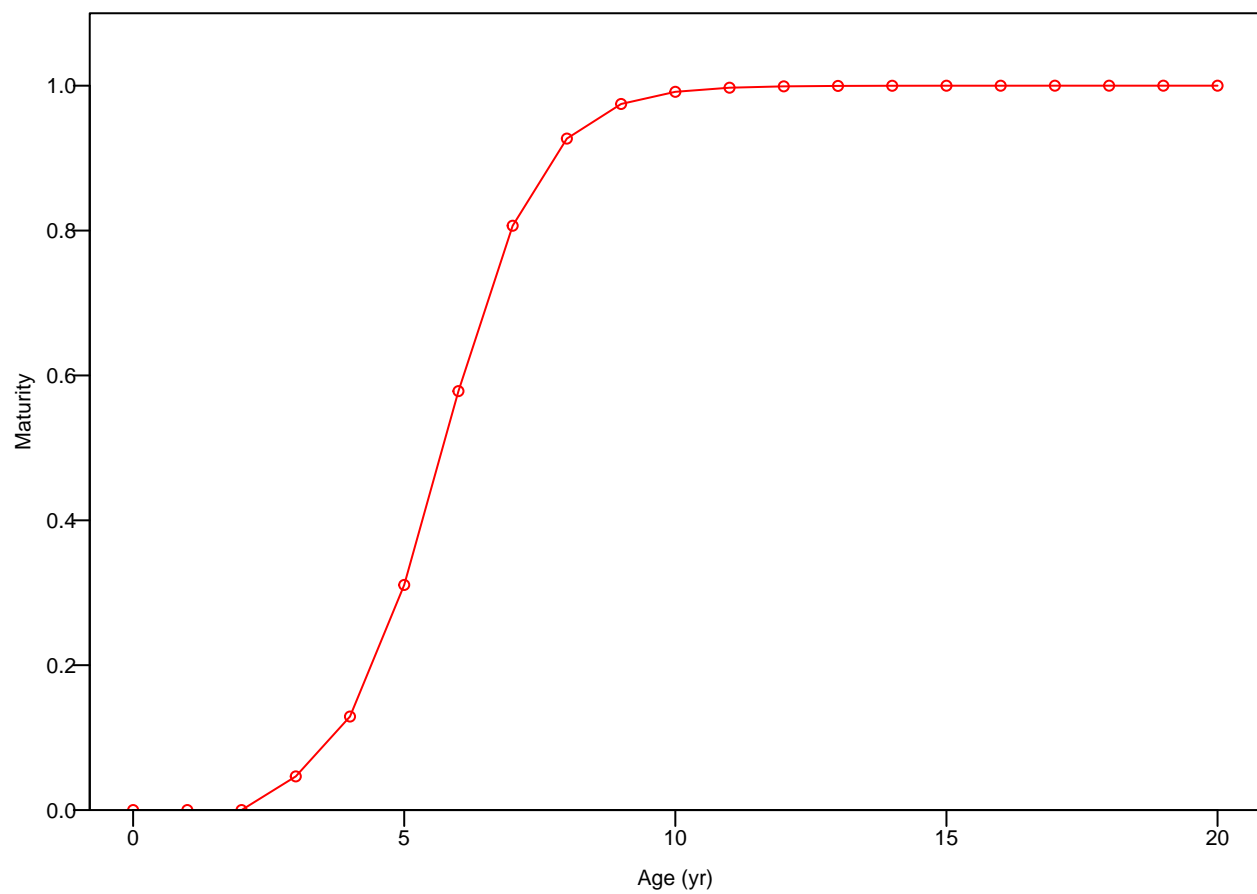


Figure 6: Assumed Maturity ogive for females.

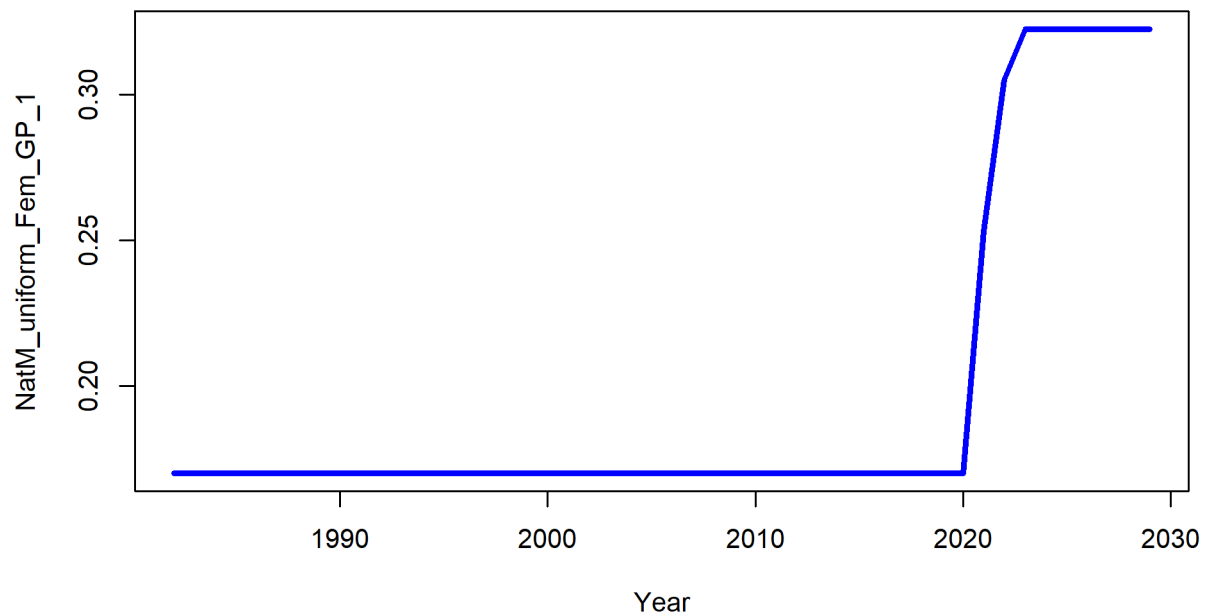


Figure 7: Estimated variation in natural mortality vectors for females of area 1

deviations and for the preceding years 1965-1982 as early recruitment deviations. Recruitment deviations were assumed to have a penalty of 0.6 on the standard deviation (σ_R).

- All fleets assumed a double-normal (dome-shaped) selectivity (option 20).
-
- Fishing mortality was modeled using the hybrid F method (Option 3). In the original model, option three was selected for the fishing mortality (F) report basis. However, for this document Option five was selected for the fishing mortality (F) report basis only for representation and to allow the use of MVLN; this option corresponded to the simple unweighted average of the F of the age classes chosen to represent the Fbar (age 5–10).

2.8 Model Diagnostics

Prepare outputs of retrospective runs (see Supplement for coding)

```
retro.idx = r4ss::SSsummarize(retroModels, verbose = F)
retro.len = ss3diags::SSretroComps(retroModels)
```

2.8.1 Survey indices

The BestCase model fitted all indices moderately well, with runs tests indicating mixed evidence for a systematic residual pattern.

```
sspar(mfrow = c(3, 2), plot.cex = 0.7)
for (i in 1:2) {
  SSplotIndices(ref, subplots = 2, fleets = i + 1)
  r = SSplotRuntest(ref, add = T, verbose = F, indexselect = i)
}
```

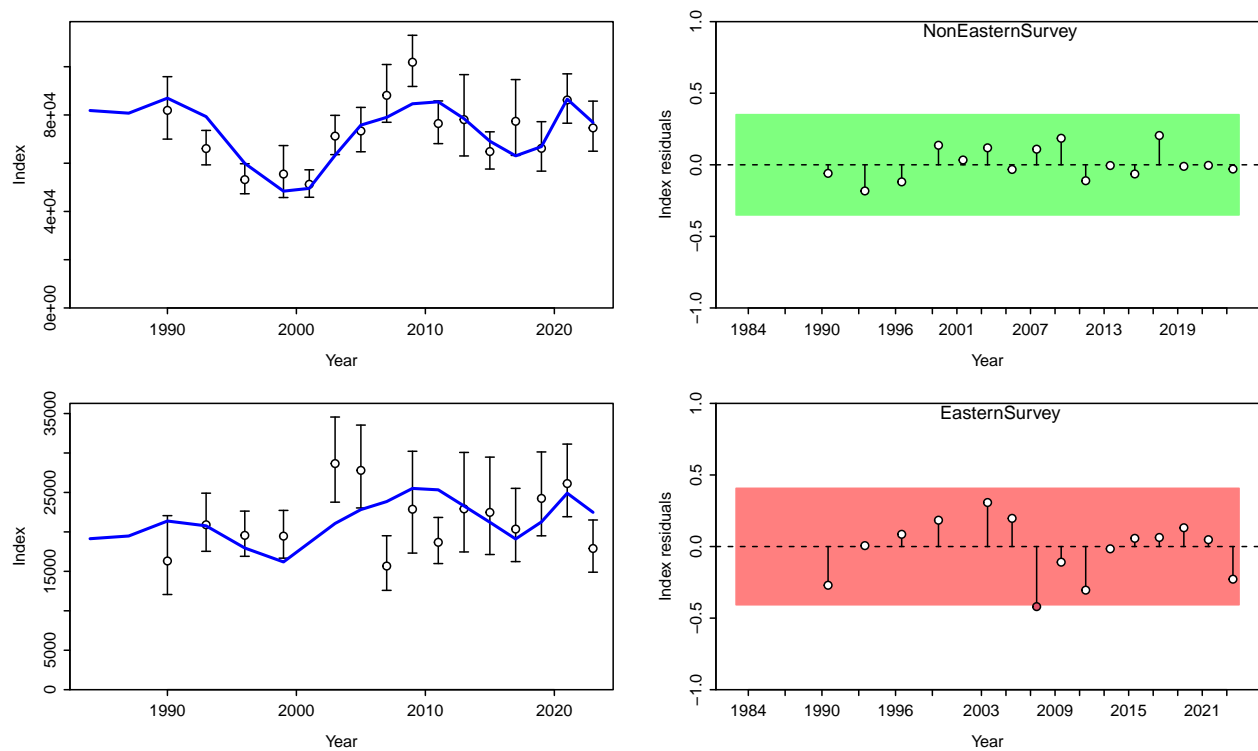


Figure 8: Fit, residual diagnostics and hindcast cross-validations for Survey Index

- Figure 8 check conflict between indices and mean length. As the value are under 30%, no major conflicts were found

```
sspar(mfrow = c(1, 2), plot.cex = 0.8)
SSplotJABBAres(ref, subplots = "cpue", add = T, col = sscol(3)[c(1,
3, 2)])
Plotting JABBA residual plot

RMSE stats by Index:
      indices RMSE.perc nobs
1 NonEasternSurvey      20.4   18
2   EasternSurvey      21.7   17
3      Combined      21.0   35
SSplotJABBAres(ref, subplots = "len", add = T, col = sscol(3)[c(1,
3, 2)])
Plotting JABBA residual plot
```

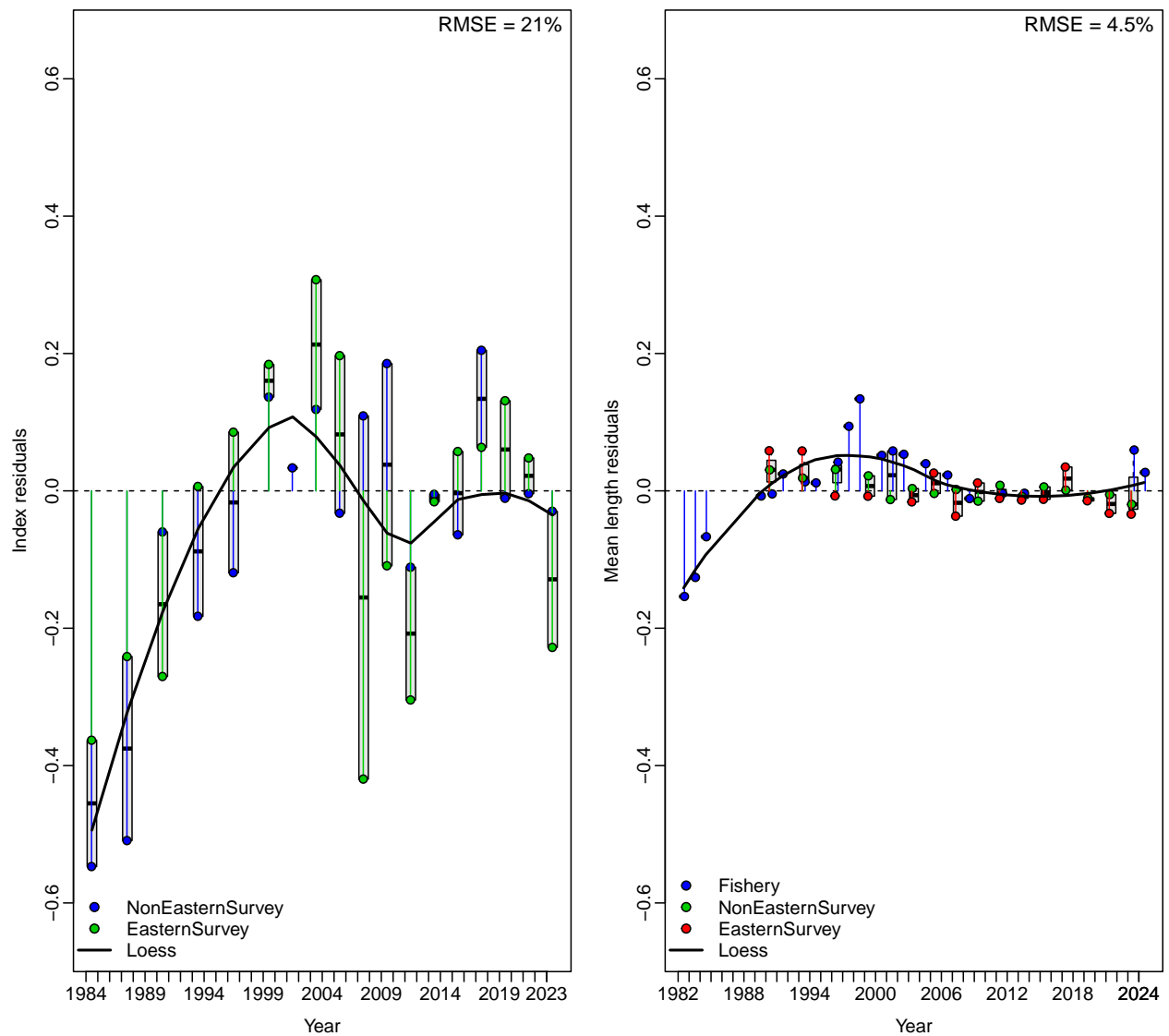


Figure 9: Joint residuals

RMSE stats by Index:

	indices	RMSE.perc	nobs
1	Fishery	6.5	21
2	NonEasternSurvey	1.5	16
3	EasternSurvey	3.0	15
4	Combined	4.5	52

2.8.2 Size composition

- The estimated selectivity curves for the BestCase are shown in Figure 9, with time-varying selectivity patterns for illustrated in Figure 10.
- The fits to the size composition (Figure 11) and conditional age-length-key data (data not shown) appeared overall adequate and only showed evidence for non-random residual patterns in few instances. Sex ratio at length by the current reference case is also satisfying (Figure 14).

```
sspar(mfrow = c(1, 1), plot.cex = 0.8)
SSplotSelex(ref, subplots = 2)
```

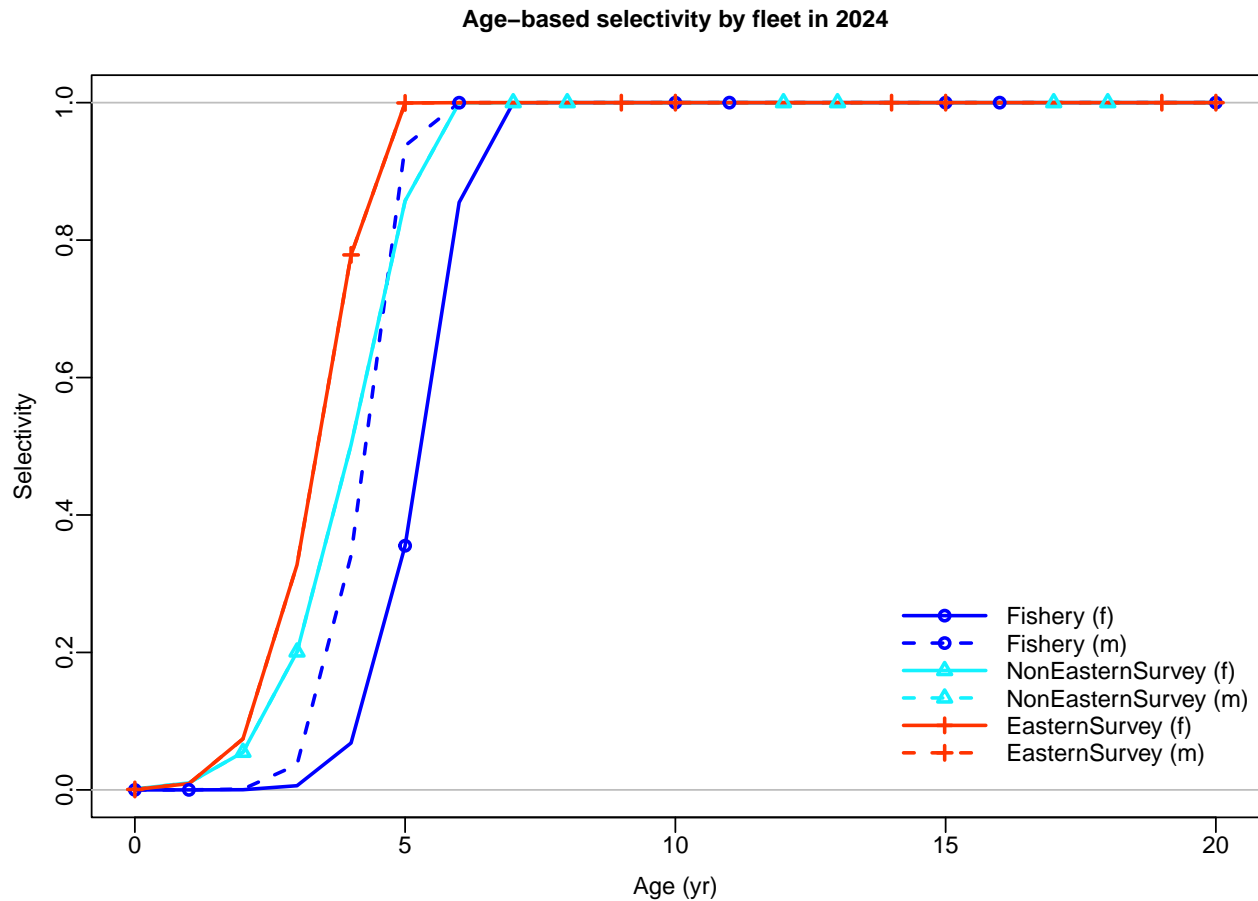


Figure 10: Estimated logistic selectivity curves by fleet

```
sspar(mfrow = c(1, 2), plot.cex = 0.7)
SSplotSelex(ref, subplots = 12)
```

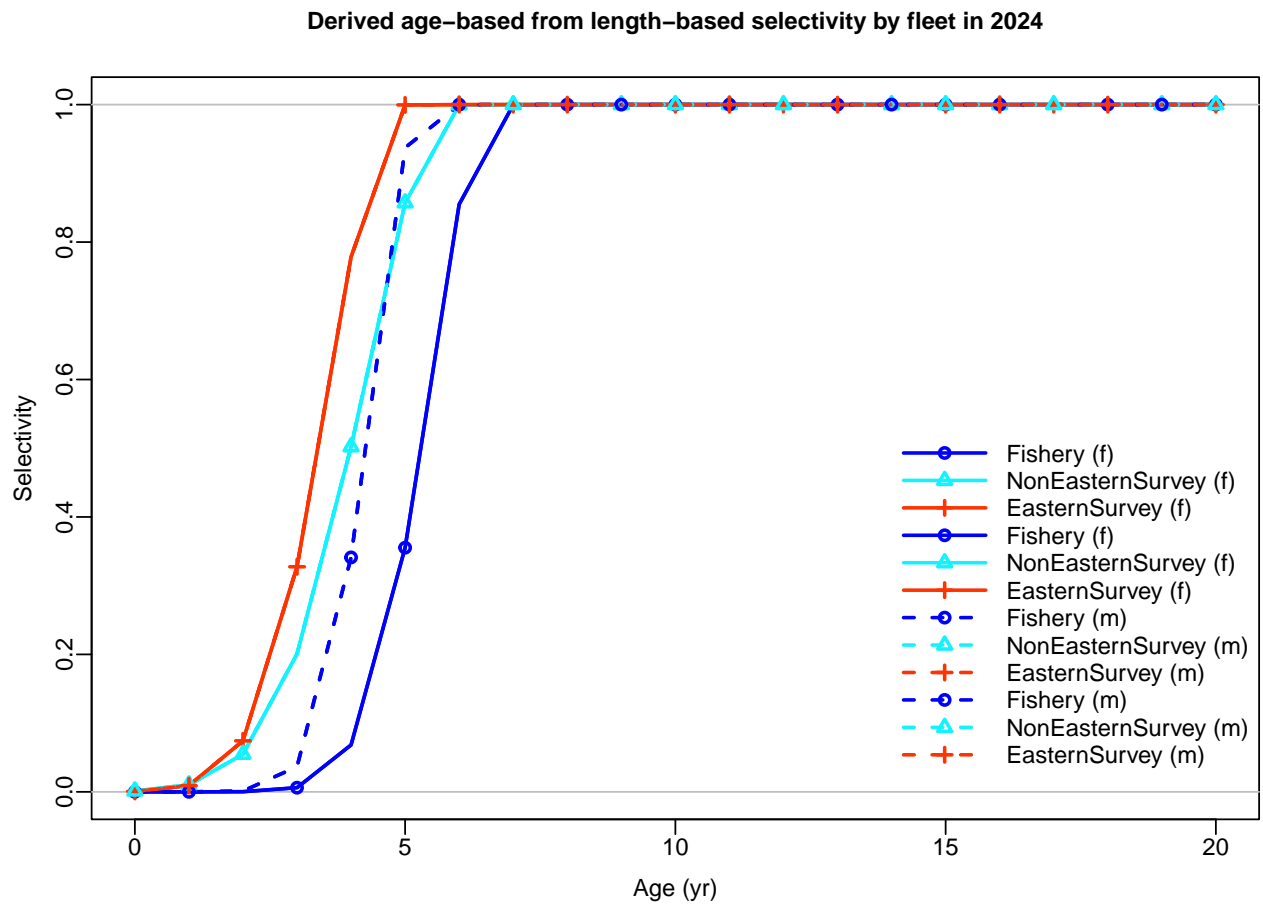


Figure 11: Estimated logistic selectivity curves by fleet

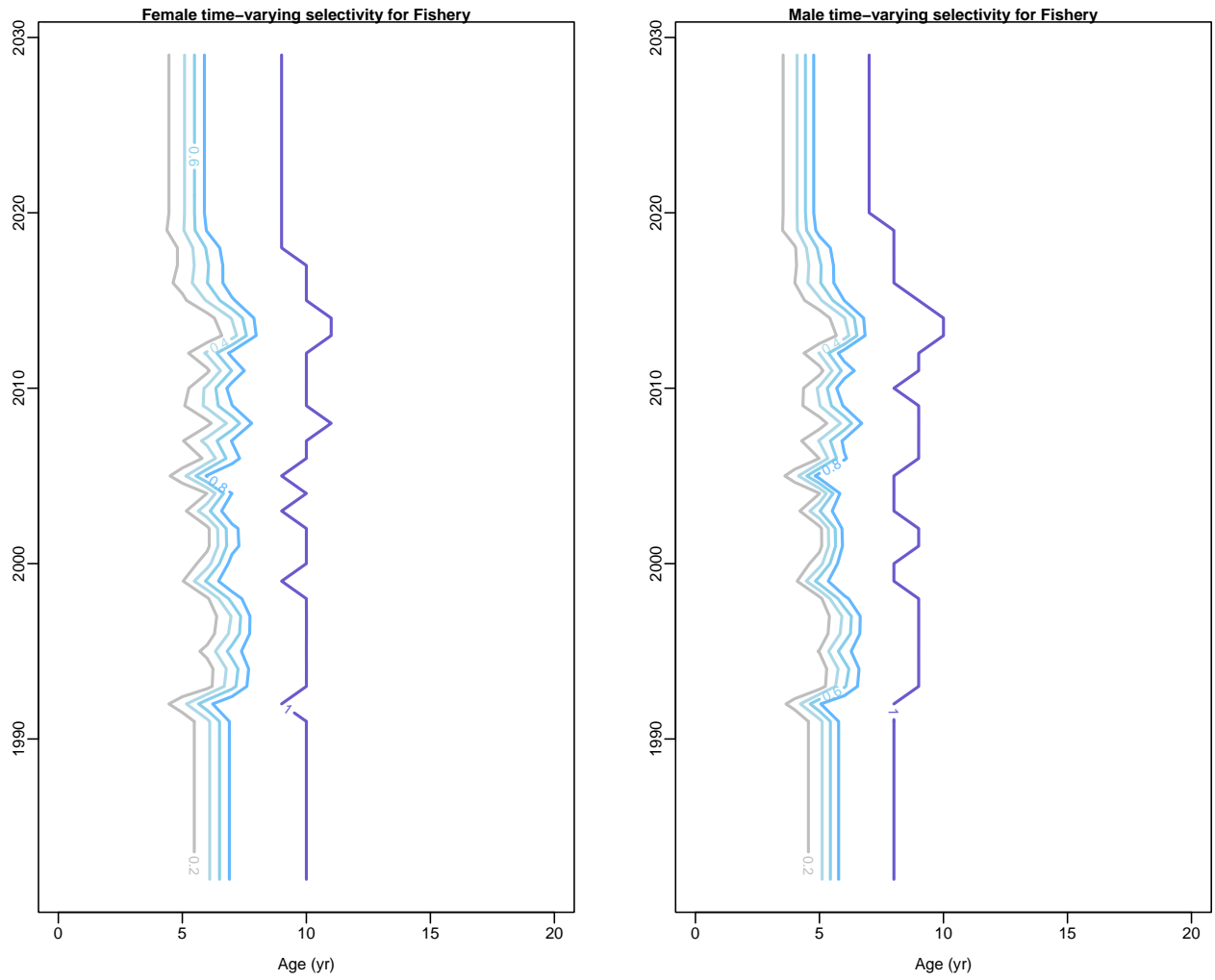


Figure 12: Plot illustrating the estimated time-varying changes in the selectivity curves for the fishery

```
SSplotComps(ref, subplots = 21)
```

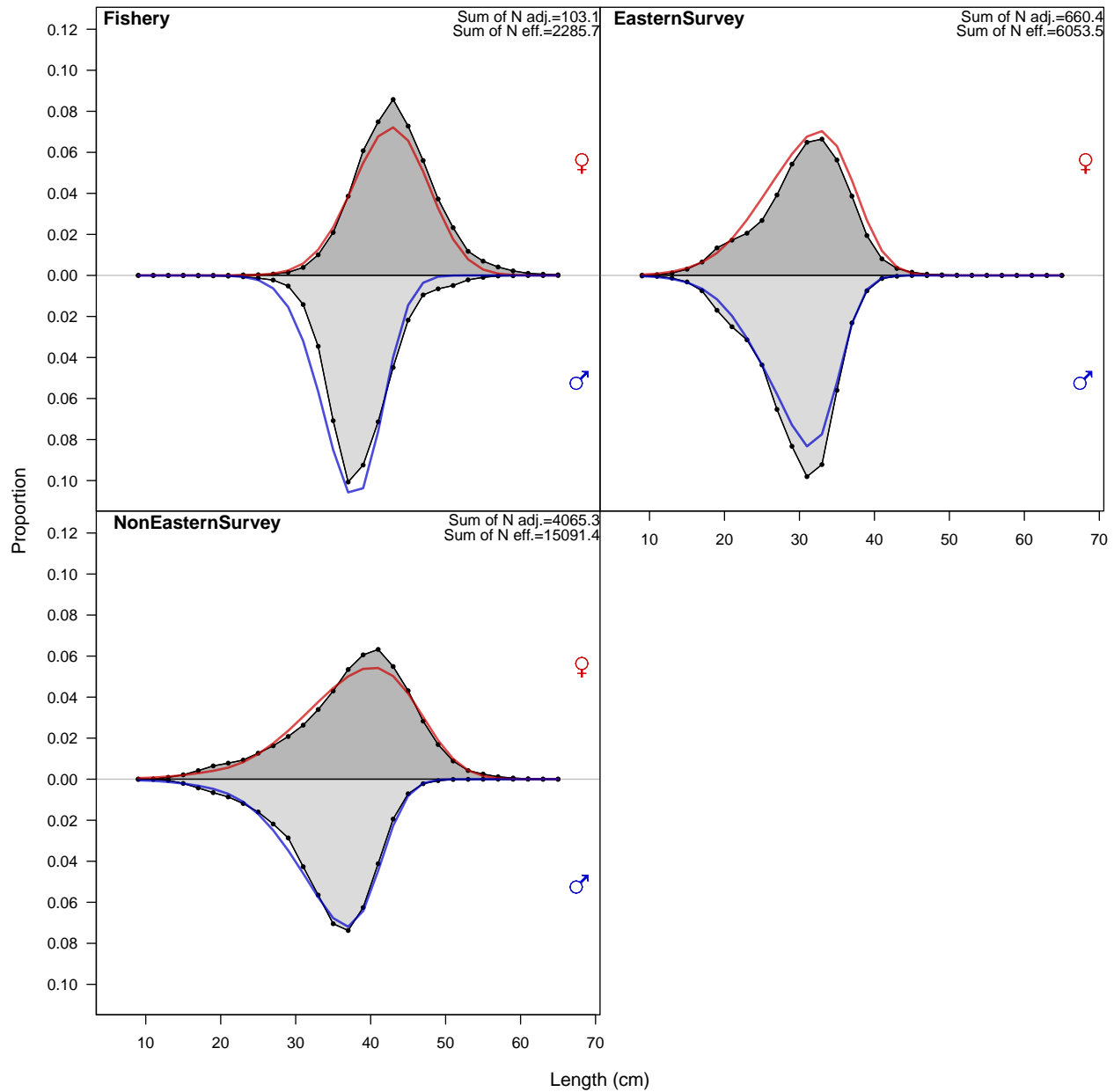


Figure 13: Summary of observed and expected composition data aggregated across years

```
SSplotComps(ref, subplots = 24)
```

```
sspar(mfrow = c(3, 3), plot.cex = 0.5)
for (i in 1:2) {
  r = SSplotRunstest(ref, subplots = "len", add = T, indexselect = i,
    verbose = F)
}
```

```
SSplotSexRatio(ref, kind = "LEN")
```

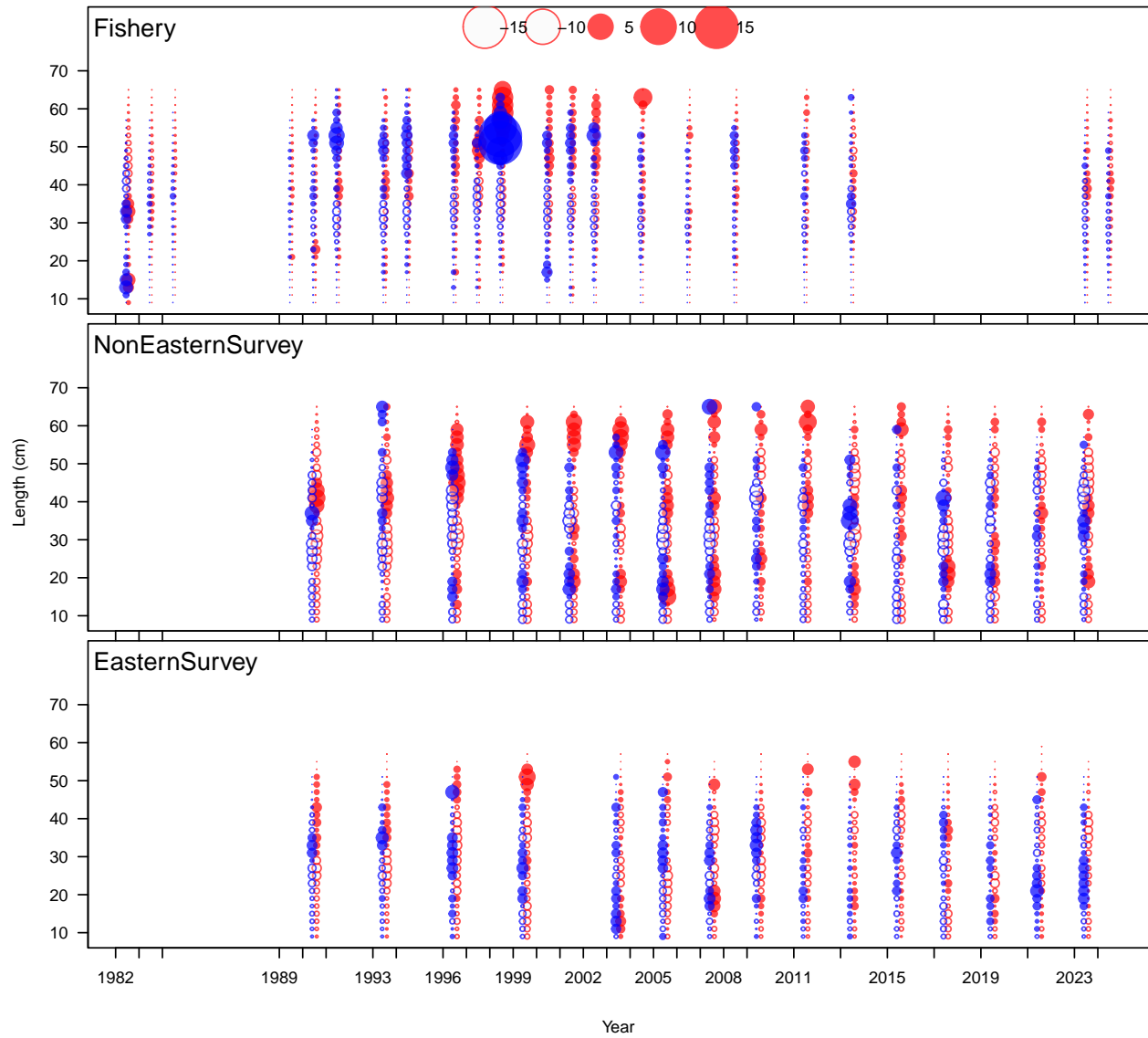


Figure 14: Residuals for mean lengths of size composition data from fishing fleets and surveys

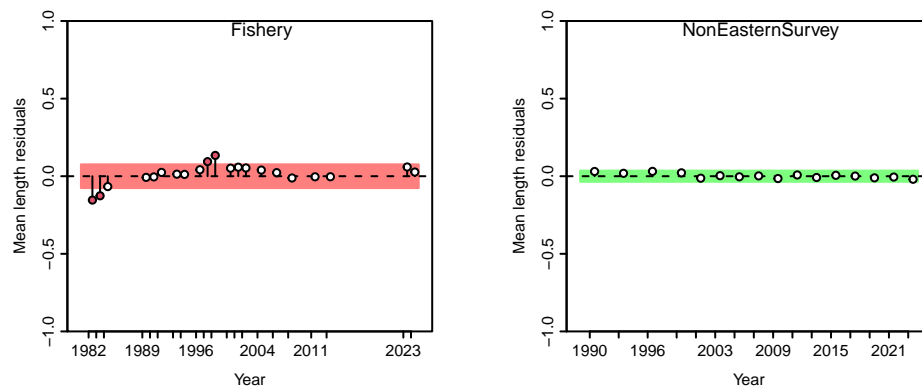


Figure 15: Run test for mean lengths of size composition data from fishing fleets and surveys

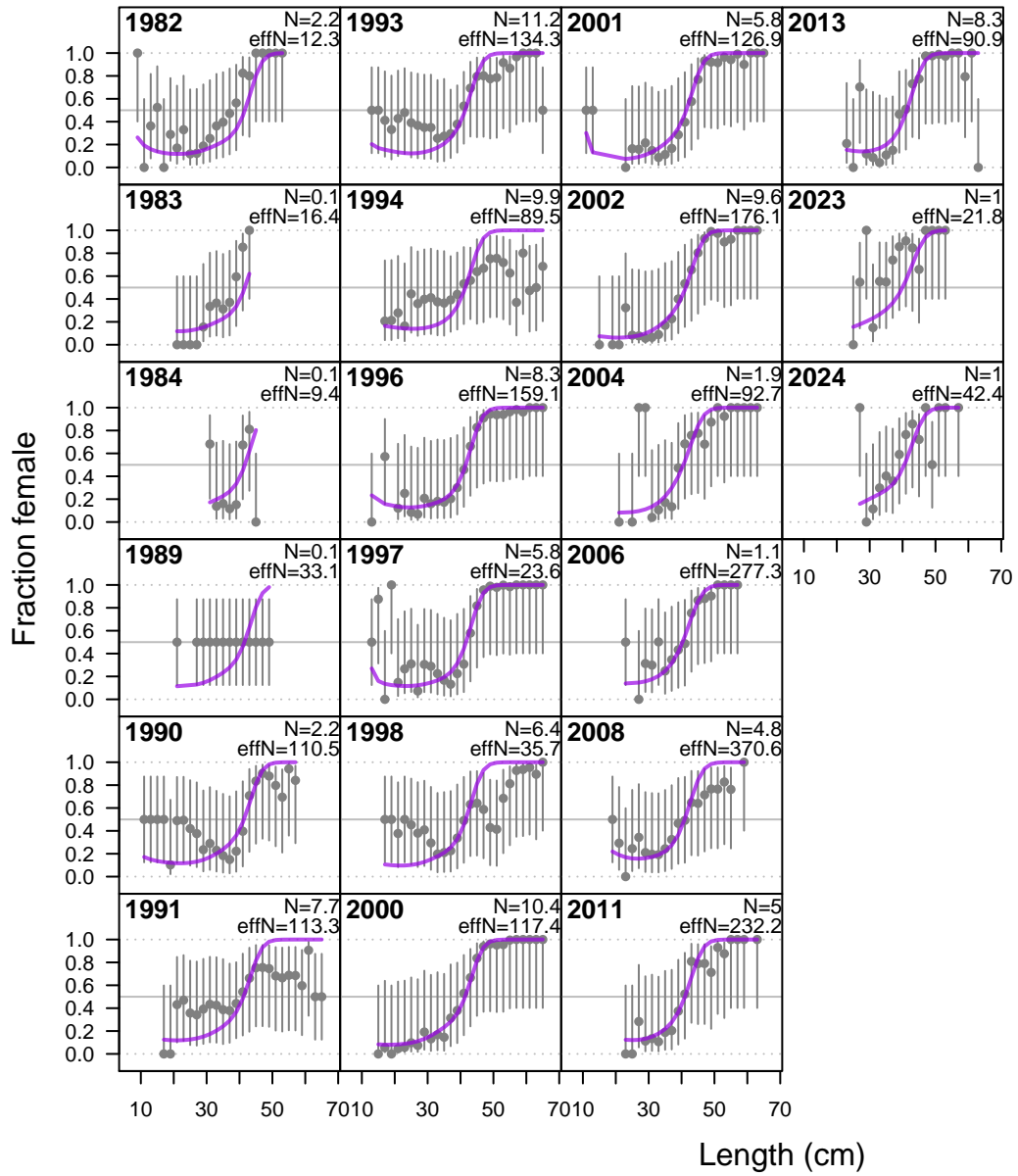


Figure 16: Observed and predicted sex ratio by length

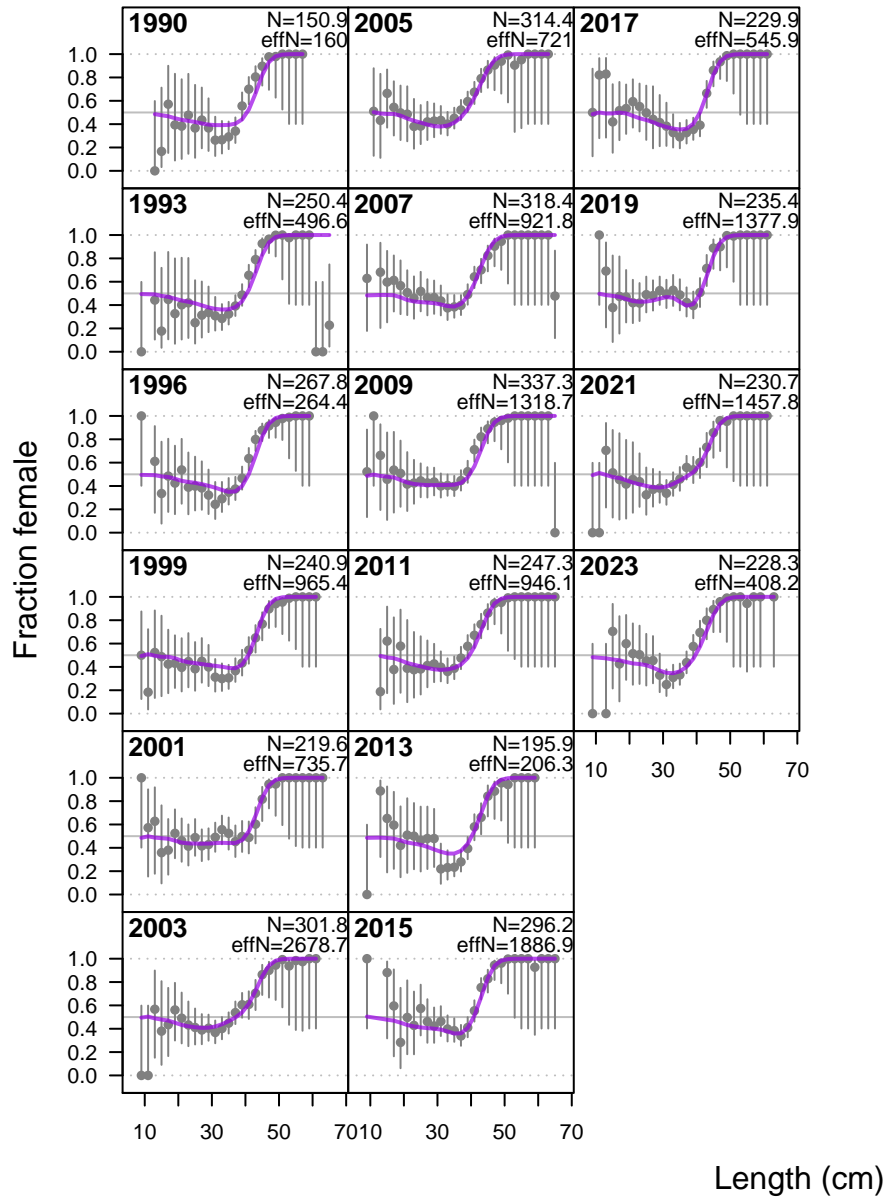


Figure 17: Observed and predicted sex ratio by length

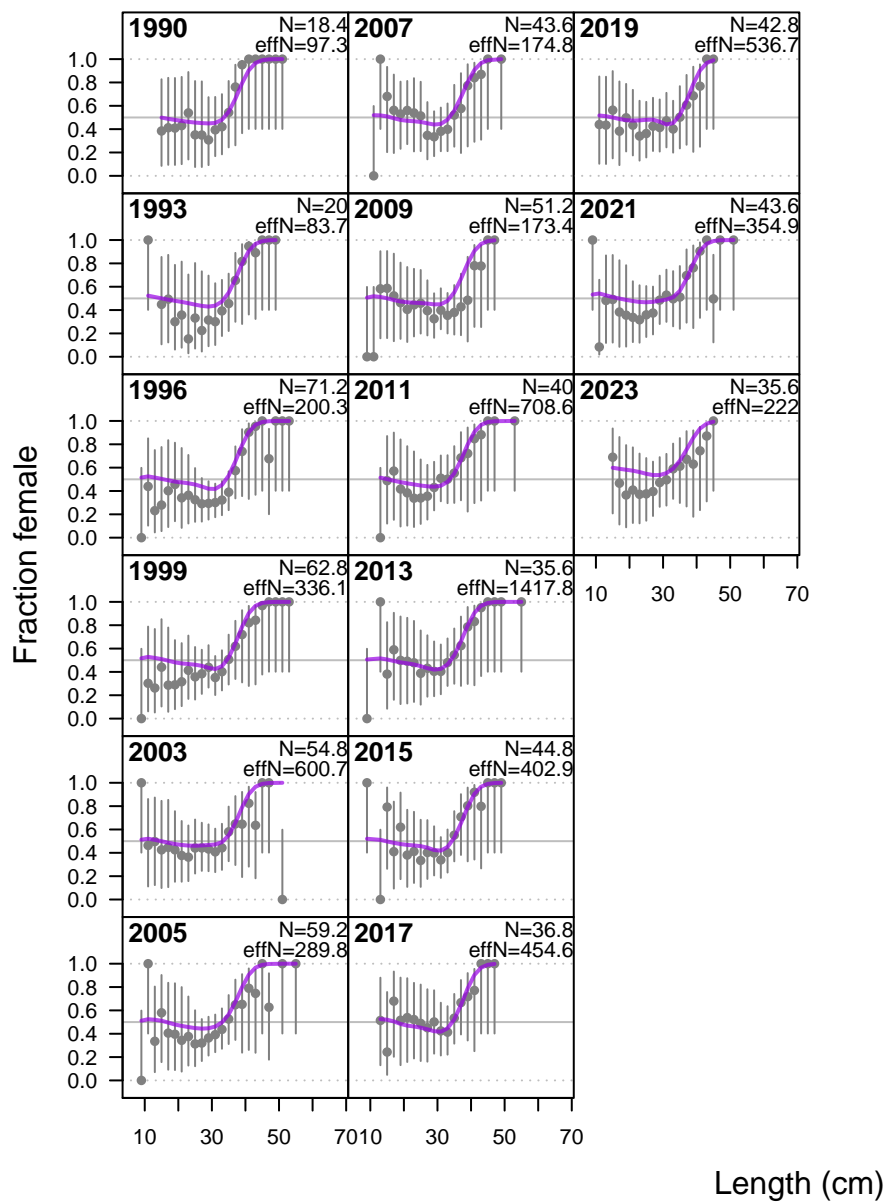


Figure 18: Observed and predicted sex ratio by length

2.8.3 Restropective Analysis

- The retrospective analysis showed a slightly retrospective pattern on SSB from Mohn's Rho of 0.04, while the F bias was -0.05. Forecast bias were similar to retrospective bias. All retrospective peels fell within the 95% confidence intervals of the full model

```
sspar(mfrow = c(2, 2), plot.cex = 0.65)
r = SSplotRetro(retro.idx, add = T, legend = F, forecast = F,
  verbose = F)
r = SSplotRetro(retro.idx, add = T, forecastrho = T, legend = F,
  verbose = F, xlim = c(2005, 2024))
r = SSplotRetro(retro.idx, subplots = "F", add = T, legend = F,
  forecast = F, verbose = F)
r = SSplotRetro(retro.idx, subplots = "F", add = T, forecastrho = T,
  legend = F, verbose = F, xlim = c(2005, 2024))
mtext(c("Retro", "Forecast"), 3, outer = T, line = -0.5, at = c(0.3,
  0.8), cex = 0.8)
```

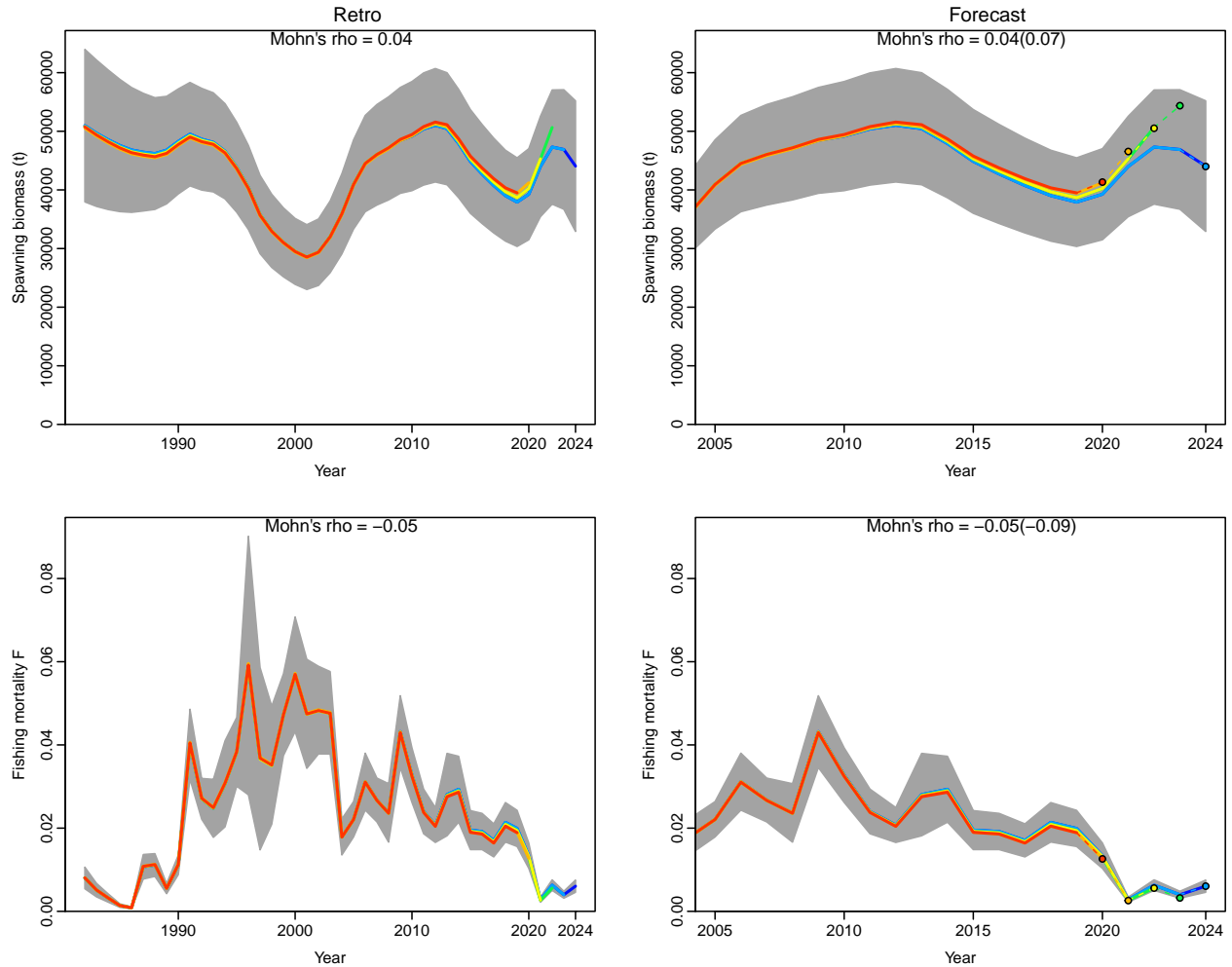


Figure 19: Retrospective analysis and retrospective forecasts for the 2025 BestCase model

2.8.4 Hindcast Cross-Validations

Hindcast cross-validations indicated that the model has moderate prediction skill for the indices. By contrast, the model indicated good prediction skill for mean lengths and age (Figure 16).

```
sspar(mfrow = c(2, 3), plot.cex = 0.5)
SSplotHCxval(retro.len, subplots = "len", add = T, verbose = FALSE)
Warning in qlnorm(0.975, meanlog = log(y), sdlog = indexSEvec): NaNs produced
Warning in qlnorm(0.025, meanlog = log(y), sdlog = indexSEvec): NaNs produced
```

	Index	Season	MASE	MAE.PR	MAE.base	MASE.adj	n.eval
1	Fishery	1	1.7244927	0.04196762	0.02433621	0.4196762	1
2	NonEasternSurvey	1	0.8529637	0.02105541	0.02468501	0.2105541	2
3	EasternSurvey	1	2.2815371	0.04040195	0.01770822	0.4040195	2

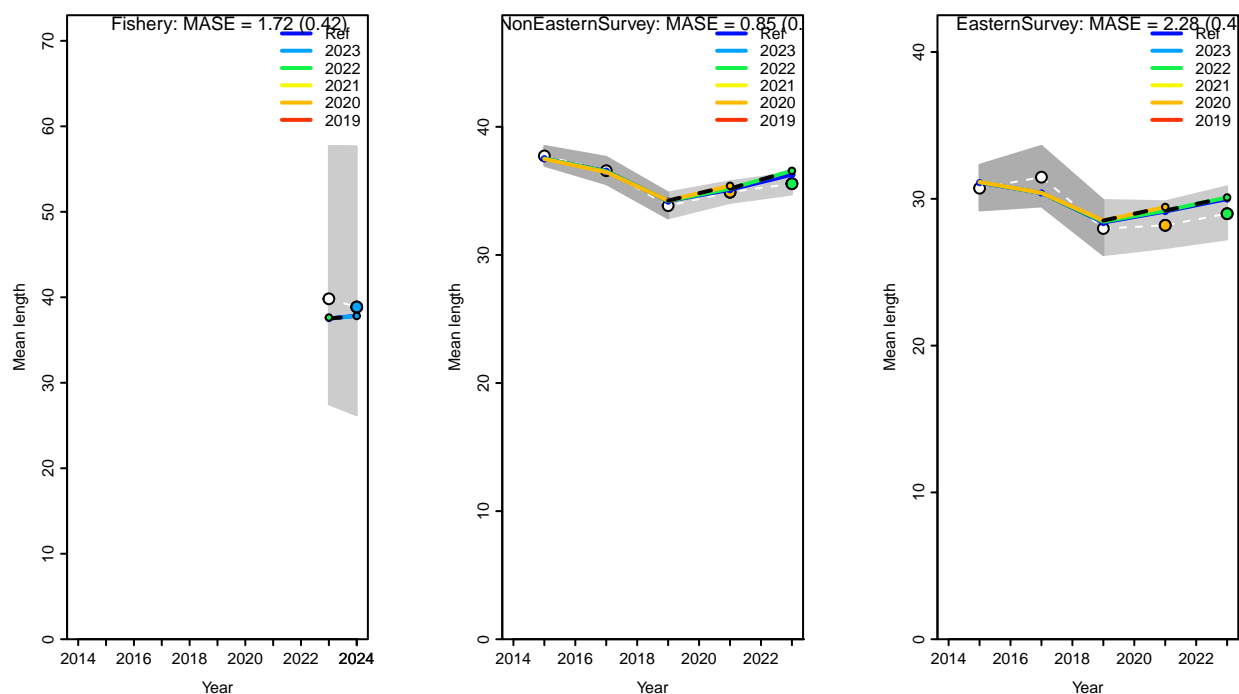


Figure 20: Fit and hindcast cross-validations for the Survey size compositions of the BestCase model

```
sspar(mfrow = c(1, 1), plot.cex = 0.5)
SSplotHCxval(retro.len, subplots = "age", add = T, verbose = FALSE)
```

	Index	Season	MASE	MAE.PR	MAE.base	MASE.adj	n.eval
1	Fishery	1	0.7303455	0.08181016	0.1120157	0.7303455	3
2	Fishery	1	0.7303455	0.08181016	0.1120157	0.7303455	3
3	Fishery	1	0.7303455	0.08181016	0.1120157	0.7303455	3

Hindcast with Cross-Validation of CPUE observations

```
sspar(mfrow = c(1, 2), plot.cex = 0.9)
SSplotHCxval(retroSummary, xmin = 2006, add = T, legendcex = 0.6)
Plotting Hindcast Cross-Validation (one-step-ahead)
```

Computing MASE with only 2 of 5 prediction residuals for Index NonEasternSurvey

Warning: Unequal spacing of naive predictions residuals may influence the interpretation of MASE

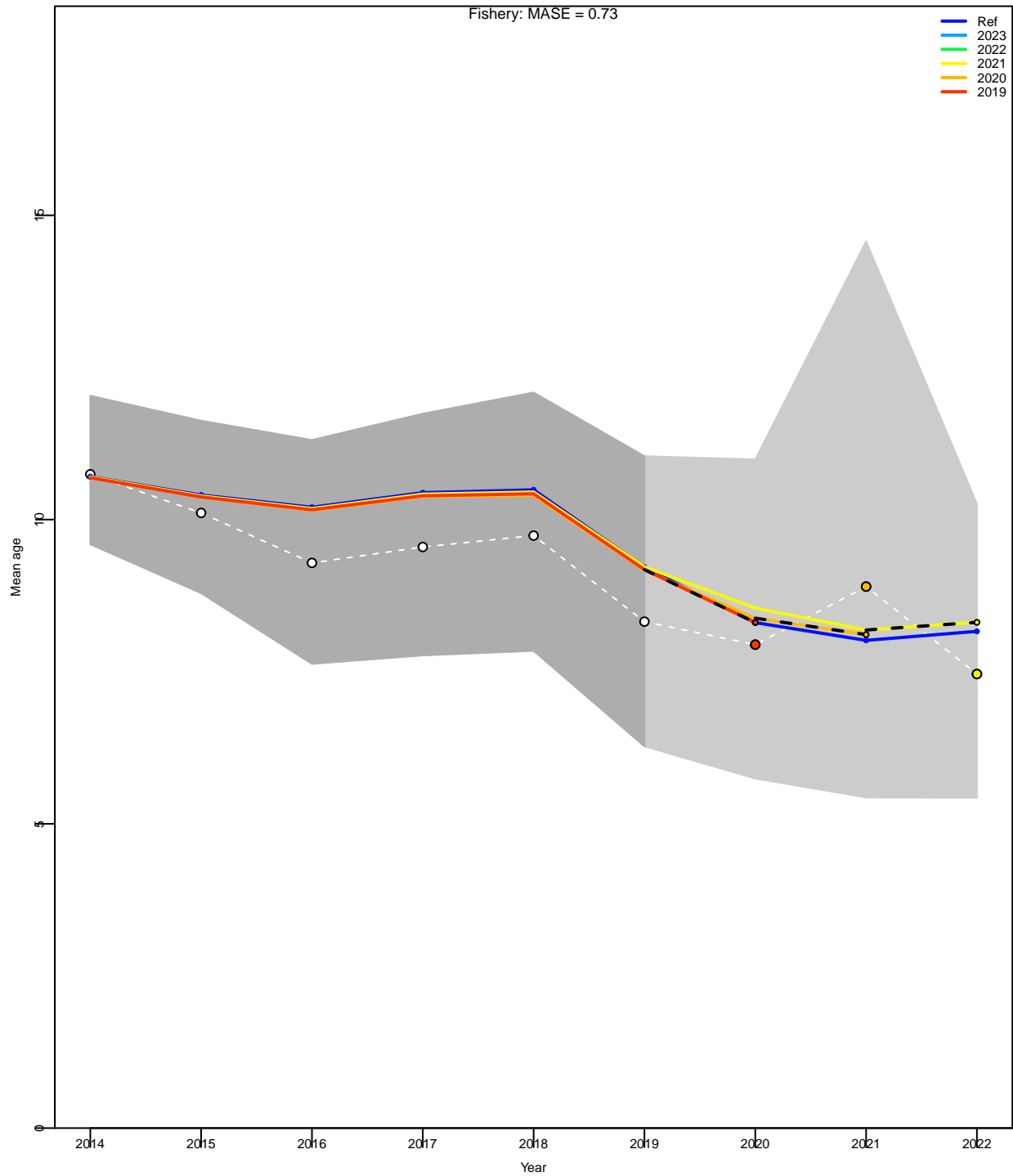


Figure 21: Fit and hindcast cross-validations for the fishery age compositions of the BestCase model

```
Computing MASE with only 2 of 5 prediction residuals for Index EasternSurvey
```

```
Warning: Unequal spacing of naive predictions residuals may influence the interpretation of MASE
```

```
MASE stats by Index:
```

	Index	Season	MASE	MAE.PR	MAE.base	MASE.adj	n.eval
1	NonEasternSurvey	1	0.8202749	0.1676188	0.2043447	0.8202749	2
2	EasternSurvey	1	1.0226579	0.2315900	0.2264589	1.0226579	2

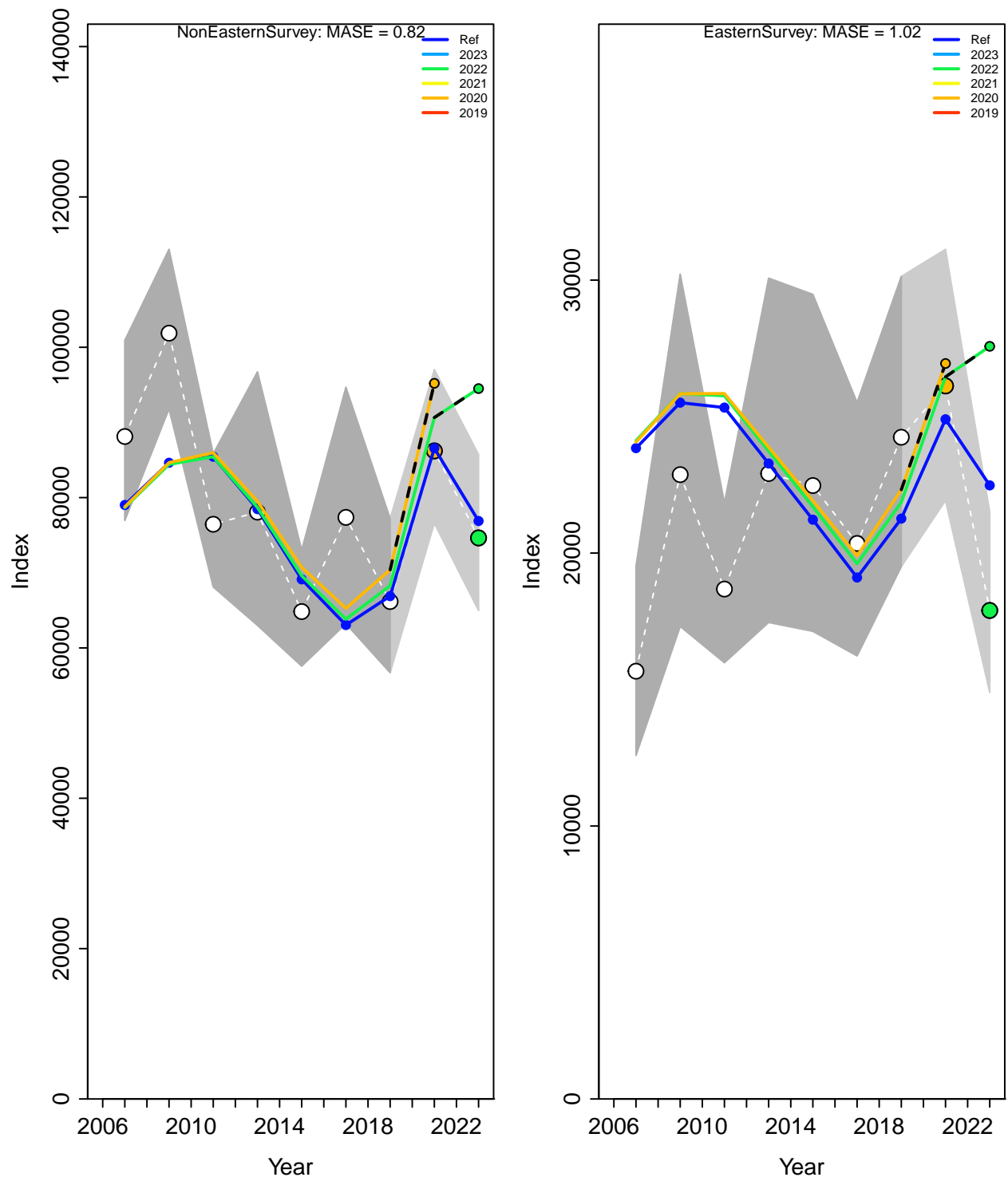


Figure 22: Hindcast with Cross-Validation of CPUE observations for the BestCase model

2.8.5 Profiling of steepness

The reference case was further evaluated through profiling it over a range of steepness values $h = 0.45 - 0.95$. Figure 22 shows that a higher range of $h = 0.90 - 0.95$ is supported by the total negative log-likelihood.

The stock estimates appear insensitive to alternative assumption about the steepness h values tested (Figure 22)

```
# summarize output
profilessummary <- SSsummarize(profilemodels, verbose = F)
results <- SSplotProfile(profilesummary, add_cutoff = TRUE, verbose = F,
  profile.string = "SR_BH_steep", profile.label = "Stock-recruit steepness (h)")

h = c(seq(0.65, 0.95, 0.05))

mvns = Map(function(x, y) {
  SSdeltaMVLN(x, add = T, run = paste0("h=", y), Fref = "Btgt",
    catch.type = "Exp", years = 1982:2024, verbose = F, plot = F)
}, x = profilemodels, y = h)

sspar(mfrow = c(3, 2), plot.cex = 0.7)
SSplotEnsemble(mvns, uncertainty = T, add = T, legendcex = 0.65,
  legendloc = "topright", verbose = F)
```

- Fishlife steepness value for the species is 0.72. The minimum likelihood is most likely an artifact of the historical trajectory of the stock, with little contrast in SSB due to the low fishing mortality. Due to asymmetric risk (Hordyk et al., 2019), it would be more appropriate to use a lower steepness value.

```
library(SPMpriors)
library(FishLife)
#####
Loading package FishLife, developed by James Thorson for the National Marine Fisheries Service
For details and citation guidance, please see http://github.com/James-Thorson-NOAA/FishLife/
#####
library(rfishbase)

Attaching package: 'rfishbase'
The following object is masked from 'package:FLCore':

  distribution
library(FLCandy)
Loading required package: plyr

Attaching package: 'plyr'
The following objects are masked from 'package:FLCore':

  desc, join
The following object is masked from 'package:ggpubr':

  mutate
Loading required package: FLlife
Warning: multiple methods tables found for 'leslie'

Attaching package: 'FLlife'
The following object is masked from 'package:FLBRP':
```

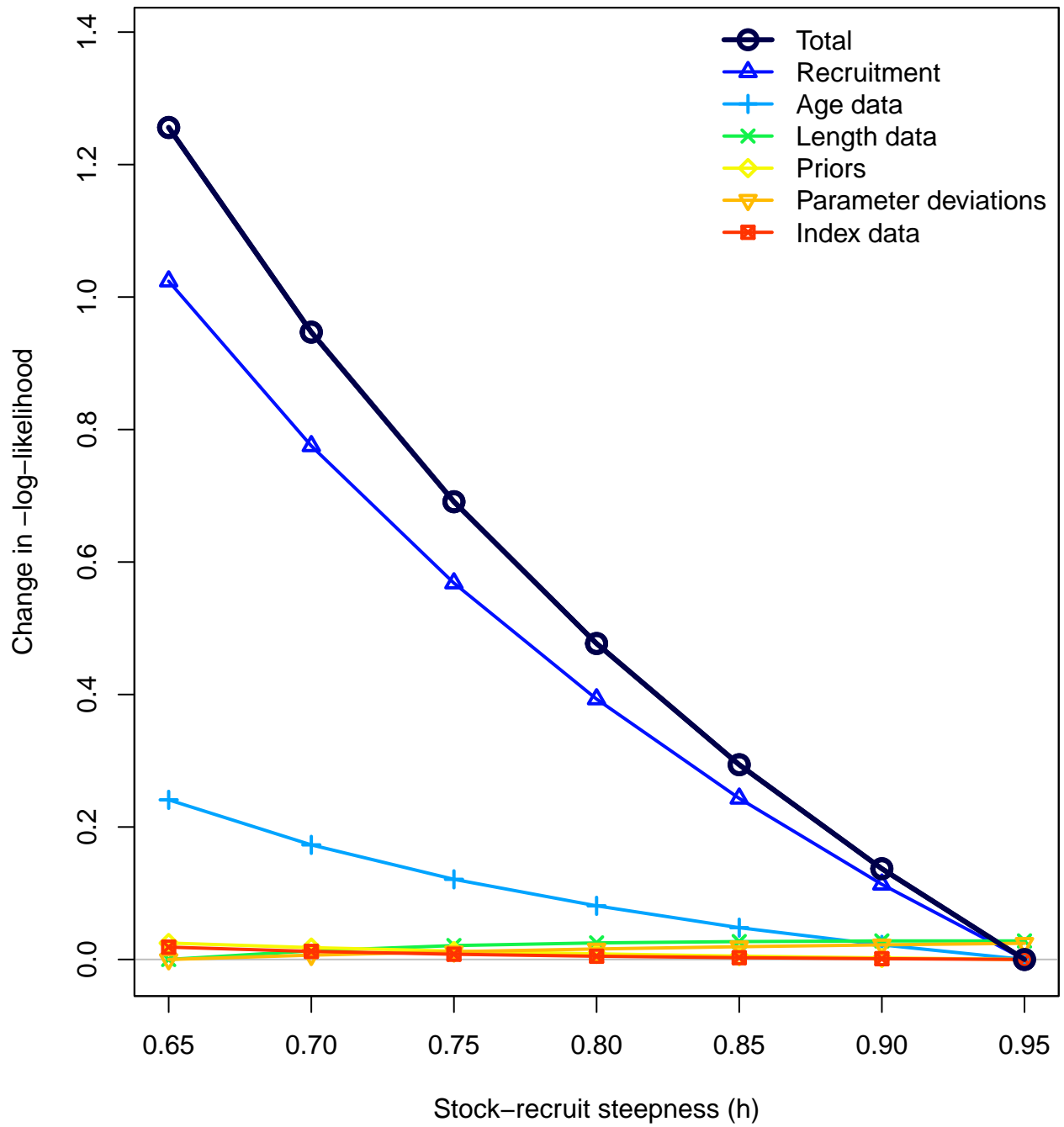


Figure 23: Negative log-likelihood profiles over a range of steepness values ($h = 0.65-0.95$)

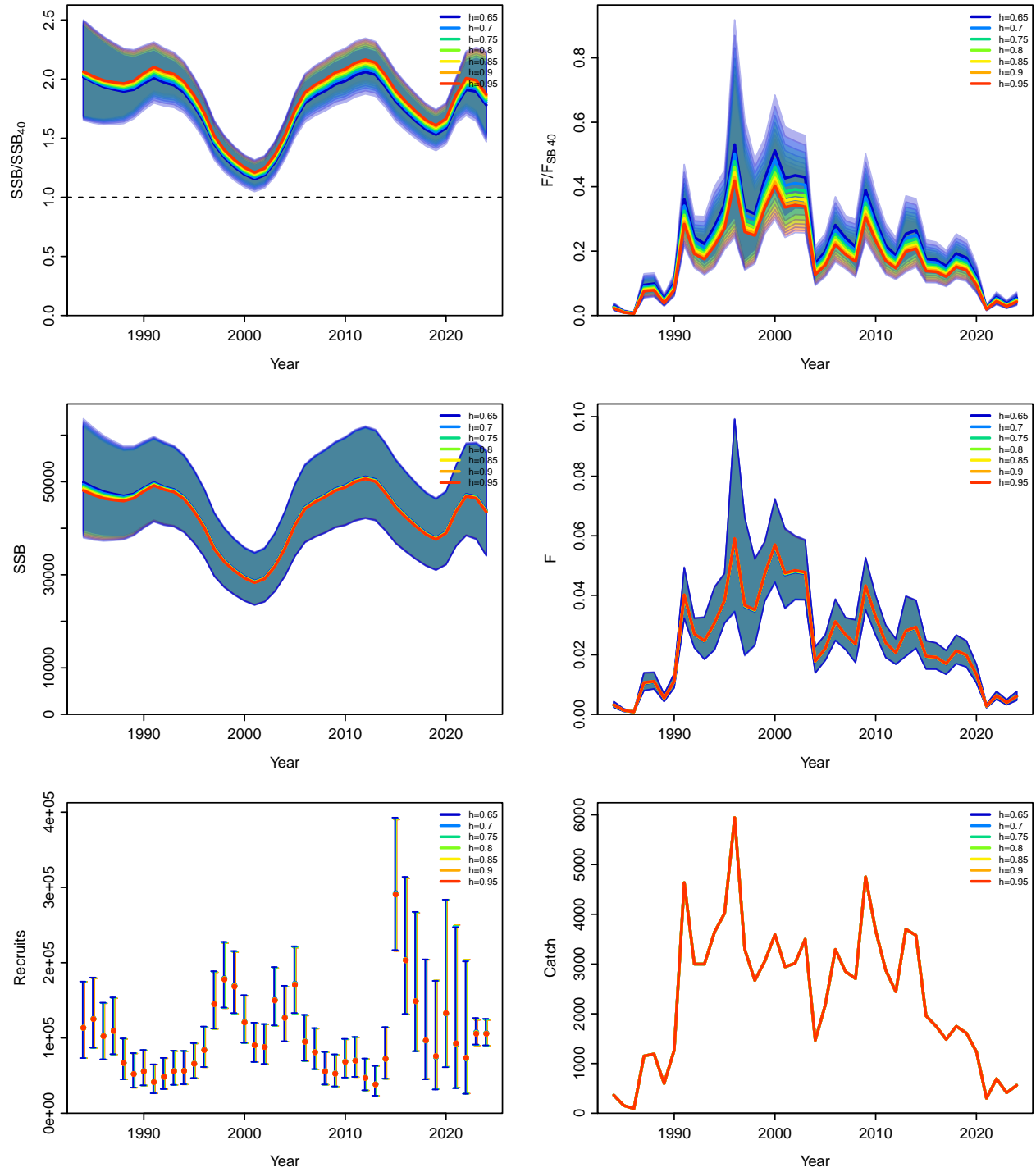


Figure 24: Comparison of stock trajectories with alternative stepness values ($h = 0.65-0.95$)

```

    leslie
The following objects are masked from 'package:FLCore':

    gompertz, leslie, lopt, richards
Loading required package: mydas
Warning: replacing previous import 'FLCore::gompertz' by 'FLife::gompertz' when
loading 'FLCandy'
Warning: replacing previous import 'FLCore::lopt' by 'FLife::lopt' when loading
'FLCandy'
Warning: replacing previous import 'FLCore::leslie' by 'FLife::leslie' when
loading 'FLCandy'
Warning: replacing previous import 'FLCore::richards' by 'FLife::richards' when
loading 'FLCandy'
Warning: replacing previous import 'FLCore::%+%' by 'ggplot2::%+%' when loading
'FLCandy'
Warning: multiple methods tables found for 'leslie'

Attaching package: 'FLCandy'
The following object is masked from 'package:mydas':

    hcrICES
The following object is masked from 'package:FLife':

    leslie
The following object is masked from 'package:FLBRP':

    leslie
The following objects are masked from 'package:FLSRTMB':

    from_logits, to_logits
The following objects are masked from 'package:FLCore':

    invALK, leslie
The following object is masked from 'package:stats':

    pt

stk_rexsole = flmvn_traits(Genus = "Glyptocephalus", Species = "zachirus",
  Loo = c(49.38, 0.1), Lm = c(32, 0.1), h = c(0.95, 0.99),
  K = c(0.18, 0.1), tmax = c(20, 0.1), M = c(0.17, 0.1), Plot = T,
  savepng = F)
Closest match: Actinopterygii_Pleuronectiformes_Pleuronectidae_Glyptocephalus_zachirus
  [,1]      [,2]
[1,] "K"      "M"
[2,] "Winfinity" "Loo"
[3,] "tmax"     "tm"
[4,] "Lm"       "Temperature"
[5,] "ln_margsd" "rho"
[6,] "logitbound_h" "ln_r"

table = stk_rexsole$traits
knitr::kable(table, "pipe", align = "lcccc", caption = "Tuned fishlife trait history for Alaska rex so.

```

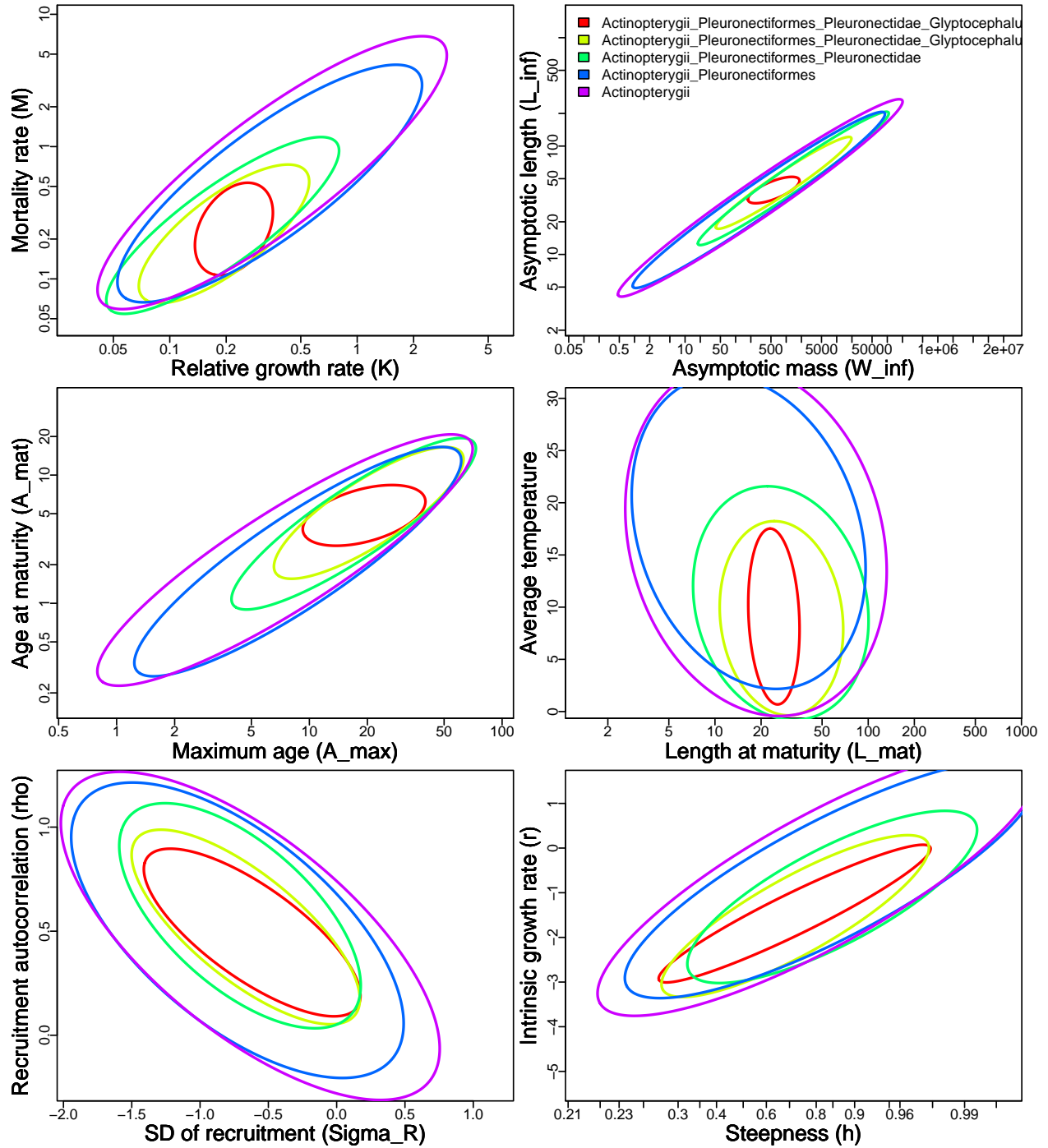


Figure 25: Tuned fishlife trait history for Alaska rex sole

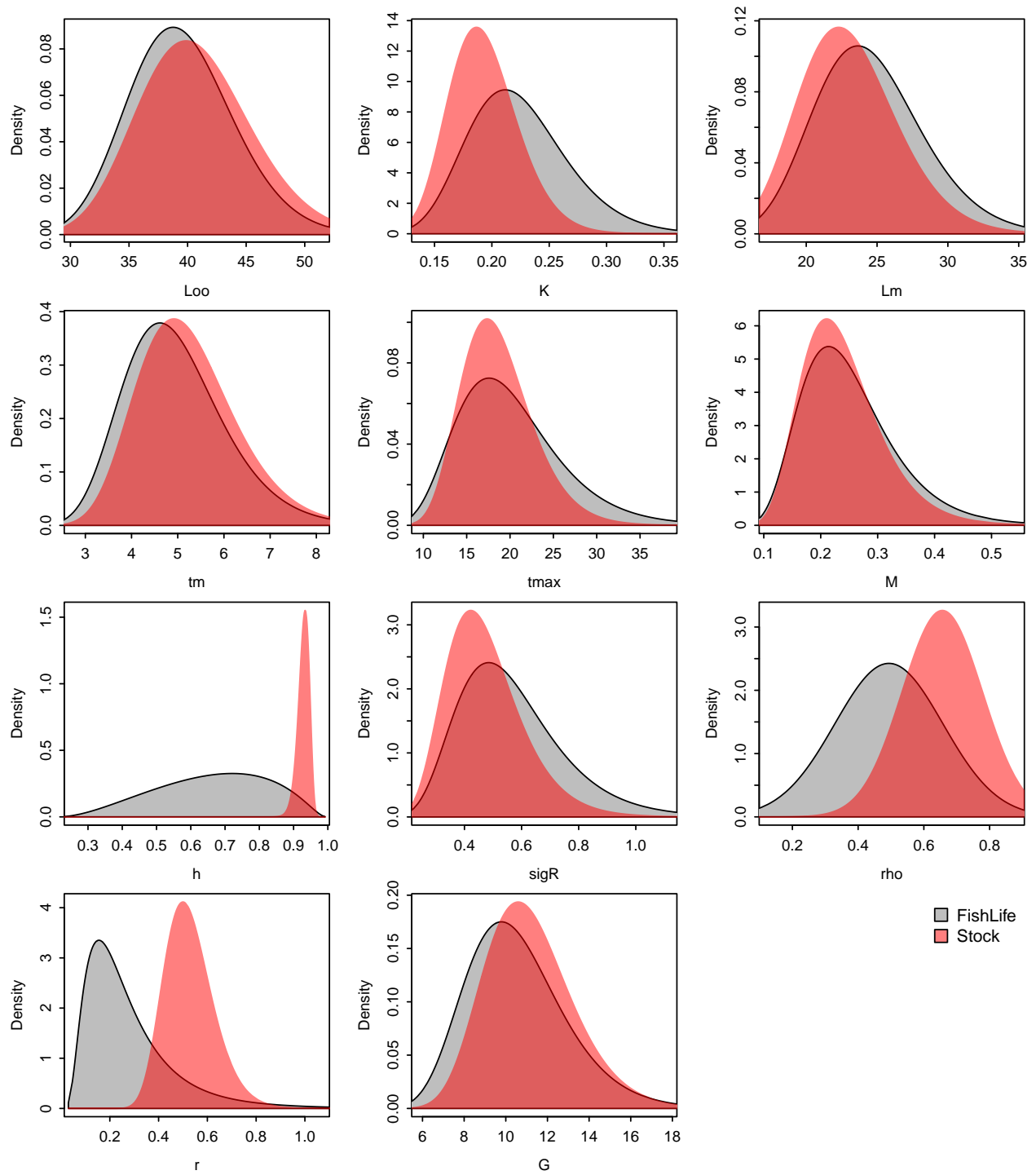


Figure 26: Tuned fishlife trait history for Alaska rex sole

Table 1: Tuned fishlife trait history for Alaska rex sole

trait	mu.sp	cv.sp	mu.stk	cv.stk	lc.stk	uc.stk	upper.quant
Loo	39.2772	0.1151	40.4418	0.1184	34.9205	46.8360	0.9
K	0.2199	0.1972	0.1912	0.1626	0.1487	0.2457	0.9
Lm	24.2392	0.1583	22.8101	0.1530	18.6401	27.9130	0.9
tm	4.8481	0.2247	5.1291	0.2065	3.8551	6.8240	0.9
tmax	19.2282	0.3060	18.2203	0.2132	12.4127	26.7452	0.9
M	0.2382	0.3376	0.2292	0.3095	0.1504	0.3495	0.9
h	0.7208	0.2647	0.9338	0.0171	0.9108	0.9512	0.9
sigR	0.5386	0.3321	0.4570	0.2891	0.3017	0.6923	0.9
rho	1.6381	0.3331	1.9274	0.1864	1.5608	2.3800	0.9
r	0.2305	0.6993	0.5182	0.1938	0.2312	1.1617	0.9
G	10.3087	0.2299	10.9892	0.1871	8.2149	14.7004	0.9

```
write.csv(table, file = "Fishlife table.csv", row.names = F)
```

3 Assessment outcome

3.1 Reference Points

- Reference points were estimated within the Stock Synthesis model, where the biomass reference points are considered as SSB for females. For illustration only, target reference points of $B_{tgt} = SSB_{40}$ (biomass equal to 40 percent of unfished biomass SSB_0) and $F_{tgt} = F_{SB40}$ (fishing mortality level at SSB_{40}) were proposed to serve a preliminary as proxies for B_{MSY} and F_{MSY} . In the light of uncertainty about the underlying stock recruitment relationship, this choice of precautionary MSY proxies is likely to reduce the asymmetric risk of overfishing, while still attaining more than 95% of the theoretical MSY at F_{MSY} and ensuring that about more 40% more SSB is left in the water to ensure both future recruitment and catch opportunities.
- When compared to the BaseCase, the BestCase improves the fit of the last year of the indices and the LFDs

```
sspar(mfrow = c(2, 2), plot.cex = 0.7)
SSplotIndices(base, subplots = 2)
SSplotIndices(ref, subplots = 2)
```

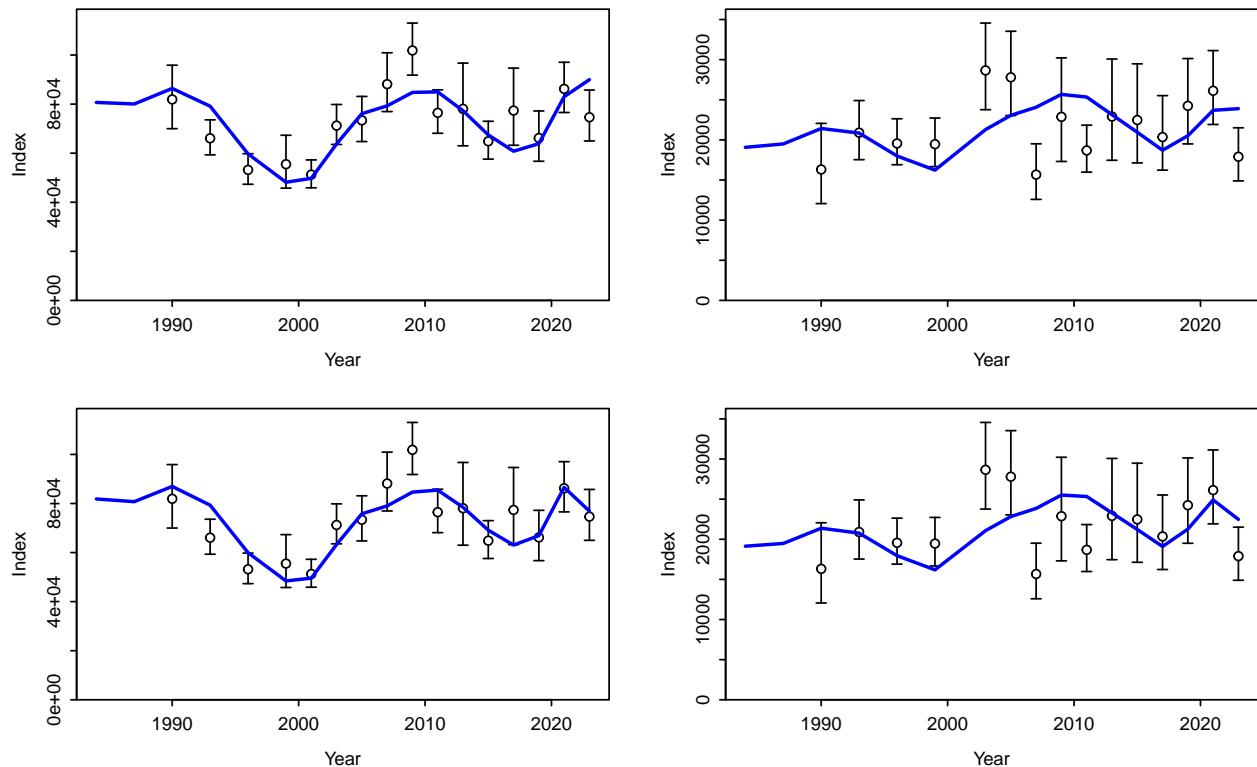


Figure 27: Comparison between BaseCase and BestCase model for the fit of survey indices

```
sspar(mfrow = c(1, 2), plot.cex = 0.7)
SSplotComps(base, subplots = 21)
```

```
SSplotComps(ref, subplots = 21)
```

```
sspar(mfrow = c(1, 2), plot.cex = 0.7)
SSplotComps(base, subplots = 21, kind = "AGE")
```

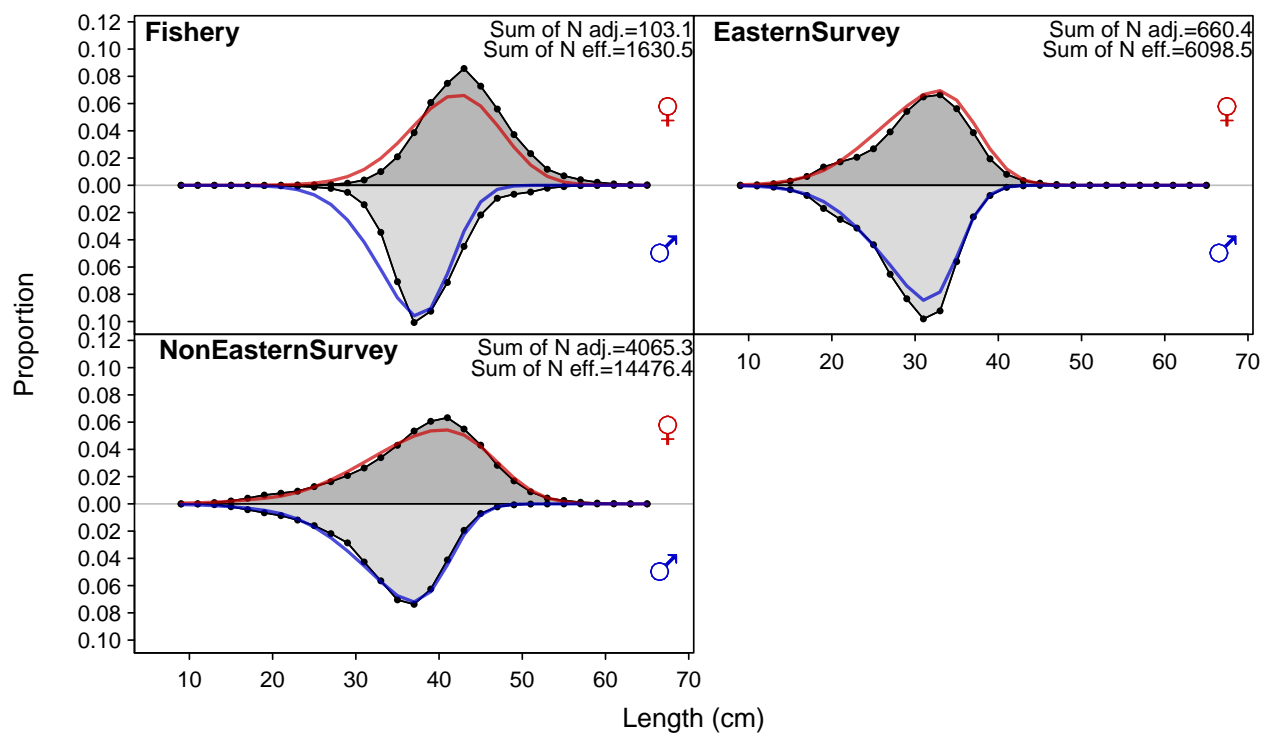



Figure 28: Comparison between BaseCase (top) and BestCase for the fit of the aggregated length compositions (down)

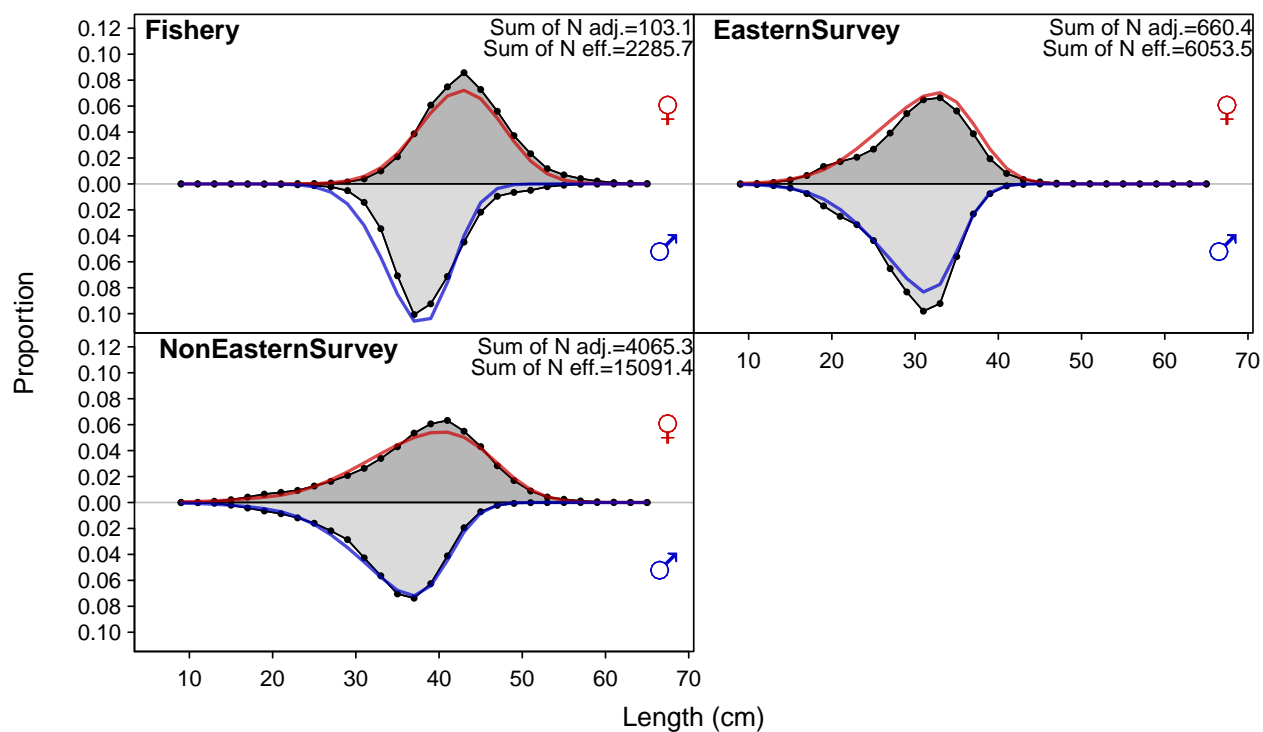


Figure 29: Comparison between BaseCase (top) and BestCase for the fit of the aggregated length compositions (down)

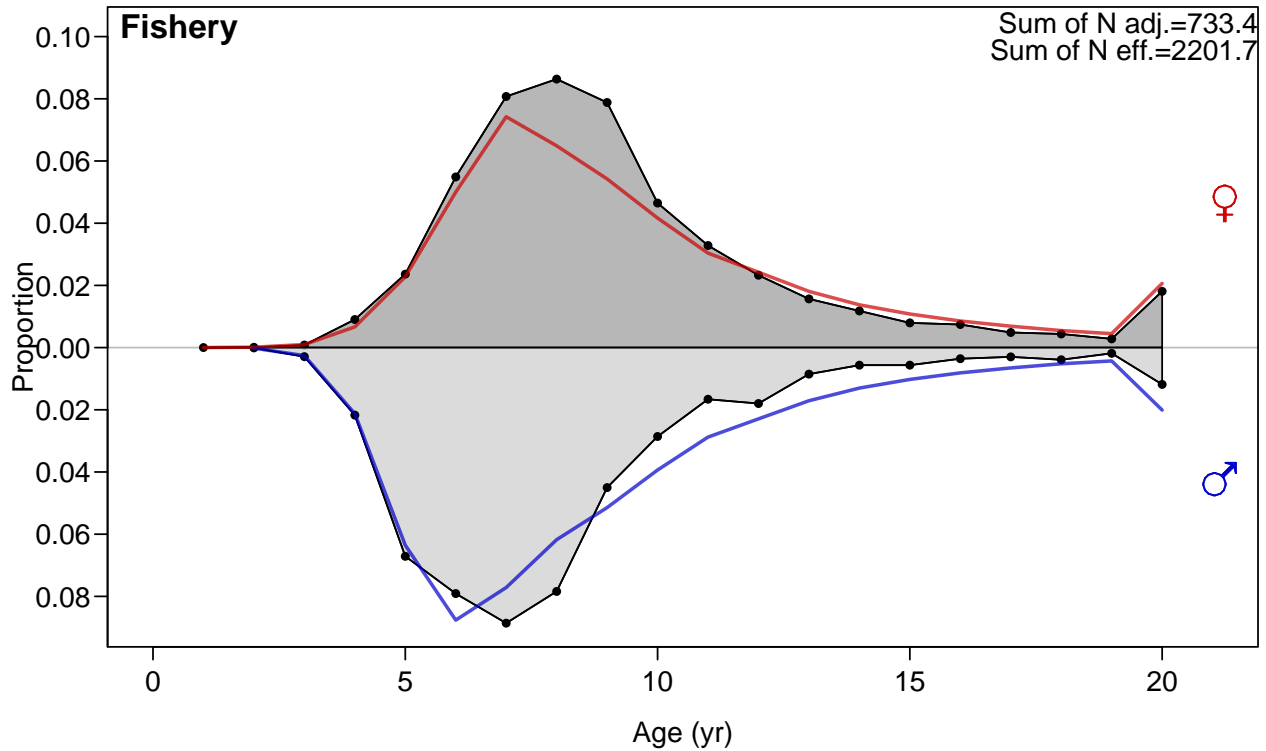


Figure 30: Comparison between BaseCase (top) and BestCase model for the fit of the aggregated age compositions (down)

```
SSplotComps(ref, subplots = 21, kind = "AGE")
```

3.2 Stock Status as estimated by the BestCase model

- Only for illustration, Blim and Btrigger were added

```
mvnbase = FLRef::ssmvlm(ref, Fref = "Btgt", years = 1982:2024,
  verbose = T)

starter.sso with Bratio: SSB/SSB0 and F: _abs_F

stk = ss2FLStockR(mvnbase)

# Add Btrigger + Blim
stk@refpts = rbind(stk@refpts, FLPar(Btrigger = stk@refpts[[2]] *
  1, Blim = stk@refpts[[2]] * 0.5))

# with uncertainty
stki = ss2FLStockR(mvnbase, output = "iters")
stki@refpts = stk@refpts

plotAdvice(stki)
```

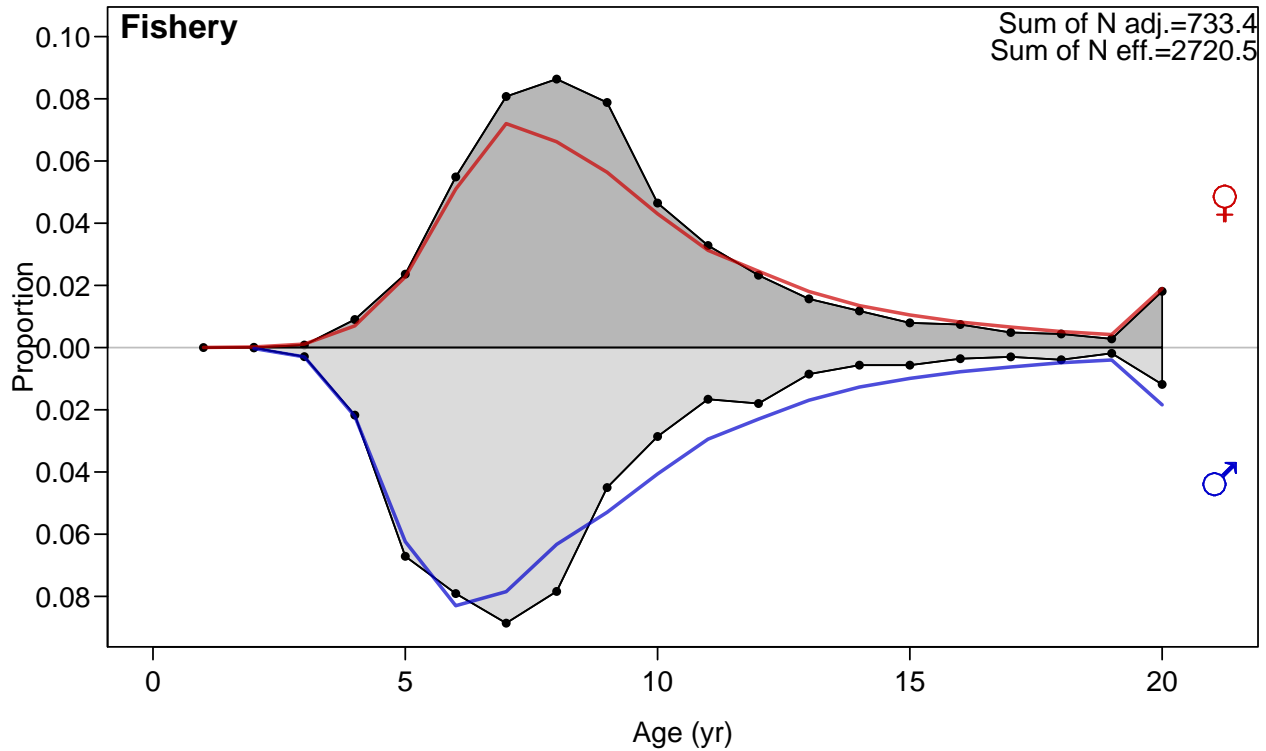


Figure 31: Comparison between BaseCase (top) and BestCase model for the fit of the aggregated age compositions (down)

3.3 Comparison with previous model

```
## Summarize output
mydir <- "~/Max/Commitees/CIE/Alaska rex sole/Ensemble/run13_sampler_and_survbio_edits/"
rdata_retros = "~/Max/Commitees/CIE/Alaska rex sole/Ensemble/rdata_retros"
rdata_runs = "~/Max/Commitees/CIE/Alaska rex sole/Ensemble/rdata_runs"
retroModels <- SSgetoutput(dirvec = file.path(mydir, "Retrospective",
  paste("retro", 0:-5, sep = "")))
i = "run13_sampler_and_survbio_edits"

retroSummary <- r4ss::SSsummarize(retroModels)
endyrvec <- endyrvec <- retroSummary$endyrs + 0:-5
save(retroSummary, retroModels, file = paste0(dir = rdata_retros,
  "/RetroModels_", i, ".Rdata"))

ss3rep = retroModels[[1]]
save(ss3rep, file = paste0(dir = rdata_runs, "/Alaska rex sole_",
  i, ".Rdata"))

load("rdata_runs/Alaska rex sole_run13_sampler_and_survbio_edits.RData",
  verbose = T)
Loading objects:
  ss3rep
bm = FLRef::ssmvln(ss3rep, Fref = "Btgt", verbose = F)
stk.bm = ss2FLStockR(bm)
```

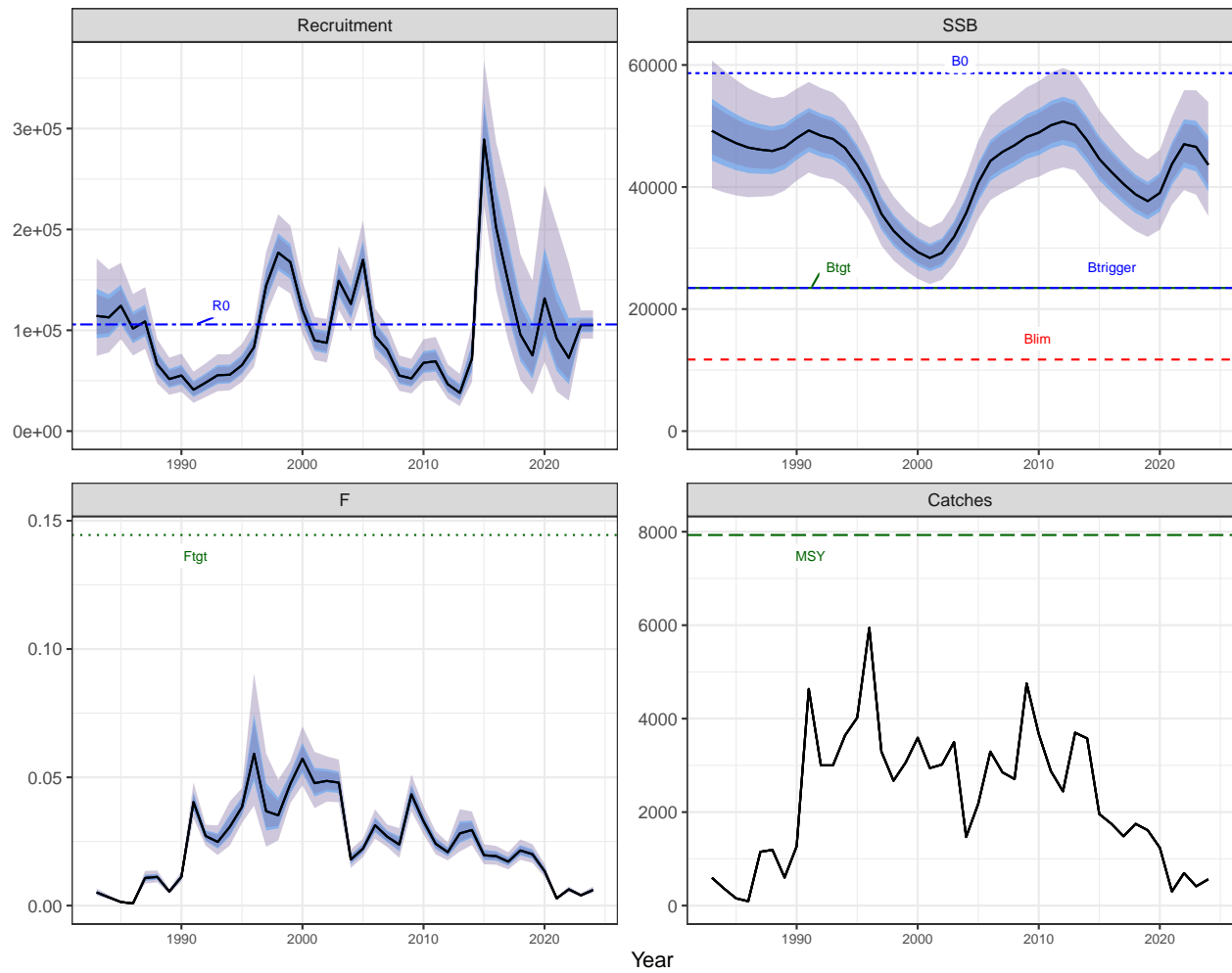


Figure 32: Estimated stock status trajectories with associated reference points for the 2025 BestCase model of Alaska rex sole

```
plot(FLStocks(New2025 = stk, Reference2021 = stk.bm)) + facet_wrap(~qname,
  scales = "free_y") + theme_bw()
```

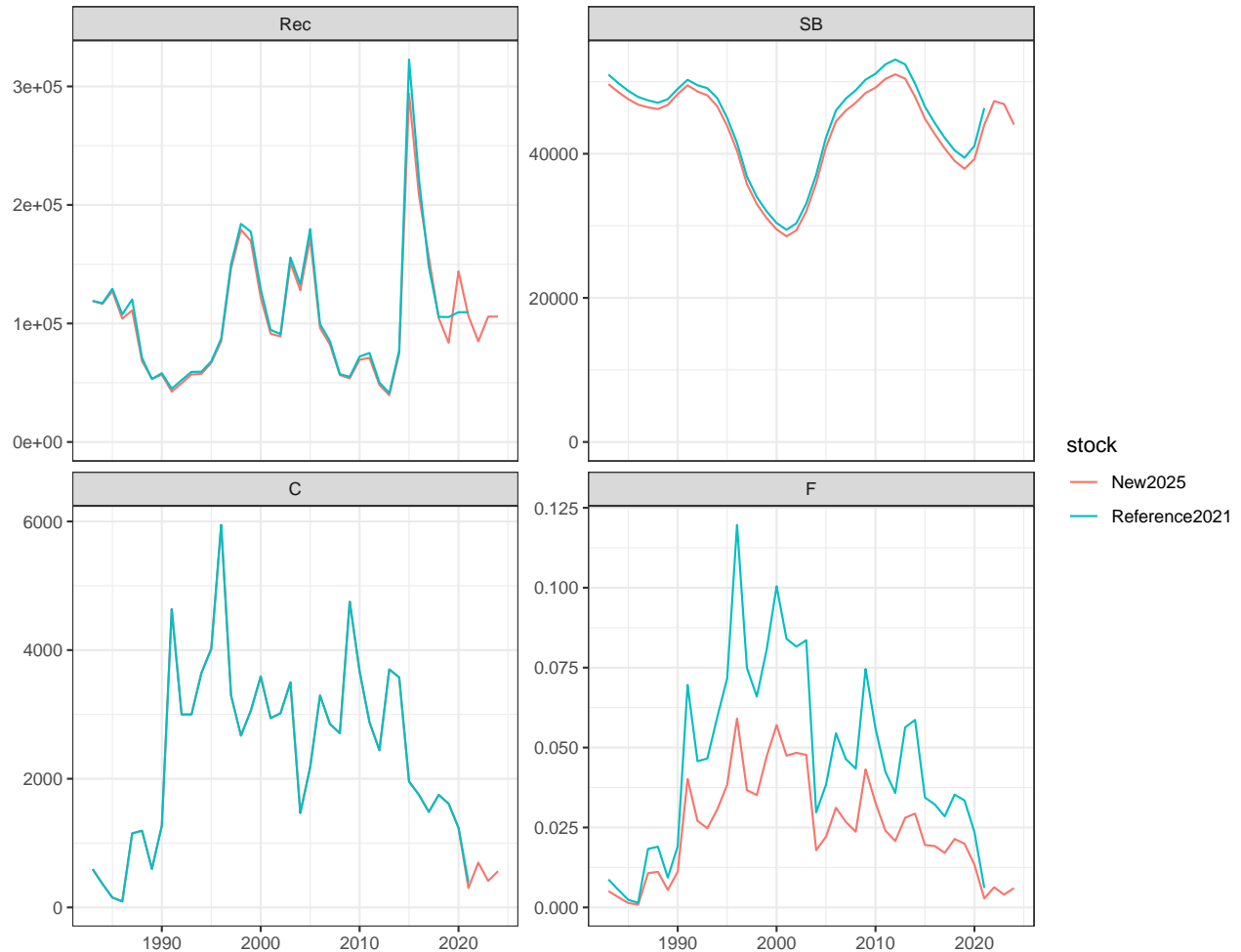


Figure 33: Comparison between the estimated stock status trajectories from the BestCase and the last previous advice model

```
stks = FLStocks(New2025 = stock2ratios(stk), Reference2021 = stock2ratios(stk.bm))
stks[[1]]@refpts = stks[[1]]@refpts[1:2]
stks[[2]]@refpts = stks[[2]]@refpts[1:2]
plotAdvice(stks)
```

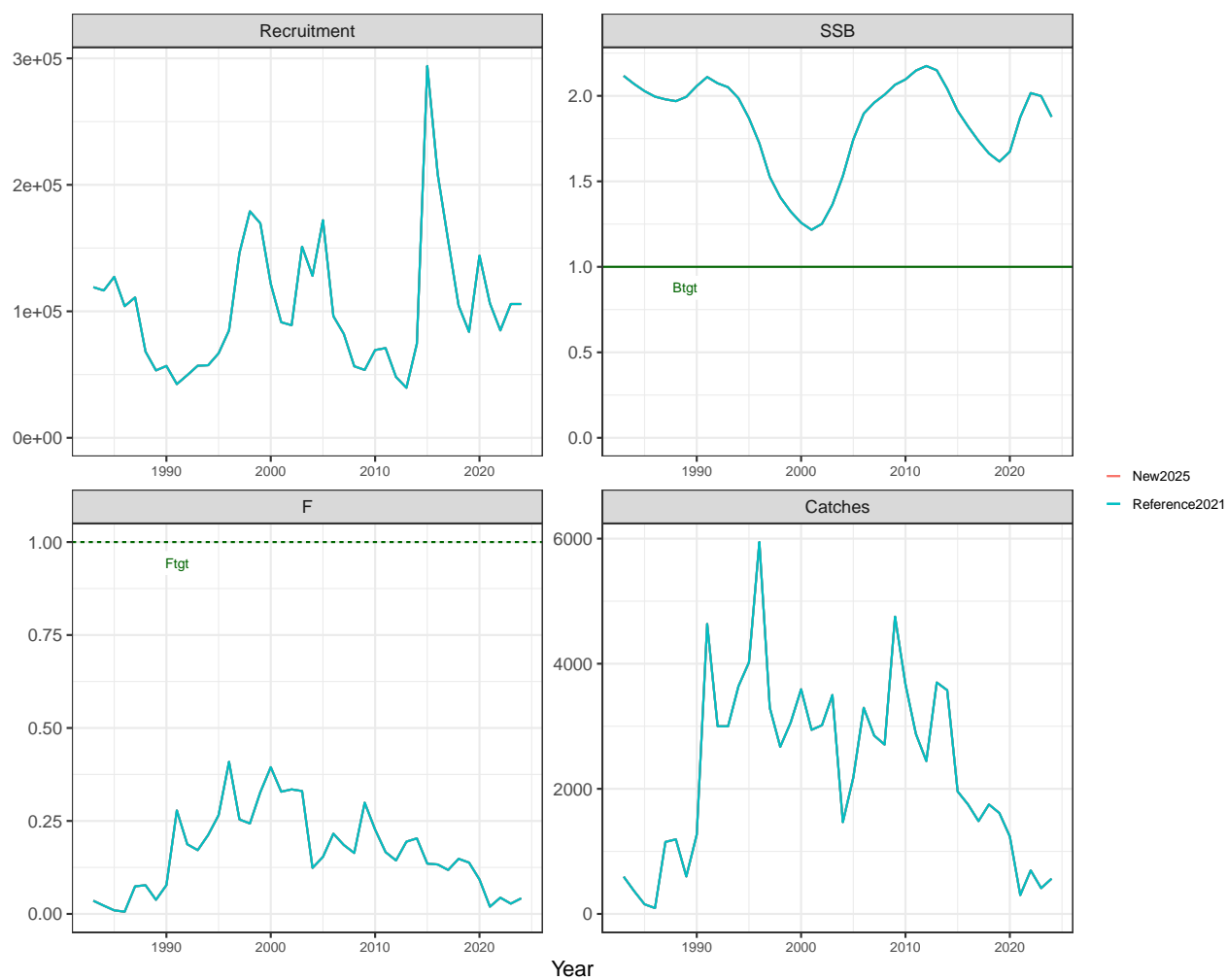


Figure 34: Comparison between the relative stock status estimates from the BestCase and the last previous advice model

```
knitr::kable(FLRef::flr2stars(stk)$refpts, "pipe", align = "lc",
  caption = "Summary of the estimated reference points
  for Alaska rex sole")
```

Table 2: Summary of the estimated reference points for Alaska rex sole

RefPoint	Value
Ftgt	0.144
Btgt	23463.126
MSY	7929.680
B0	58657.900
R0	105786.000
Btrigger	23463.126
Blim	11731.563
Fcur	0.006
Bcur	44055.300
B0.33	41875.477
B0.66	47100.658

```
knitr::include_graphics("MLVN_Compare.jpg")
```

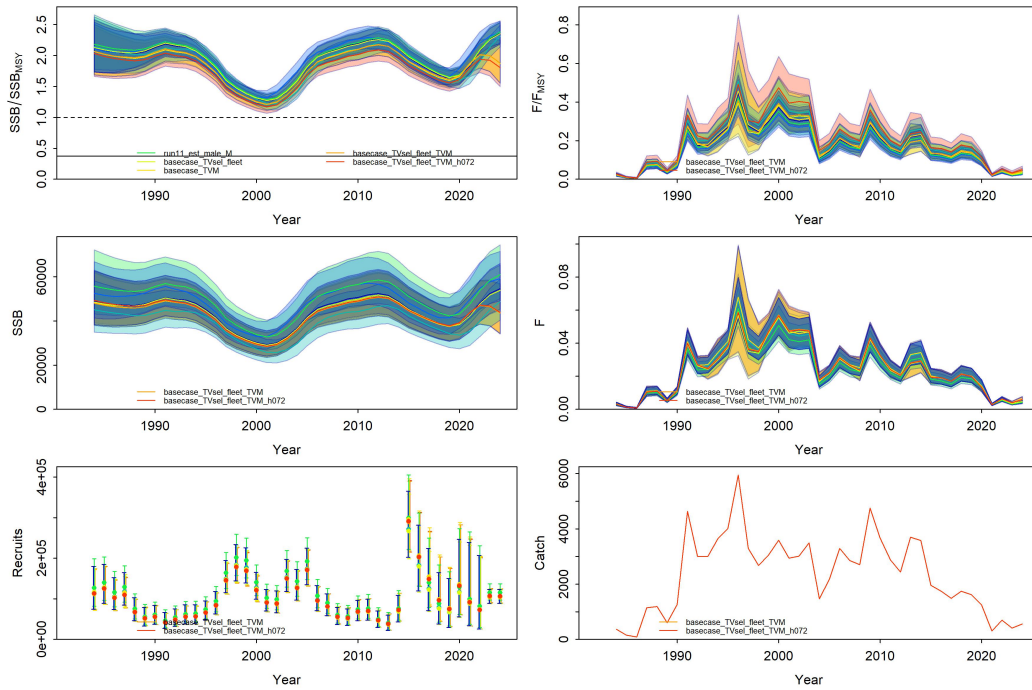


Figure 35: Comparison of alternative models

- Diagnostic table for Alaska rex sole. Models with time varying fleet selectivity and M achieved highest score.

```
knitr::include_graphics("./Diags_table.pdf")
```


Run	BaseCase	run9_early_maturity	run10_late_maturity	run11_est_male_M	BaseCase_TVsel_fleet	BaseCase_TVM	BaseCase_TVsel_fleet_TVM	BaseCase_TVsel_fleet_TVM_h072
Convergence	8.42E-05	1.24E-04	9.41E-04	8.80E-05	1.22E-04	4.26E-05	1.29E-04	8.75E-05
Total_LL	614	614	614	696	597	606	590	591
N_Params	89	89	89	91	153	98	156	156
Runs_test_cpue1	Passed	Passed	Passed	Passed	Passed	Passed	Passed	Passed
Runs_test_cpue2	Passed	Passed	Passed	Passed	Passed	Passed	Passed	Passed
Runs_test_len1	Failed	Failed	Failed	Failed	Failed	Failed	Failed	Failed
Runs_test_len2	Passed	Passed	Passed	Passed	Passed	Passed	Passed	Passed
Runs_test_len3	Passed	Passed	Passed	Passed	Passed	Passed	Passed	Passed
Runs_test_age1	Passed	Passed	Passed	Passed	Passed	Passed	Passed	Passed
RMSE_Perc	21.5	21.5	21.5	21.8	21.6	21	21	21.3
RMSE_Perc_1	5.1	5.1	5.1	4.9	4.5	5.1	4.5	4.5
RMSE_Perc_2	10.6	10.6	10.6	10.5	7.6	9.9	7.5	7.5
Retro_Rho_SSB	0.05	0.06	0.04	0.06	0.05	0.04	0.04	0.04
Forecast_Rho_SSB	0.07	0.08	0.06	0.08	0.07	0.07	0.07	0.07
Retro_Rho_F	-0.06	-0.06	-0.06	-0.06	-0.05	-0.06	-0.05	-0.05
Forecast_Rho_F	-0.07	-0.07	-0.07	-0.08	-0.08	-0.10	-0.09	-0.09
MASE_cpue1	1.10	1.10	1.10	1.09	1.01	0.88	0.82	0.80
MASE_cpue2	1.05	1.05	1.05	1.05	0.99	1.08	1.02	1.01
MASE_cpue3	1.07	1.07	1.07	1.07	1.00	0.98	0.93	0.91
MASE_len1	0.44	0.44	0.44	0.40	0.62	0.44	0.57	0.56
MASE_len2	0.20	0.20	0.20	0.22	0.21	0.21	0.21	0.22
MASE_len3	0.40	0.40	0.40	0.39	0.41	0.40	0.40	0.41
MASE_len4	0.33	0.33	0.33	0.32	0.37	0.33	0.36	0.37
MASE_age1	0.97	0.97	0.97	0.97	0.79	0.97	0.73	0.73
Weight	0.81	0.81	0.81	0.81	0.90	0.90	0.90	0.90

Figure 36: Diagnostic table for Alaska rex sole

```

out = FLRef::flr2stars(stki)$timeseries
mles = FLRef::flr2stars(stk)

# replace medians with mles
out$Rec = mles$timeseries$Rec
out$SSB = mles$timeseries$SSB
out$Bratio = mles$timeseries$Bratio
out$F = mles$timeseries$F
out$Fratio = mles$timeseries$Fratio

write.csv(out, file = "Alaska rex sole.stars.csv", row.names = F)
write.csv(mles$refpts, file = "Alaska rex sole.refpts.stars.csv",
  row.names = F)

```

4 **Supplement: R code to run additional diagnostics (to be run outside the script)**

```
# ><><><><><><><><><><><><><><><><><><><><><><><><><><><>
# > Stock Synthesis reference model summary and diagnostic
# for Alaska rex sole > @author Henning Winker (GFCM),
# modified by Max Cardinale (SLU) > henning.winker@fao.org

# ><><><><><><><><><><><><><><><><><><><><><><><><><><><> Load packages
library("r4ss")
library("ss3diags")

# First set working directory to the R file location with
# ss3 subfolders

# Define run name of folder
run = "basecase_TVsel_fleet_TVM"

# Load the model run
ss3rep = SS_output(run)

# Plot the model run
r4ss::SS_plots(ss3rep)

# Save the r4ss object as rdata
dir.create("rdata")
save(ss3rep, file = file.path("rdata", paste0("Alaska rex sole_",
run, ".rdata")))

# approximate uncertainty and produce Kobe Plot
sspar(mfrow = c(1, 1))
mvn = SSdeltaMVLN(ss3rep, Fref = "Btgt", run = "ref", catch.type = "Exp")

# Plot trajectories with CIs
sspar(mfrow = c(3, 2))
SSplotEnsemble(list(mvn), add = T)

# ><><><><><><><><><><><><><><><><><><><><><><><><><><><>
# Compile results summary
# ><><><><><><><><><><><><><><><><><><><><><><><><><><><>

# Make output PDF
pdf(paste0("Alaska rex sole_", run, ".pdf"))
sspar(mfrow = c(2, 2), plot.cex = 0.7)
SSplotBiology(ss3rep, mainTitle = F, subplots = c(1))
SSplotBiology(ss3rep, mainTitle = F, subplots = c(2))
SSplotBiology(ss3rep, mainTitle = F, subplots = c(6))
SSplotBiology(ss3rep, mainTitle = F, subplots = c(9))

sspar(mfrow = c(2, 2), plot.cex = 0.7)
SSplotBiology(ss3rep, mainTitle = F, subplots = c(4))

# Recruitment
```

```

sspar(mfrow = c(2, 2), plot.cex = 0.7)
SSplotRecdevs(ss3rep, subplots = 1)
SSplotRecdevs(ss3rep, subplots = 2)
SSplotSpawnrecruit(ss3rep, subplots = 1)
SSplotSpawnrecruit(ss3rep, subplots = 3)

par(mfrow = c(1, 1))
SSplotDynamicB0(ss3rep)

sspar(mfrow = c(1, 3), plot.cex = 0.7)
SSplotIndices(ss3rep, subplots = 2)

sspar(mfrow = c(1, 1), plot.cex = 0.7)
SSplotIndices(ss3rep, subplots = 9)

sspar(mfrow = c(2, 3), plot.cex = 0.9)
SSplotHCxval(retro.idx, xmin = 2006, add = T, legendcex = 0.6,
  Season = 1)
SSplotHCxval(retro.len, add = T, subplots = "len", legendloc = "topleft",
  indexUncertainty = TRUE, legendcex = 0.6)

par(mfrow = c(1, 1))
SSplotSelex(ss3rep, subplots = 1)

SSplotComps(ss3rep, subplots = 21)

SSplotComps(ss3rep, subplots = 1)

SSplotSexRatio(ss3rep, kind = "LEN")

# Bubble
SSplotComps(ss3rep, subplots = 24)

# ALK
SSplotComps(ss3rep, kind = "cond", subplots = 3)

# Runs Fleets
sspar(mfrow = c(3, 3), plot.cex = 0.5)
SSplotRunstest(ss3rep, subplots = "len", add = T)

# HC Fleets
sspar(mfrow = c(3, 2), plot.cex = 0.5)
for (i in 1:2) {
  SSplotHCxval(retro.len, subplots = "len", add = T, legendloc = "topleft",
    indexselect = i)
}

# Retro
sspar(mfrow = c(2, 2), plot.cex = 0.65)
SSplotRetro(retro.idx, add = T, legend = F, forecast = F)
SSplotRetro(retro.idx, add = T, forecastrho = T, legend = F)
SSplotRetro(retro.idx, subplots = "F", add = T, legend = F, forecast = F)
SSplotRetro(retro.idx, subplots = "F", add = T, forecastrho = T,

```

```

    legend = F)
mtext(c("Retro", "Forecast"), 3, outer = T, line = -0.5, at = c(0.3,
    0.8), cex = 0.8)

par(mfrow = c(1, 1))
SSplotYield(ss3rep, subplots = 2)

sspar(mfrow = c(1, 1))
mvn = SSdeltaMVLN(ss3rep, Fref = "Btgt", catch.type = "Exp",
    run = "ref")

sspar(mfrow = c(3, 2))
SSplotEnsemble(list(mvn), add = T)

dev.off()

```

4.0.1 Literature cited

- Felipe Carvalho, Henning Winker, Dean Courtney, Laurence Kell, Maia Kapur, Massimiliano Cardinale, Michael Schirripa, Toshihide Kitakado, Dawit Y. Ghebrehiwet, Kevin R. Piner, Mark N. Maunder, Rick Methot, 2021. A Cookbook for Using Model Diagnostics in Integrated Stock Assessments. Fisheries Research, <https://doi.org/10.1016/j.fishres.2021.105959>.
- Adrian R. Hordyk, Quang C. Huynh, Thomas R. Carruthers 2019. Misspecification in stock assessments: Common uncertainties and asymmetric risks. Fish and Fisheries, <https://doi.org/10.1111/faf.12382> “