Introduction to Bioinformatics and Computational Biology

2021-01-18

Contents

1	Cou	urse information	11
	1.1	Contributors	11
	1.2	Supporting staff	11
2	Inti	roduction	13
	2.1	Welcome	14
	2.2	Brief history of bioinformatics	14
	2.3	Should I take this course?	14
	2.4	Course information	14
3	Hig	h throughput sequencing	15
	3.1	Three generations of sequencing technologies	15
	3.2	FASTQ and FASTQC	15
	3.3	Early sequence alignment $(1 \text{ with } 1) \dots \dots \dots \dots$	15
	3.4	Sequence search algorihtms (1 with many)	16
	3.5	Borrow-Wheeler Aligner (many with many)	16
	3.6	Alignment output	16
4	$\mathbf{R}\mathbf{N}$	A-seq quantification	17
	4.1	RNA-seq & RNA QC (DV200)	17
	4.2	RNA-seq replicates and study design	17
	4.3	STAR and PE BAM file	17
	4.4	RNA-seq QC	17
	4.5	FPKM vs TPM	17
	4.6	RSEM & Salmon	17

4 CONTENTS

5	Diff	ferential expression, False discovery rate, Gene ontology	19
	5.1	RNA-seq NB distribution	19
	5.2	DESeq2 and variance stabilization	19
	5.3	Multiple hypotheses testing and FWER $\ \ldots \ \ldots \ \ldots \ \ldots$	19
	5.4	FDR	19
	5.5	GO	19
6	GS	EA, Clustering	21
	6.1	GSEA	21
	6.2	Heatmap and clustering quality	21
	6.3	H-cluster	21
	6.4	K-means	21
	6.5	Pick K and consensus clustering	21
	6.6	Batch effect removal	21
7	Din	nension Reduction	23
	7.1	MDS	23
	7.2	LDA	23
	7.3	PCA	23
8	Cla	ssification	2 5
	8.1	Intro to machine learning	25
	8.2	Cross validation	25
	8.3	Regression	25
	8.4	Regularization	25
	8.5	KNN	25
	8.6	Decision trees	25
	8.7	Random forest	25
	8.8	SVM	25
9	Mo	dule I Review	27
	9.1	Gene Expression Module Summary	27
	9.2	Gene Expression Analysis Scenarios	27

CONTENTS	F
CONTENIOS	5
CONTENIO	· ·

10	Transcription Factor Motif Finding		29
	10.1 Transcription regulation		29
	10.2 Motif representation		29
	10.3 EM		29
	10.4 Gibbs sampler		29
	10.5 Gibbs intuition		29
	10.6 Motif finding in eukaryotes		29
	10.7 Known motif database	•	29
11	ChIP-seq, Expression Integration		31
	11.1 ChIP-seq		31
	11.2 BWA and MACS		31
	11.3 ChIP-seq QC		31
	11.4 TF interactions (motif)		31
	11.5 TF target genes (expression integration)	•	31
12	2 Epigenetics, DNA Methylation		33
	12.1 Epigenetics		33
	12.2 DNA methylation		33
	12.3 Promoter function		33
	12.4 Gene body function		33
	12.5 Enhancer function		33
	12.6 Repetitive region function		33
	12.7 Early cancer detection	•	33
13	B Histone Modifications, Chromatin Accessibility		35
	13.1 Nucleosome positions		35
	13.2 Histone modification		35
	13.3 Promoters (bivalent)		35
	13.4 Genes (K36me3, new genes)		35
	13.5 Enhancers (K27ac)		35
	13.6 Super-enhancers		35
	13.7 DNase-seq		35
	13.8 ATAC-seq		35

6 CONTENTS

14	Long Range Chromatin Interactions	37
	14.1 Chromatin interactions	37
	14.2 HiC	37
	14.3 HiC contact map	37
	14.4 HiC normalization	37
	14.5 Fractal globule	37
	14.6 Loops	37
	14.7 Domains	37
	14.8 Compartments	37
	14.9 Phase separation	37
15	Hidden Markov Model	39
	15.1 Intro to HMM	39
	15.2 Pb1: Forward & backward procedure	39
	15.3 Pb2: Viterbi algorithm	39
	15.4 Pb3: Parameter estimation	39
	15.5 HMM application	39
16	6 Module II Review	41
	16.1 Module II Review	41
	16.2 Practive Questions	41
17	SNP and GWAS	43
	17.1 SNP and LD	43
	17.2 Family-based vs case-control association studies	43
	17.3 GWAS studies and catalog	43
	17.4 GTEx and eQTL	43
18	GWAS and Epigenomics	45
	18.1 Find tissue / cell type	45
	18.2 Identify causal SNPs and genes	45
	18.3 Predict phenotypes	45

CONTENTS	7
----------	---

19	Sing	ele-cell RNA-seq (1)	47
	19.1	Intro to scRNA-seq	47
	19.2	Smart, Droplet, microwell, SCI-based	47
	19.3	QC	47
	19.4	Normalization	47
	19.5	Imputation	47
	19.6	Dimension reduction	47
	19.7	Clustering	47
	19.8	t-SNE and UMAP	47
20	Sing	ele-cell RNA-seq (2)	49
	20.1	Annotate scRNA-seq clusters	49
	20.2	Differential expression	49
	20.3	Batch effect removal	49
	20.4	Pseudotime	49
	20.5	Overload 10X	49
	20.6	Other applications (CITE-seq, multi-seq, spatial transcriptomics)	49
21		Other applications (CITE-seq, multi-seq, spatial transcriptomics) FAC-seq	49 51
21	\mathbf{scA}^{r}	,	
21	scA'.	ΓAC-seq	51
21	scA ^r . 21.1 21.2	ΓAC-seq Intro to scATAC-seq	51 51
21	scA7 21.1 21.2 21.3	FAC-seq Intro to scATAC-seq	51 51
21	scA7 21.1 21.2 21.3 21.4	FAC-seq Intro to scATAC-seq	51515151
	scA7 21.1 21.2 21.3 21.4 21.5	FAC-seq Intro to scATAC-seq	5151515151
	21.1 21.2 21.3 21.4 21.5 Mod	Intro to scATAC-seq	51 51 51 51 51 51
22	21.1 21.2 21.3 21.4 21.5 Mod 22.1	Intro to scATAC-seq	5151515151515153
22	scA. 21.1 21.2 21.3 21.4 21.5 Mod 22.1 Can	Intro to scATAC-seq	 51 51 51 51 51 51 51 53 53
22	21.1 21.2 21.3 21.4 21.5 Mod 22.1 Can 23.1	Intro to scATAC-seq	 51 51 51 51 51 51 53 53 55
22	scA7. 21.1 21.2 21.3 21.4 21.5 Mod 22.1 Can 23.1 23.2	Intro to scATAC-seq	51 51 51 51 51 51 51 53 53 55 55

8 CONTENTS	
------------	--

	23.5 Interpret tumor mutations	55
	23.6 Find cancer genes	55
	23.7 Summary and future	55
24	Cancer Subtyping, Survival Analyses	57
	24.1 TCGA expression	57
	24.2 Tumor subtypes	57
	24.3 Survival analysis	57
	24.4 GoF Oncogenes and LoF TS $\ \ldots \ \ldots \ \ldots \ \ldots \ \ldots$	57
	24.5 Chromatin regulator mutations in cancer $\dots \dots \dots \dots$	57
	24.6 DNA methylation and CIMP	57
25	Targeted Therapy, Drug Resistance, Compound and Genetic Screens	59
	25.1 Hallmarks of cancer	60
	25.2 Chemo vs targeted the rapy	60
	25.3 Drug resistance	60
	25.4 Synthetic lethality	60
	25.5 Precision medicine	60
	25.6 Tumor (bulk vs scRNA-seq), mice, cell lines	60
	25.7 Compound screens	60
	25.8 Genetic screens	60
	25.9 Tumor heterogeneity	60
2 6	Cancer Immunotherapy (1)	61
	26.1 Systemic immunotherapy	61
	26.2 Personalized immunotherapy	61
	26.3 HLA and neoantigens	61
	26.4 Tumor immune deconvolution	61
	26.5 T cell signaling (PD1/PDL1, etc)	61
	26.6 Other immune-cells (scRNA-seq) $\ \ldots \ \ldots \ \ldots \ \ldots \ \ldots$	61

CONTENTS	9
----------	---

27	Cancer Immunotherapy (2)	63
	27.1 TCR analysis	63
	27.2 BCR analysis	63
	27.3 Microbiome	63
	27.4 Immunotherapy response biomarkers	63
	27.5 Targeted the rapy as immune-modulators	63
	27.6 Epigenetic therapy as immune-modulators	63
28	CRISPR Screens	65
	28.1 CRISPR and KO	65
	28.2 CRISPRa and CRISPRi	65
	28.3 CRISPR design and outcome	65
	28.4 CRISPR screens & DepMap	65
	28.5 CRISPR screen analysis	65
	28.6 CRISPR screens in drug response	65
	28.7 CRISPR screens in immunology	65
	28.8 Enhancer CRISPR screen	65
	28.9 CRISPR screens + scRNA-seq \dots	65
29	Module IV Review and Course Review	67
	29.1 Module IV Review	67
	29.2 Course Review	67

10 CONTENTS

Course information

This is the course material for STAT115/215 BIO/BST282 at Harvard University.

1.1 Contributors

Xiaole Shirley Liu (lead instructor) Joshua Starmer Martin Hemberg Ting Wang Feng Yue Ming Tang etc.

1.2 Supporting staff

Jack Kang Scarlett Ge etc.

Introduction

- 2.1 Welcome
- 2.2 Brief history of bioinformatics
- 2.2.1 Protein structure wave
- 2.2.2 Gene expression wave
- 2.2.3 Genome sequencing wave
- 2.2.4 High throughput sequencing
- 2.2.5 Big data challenge from sequencing
- 2.3 Should I take this course?
- 2.3.1 Bioinformatics vs computational biology
- 2.3.2 Levels of bioinformatics
- 2.3.3 Is this class for me?
- 2.3.4 All biology is computational biology
- 2.4 Course information
- 2.4.1 Course organization and material
- 2.4.2 Course instructor and TAs
- 2.4.3 Homework and grading
- 2.4.4 Lab and Odyssey sign up

High throughput sequencing

3.1 Three generations of sequencing technologies

First generation sequencing is Sanger sequencing. It is the technology that was used to obtain the first human genome sequence.

Second generation sequencing is also called next generation sequencing (NGS) and is the start of high throughput sequencing. It is what scientists use most often nowadays, and Illumina is the market leader. Most of the rest of this course will cover data analysis using second generation sequencing.

Third generation sequencing is single-molecule sequencing. There are many new technologies still under active development, although none has reached market penetration.

3.2 FASTQ and FASTQC

NGS generates FASTQ files. FASTQC is an computational approach to evaluate the quality of your NGS data.

3.3 Early sequence alignment (1 with 1)

In the early days (1970s), scientists were not worried about having to align too many sequences. They wanted to find the best alignment between two sequences. Many bioinformatics courses start with learning these, although it is not the main focus of our course. We included two videos in case you are interested.

The Needlemen-Wunsch algorithm is the earliest algorithm to find the alignment between two sequences and score their similarity.

When two sequences are long, and only a portion of them can align well with each other, the Smith-Waterman algorithm can find the best local sequence alignment. It is still considered the best alignment approach, although it is slow.

3.4 Sequence search algorithms (1 with many)

With more and more sequences available in the public in the 1980s, scientists were interested in finding whether their newly sequenced string has been sequenced before in the public database. Therefore, the fast search algorithm BLAST was developed, using one sequence as the query to find similar sequences from a database.

3.5 Borrow-Wheeler Aligner (many with many)

With NGS, scientists need much faster search (aka mapping) algorithms in order to align the millions of sequences to the reference genome. The current best algorithm is called Borrow-Wheeler Aligner or BWA.

In order to understand BWA, we first need to introduce Borrows-Wheeler transformation and LF mapping

The basic idea of Borrows-Wheeler alignment

3.6 Alignment output

NGS raw data is in FASTQ. Alignment gives you SAM (alignment) or BAM (binary version of SAM) files which contain the sequence information in FASTQ and the mapping locations. BED file is the simpliest, although there is information loss.

RNA-seq quantification

- 4.1 RNA-seq & RNA QC (DV200)
- 4.2 RNA-seq replicates and study design
- 4.3 STAR and PE BAM file
- 4.4 RNA-seq QC
- 4.5 FPKM vs TPM
- 4.6 RSEM & Salmon

Differential expression, False discovery rate, Gene ontology

- 5.1 RNA-seq NB distribution
- 5.2 DESeq2 and variance stabilization
- 5.3 Multiple hypotheses testing and FWER
- 5.4 FDR
- 5.5 GO

 $20 CHAPTER \ 5. \ DIFFERENTIAL \ EXPRESSION, FALSE \ DISCOVERY \ RATE, \ GENE \ ONTOLOGY$

GSEA, Clustering

- 6.1 **GSEA**
- 6.2 Heatmap and clustering quality
- 6.3 H-cluster
- 6.4 K-means
- 6.5 Pick K and consensus clustering
- 6.6 Batch effect removal

Dimension Reduction

- 7.1 MDS
- 7.2 LDA
- 7.3 PCA

Classification

- 8.1 Intro to machine learning
- 8.2 Cross validation
- 8.3 Regression
- 8.4 Regularization
- 8.5 KNN
- 8.6 Decision trees
- 8.7 Random forest
- 8.8 SVM

Module I Review

- 9.1 Gene Expression Module Summary
- 9.2 Gene Expression Analysis Scenarios

Transcription Factor Motif Finding

- 10.1 Transcription regulation
- 10.2 Motif representation
- 10.3 EM
- 10.4 Gibbs sampler
- 10.5 Gibbs intuition
- 10.6 Motif finding in eukaryotes
- 10.7 Known motif database

ChIP-seq, Expression Integration

- 11.1 ChIP-seq
- 11.2 BWA and MACS
- 11.3 ChIP-seq QC
- 11.4 TF interactions (motif)
- 11.5 TF target genes (expression integration)

Epigenetics, DNA Methylation

- 12.1 Epigenetics
- 12.2 DNA methylation
- 12.3 Promoter function
- 12.4 Gene body function
- 12.5 Enhancer function
- 12.6 Repetitive region function
- 12.7 Early cancer detection

Histone Modifications, Chromatin Accessibility

- 13.1 Nucleosome positions
- 13.2 Histone modification
- 13.3 Promoters (bivalent)
- 13.4 Genes (K36me3, new genes)
- 13.5 Enhancers (K27ac)
- 13.6 Super-enhancers
- 13.7 DNase-seq
- 13.8 ATAC-seq

Long Range Chromatin Interactions

- 14.1 Chromatin interactions
- 14.2 HiC
- 14.3 HiC contact map
- 14.4 HiC normalization
- 14.5 Fractal globule
- **14.6** Loops
- 14.7 Domains
- 14.8 Compartments
- 14.9 Phase separation

Hidden Markov Model

- 15.1 Intro to HMM
- 15.2 Pb1: Forward & backward procedure
- 15.3 Pb2: Viterbi algorithm
- 15.4 Pb3: Parameter estimation
- 15.5 HMM application

Module II Review

- 16.1 Module II Review
- 16.2 Practive Questions

SNP and GWAS

- 17.1 SNP and LD
- 17.2 Family-based vs case-control association studies
- 17.3 GWAS studies and catalog
- 17.4 GTEx and eQTL

GWAS and Epigenomics

- 18.1 Find tissue / cell type
- 18.2 Identify causal SNPs and genes
- 18.3 Predict phenotypes

Single-cell RNA-seq (1)

- 19.1 Intro to scRNA-seq
- 19.2 Smart, Droplet, microwell, SCI-based
- 19.3 QC
- 19.4 Normalization
- 19.5 Imputation
- 19.6 Dimension reduction
- 19.7 Clustering
- 19.8 t-SNE and UMAP

Single-cell RNA-seq (2)

- 20.1 Annotate scRNA-seq clusters
- 20.2 Differential expression
- 20.3 Batch effect removal
- 20.4 Pseudotime
- 20.5 Overload 10X
- 20.6 Other applications (CITE-seq, multi-seq, spatial transcriptomics)

scATAC-seq

- 21.1 Intro to scATAC-seq
- 21.2 Sample and cell QC
- 21.3 Dimension reduction, clustering & visualization
- 21.4 Differential peaks and annotations
- 21.5 Integration with scRNA-seq

Module III Review

22.1 Module III Review

Cancer Genome Sequencing, Mutation analyses

- 23.1 Intro to TCGA
- 23.2 Cancer mutation characterization
- 23.3 Cancer mutation patterns
- 23.4 Tumor purity and clonality
- 23.5 Interpret tumor mutations
- 23.6 Find cancer genes
- 23.7 Summary and future

Cancer Subtyping, Survival Analyses

- 24.1 TCGA expression
- 24.2 Tumor subtypes
- 24.3 Survival analysis
- 24.4 GoF Oncogenes and LoF TS
- 24.5 Chromatin regulator mutations in cancer
- 24.6 DNA methylation and CIMP

Targeted Therapy, Drug Resistance, Compound and Genetic Screens

- 25.1 Hallmarks of cancer
- 25.2 Chemo vs targeted therapy
- 25.3 Drug resistance
- 25.4 Synthetic lethality
- 25.5 Precision medicine
- 25.7 Compound screens
- 25.8 Genetic screens
- 25.9 Tumor heterogeneity

Cancer Immunotherapy (1)

- 26.1 Systemic immunotherapy
- 26.2 Personalized immunotherapy
- 26.3 HLA and neoantigens
- 26.4 Tumor immune deconvolution
- 26.5 T cell signaling (PD1/PDL1, etc)
- 26.6 Other immune-cells (scRNA-seq)

Cancer Immunotherapy (2)

- 27.1 TCR analysis
- 27.2 BCR analysis
- 27.3 Microbiome
- 27.4 Immunotherapy response biomarkers
- 27.5 Targeted therapy as immune-modulators
- 27.6 Epigenetic therapy as immune-modulators

CRISPR Screens

28.1	CRISPR	and KC
40.1	CUISEL	and NU

- 28.2 CRISPRa and CRISPRi
- 28.3 CRISPR design and outcome
- 28.4 CRISPR screens & DepMap
- 28.5 CRISPR screen analysis
- 28.6 CRISPR screens in drug response
- 28.7 CRISPR screens in immunology
- 28.8 Enhancer CRISPR screen
- 28.9 CRISPR screens + scRNA-seq

Module IV Review and Course Review

- 29.1 Module IV Review
- 29.2 Course Review