

Catalogue of offering - transcriptome

01 December 2025
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Cat No	Bulk Transcriptome	Description and application	Typical applications	Current Capability	Platform	Data	Pre-Project engagement (Optional)	Sample requirements	KeyQC metrics (Data Analysis)	BOM	Analysis Scope	Link to the sample report	Standard Deliverables	Tool versions	Estimated Time
	RNA-seq Eukaryotic poly A model (reference available)	<p>Pros High sensitivity profiling of gene expression changes; sensitive of gene expression with changes gene are in high expression Gene expression over broad dynamic range Suitable for studies focusing on cell-line changes in treatment studies and scale assays is a cost assay to profile differential Can be applied where RNA is limiting and low sample inputs</p> <p>Limitations Profiles the gene expression is gene transcripts captured with a PolyA; non -poly A transcripts not profiled RNA integrity should be high and consistent > RIN 7</p>	Evaluate differential gene expression for treatment Profile genes for perturbations on treatments	Yes	Illumin 150X2	17million* 2 (currently we are asking for 25 millions* 2) is that required?	Sample shipping and preparation on SOP Power analysis for RNA-seq Design review analysis	Number of cells - specify range Amount of of tissue - amount Sample type - Cells, Tissue frozen; Tissue in RNA-later; FFPE Extracted RNA > conc; Quality	Phred Score >30, % of reads retained after trimming >80 % %, % of reads belonging to host > 95%, Mapping rate >95%, Exonic rate > 80%	Sample extraction (12) Sample QC (18) Library prep A (Components) (6 /12) Library QC (14) Sequencing - 6 / 12 *17 million reads Data handling and storage Time for data analysis	Primary processing - FastQC, check for data quality and Pre-processing reads and adapter removal Reference alignment, mapped count file generation Typical comparisons Type 1 - pair of samples controls vs treatment Type 2 - Groups of controls vs treatments Type 3 - Control - treatment vs Modified - treatment (four samples) Data normalization, Differential gene expression	https://workdrive.zohoexternal.com/external/be95ce22e70c96bf19b7f0f391625182e3353a2d8727db6dc90b833613a1249	<p>Raw & QC Trimmed data (.fastq files) and QC reports per sample</p> <p>Mapped data (.BAM) files sample and related QC reports</p> <p>Gene expression matrix (.csv/xlsx format) : Raw counts, Normalized counts</p> <p>Sample clustering reports</p> <p>Differential gene expression report</p> <p>Functional enrichment reports (Gene ontology – Biological process, Cellular Component, Molecular Function & KEGG/REACTOME pathways)</p> <p>Supporting visualization each step (PCA/UMAP plots, Heatmaps, Volcano plots, Functional enrichment gradient plots, KEGG pathview maps*)</p> <p><i>*KEGG Pathview maps for pathways selected according to the interest of the study</i></p>	<p>1.Fastqc - FastQC v0.11.9</p> <p>2.multiQC - multiqc, version 1.27.1</p> <p>3.Kraken - Kraken 1.1.1</p> <p>4.Cutadapt - 5.0</p> <p>5.STAR - 2.7.11b</p> <p>6.Samtools - samtools 1.21</p> <p>7.Qualimap - QualiMap v.2.3</p> <p>8.Rseqc - RSeQC-5.0.1</p> <p>9.FeatureCounts - Version 2.1.1</p> <p>10.Python - Python 3.12.2</p> <p>11.R - version 4.4.1</p> <p>###Downstream Analysis</p> <p>12.DESeq2 - 1.46.0</p> <p>13.edgeR - 4.4.2</p> <p>14.ClustalPro - 4.14.6</p> <p>15.GSEABase - 1.68.0</p> <p>16.fgsea - 1.32.4</p> <p>17.ggplot2 - 4.0.1</p> <p>18.pheatmap 1.0.13</p> <p>19.Cytoscape 3.10.3</p>	Hand on-4 days Machine Run-1 week (based on sample count - n=30-40)
	Eukaryotic poly A non-model (reference with denovo) [What is the requirement of reference with denovo?]	<p>Pros Exploratory profiling to create a transcriptome and evaluate gene expression profiles for a fungal/plant/mo del organism with limited reference genome information in data bases High sensitivity profiling of gene expression changes; sensitive of gene expression with changes gene are in high expression Gene expression</p>	Profile and generate a first reference transcriptome, when a reference is not readily available First line investigation of host-pathogen interactions; where the host OR pathogen reference genome is available Typically suitable for fungal transcriptome where there is considerable strain divergence		Illumin 150X2	25million* 2					Ref alignment. DGE, pathway analysis				

		over broad dynamic range Suitable for studies focusing on cell-line changes in treatment studies and scale assays is a cost assay to profile differential Can be applied where RNA is limiting and low sample inputs Limitations Profiles the gene expression is gene transcripts captured with a PolyA; non -poly A transcripts not profiled RNA integrity should be high and consistent > RIN 7												
	Total RNAseq human/mouse/rat	Pros Deeper transcriptome profiling where there is interest in the total transcript complement to include the transcripts lacking poly A; Suitable to studies where total transcript signature is needed, that include circular RNA RNA is lower quality and is degraded partially with RIN > 4 Limitations Requires higher read depth to compensate for ribosomal and structure RNA carry over post ribodepletion - Needs for 20-30% higher read generated to achieve the same coverage as polyA based library for core gene set RNA requirement in quantity needed to support ribodepletion Higher input costs in the overall of read generation and analysis	Total transcript profiling to when interest is in non-polyA transcripts, circular RNA RNA is low RIN and cannot be processed using polyA methods	Yes	ILLumin 150X2	30million* 2 [current depth is 25*2 - why the increase in depth?		Phred Score >30, % of reads retained after trimming >80 %, % of reads belonging to host > 95%, Mapping rate >95%, Exonic rate > 30%		Ref alignment. DGE, pathway analysis, lncRNA, circRNA	https://workdrive.zohoexternal.com/external/a0cf74094cac2ae0575113ccc9898796c301565c83fac421bc8325b447e347	1) Raw Data and Raw Data counts across all samples. 2) Normalized read count for each samples along with Sample Grouping/Clustering and QC related plots : PCA plot, Heatmap based on top 2000 variant genes, dendrogram and Sample-Sample correlation plot. 3) Differential expression reports for each comparison along with corresponding visualization – Volcano plots. 4) Functional over-representation (enrichment) analysis per comparison, along with corresponding visualization – Gradient Bar plots for top 20 enriched GO: BP, MF, CC and Reactome Pathways. 5) Raw Count and Normalized count of lncRNA for each samples along with Sample Grouping/Clustering and QC related plots : PCA plot, Heatmap on top 2000 variant genes, dendrogram and Sample-Sample correlation plot.	1.Fastqc - FastQC v0.11.9 2.multiQC - multiqc, version 1.27.1 3.Kraken - 1.1.1 4.Cutadapt - 5.0 5.STAR - 2.7.11b 6.Samtools - samtools 1.21 7.Qualimap - QualiMap v.2.3 8.Rseqc - RSeQC-5.0.1 9.FeatureCounts - Version 2.1.1 10.Python - Python 3.12.2 11.R - version 4.4.1 ###Downstream Analysis 12.DESeq2 - 1.46.0 13.edgeR - 4.4.2 14.ClustalPro - 4.14.6 15.GSEABase - 1.68.0 16.fgsea - 1.32.4 17.ggplot2 - 4.0.1 18.pheatmap 1.0.13 19.Cytoscape 3.10.3 CircRNA STAR version 2.7.11b circExplorer2 2.3.8 CircRNAprofilr = 1.20.0 Deseq2 = 1.46.0 Clusterprofile = 4.14.6	Hands on time - 1-2 weeks and Machine time - 1 week

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		RNA Discover miRNA-gene networks Limitations NGS methods are sampling - some species may not be detected non-target miRNA may also be detected that may increase False discovery	might be required for discovery?]																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																								</
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													expression reports for each comparison along with corresponding visualization – Volcano plots. 5)Validated targets of Known miRNA identified by miRTarbase 6)Co-expressed * targets of Known miRNA identified by Co-expression analysis (Pearson correlation) [Possible if mRNAseq data available for same set of samples*] 7)Functional over-representation (enrichment) analysis per comparison, along with corresponding visualization – Dot plots for top 20 enriched GO: BP, MF, CC and Reactome Pathways. 8) Coexpression network for miRNA and mRNA (Validated and Co-expressed)"			
	miRNAseq-mRNA integrated analysis non-human	Pro Profile small and RNA expression - measure differential expression profiles Discover miRNA-gene networks and report perturbations Limitations non-target miRNA may also be detected that may increase False discovery miRNA - RNA is a computed network Some miRNA many not be annotated and could be due to current state of information during analysis of data	Discovery and analysis of gene regulatory-networks; combines the analysis of differential RNA analysis and RNA in a sample. Often can be used to interrogate group of controls vs treatments to map perturbations	Yes	Illumin 150X2											
	Targeted fusion Seq (NGS)	Pros Targets specific RNA fusions and can achieve sensitive detection Limitations Investigation is limited to the targeted content	Detection of tumour fusion RNA transcripts Analyse presence of fusions from bulk RNA derived from tumours		Illumin 150X2											
	Bulk repertoire sequencing Bcell/Tcell (Lonf read)	Pros Profiles the antibody VH and VL or TCR heavy and light chains Limitations Depth of reads needed to saturate repertoire	Profiling immune repertoires for antibody and TCR diversity and clonal lineages Assessing repertoires pre-post immunization/treatment/infections		ONT											

RNA seq denovo/gen e annotation, pathway mining and metabolite mapping; Gene ranking and query														
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