Class 7: Machine Learning 1

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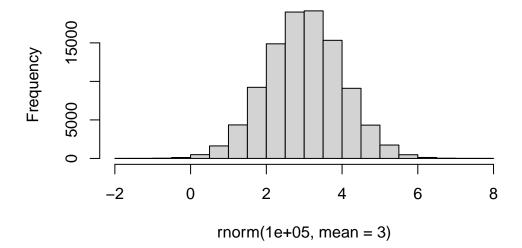
Today we will start our multi-part exploration of some key machine leaning methods. We will begin with clustering - finding groupings in data, and then dimensionallity reduction.

Clustering

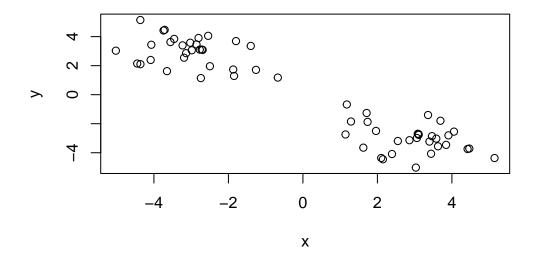
Let's start with "k-means" clustering. The main function in base R for this kmeans().

```
# Make up some data
hist(rnorm(100000, mean=3) )
```

Histogram of rnorm(1e+05, mean = 3)



```
tmp <- c(rnorm(30, -3), rnorm(30, +3))
x <- cbind(x=tmp, y=rev(tmp))
plot(x)</pre>
```



Now let's try kmeans()

```
km <- kmeans(x, centers=2)
km</pre>
```

K-means clustering with 2 clusters of sizes 30, 30

Cluster means:

```
x y
1 2.949726 -3.016980
2 -3.016980 2.949726
```

Clustering vector:

Within cluster sum of squares by cluster:

[1] 61.53043 61.53043 (between_SS / total_SS = 89.7 %)

Available components:

- [1] "cluster" "centers" "totss" "withinss" "tot.withinss"
- [6] "betweenss" "size" "iter" "ifault"

attributes(km)

\$names

[1] "cluster" "centers" "totss" "withinss" "tot.withinss"

[6] "betweenss" "size" "iter" "ifault"

\$class

- [1] "kmeans"
 - Q1. How many points in each cluster?

km\$size

- [1] 30 30
 - Q2. What component of your resut object detils cluster assignment/membership?

km\$cluster

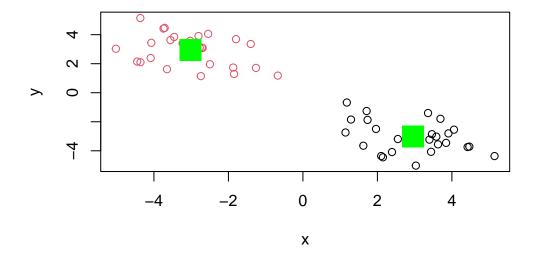
- - Q3. What are the centers/mean values of each cluster?

km\$centers

X

- 1 2.949726 -3.016980
- 2 -3.016980 2.949726
 - Q4. Make a plot of your data showing your clustering results.

```
plot(x, col=km$cluster)
points(km$centers, col="green", pch=15, cex=3)
```

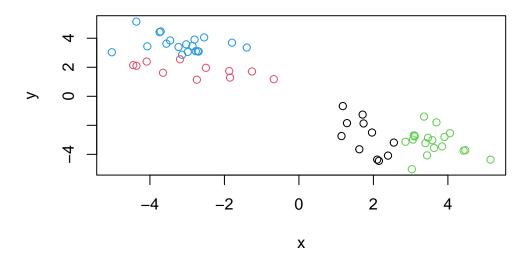


$$c(1:5) + 100$$

[1] 101 102 103 104 105

Q5. Run kmeans() again and cluster in 4 groups and plot the results.

```
km4 <- kmeans(x, centers=4)
plot(x, col=km4$cluster)</pre>
```



Hierarchical Clustering

This form of clustering aims to reveal the structure in your data by progressively grouping points into a ever smaller number of clusters.

The main function in base R for this called hclust(). This function does not take our input data directly but wants a "distance matrix" that details how (dis)similar all our input points are to each other.

```
hc <- hclust(dist(x))
hc</pre>
```

Call:

hclust(d = dist(x))

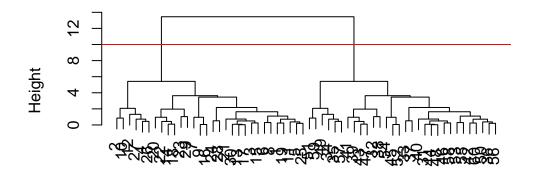
 $\begin{array}{lll} \hbox{\tt Cluster method} & : & \hbox{\tt complete} \\ \hbox{\tt Distance} & : & \hbox{\tt euclidean} \end{array}$

Number of objects: 60

The print out above is not very useful (unclick that from kmeans) but there is a useful plot() method.

```
plot(hc)
abline( h=10, col="red")
```

Cluster Dendrogram

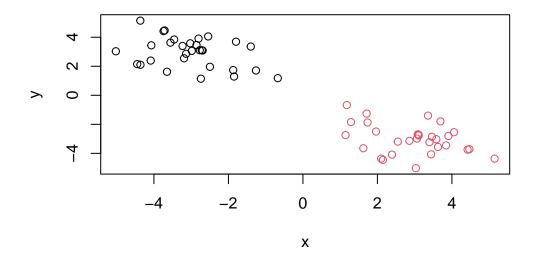


dist(x)
hclust (*, "complete")

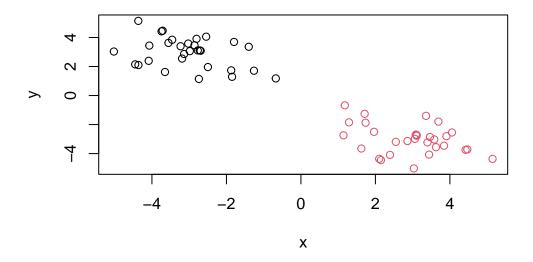
To get my main result (my cluster embership vector) I need to "cut" my tree using the function cutree()

```
grps <- cutree(hc, h=10)
grps</pre>
```

```
plot(x, col=grps)
```



plot(x, col=cutree(hc, h=6))



Principal Component Analysis (PCA)

The goal of PCA is to reduce the dimensionality of a dataset down to some smaller subset of new variables (called PCs) that are a useful bases for further analysis, like visualization, clustering, etc.

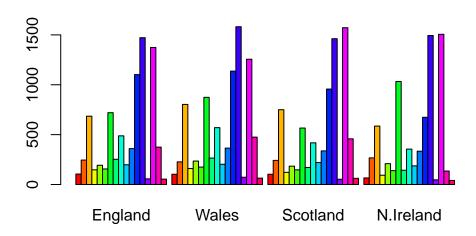
Dara Import

Read data about crazy eating trends in the UK and N. Ireland

```
url <- "https://tinyurl.com/UK-foods"
x <- read.csv(url, row.names = 1)
x</pre>
```

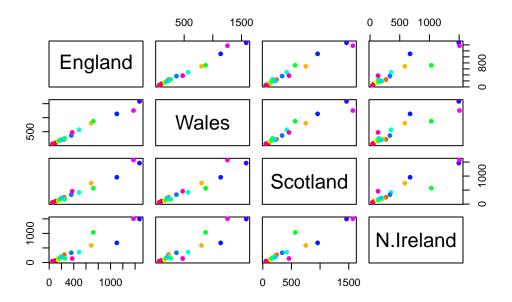
	England	Wales	${\tt Scotland}$	N.Ireland
Cheese	105	103	103	66
Carcass_meat	245	227	242	267
Other_meat	685	803	750	586
Fish	147	160	122	93
Fats_and_oils	193	235	184	209
Sugars	156	175	147	139
Fresh_potatoes	720	874	566	1033
Fresh_Veg	253	265	171	143
Other_Veg	488	570	418	355
Processed_potatoes	198	203	220	187
Processed_Veg	360	365	337	334
Fresh_fruit	1102	1137	957	674
Cereals	1472	1582	1462	1494
Beverages	57	73	53	47
Soft_drinks	1374	1256	1572	1506
Alcoholic_drinks	375	475	458	135
Confectionery	54	64	62	41

```
barplot(as.matrix(x),beside=2, col=rainbow(nrow(x)))
```



The so-called "pairs" plot can be useful for small datasets:

```
#rainbow(nrow(x))
pairs(x, col=rainbow(nrow(x)),pch=16 )
```



So the paris plot is useful for small datasets but it can be lots of work to interpret and gets intractable for larger datasets.

So PCA to the rescue...

The main function to do PCA in base R is called prcomp(). This function wants the transpose of our data in this case.

```
pca <- prcomp(t(x))
summary(pca)</pre>
```

Importance of components:

```
attributes(pca)
```

\$names

[1] "sdev" "rotation" "center" "scale" "x"

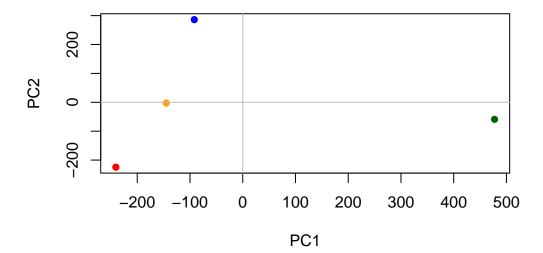
```
$class
[1] "prcomp"
```

```
pca$x
```

```
PC1
                             PC2
                                        PC3
                                                      PC4
          -144.99315
                       -2.532999 105.768945 -4.894696e-14
England
Wales
          -240.52915 -224.646925 -56.475555
                                            5.700024e-13
Scotland
                      286.081786 -44.415495 -7.460785e-13
           -91.86934
N.Ireland 477.39164
                      -58.901862
                                 -4.877895
                                            2.321303e-13
```

A major PCA result viz is called a "PCA plot" (a.k.a a score plot, biplot, PC1 vs PC2 plot, ordination plot)

```
mycols <- c("orange", "red", "blue", "darkgreen")
plot(pca$x[,1], pca$x[,2], col=mycols, pch=16,
    xlab="PC1", ylab="PC2")
abline(h=0, col="grey")
abline(v=0, col="grey")</pre>
```



Another important output from PCA is called "loadings" vector or the "rotation" component - this tells us how much the original variables (the food in this case) contribute to the new PCs

pca\$rotation

	PC1	PC2	PC3	PC4
Cheese	-0.056955380	0.016012850	0.02394295	-0.694538519
Carcass_meat	0.047927628	0.013915823	0.06367111	0.489884628
Other_meat	-0.258916658	-0.015331138	-0.55384854	0.279023718
Fish	-0.084414983	-0.050754947	0.03906481	-0.008483145
Fats_and_oils	-0.005193623	-0.095388656	-0.12522257	0.076097502
Sugars	-0.037620983	-0.043021699	-0.03605745	0.034101334
Fresh_potatoes	0.401402060	-0.715017078	-0.20668248	-0.090972715
Fresh_Veg	-0.151849942	-0.144900268	0.21382237	-0.039901917
Other_Veg	-0.243593729	-0.225450923	-0.05332841	0.016719075
Processed_potatoes	-0.026886233	0.042850761	-0.07364902	0.030125166
Processed_Veg	-0.036488269	-0.045451802	0.05289191	-0.013969507
Fresh_fruit	-0.632640898	-0.177740743	0.40012865	0.184072217
Cereals	-0.047702858	-0.212599678	-0.35884921	0.191926714
Beverages	-0.026187756	-0.030560542	-0.04135860	0.004831876
Soft_drinks	0.232244140	0.555124311	-0.16942648	0.103508492
Alcoholic_drinks	-0.463968168	0.113536523	-0.49858320	-0.316290619
Confectionery	-0.029650201	0.005949921	-0.05232164	0.001847469

PCA looks to be a super useful method for gaining some insight into high dimensional data that is difficult to examine in other ways.

PCA of RNASeq data

```
url2 <- "https://tinyurl.com/expression-CSV"</pre>
  rna.data <- read.csv(url2, row.names=1)</pre>
  head(rna.data)
       wt1 wt2
                wt3 wt4 wt5 ko1 ko2 ko3 ko4 ko5
gene1
       439 458
                408
                     429 420
                               90
                                  88
                                      86
                                           90
                                               93
gene2 219 200
                204
                    210 187 427 423 434 433 426
gene3 1006 989 1030 1017 973 252 237 238 226 210
gene4 783 792
                829 856 760 849 856 835 885 894
```

```
gene5 181 249 204 244 225 277 305 272 270 279
gene6 460 502 491 491 493 612 594 577 618 638
  ##Again we have to take the transpose of our data
  pca <- prcomp(t(rna.data), scale=TRUE)</pre>
  summary(pca)
Importance of components:
                          PC1
                                 PC2
                                         PC3
                                                  PC4
                                                          PC5
                                                                  PC6
                                                                          PC7
Standard deviation
                       9.6237 1.5198 1.05787 1.05203 0.88062 0.82545 0.80111
Proportion of Variance 0.9262 0.0231 0.01119 0.01107 0.00775 0.00681 0.00642
Cumulative Proportion 0.9262 0.9493 0.96045 0.97152 0.97928 0.98609 0.99251
                           PC8
                                   PC9
                                             PC10
                       0.62065 0.60342 3.457e-15
Standard deviation
Proportion of Variance 0.00385 0.00364 0.000e+00
Cumulative Proportion 0.99636 1.00000 1.000e+00
     Q. How many genes in this dataset?
  nrow(rna.data)
[1] 100
  attributes(pca)
$names
[1] "sdev"
               "rotation" "center"
                                      "scale"
$class
[1] "prcomp"
  pca$x
          PC1
                     PC2
                                PC3
                                            PC4
                                                       PC5
                                                                  PC6
wt1 -9.697374 1.5233313 -0.2753567 0.7322391 -0.6749398
                                                           1.1823860
wt2 -9.138950  0.3748504  1.0867958 -1.9461655  0.7571209 -0.4369228
```

```
wt3 -9.054263 -0.9855163 0.4152966 1.4166028 0.5835918 0.6937236
wt4 -8.731483 -0.7468371 0.5875748 0.2268129 -1.5404775 -1.2723618
wt5 -9.006312 -0.2945307 -1.8498101 -0.4303812 0.8666124 -0.2496025
    8.846999 2.2345475 -0.1462750 -1.1544333 -0.6947862 0.7128021
    9.213885 -3.2607503 0.2287292 -0.7658122 -0.4922849
                                                    0.9170241
    9.458412 -0.2636283 -1.5778183 0.2433549 0.3654124 -0.5837724
    8.883412 0.6339701 1.5205064 0.7760158 1.2158376 -0.1446094
ko5
    9.225673 0.7845635 0.0103574 0.9017667 -0.3860869 -0.8186668
                     PC8
                                PC9
          PC7
                                           PC10
wt1 -0.24446614
              1.03519396
                         0.07010231 3.073930e-15
wt2 -0.03275370  0.26622249  0.72780448  1.963707e-15
wt3 -0.03578383 -1.05851494 0.52979799 2.893519e-15
wt4 -0.52795595 -0.20995085 -0.50325679 2.872702e-15
wt5 0.83227047 -0.05891489 -0.81258430 1.693090e-15
ko1 -0.07864392 -0.94652648 -0.24613776 4.052314e-15
ko2 0.30945771 0.33231138 -0.08786782 3.268219e-15
ko4 -0.35073859 0.30381920 -0.87353886 3.615164e-15
   1.56584821
```

head(pca\$x)

```
PC1
                  PC2
                            PC3
                                       PC4
                                                 PC5
                                                          PC6
wt1 -9.697374
            1.5233313 -0.2753567 0.7322391 -0.6749398
                                                    1.1823860
wt2 -9.138950  0.3748504  1.0867958 -1.9461655  0.7571209 -0.4369228
wt4 -8.731483 -0.7468371 0.5875748 0.2268129 -1.5404775 -1.2723618
wt5 -9.006312 -0.2945307 -1.8498101 -0.4303812 0.8666124 -0.2496025
ko1 8.846999 2.2345475 -0.1462750 -1.1544333 -0.6947862 0.7128021
          PC7
                     PC8
                                PC9
                                           PC10
wt1 -0.24446614 1.03519396 0.07010231 3.073930e-15
wt2 -0.03275370 0.26622249 0.72780448 1.963707e-15
wt3 -0.03578383 -1.05851494 0.52979799 2.893519e-15
wt4 -0.52795595 -0.20995085 -0.50325679 2.872702e-15
wt5 0.83227047 -0.05891489 -0.81258430 1.693090e-15
ko1 -0.07864392 -0.94652648 -0.24613776 4.052314e-15
```

I will make a main result figure use ggplot:

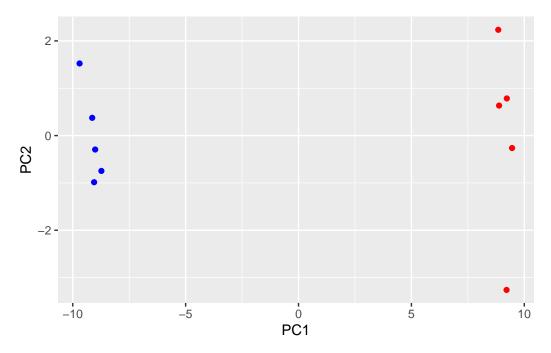
```
library(ggplot2)
```

```
res <- as.data.frame(pca$x)

mycols <- c(rep("blue", 5), rep("red", 5))
mycols

[1] "blue" "blue" "blue" "blue" "blue" "red" "red" "red" "red"

ggplot(res) +
   aes(x=PC1, y=PC2, label=row.names(res)) +
   geom_point(col=mycols)</pre>
```



```
colnames(rna.data)
```

```
[1] "wt1" "wt2" "wt3" "wt4" "wt5" "ko1" "ko2" "ko3" "ko4" "ko5" kmeans(pca$x[,1], centers =2)
```

 $K ext{-means}$ clustering with 2 clusters of sizes 5, 5

Cluster means:

[,1]

1 -9.125676

2 9.125676

Clustering vector:

wt1 wt2 wt3 wt4 wt5 ko1 ko2 ko3 ko4 ko5 1 1 1 1 1 2 2 2 2 2

Within cluster sum of squares by cluster:

[1] 0.5017505 0.2648467

(between_SS / total_SS = 99.9 %)

Available components:

[1] "cluster" "centers" "totss" "withinss" "tot.withinss"

[6] "betweenss" "size" "iter" "ifault"