

What is “Genetic Draft”?

It's not a fundamental “force” like mutation, selection, and drift.

It's an **effect** of mutation at a selected locus, that reduces variation at nearby (linked) loci, thereby reducing the **apparent N**.

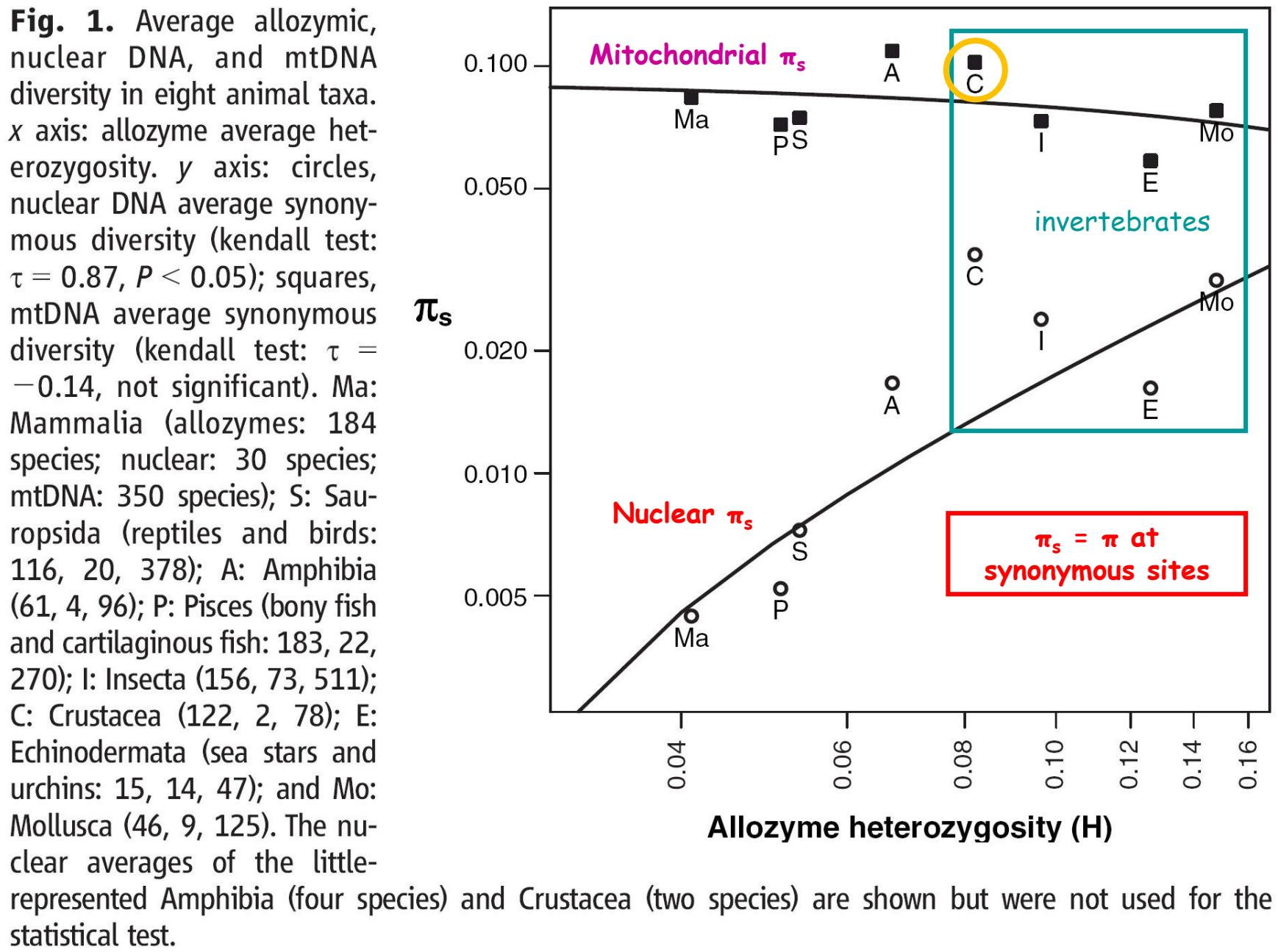
Why should we care?

Neutral theory predicts that at **mutation-drift equilibrium**, DNA polymorphism should be proportional to N:

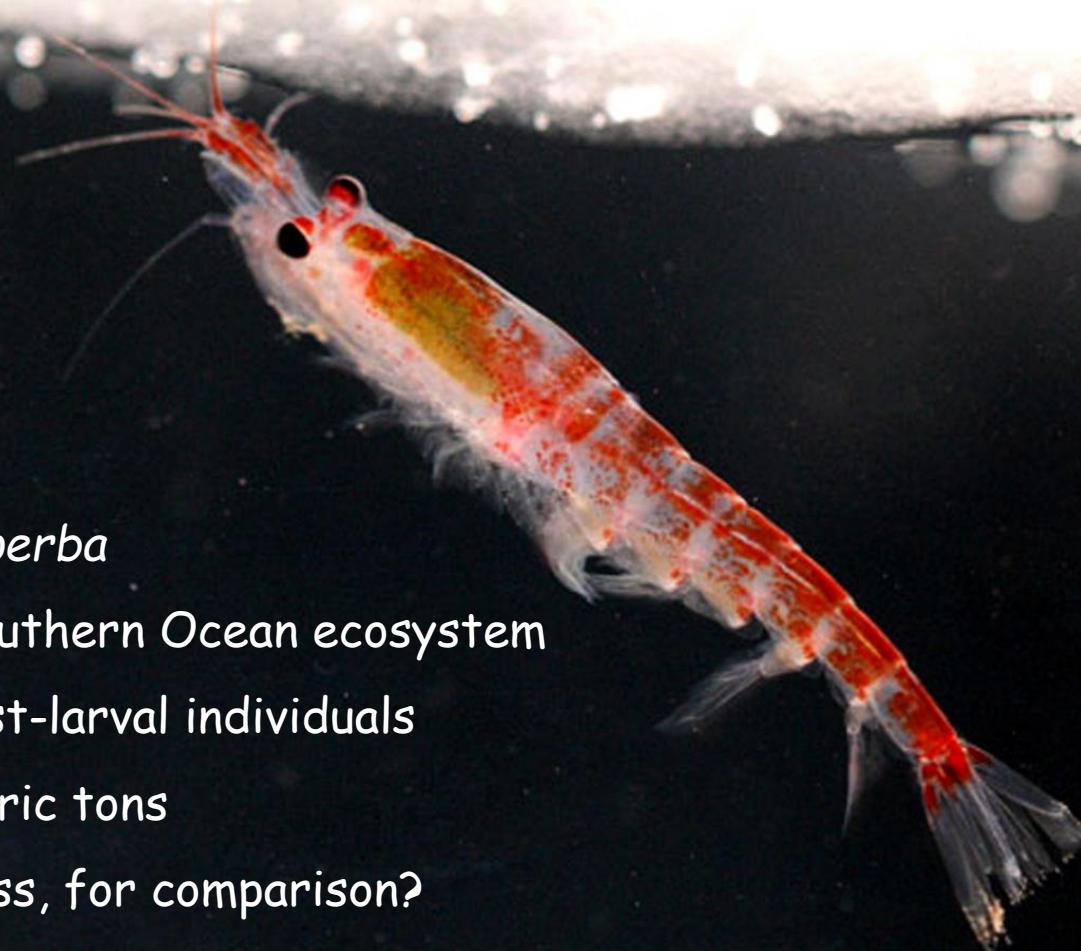
$$\pi \approx \Theta \approx 4N\mu \quad (\text{at a diploid locus})$$

But it's **not!** (Not even approximately, especially for mitochondria.)

If we can figure out **why** the neutral theory fails, we'll learn something important about genetics and ecology (i.e., mutation and selection).



The most abundant animal on Earth?



Euphausia superba

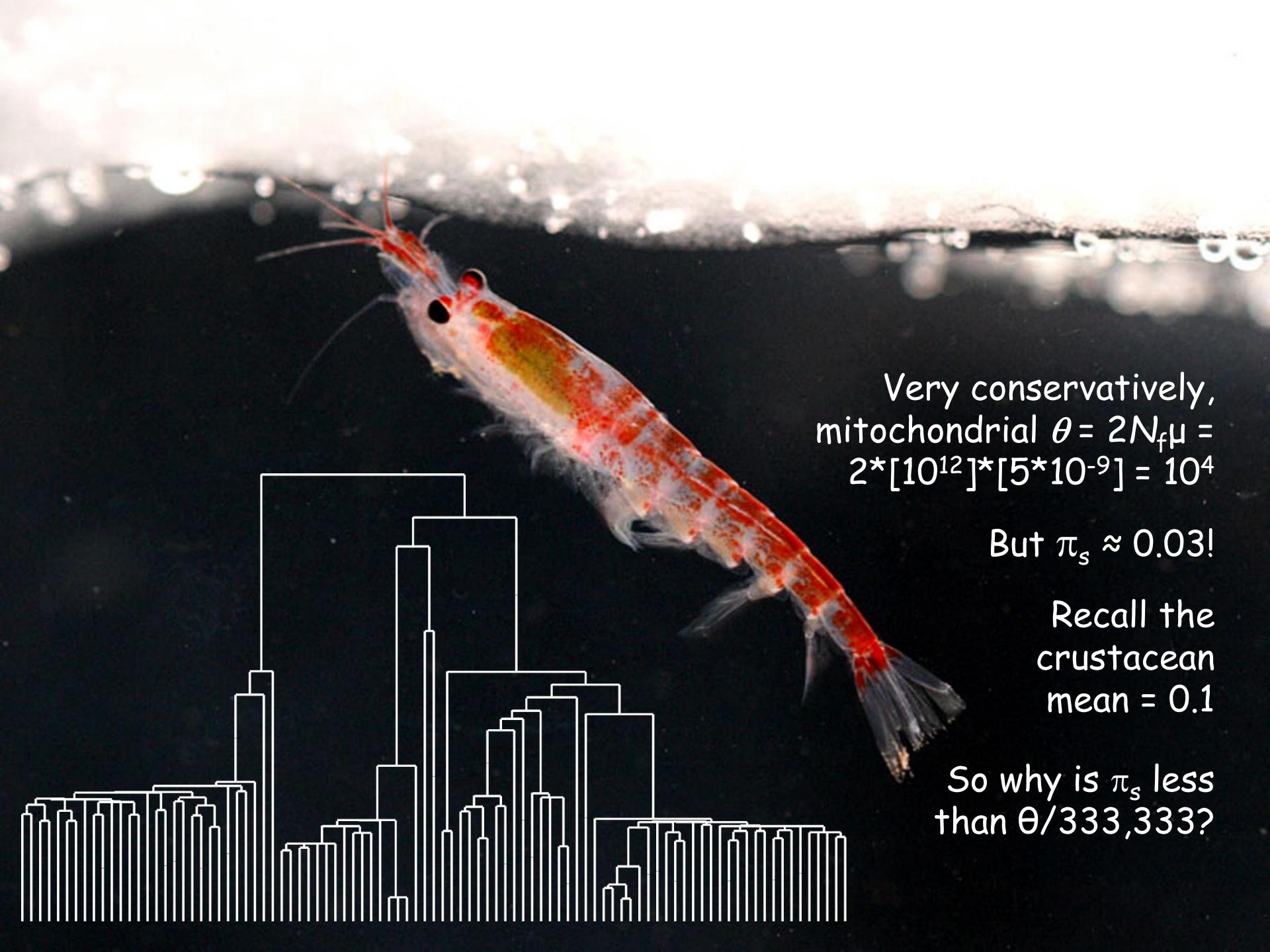
Key link in Southern Ocean ecosystem

$10^{14} - 10^{15}$ post-larval individuals

$1-4 \times 10^8$ metric tons

Human biomass, for comparison?

$7 \times 10^9 \times 0.04$ m.t. = 2.8×10^8 m.t.



Very conservatively,
mitochondrial $\theta = 2N_f\mu =$
 $2*[10^{12}]*[5*10^{-9}] = 10^4$

But $\pi_s \approx 0.03!$

Recall the
crustacean
mean = 0.1

So why is π_s less
than $\theta/333,333?$

The draft model: Locus "A" is selected, locus "B" is neutral

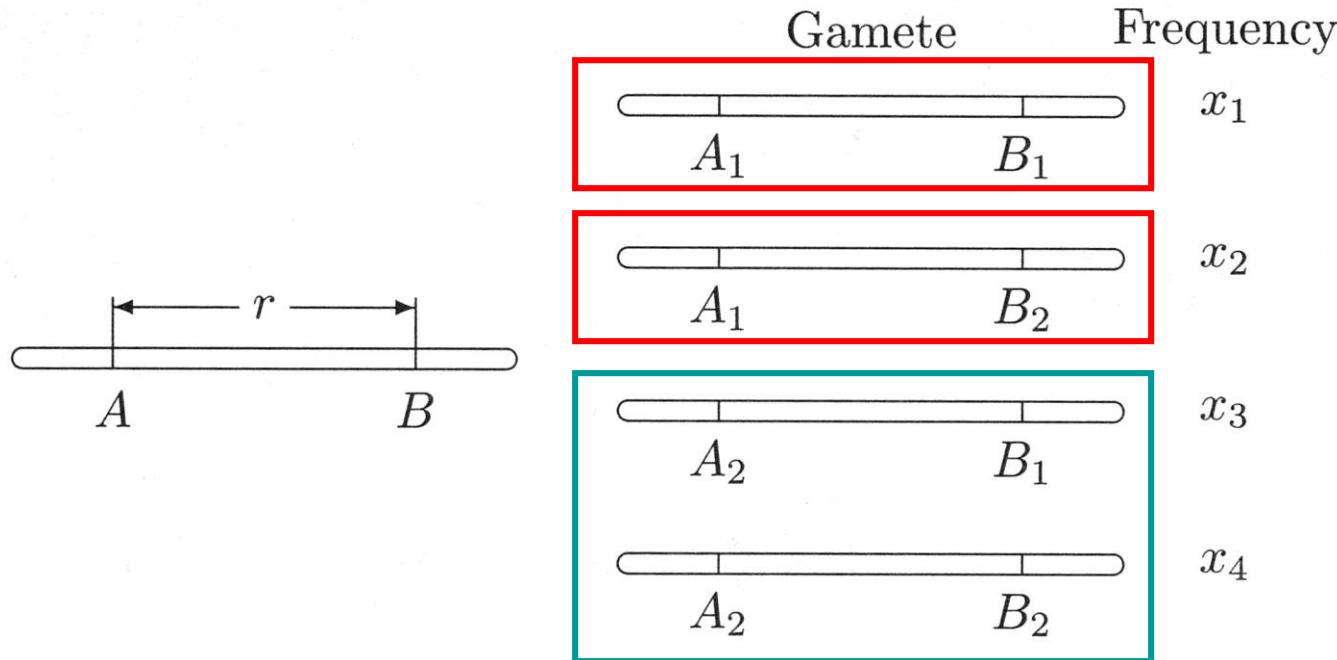


Figure 4.1: The chromosome on the left shows the position of the A and B loci. The right side illustrates the four possible gametes with their frequencies.

Step 0: The population is fixed for A_2 , polymorphic for B_1 and B_2 .

Frequency of $B_1 = p_B$, frequency of $B_2 = q_B = (1-p_B)$.

Step 1: A mutation to the selectively favored allele A_1 occurs.

But on which genetic background?

B_1 ?

B_2 ?

Step 2: The lucky B-allele “hitches a ride” with A_1 ...

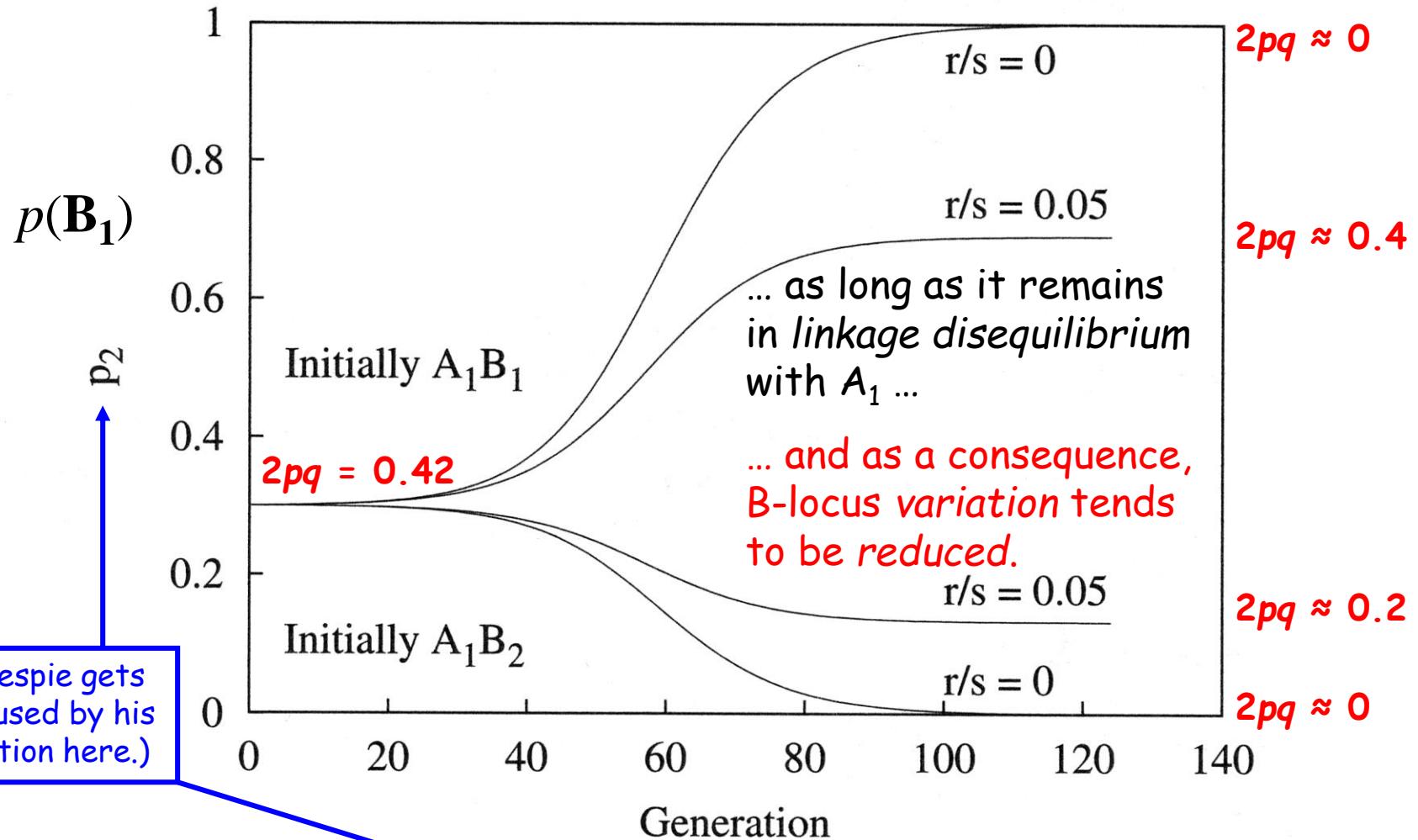


Figure 4.5: The frequency of the B_2 allele under different hitchhiking scenarios. For the upper two curves, the A_1 allele is initially linked to the B_1 allele; in the bottom two, it is linked to the B_2 allele. $s = 0.2$ for all trajectories.

An important consequence: hitchhiking “sweeps away” variation near the selected locus

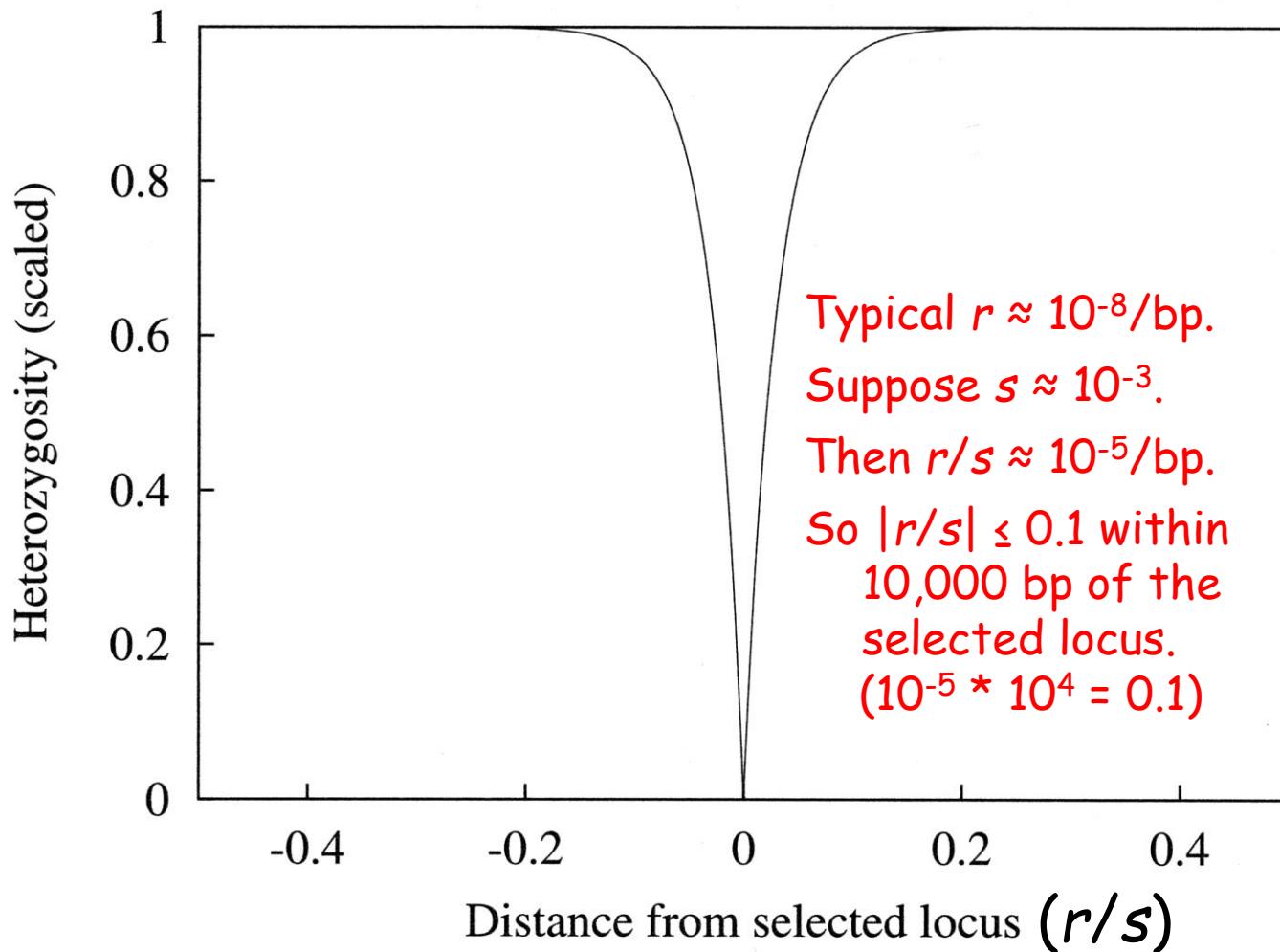
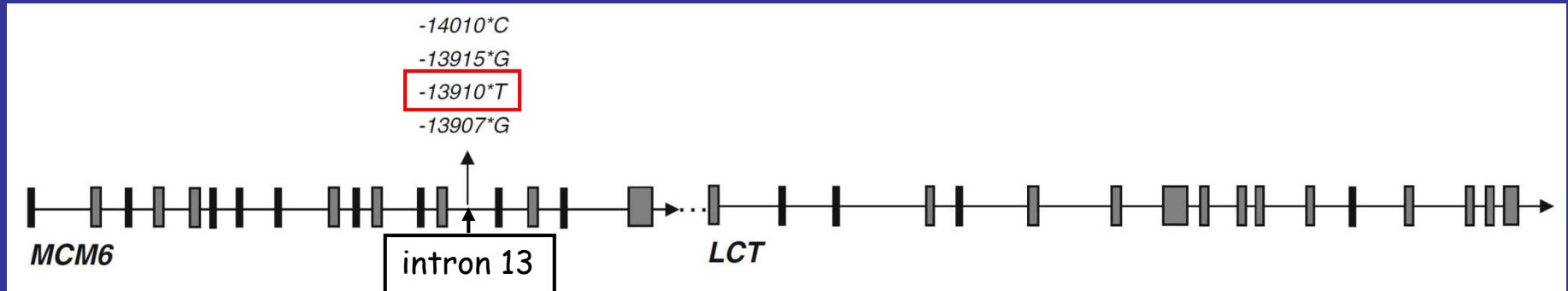


Figure 4.4: The ratio of the final to initial heterozygosity at a neutral locus as a function of the distance from the selected locus as measured by r/s . Negative values of r/s are left of the selected locus, positive values are to the right.

LCT region and putative lactase persistence mutations



-14133 TTTATGTAAGTGTGAATGCTCATACGACCATGGAATTCTCCCTTAAAGAGCTGGTAAGCATTGAGTGTAGTTGTTAGACGGAGACGATCACGTC
Cdx-2

-14034 ATAGTTTATAGAGTGCATAAAGACCGTAAGTTACCATTAAACCTTCATTCAAGGAAAATGTACTTAGACCCACAATGTACTAGTAGGCCTCTGCGCT

-13934 GGCAATAAGATAAGATAAAGTACCCCTGGCCTCAAAGGAACTCTCCTTAGGTTGCATTGTATAATGTTGATTTTAGATTGTTCTTGAGCCCT
HNF3 α /Fox HNF4 α
GATA6 Oct-1

-13833 GCATTCCACGAGGATAGGTCAGTGGTATTAACGAGGTAAAGGGGAGTAGTACGAAAGGGCATTCAAGCGTCCCATCTCGCTCAACCAAAGCAGCCC

-13733 TGCTTTTCCTAGTTTATTAAGGTTGATGTAAGGTCGTCTTGAAA -13684

Lactase in Utah, again!

The first 26 chromosomes share the consensus sequence (as do 60 others omitted to save space).

The other 34 chromosomes are shown as differences from the consensus. (And sorted by their number of differences from the consensus.)

These 101 variable sites are embedded in a region of roughly 140,000 base pairs.

In what sense are the consensus chromosomes “younger” than the others?

Are their mutations younger? Or is it just the combination of mutations?

Enlightening exercise:
Identify and describe a few recombination events.

cons aaggaggcgacattccgcttcaggcattcctatctaaacagaccaacgta**A**gggtacaatgcctaaccagacgttcaactctggctgttattcctcgat
01
02
03
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24
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27
28
29
30
31 gg.a.ca.ag.g.gt.....
32
33
34
35
36
37
38
39
40 gg.a..at.gt.c.t...tcc...agtag.t.cat.....
41
42
43 ..aa..at.gt.c.t...tcc...agtag.t.cat.....
44 ..aa..at.gt.c.t...tcc...agtag.t.cat.....
45 ..aa..at.gt.c.t...tcc...agtag.t.cat.....
46
47 gg.a..at.gt.c.t...tcc...agtag.t.cat.....
48 gg.a..at.gt.c.t...tcc...agtag.t.cat.....
49 gg.a..at.gt.c.t...tcc...agtag.t.cat.....
50
51 gg.a.ca.ag.g.gtta.ccgga.....
52 gg.a.ca.ag.g.gtta.ccgga.....
53 gg.a.ca.ag.g.gtta.ccgga.....
54 gg.a.ca.ag.g.gtta.ccgga.....
55 gg.a.ca.ag.g.gtta.ccgga.....
56 gg.a.ca.ag.g.gtta.ccgga.....
57 gg.a.ca.ag.g.gtta.ccgga.....
58 gg.a.ca.ag.g.gtta.ccgga.....
59 gg.a.ca.ag.g.gtta.ccgga.....
60 gg.a.ca.ag.g.gtta.ccgga.....

Homo sapiens (human) Build 37.1 (Current)

Chromosome: 1 | 2 | 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 X Y

Query: LCT [clear]

Master Map: Genes On Sequence

Region Displayed: 135,360K-137,790K bp

Ideogram Contig

Region Shown:

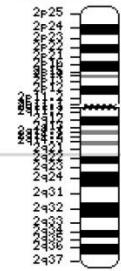
135,360K

137,790K

out
 zoom
 in

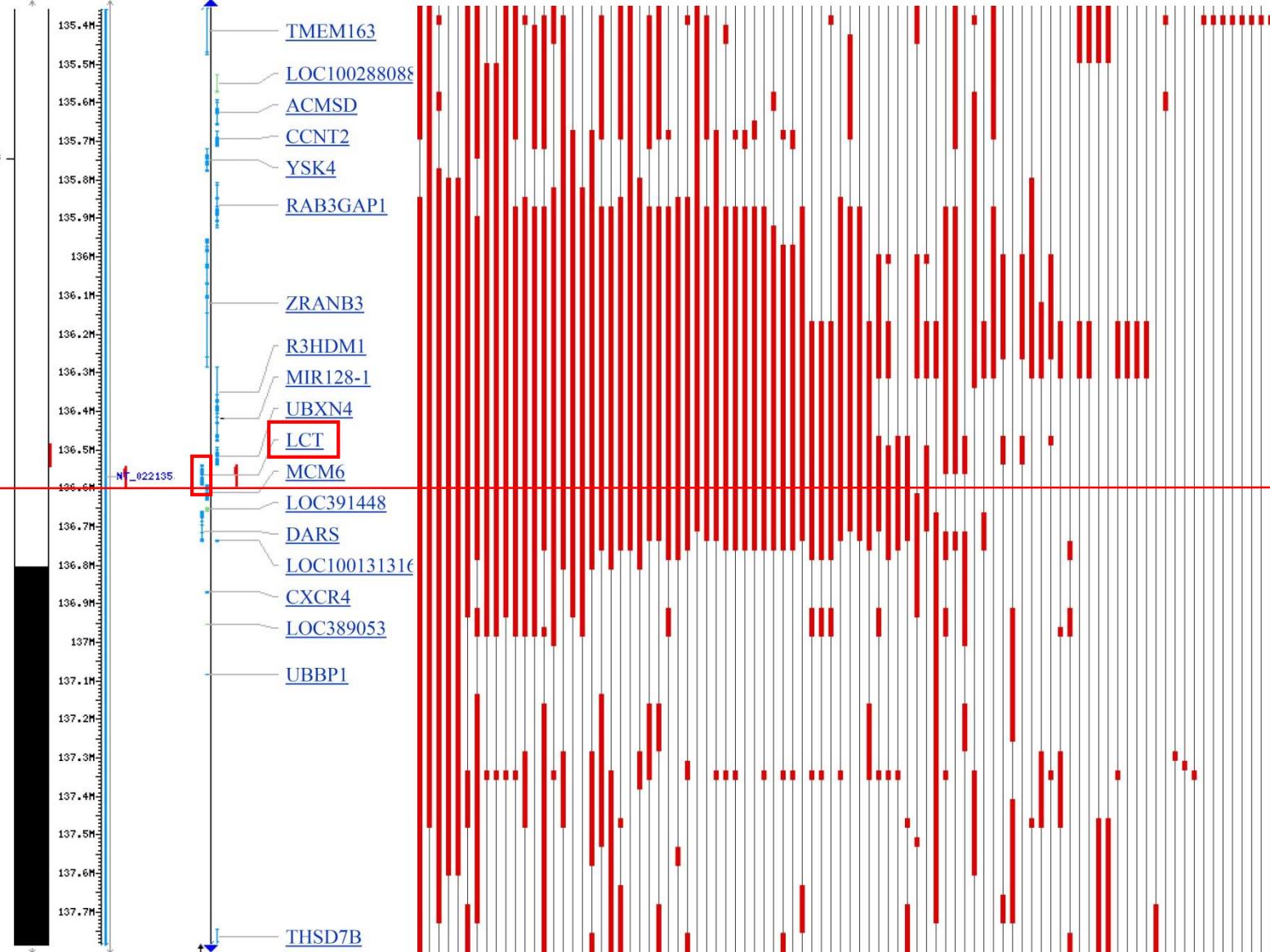
You are here:

Ideogram



LCT-13190*T

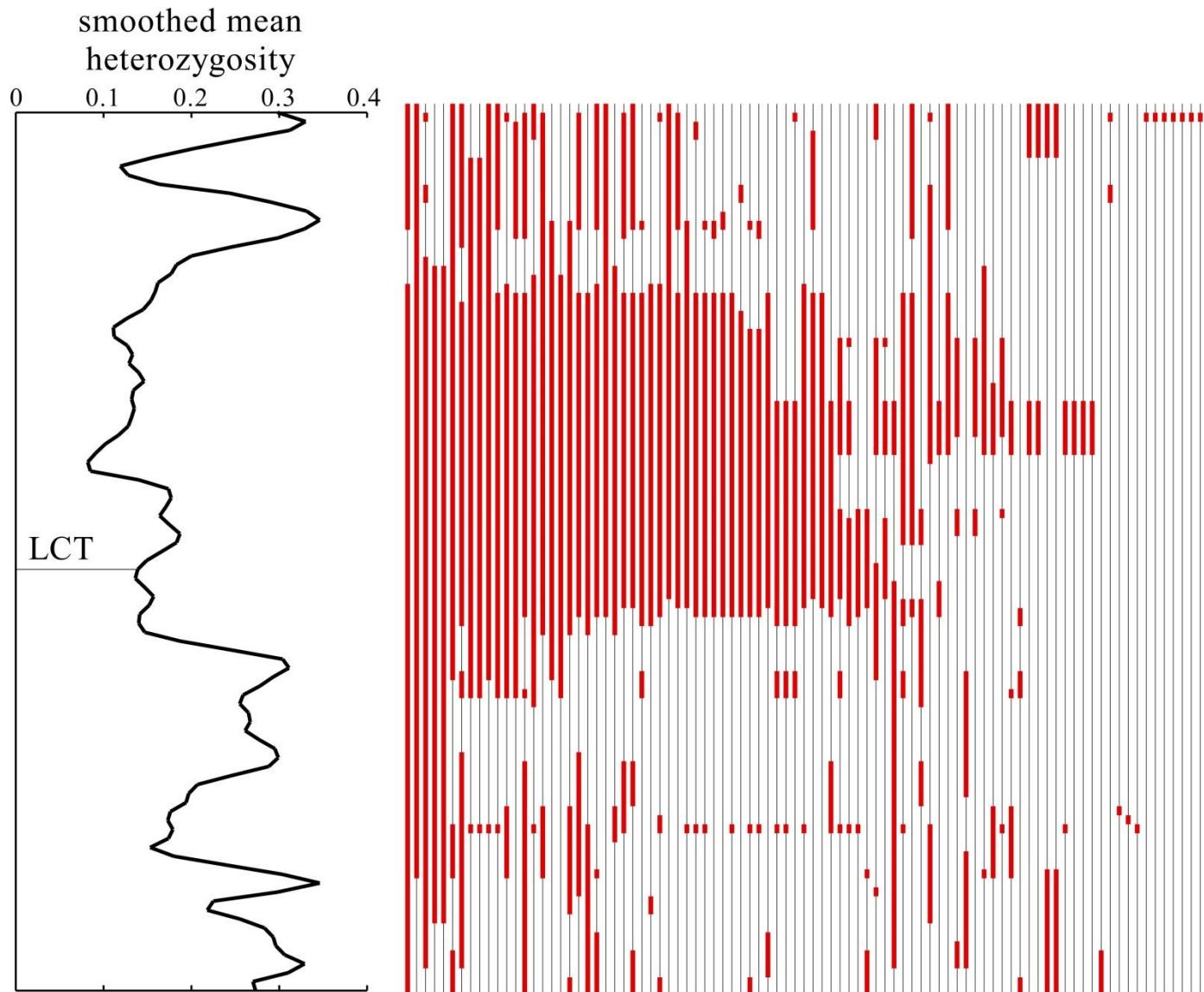
2.4 Mbp
19 genes



Homozygous intervals of
chromosome 2 in 90 Utahns

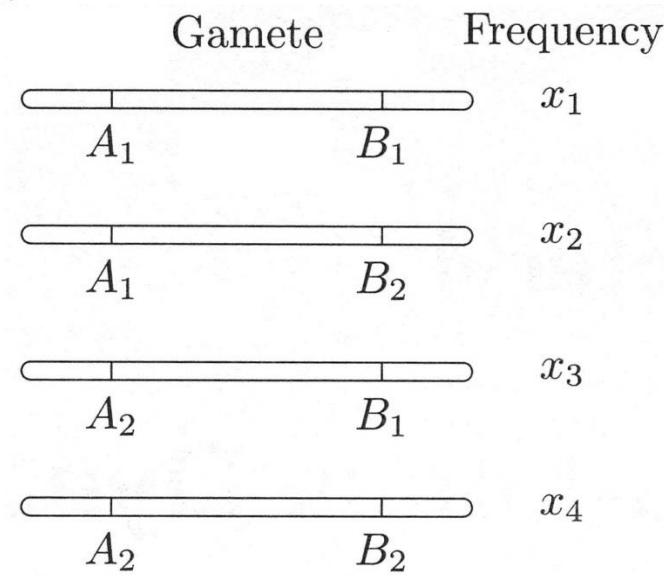
Overall heterozygosity is “drafted down” at HapMap SNP loci.

But why so modestly?



4.3 The following program, written in Python, will print out the ratio of the final to starting heterozygosities at the *B* locus.

```
s, r, N = 0.1, 0.001, 5000
eps = 1.0 / ( 2 * N)
x1, x2 = eps, 0.0
x3, x4 = 0.5 - x1, 0.5
while x1 + x2 < 1.0 - eps:
    p1 = x1 + x2
    q1 = 1.0 - p1
    wBar1 = 1.0 - q1 * s / 2.0
    wBar3 = 1.0 - p1 * s / 2.0 - q1 * s
    wBar = 1.0 - q1 * s
    rWD = r * (1 - s / 2.0) * (x1 * x4 - x2 * x3)
    x1 = (x1 * wBar1 - rWD) / wBar
    x2 = (x2 * wBar1 + rWD) / wBar
    x3 = (x3 * wBar3 + rWD) / wBar
    x4 = (x4 * wBar3 - rWD) / wBar
    p2 = x1 + x3
    q2 = 1.0 - p2
    print 2.0 * p2 * q2 / 0.5
```



Exercise: Do this for
 $s = 0.001, 0.01, 0.1$
 $r = 0.0001, 0.003, 0.001$
And consider initial *B* locus allele frequencies other than 0.5!

So, what are the predictions of this model?

Less variation where sweeps are more frequent.

Less variation where recombination rates are lower.

(Loci are strongly affected where $|r/s| < 0.1$.)

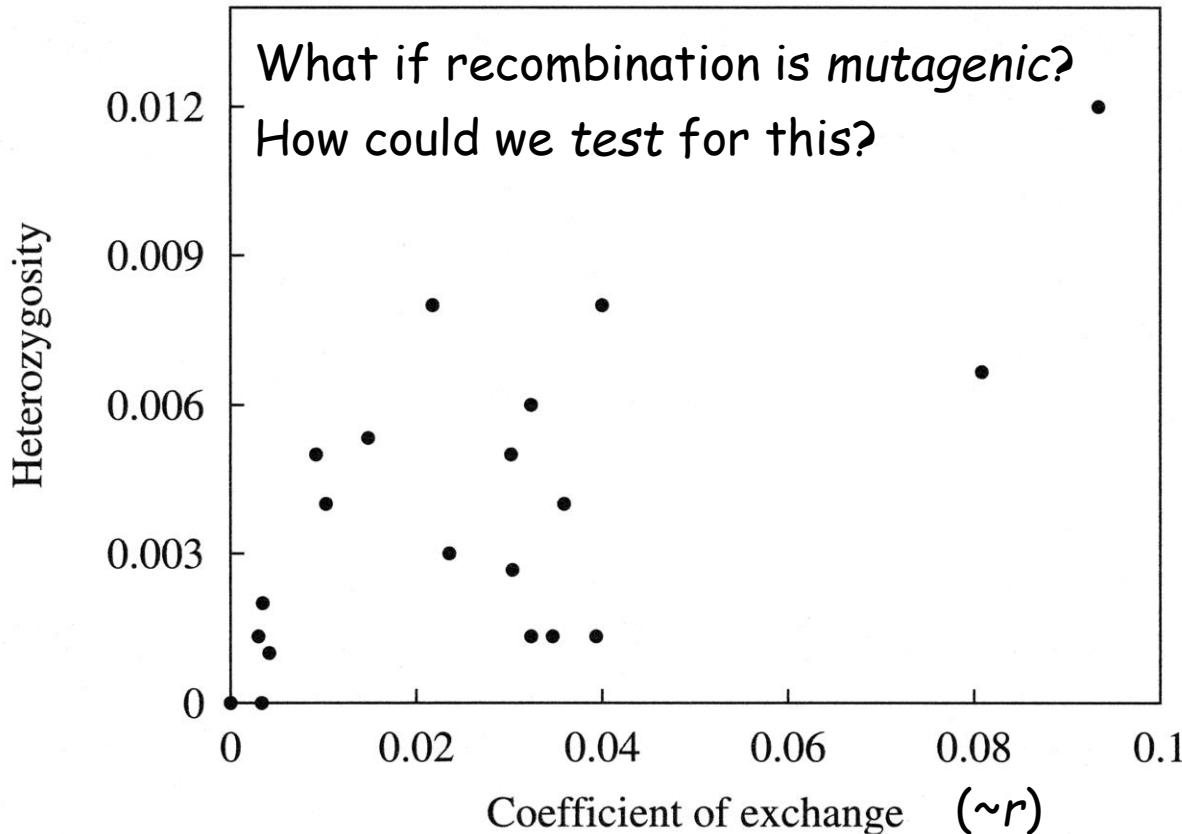


Figure 4.3: The observed silent heterozygosity on the X chromosome of *Drosophila melanogaster* as a function of the local rate of recombination. The data are from Begun and Aquadro (1992).

What determines the frequency of adaptive sweeps (ρ)?

1. The rate of environmental change.
2. The rate at which adaptive mutations occur in the population.

N_e
and
 π

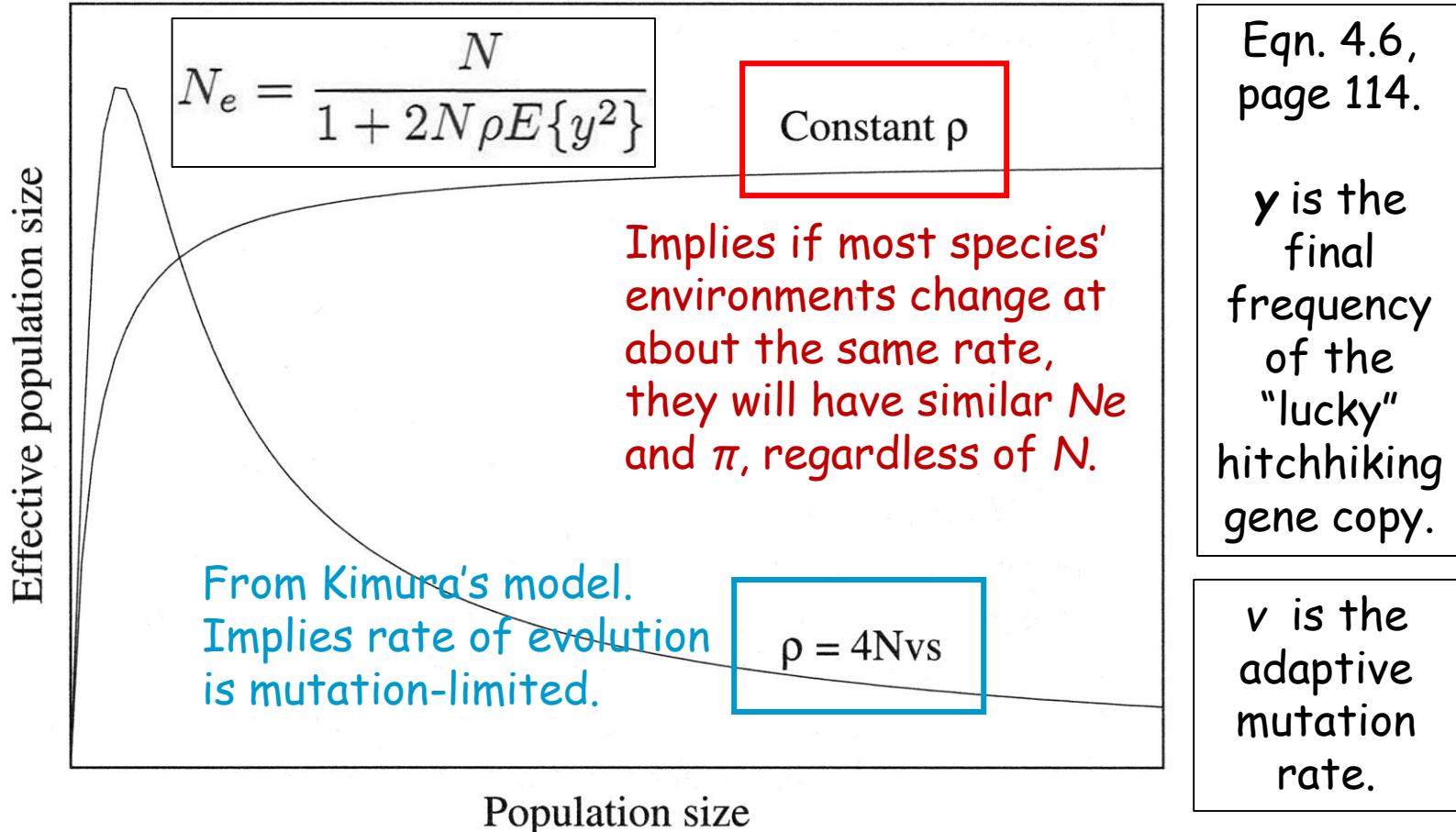


Figure 4.6: The relationship between the population size and the effective population size under genetic draft.

Mitochondria have no recombination, so $E\{y^2\} = 1$

$$N_e = \frac{N}{1 + 2N\rho E\{y^2\}}$$



Krill live "naked" in the Southern Ocean which is always changing (e.g. in temperature).

So their rate of adaptation could be mutation-limited.

$$\rho = 2N_f vs$$

So ρ could increase and N_e decrease with N !

But is the rate of environmental change similar for all genetic loci, in addition to being similar for most species?

π varies overall by roughly a factor of 100, which is really not enough.

Most values fall between 0.002 and 0.02, and are much smaller than $4N\mu$!

TABLE 2. Estimates of nucleotide diversity (π ; heterozygosity at the nucleotide level)

DNA or gene region	Organism	Method	n	bp	π
mtDNA	Human	R	100	16,500	0.004
mtDNA	<i>D. melanogaster</i>	R	10	11,000	0.008
β -Globin	Human	R	50	35,000	0.002
Growth hormone	Human	R	52	50,000	0.002
Notch gene region	<i>D. melanogaster</i> (1)	R	37	60,000	0.005
White locus region	<i>D. melanogaster</i> (2)	R	38	45,000	0.011
Factor IX	Human (3)	S	22	2460	0.0002
<i>Adh</i> locus (C)	<i>D. melanogaster</i>	S	11	765	0.006
Prochymosin (C)	Bovine (4)	S	8	1146	0.004
Growth hormone (C)	Pig (4)	S	6	651	0.007
Class I MHC (<i>HLA-A</i>)	Human (5)	S	5	274	0.043
Class I MHC (<i>H2-K</i>)	Mouse (5)	S	4	273	0.077

Summary

Genetic "draft" is the effect of a selective sweep at one locus on the variation at nearby (linked) loci.

On average it *reduces variation* at those linked loci.

In this way it reduces the apparent or effective N.

Gillespie calls it a "new stochastic force in evolution", arising from the stochasticity of the *initial conditions*.

That is, when and where (on which chromosome) a favored mutation occurs.

Whether we consider it a new *force*, or just an *effect*, it's important!

Apparently we don't yet understand the whole story.

For example, could it help explain the evolution of recombination rates?