## Coexistence of alleles: insights of Modern

# Coexistence Theory into the maintenance of

# genetic diversity

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### <sub>9</sub> 1 Introduction

The question of how genetic variation is maintained, despite the effects of selection and drift, continues to be central to the study of evolutionary biology (Walsh & Lynch, 2018).

Classical explanations include overdominance (heterozygote advantage) or frequencydependent selection, but in the modern era of genomic data, all patterns of variation that
exceed the expected variation under neutrality tend to be categorized broadly as balancing selection, regardless of the evolutionary mechanism (Mitchell-Olds *et al.*, 2007). One
of the evolutionary mechanisms coined under balancing selection is sexually antagonistic selection, which occurs when the direction of natural selection on traits or loci differs
between the sexes (Arnqvist & Rowe, 2013).

Sexually antagonistic selection has been identified as a powerful engine of speciation that in some cases can mantain polymorphisms in a population (Gavrilets, 2014).

This sexual conflict can result in phenotypically distinct sexes that express morphological, physiological, and behavioral traits to different degrees (Mori *et al.*, 2017; Connallon
& Hall, 2018). However, sexually antagonistic selection can only mantain polymorphism
in specific scenarios, as classical predictions show that sexual antagonism often results in
the fixation of one of the alleles (Connallon *et al.*, 2018). Importantly, the effect of sexually antagonistic selection, has been generally studied under strong simplifying assumptions such as constant population sizes and homogeneous environments (e.g., Kidwell *et al.* (1977); Pamilo (1979); Immler *et al.* (2012)). Few studies have explored the effect of

assumptions. Excepctions include Connallon *et al.* (2018) who found that classical predictions break down when fluctuations in the environment combined with life-history traits allow local adaptations and promote the maintenance of genetic diversity. The effect of environmental fluctuations without local adaptation, however, has not been studied in the context of sexually antagonistic selection to the best of our knowledge.

The contribution of environmental fluctuations to genetic variability remains a debated issue in evolutionary biology. Classic theoretical models predict that temporal fluctuations in environmental conditions are unlikely to maintain a genetic polymorphism

(Hedrick, 1974; 1986). However, other studies have found that fluctuating selection can
maintain genetic variance on sex-linked traits (Reinhold, 2000), or in populations where
generations overlap (Ellner & Hairston Jr, 1994; Ellner & Sasaki, 1996). Similarly, temporal changes in population sizes have been shown to mitigate the effect of genetic drift in
small populations (Pemberton *et al.*, 1996), and in annual plant systems (Nunney, 2002).
Thus, both fluctuations in selection and population sizes could dramatically change the
effect of sexually antagonistic selection in the maintenance of genetic diversity.

Importantly, progress requires more than just identifying if environmental fluctuations can maintain genetic diversity in a population, but to quantify how exactly they
contribute to its maintenance (Ellner *et al.*, 2016). Modern coexistence theory (MCT) provides a powerful conceptual framework to do so (Chesson, 2000b; 1994; Barabás *et al.*,
2018). Although its core ideas were formalized in an ecological context (Chesson, 1994;
2000a), this framework provides the necessary tools to examine the relative contributions
of fluctuations to diversity maintenance, which can also be applied to evolutionary con-

texts (Ellner & Sasaki, 1996; Reinhold, 2000). From an ecological perspective, polymorphism of sexually antagonistic alleles is equivalent to the coexistence of species, and the fixation of either one of the alleles in a population is equivalent to competitive exclusion. The coexistence of alleles, thus, can be examined through the same lens as the coexistence

of competing species.

Here, we seek to explicitly apply recent advances in MCT to the question of how polymorphism is maintained under sexually antagonistic selection. We examined how fluctuations in selection values, fluctuations in population sizes, and their interactions can stabilize or hinder the coexistence of alleles. In particular, we examined i) Can fluctuations in
population sizes and selection values allow sexually antagonistic alleles to coexist when
differences in their fitness would typically not allow them to? and ii) What is the relative
contribution of different types of fluctuations that allow two sexually antagonistic alleles
to be maintained in a population? Our study provides the tools to analyze evolutionary
dynamics from a novel perspective and contributes to answering long-lasting questions
regarding the effect of non-constant environments on genetic diversity.

## 67 2 Methods

We first present a model that describes the evolutionary dynamics of sexually antagonistic alleles and show how changes in allele frequencies can be expressed in terms of growth rates, a necessary condition for analyses done using modern coexistence theory. We continue by simulating different scenarios of alleles invading a population, where we allowed population sizes, selection values, both, or neither to vary. Finally, we examine the results of our simulations through a MCT lens by calculating the contribution of each of these fluctuations in the coexistence of alleles.

### Population dynamics of sexually antagonistic alleles

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Our model considered evolution at single, biallelic locus with frequency and density independent effects on the relative fitness of females and males. We examined the dynammics of two sexually antagonistic alleles, j and k, that affect fitness in the haploid state. We assumed allele j always has a high fitness in females ( $w_{jf}=1$ ), but variable fitness in males ( $w_{jm}<1$ ); and allele k has a high fitness in males ( $w_{km}=1$ ), but variable fitness in females ( $w_{kf}<1$ ). The selection against allele k in males is therefore k0 and the selection against allele k1 in females is k1 females is k2.

The frequency of each allele in each sex at the beginning of a life-cycle at time t is given by:

$$p_{jm,t} = \frac{n_{jm,t}}{N_{m,t}} \tag{1}$$

 $p_{jf,t} = \frac{n_{jf,t}}{N_{f,t}} \tag{2}$ 

$$p_{km,t} = \frac{N_{m,t} - n_{jm,t}}{N_{m,t}} \tag{3}$$

$$p_{kf,t} = \frac{N_{f,t} - n_{jf,t}}{N_{f,t}} \tag{4}$$

where  $N_{m,t}$  and  $N_{t,t}$  are the numbers of males and females in a population at time t,  $n_{jf,t}$  is the number of females f with allele j, and  $n_{jm,t}$  is the number of males m with allele j at time t, respectively.

The individuals in the population mate at random before selection occurs, and therefore the frequency of offspring with allele j after mating,  $p'_{j,t}$  can be expressed as:

$$p'_{j,t} = \frac{n_{jf}}{N_f} \frac{n_{jm}}{N_m} + \frac{1}{2} \frac{n_{jf}}{N_f} \frac{(N_m - n_{jm})}{N_m} + \frac{1}{2} \frac{(N_f - n_{jf})}{N_f} \frac{n_{jm}}{N_m},$$
 (5)

which upon rearranging and simplifying gives:

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$$p'_{j,t} = \frac{(N_{m,t}n_{jf,t} + N_{f,t}n_{jm,t})}{2N_f N_m}.$$
 (6)

Selection acts upon these offspring in order to determine the allelic frequencies in females and males in the next generation, t + 1. As an example the frequency of females with allele j after selection is given by:

$$p'_{jf,t+1} = \frac{n_{jf,t+1}}{N'_{f,t+1}} = \frac{p'_j w_{jf}}{p'_j w_{jf} + (1 - p'_j) w_{kf}}$$
(7)

The logarithmic growth rate of j in females, is therefore given by the number of females with allele j after selection, divided by the original number of females carrying allele j:

$$r_{jf,t} = \ln\left(\frac{n'_{jf,t+1}}{n_{jf,t}}\right) \tag{8}$$

An equivalent expression for the per capita growth rate of allele j in males m can be obtained by exchanging f for m across the various subscripts in this expression.

Allelic coexistence in a sexual population, however, is ultimately influenced by growth

and establishment of an allele across both sexes. Therefore, the full growth rate of allele j across the entire population of females *and* males is given by:

$$r_{j} = \ln \left( \frac{n'_{jf,t+1} + n'_{jm,t+1}}{n_{jf,t} + n_{jf,t}} \right) . \tag{9}$$

- An equivalent expression describes  $r_k$ , the growth rate of allele k.
- Selection mantains both alleles in the population under the condition that:

$$\frac{S_m}{1 + S_m} < S_f < \frac{S_m}{1 - S_m} \tag{10}$$

(Kidwell *et al.*, 1977; Pamilo, 1979; Connallon & Hall, 2018) Thus, the maintenance of polymorphism of sexually antagonistic alleles is solely determined by the values of  $S_m$  and  $S_f$ . Note that in our model, the values  $S_m$  and  $S_f$  are bounded from 0 to 1. Therefore the **parameter space of sexually antagonistic selection** is within the range  $0 < S_m, S_f < 1$ . Classic theoretical models predict that in constant environments, only in  $\approx 0.38$  of the selection parameter space alleles can coexist (Kidwell *et al.*, 1977; Pamilo, 1979; Connallon *et al.*, 2018). If fluctuations in population sizes or selection values have an effect on the coexistence of sexually antagonistic alleles, it would be reflected in increases or decreases of the proportion of the parameter space of selection where polymorphism is maintained.

## 116 Simulations

Typically, MCT would require decomposing alleles' growth rates (e.g., Eqn. 9) analytically to examine the relative contributions of different types of fluctuations to their coexistence

(Barabás *et al.*, 2018). Although we present an analytical approach in the Supporting Information, our general solution is not easily interpretable and soon becomes mathematically intractable (S1 Supporting Information). Thus, we opted for an extension of MCT that provides the flexibility to examine the contributions of different processes to coexistence using simulations (Ellner *et al.*, 2019; Shoemaker *et al.*, 2020).

For each simulation, we examined coexistence outcomes across the selection param-124 eter space of sexually antagonistic selection (0  $< S_m, S_f < 1$ ). To do so, we partitioned the parameter space into a grid of  $50 \times 50$ , which yielded 2500 pairwise combinations 126 of different  $w_{im}$  and  $w_{kf}$  values. For each pairwise combination of  $w_{im}$  and  $w_{kf}$ , as we 127 detail in the next sections, we iterated our model while controlling the effect size of fluctuations in selection  $(\sigma_w)$  and their correlation  $(\rho_w)$ , as well as fluctuations in population 129 sizes  $(\sigma_g)$  and their correlation  $(\rho_g)$ . Then, we performed "invasion simulations" of each 130 allele invading a population, evaluated coexistence outcomes, and determined the rela-131 tive contribution of each type of fluctuation. Finally, we calculated for each simulation the proportion of the parameter space that allowed alleles to coexist. 133

We explored all of the combinations of low, intermediate, and high fluctuations in fitness values and population sizes, with different extents of correlations between fluctuations (Table 1). As a control simulation, we set  $\sigma_w = 0.001$  and  $\sigma_g = 0.001$ , with no correlation between fluctuations. We ran ten replicates per parameter combination, which resulted in 3780 simulations.

#### 39 Timeseries

To incorporate the effects of fluctuations into our population dynamics model we generated independent timeseries of fluctuations in selection and population sizes. In the case of fluctuations in selection values, for a given value of  $w_{jm}$  and  $w_{kf}$  (i.e., a fixed point in the selection parameter space), we generated a timeseries of 500 timesteps made up of correlated fluctuations of  $w_{jm}$  and  $w_{kf}$ . We controlled the effect size of fluctuations in selection ( $\sigma_w$ ) and its correlation ( $\rho_w$ ) by using the Cholesky factorization of the variance-

$$C_w = \begin{bmatrix} \sigma_w^2 & \rho_w \sigma_w^2 \\ \rho_w \sigma_w^2 & \sigma_w^2 \end{bmatrix} \tag{11}$$

We multiplyed Eqn. 11 by a  $(2 \times 500)$  matrix of random numbers from a normal distribution with mean 0 and unit variance, which yielded  $\gamma_j$  and  $\gamma_k$ . Then, we calculated the new fitness values at time t+1 as  $w_{jm,t+1}=w_{jm}^{\gamma_{j,t}}$  and  $w_{kf,t+1}=w_{kf}^{\gamma_{k,t}}$ .

Similarly, we generated a timeseries of 500 timesteps made up of correlated fluctuations in population sizes. We chose values of  $N_m=200$  and  $N_f=200$  as the initial value of population sizes throughout our simulations. We performed a Cholesky factorization of the variance-covariance matrix, controlling the effect size of fluctuations in population sizes with  $\sigma_g$  and their correlation with  $\rho_g$ . Similar to our previous approach, we multiplied this factorization by a random matrix of uncorrelated random variables, which yielded  $\gamma_m$  and  $\gamma_f$ . Finally, we calculated the number of males and females in the population at time t+1 as  $N_{m,t+1}=N_m+\gamma_{m,t}$  and  $N_{f,t+1}=N_f+\gamma_{f,t}$ . Therefore, the

population sizes in each timestep differed from the inital value of 200 individuals on the order of  $\rho_g$ . Note that the scales of  $\sigma_g$  and  $\sigma_w$  are different from each other. While  $\sigma_w$  controls the exponential change in fitness values in each timestep,  $\sigma_g$  controls the number of individuals added to a population in each timestep.

Finally, we performed simulations where our population dynamics model (Eqns. 1 to 9) iterated over 500 timesteps while allowing selection values and population sizes to fluctuate in each timestep. We started each simulation with the initial values of  $N_m$  and  $N_f$  described before and equal frequencies of allele j and allele k in each sex. For each timestep t in our simulations, the values of  $w_{jm}$   $w_{kf}$ ,  $N_m$  and  $N_f$  used to calculate allele's frequencies in timestep t (e.g., Eqn. 7), corresponded to the t values calculated in each timeseries, as described previously. This approach yielded a final timeseries that captured the dynamics of sexually antagonistic alleles, with fluctuating values of selection and population sizes.

#### 171 Invasion simulations

Modern coexistence theory has shown that coexistence is promoted by mechanisms that give species a population growth rate advantage over other species when they become rare (Chesson, 1982; 2003; Barabás  $et\ al.$ , 2018). Typically, one species is held at its resident state, as given by its steady-state abundances while the rare species is called the invader. In the context of alleles in a population, an allele is an invader when a mutation occurs that introduces that allele into a population in which it is absent (e.g., if in a population with only k alleles, a random mutation made one individual carry the j al-

lele). Within sexually antagonistic selection, each allele has two pathways of invasion, depending on whether the mutation arises in a female or in a male. If an alleles' invasion 180 growth rate (or the average instantaneous population growth rate when rare) is positive, 181 it buffers it against extinction, maintaining its persistence in the population. Coexistence, and hence polymorphism, occurs when both alleles have positive invasion growth rates. 183 We used the timeseries that captured the dynamics of our population model as a tem-184 plate to perform invasion simulations of both alleles. We performed 500 independent invasion simulations, one for each timestep in our timeseries. We explored all four po-186 tential combinations of each allele invading through each pathway (e.g., allele *j* invading 187 through males, and allele k invading through females, and so on). To simulate invasion, we set the density of the invading allele to one individual. For example, if allele j was 189 invading via males, then we would set  $n_{jm,i} = 1$  and  $n_{jf,i} = 0$ . Note that each inva-190 sion simulation was independent of the iteration that we used to generate the timeseries, therefore we denoted the initial timestep in an invasion simulation with the subscript *i*. 192 We also set the resident allele, in this case k, to the corresponding value of the timeseries 193 minus one individual,  $n_{km,i} = N_{m,t} - 1$  and  $n_{kf,i} = N_{f,t}$ . Then, we iterated our model one timestep, i + 1, and calculated the logarithmic growth rate of j allele invading as: 195

$$r_j = \ln\left(\frac{n_{jm,i+1} + n_{jf,i+1}}{1}\right) \tag{12}$$

Correspondingly, the logarithmic growth rate of the *k* allele as a resident would be

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197 given by:

$$r_k = \ln\left(\frac{n_{km,i+1} + n_{kf,i+1}}{n_{km,i} + n_{kf,i}}\right)$$
(13)

Following the approach of Shoemaker *et al.* (2020), we treated each invasion simulation independently, and hence we performed 500 invasion simulations. We then calculated, for each allele invading via a different pathway, its mean invasion growth rate as the average of the 500 invasion growth rates. We also calculated the mean growth rate of the resident allele as the average of the 500 resident growth rates. We determined alleles to be coexisting if both of alleles had positive mean invasion growth rates, which is often referred to as the mutual invasibility criterion (Barabás *et al.*, 2018).

#### 205 Functional decompostion

Our invasion simulations tell us whether or not sexually antagonistic alleles can coexist in a determined point of the selection parameter space. However, we also quantified
the relative contributions of fluctuating selection and population sizes into the predicted
coexistence outcome using a *functional decomposition* approach (Ellner *et al.*, 2016; 2019;
Shoemaker *et al.*, 2020).

We applied the functional decomposition approach by breaking up the average growth rate of each allele into a null growth rate in the absences of fluctuations in all selected variables, a set of main effect terms that represent the effect of only one variable fluctuating, and a set of two-way interaction terms representing the effect of variables fluctuating simultaneously (Ellner *et al.*, 2019). In our simulations, this is a function of four variables: the number of males in the population  $(N_m)$ , the number of females in the population

 $(N_f)$ , the fitness of allele j in males  $(w_{jm})$ , and the fitness of allele k in females  $(w_{kf})$ . As an example, if only  $N_m$  and  $N_f$  were fluctuating, the growth rate of allele j when it is the invader at timestep t could be decomposed into:

$$r_{j,t}(N_m, N_f) = \mathcal{E}_j^0 + \mathcal{E}_j^{N_m} + \mathcal{E}_j^{N_f} + \mathcal{E}_j^{N_m N_f}$$
(14)

Where  $\mathcal{E}^0$  is the null growth rate when  $N_m$  and  $N_f$  are set to their averages. Terms with superscripts represent the marginal effects of letting all superscripted variables vary while fixing all the other variables at their average values. For example, the term  $\mathcal{E}^{N_m}$  expresses the contribution of fluctuations in  $N_m$  when  $N_f$  is at its average, without the contribution when both variables are set to their averages:

$$\mathcal{E}_j^{N_m} = r_{j,t}(N_m, \overline{N_f}) - \mathcal{E}_j^0 \tag{15}$$

If we average Eqn. 14 across the timesteps in our simulation, we get a partition of the average population growth rate into the variance–free growth rate, the main effects of variability in  $N_m$ , the main effects of variability in  $N_f$ , and the interaction between variability in  $N_m$  and  $N_f$ 

$$\overline{r}_j = \mathcal{E}_j^0 + \overline{\mathcal{E}_j}^{N_m} + \overline{\mathcal{E}_j}^{N_f} + \overline{\mathcal{E}_j}^{N_m N_f}$$
(16)

However, in our simulations  $w_{jm}$  and  $w_{kf}$  also fluctuated, therefore the full functional decomposition of the growth rate of allele j as an invader is found in Table 2, as well as a brief description of the meaning of each term. The implementation and interpretation

of the functional decomposition of the invasion growth rates of each allele are identical to each other. Note that Table 2 does not include three or four–way interactions (e.g.,  $\overline{\mathcal{E}}_{j}^{N_{m}N_{f}w_{jm}w_{fk}}$ ). This is because in our simulations, we did not allow fluctuations in selection and population sizes to be correlated, therefore their effects are solely captured by the terms in Table 2. We calculated the value of each of the terms in Table 2 by performing another set of invasion simulations as described previously, but instead of allowing all variables to fluctuate, systematically setting the required variables to their means and subtracting the corresponding  $\mathcal{E}$  values.

The functional decomposition approach further requires the *comparison* of each term, to understand if how it affects invaders and residents. This is because fluctuations can promote coexistence by helping whichever allele is rare, or they can hurt whichever allele is common. Therefore, to understand the role of each type of fluctuation, it is necessary to compare how it affects invader *and* resident growth rates. In the example presented in Eqn. 16, if allele j is invading, then allele k is at it's resident state and there exists an analogue decomposition of  $\bar{r}_k$  with the exact same terms as Eqn. 16. Therefore we can express the difference between contributions of fluctuations in  $N_m$  as:

$$\Delta_j^{N_m} = \overline{\mathcal{E}}_j^{N_m} - \overline{\mathcal{E}}_k^{N_m} \tag{17}$$

If  $\Delta_j^{N_m}$  is positive, then fluctuations in the male population benefit allele j when it is rare more than what they benefit k as a resident. If  $\Delta_j^{N_m}$  is negative, then fluctuations benefit k as a resident more than j as an invader, and if it is minimal, then fluctuations have an equal effect in j and k. Therefore, for each allele invading via a different pathway, we calculated 11  $\Delta$  values, one for each one of the  $\mathcal{E}$  terms in Table 2. However, since the magnitude of each one of these values could vary considerably across the parameter space of selection, to make them comparable, we normalized each  $\Delta$  value by dividing it by the square root of the sum of the squares of the 11  $\Delta$  values. For example, the normalized value of Eqn. 17 would be given by:

$$\delta_j^{N_m} = \frac{\Delta_j^{N_m}}{\sqrt{\sum_{d=1}^{11} (\Delta_d)^2}}$$
 (18)

This normalization bounded  $\delta$  values from -1 to 1.

## 258 3 Results

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Our results showed that both fluctuations in selection and in population sizes can substantially increase the expected genetic variability under sexually antagonistic selection.

The average proportion of coexistence in the selection parameter space increased with the
effect size of fluctuations when fluctuations were large enough (Fig. 1). Fluctuations with
small effect sizes either decreased or matched the average proportion of the parameter
space of allelic coexistence compared to the control simulation (Fig. 1). Increments in allelic coexistence were more likely when fluctuations were large, and fluctuations in population sizes were negatively correlated, while fluctuations in selection were positively
correlated (Fig. 1).

Fluctuations increased coexistence by allowing both alleles to have a positive inva-

sion growth rate in instances where selection would typically not allow them to (i.e., they made parts of the parameter space "flip" into coexistence). As a baseline, we show in Fig. 2A the outcome of the control simulation, which matches previous findings that without fluctuations, alleles can coexist in only  $\approx 0.38$  of the selection parameter space (Connallon & Hall, 2018). These "flips" occurred with both types of fluctuations and were more common with larger fluctuations and strongly correlated effects, for which we show examples in Fig. 2A . However, note that there are also parts of the parameter space where coexistence is lost compared to the control simulation, which was more likely when population sizes were fluctuating (Fig. 2A).

Alleles had positive invasion growth rates when positive contributions of fluctuations 278 outweighted the negative contributions of fluctuations. As an example in Fig. 2B we 279 show the functional decomposition of both alleles invading via their favoured pathway 280 in parts of the parameter space that "flipped" into coexistence and competitive exclusion 28 (which correspond to the square and triangle in Fig. 2A). Note that each type of fluctation 282 made similar contributions to each allele, both when they were coexisting or experiencing 283 competitive exclusion (Fig. 2B). However,  $\delta_0$ , which captures the effect of fluctuations 284 set to their averages, captured both positive and negative contributions for both alleles 285 (Fig. 2B). 286

The relative contribution of fluctuations in population sizes depended on the sex where invasion occurred (Fig. 2B and Fig. 3).

## Figures and tables

Table 1: Parameters used in our simulations to control the effect size of fluctuations in population sizes  $(\sigma_g)$  and selection values  $(\sigma_w)$ , as well as their respective correlations  $(\rho_g)$  and  $(\rho_w)$ . We ran ten replicates for each one of the factorial combinations of the following parameters.

Parameter	Values	Description
$\sigma_w$	0.001, 0.1, 0.3, 0.5, 0.7, 0.9	Effect size of fluctuations in fitness values
$\sigma_g$	0.001, 1, 10, 20, 30, 50	Effect size of fluctuations in population sizes
$ ho_w$	-0.75, 0, 0.75	Correlation between fluctuations in fitness values
$ ho_{\mathcal{g}}$	-0.75, 0, 0.75	Correlation between fluctuation in population sizes

Table 2: Functional decomposition of the growth rate of allele j. As we show in Eqn. 16, each term captures the contribution of fluctuations to an alleles' invasion growth rate.

Term	Formula	Meaning
$\mathcal{E}_{j}^{0}$	$\overline{r_j}(\overline{N_m},\overline{N_f},\overline{w_{jm}},\overline{w_{kf}})$	Growth rate at mean population size and fitness values.
$\overline{\mathcal{E}}_{j}^{N_{m}}$	$\overline{r}_j(N_m\overline{N_f},\overline{w_{jm}},\overline{w_{kf}})-\mathcal{E}_j^0$	Main effect of fluctuations in $N_m$
$\overline{\mathcal{E}}_{i}^{N_{f}}$	$\overline{r_j}(\overline{N_m},N_f,\overline{w_{jm}},\overline{w_{kf}})-\mathcal{E}_j^0$	Main effect of fluctuations in $N_f$
$\overline{\mathcal{E}}_{j}^{w_{jm}}$	$\overline{r_j}(\overline{N_m},\overline{N_f},w_{jm},\overline{w_{kf}})-\mathcal{E}_j^0$	Main effect of fluctuations in $w_{jm}$
$\overline{\mathcal{E}}_{j}^{w_{kf}}$	$\overline{r_j}(\overline{N_m},\overline{N_f},\overline{w_{jm}},w_{kf})-\mathcal{E}_j^0$	Main effect of fluctuations in $w_{kf}$
$ \frac{\mathcal{E}_{j}^{0}}{\mathcal{E}_{j}^{N_{m}}} $ $ \frac{\mathcal{E}_{j}^{N_{f}}}{\mathcal{E}_{j}^{w_{jm}}} $ $ \frac{\mathcal{E}_{j}^{w_{f}}}{\mathcal{E}_{j}^{w_{hf}}} $ $ \frac{\mathcal{E}_{j}^{w_{hf}}}{\mathcal{E}_{j}^{N_{m},N_{f}}} $	$\overline{r_j}(N_m, N_f, \overline{w_{jm}}, \overline{w_{kf}}) - [\mathcal{E}_j^0 + \overline{\mathcal{E}}_j^{N_m} + \overline{\mathcal{E}}_j^{N_f}]$	Interaction of fluctuations in $N_m$ and $N_f$
$\overline{\mathcal{E}}_{i}^{a_{jm},a_{kf}}$	$\overline{r_j}(\overline{N_m},\overline{N_f},w_{jm},w_{kf})-[\mathcal{E}_j^0+\overline{\mathcal{E}}_j^{w_{jm}}+\overline{\mathcal{E}}_j^{w_{kf}}]$	Interaction of fluctuations in $w_{jm}$ and $w_{kf}$
$\overline{\mathcal{E}}_{j}^{N_m w_{jm}}$	$\overline{r_j}(N_m, \overline{N_f}, w_{jm}, \overline{w_{kf}}) - [\mathcal{E}_j^0 + \overline{\mathcal{E}_j}^{N_m} + \overline{\mathcal{E}_j}^{w_{jm}}]$	Interaction of fluctuations in $N_m$ and $w_{jm}$
$\overline{\mathcal{E}}_{j}^{N_{m}w_{kf}}$	$\overline{r_j}(N_m, \overline{N_f}, \overline{w_{jm}}, w_{kf}) - [\mathcal{E}_j^0 + \overline{\mathcal{E}}_j^{N_m} + \overline{\mathcal{E}}_j^{w_{kf}}]$	Interaction of fluctuations in $N_m$ and $w_{kf}$
$\overline{\mathcal{E}}_{j}^{N_{f}w_{jm}}$	$\overline{r_j}(\overline{N_m}, N_f, w_{jm}, \overline{w_{kf}}) - [\mathcal{E}_j^0 + \overline{\mathcal{E}}_j^{N_f} + \overline{\mathcal{E}}_j^{w_{jm}}]$	Interaction of variation in $N_f$ and $w_{jm}$
$\overline{\mathcal{E}}_{j}^{N_{f}w_{fk}}$	$\overline{r_j}(\overline{N_m}, N_f, \overline{w_{jm}}, w_{kf}) - [\mathcal{E}_j^0 + \overline{\mathcal{E}_j^{N_f}} + \overline{\mathcal{E}_j^{w_{kf}}}]$	Interaction of fluctuations $N_f$ and $w_{kf}$

#### Correlation in fluctuations in selection, $\rho_w$ -0.750 0.75 70 50 30 0.75 20 Correlation in fluctuations in population sizes, $\rho_{\text{g}}$ 10 Fluctuations in population sizes, $\sigma_{g}$ 0.001 70 50 30 20 10 0.001 70 50 30 20 10 0.001 0.3 0.9 0.001 0.1 0.3 0.5 0.7 0.9 0.001 0.1 0.5 0.7 0.5 Fluctuations in selection, $\sigma_{\rm w}$ Average proportion of coexistence .3 .38 .5

Figure 1: Average proportion of coexistence for different parameter combinations. We show, for all factorial combinations of values of  $\sigma_g$ ,  $\sigma_w$ ,  $\rho_g$  and  $\rho_w$ , the average proportion of coexistence of in our simulations. Each panel corresponds to a different combination of correlations between fluctuations. Labels on top indicate the correlation between fluctuations in selection  $\rho_w$ , while labels on the right show the correlation in fluctuations between fluctuations in population sizes  $\rho_g$ . As a basis of comparison, we show the expected proportion of coexistence (0.38) as the midpoint in our color scheme.

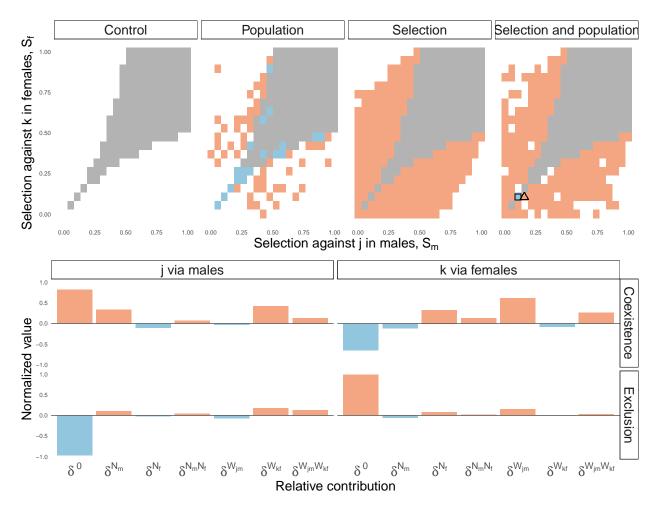


Figure 2: Coexistence outcomes and functional decomposition of one simulation. We show in A) for one of the replicates highlighted with a white square in Fig. ?? ( $\sigma_g = 70$ ,  $\sigma_w = 0.9$ ,  $\rho_g = -0.75$  and  $\rho_{w=0.75}$ ,) the coexistence outcomes when j invades via females and k invades via males. Grey areas denote points in the parameter space where alleles can coexist, while white areas are points where at least one allele does not have a positive invasion growth rate. We highlight in black a point in the parameter space where coexistence is lost compared to the control simulation. In B) we show for the same simulation, the functional decomposition of each allele across the parameter space of selection. The highlighted black points correspond to the same point shown in A).

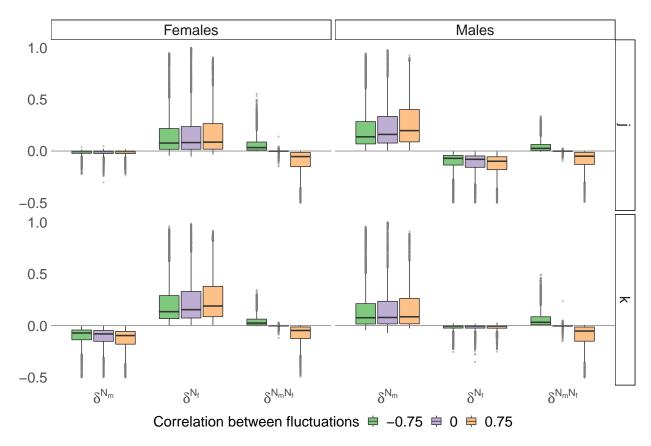


Figure 3: Boxplots of  $\delta$  values. In A) we show the box plots of  $\delta$  values that capture the relative contributions of fluctuations in population sizes, for all of the replicates in our simulation in which  $\sigma_g=70$  and  $\sigma_w=0.001$ . Labels on top indicate the correlation between fluctuations in population sizes,  $\rho_g$ . Each color corresponds to a different allele and invasion pathway in our simulations. In B) we show the box plots of  $\delta$  values that capture the contributions of fluctuations in selection, for all of the replicates in our simulation in which  $\sigma_w=0.9$  and  $\sigma_g=0.001$ . Labels on top indicate the correlation between fluctuations in selection,  $\rho_w$ , with the same color nomenclature as in A). Each box plot extends from the first to third quantiles of the corresponding posterior distribution of parameter values, and the line inside the the box indicates the median. The upper whisker extends to the largest value no further than 1.5 times the inter-quantile range (IQR, or the distance between the first and third quartiles); the lower whisker extends to the smallest value at most 1.5 times the IQR. Data beyond the end of the whiskers are determined to be outliers and are plotted individually with solid grey points.

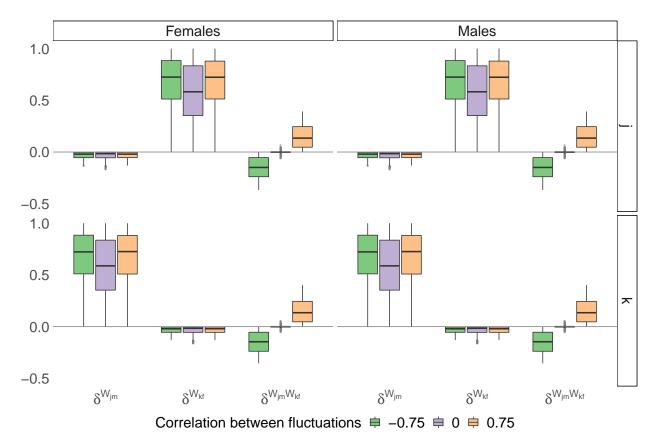


Figure 4: Boxplots of  $\delta$  values. In A) we show the box plots of  $\delta$  values that capture the relative contributions of fluctuations in population sizes, for all of the replicates in our simulation in which  $\sigma_g=70$  and  $\sigma_w=0.001$ . Labels on top indicate the correlation between fluctuations in population sizes,  $\rho_g$ . Each color corresponds to a different allele and invasion pathway in our simulations. In B) we show the box plots of  $\delta$  values that capture the contributions of fluctuations in selection, for all of the replicates in our simulation in which  $\sigma_w=0.9$  and  $\sigma_g=0.001$ . Labels on top indicate the correlation between fluctuations in selection,  $\rho_w$ , with the same color nomenclature as in A). Each box plot extends from the first to third quantiles of the corresponding posterior distribution of parameter values, and the line inside the the box indicates the median. The upper whisker extends to the largest value no further than 1.5 times the inter-quantile range (IQR, or the distance between the first and third quartiles); the lower whisker extends to the smallest value at most 1.5 times the IQR. Data beyond the end of the whiskers are determined to be outliers and are plotted individually with solid grey points.

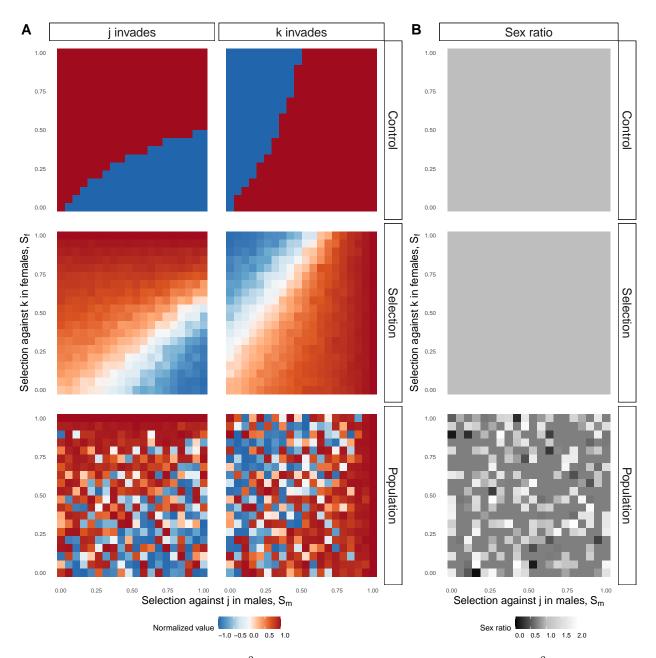


Figure 5: The relationship between  $\delta^0$  and sex ratios. In A) we show the values of  $\delta^0$  accross the selection parameter space for different types of simulations when j invades via males and k invades via females. Each pannel corresponds to an allele invading a population in a replicate of different simulations, as labels on the right indicate: Control denotes the control simulation, Selection denotes a simulation where  $\sigma_w = 0.9$  and  $\sigma_g = 0.001$ , and Population indicates a simulation where  $\sigma_w = 0.001$  and  $\sigma_g = 70$ . For simplicity we kept both correlations equal to zero. In B), we show for the same replicates shown in A), the sex ratios, calculated as  $\frac{N_f}{N_m}$ , accross the selection parameter space.

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