

1 Coexistence of sexually antagonistic alleles:
2 insights of Modern Coexistence Theory into the
3 maintenance of genetic diversity

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1 Abstract

Sexually antagonistic selection (SAS) occurs when the selection in the traits or loci differs between the sexes. This sexual conflict offers the opportunity for maintaining polymorphism in a population, but it often results in the eventual fixation of the fitter allele. However, the effects of SAS have generally been studied under strong simplifying assumptions, such as constant populations and homogeneous environments, which could considerably change the expected outcomes of SAS. Thus, in this study, we examined how fluctuations in selection and population sizes contributed to the coexistence of sexually antagonistic alleles by adopting an ecological framework that allowed us to examine evolutionary dynamics through the same lens as the coexistence of competing species. We performed simulations of alleles invading a population while allowing selection and populations sizes to fluctuate over time. Then, we quantified coexistence outcomes and the relative contribution of each type of fluctuation to each alleles' invasion growth rate. Our results showed that environmental fluctuations can dramatically increase the expected genetic variation under SAS. The positive contribution of fluctuations, however, depended on the sex and allele where invasion occurred. This study contributes to the growing body of work that shows the importance of non-constant environments on the maintenance of genetic diversity.

2 Introduction

The question of how genetic variation is maintained, despite the effects of selection and drift, continues to be central to the study of evolutionary biology (Walsh & Lynch, 2018). Classical explanations include overdominance (heterozygote advantage) or frequency-dependent selection, but in the modern era of genomic data, all patterns of variation that exceed the expected variation under neutrality tend to be categorized broadly as balancing selection, regardless of the evolutionary mechanism (Mitchell-Olds *et al.*, 2007). One of the evolutionary mechanisms coined under balancing selection is sexually antagonistic selection, which occurs when the direction of natural selection on traits or loci differs between the sexes (Lande, 1980; Arnqvist & Rowe, 2013).

Sexually antagonistic selection can in some cases can maintain polymorphisms of otherwise dis-advantageous alleles in a population (Gavrilets, 2014), which in turn can result in phenotypically distinct sexes that express morphological, physiological, and behavioral traits to different degrees (Mori *et al.*, 2017; Connallon & Hall, 2018). However, sexually antagonistic selection can only maintain polymorphism in specific scenarios, as classical predictions show that sexual antagonism often results in the fixation of one of the alleles (Curtisinger *et al.*, 1994; Connallon *et al.*, 2018). Importantly, the effect of sexually antagonistic selection, has been generally studied under strong simplifying assumptions such as constant population sizes and homogeneous environments (e.g., Kidwell *et al.* (1977); Pamilo (1979); Immler *et al.* (2012)). Few studies have explored the effect of sexually antagonistic selection on the maintenance of polymorphism with more realistic

assumptions. Exceptions include Connallon *et al.* (2018) who found that classical predictions break down when fluctuations in the environment combined with life-history traits allow local adaptations and promote the maintenance of genetic diversity. The effect of environmental fluctuations without local adaptation, however, has not been studied in the context of sexually antagonistic selection to the best of our knowledge.

The contribution of environmental fluctuations to genetic variability remains a debated issue in evolutionary biology. Classic theoretical models predict that temporal fluctuations in environmental conditions are unlikely to maintain a genetic polymorphism (Hedrick, 1974; 1986). However, other studies have found that fluctuating selection can maintain genetic variance on sex-linked traits (Reinhold, 2000), or in populations where generations overlap (Ellner & Hairston Jr, 1994; Ellner & Sasaki, 1996). Similarly, temporal changes in population sizes have been shown to mitigate the effect of genetic drift in small populations (Pemberton *et al.*, 1996), and in annual plant systems (Nunney, 2002). Thus, both fluctuations in selection and population sizes could dramatically change the effect of sexually antagonistic selection in the maintenance of genetic diversity.

Importantly, progress requires more than just identifying if environmental fluctuations can maintain genetic diversity in a population, but to quantify how exactly they contribute to its maintenance (Ellner *et al.*, 2016). Modern coexistence theory (modern coexistence theory) provides a powerful conceptual framework to do so (Chesson, 2000b; 1994; Barabás *et al.*, 2018). Although its core ideas were formalized in an ecological context (Chesson, 1994; 2000a), this framework provides the necessary tools to examine the relative contributions of fluctuations to diversity maintenance, which can also be applied

to evolutionary contexts (Ellner & Sasaki, 1996; Reinhold, 2000). From an ecological perspective, polymorphism of sexually antagonistic alleles is equivalent to the coexistence of species, and the fixation of either one of the alleles in a population is equivalent to competitive exclusion. The coexistence of alleles, thus, can be examined through the same lens as the coexistence of competing species.

Here, we seek to explicitly apply recent advances in modern coexistence theory to the question of how polymorphism is maintained under sexually antagonistic selection. We examined how fluctuations in selection values, fluctuations in population sizes, and their interactions can stabilize or hinder the coexistence of alleles. In particular, we examined

- i) Can fluctuations in population sizes and selection values allow sexually antagonistic alleles to coexist when differences in their fitness would typically not allow them to? and
- ii) What is the relative contribution of different types of fluctuations that allow two sexually antagonistic alleles to be maintained in a population?

Our study provides the tools to analyze evolutionary dynamics from a novel perspective and contributes to answering long-lasting questions regarding the effect of non-constant environments on genetic diversity.

3 Methods

We first present a model that describes the evolutionary dynamics of sexually antagonistic alleles and show how changes in allele frequencies can be expressed in terms of growth rates, a necessary condition for analyses done using modern coexistence theory. We continue by simulating different scenarios of alleles invading a population, where we

allowed population sizes, selection values, both, or neither to vary. Finally, we examine the results of our simulations through a modern coexistence theory lens by calculating the contribution of each of these fluctuations in the coexistence of alleles.

Population dynamics of sexually antagonistic alleles

Our model considered evolution at single, biallelic locus with frequency and density independent effects on the relative fitness of females and males. We examined the dynamics of two sexually antagonistic alleles, j and k , that affect fitness in the haploid state. We assumed allele j always has a high fitness in females ($w_{jf} = 1$), but variable fitness in males ($w_{jm} < 1$); and allele k has a high fitness in males ($w_{km} = 1$), but variable fitness in females ($w_{kf} < 1$). The selection against allele j in males is therefore $S_m = 1 - w_{jm}$, and the selection against allele k in females is $S_f = 1 - w_{kf}$.

The frequency of each allele in each sex at the beginning of a life-cycle at time t is given by:

$$p_{jm,t} = \frac{n_{jm,t}}{N_{m,t}} \quad (1)$$

$$p_{jf,t} = \frac{n_{jf,t}}{N_{f,t}} \quad (2)$$

$$p_{km,t} = \frac{N_{m,t} - n_{jm,t}}{N_{m,t}} \quad (3)$$

$$p_{kf,t} = \frac{N_{f,t} - n_{jf,t}}{N_{f,t}} \quad (4)$$

where $N_{m,t}$ and $N_{f,t}$ are the numbers of males and females in a population at time t , $n_{jf,t}$ is the number of females f with allele j , and $n_{jm,t}$ is the number of males m with allele j at

time t , respectively.

The individuals in the population mate at random before selection occurs, and therefore the frequency of offspring with allele j after mating, $p'_{j,t}$ can be expressed as:

$$p'_{j,t} = \frac{n_{jf}}{N_f} \frac{n_{jm}}{N_m} + \frac{1}{2} \frac{n_{jf}}{N_f} \frac{(N_m - n_{jm})}{N_m} + \frac{1}{2} \frac{(N_f - n_{jf})}{N_f} \frac{n_{jm}}{N_m}, \quad (5)$$

which upon rearranging and simplifying gives:

$$p'_{j,t} = \frac{(N_{m,t}n_{jf,t} + N_{f,t}n_{jm,t})}{2N_fN_m}. \quad (6)$$

Selection acts upon these offspring in order to determine the allelic frequencies in females and males in the next generation, $t + 1$. As an example the frequency of females with allele j after selection is given by:

$$p'_{jf,t+1} = \frac{n_{jf,t+1}}{N'_{f,t+1}} = \frac{p'_j w_{jf}}{p'_j w_{jf} + (1 - p'_j) w_{kf}} \quad (7)$$

The logarithmic growth rate of j in females, is therefore given by the number of females with allele j after selection, divided by the original number of females carrying allele j :

$$r_{jf,t} = \ln \left(\frac{n'_{jf,t+1}}{n_{jf,t}} \right) \quad (8)$$

An equivalent expression for the per capita growth rate of allele j in males m can be obtained by exchanging f for m across the various subscripts in this expression.

Allelic coexistence in a sexual population, however, is ultimately influenced by growth and establishment of an allele across both sexes. Therefore, the full growth rate of allele j across the entire population of females *and* males is given by:

$$r_j = \ln \left(\frac{n'_{jf,t+1} + n'_{jm,t+1}}{n_{jf,t} + n_{jm,t}} \right). \quad (9)$$

An equivalent expression describes r_k , the growth rate of allele k .

Selection maintains both alleles in the population under the condition that:

$$\frac{S_m}{1 + S_m} < S_f < \frac{S_m}{1 - S_m} \quad (10)$$

(Kidwell *et al.*, 1977; Pamilo, 1979; Connallon & Hall, 2018) Thus, the maintenance of polymorphism of sexually antagonistic alleles is solely determined by the values of S_m and S_f . Note that in our model, the values S_m and S_f are bounded from 0 to 1. Therefore the **parameter space of sexually antagonistic selection** is within the range $0 < S_m, S_f < 1$. Classic theoretical models predict that in constant environments, only in ≈ 0.38 of the selection parameter space alleles can coexist (Kidwell *et al.*, 1977; Pamilo, 1979; Connallon *et al.*, 2018). If fluctuations in population sizes or selection values have an effect on the coexistence of sexually antagonistic alleles, it would be reflected in increases or decreases of the proportion of the parameter space of selection where polymorphism is maintained.

135 Simulations

136 Typically, would require decomposing alleles' growth rates (e.g., Eqn. 9) analytically to
137 examine the relative contributions of different types of fluctuations to their coexistence
138 (Barabás *et al.*, 2018). Although we present an analytical approach in the Supporting In-
139 formation, our general solution is not easily interpretable and soon becomes mathemati-
140 cally intractable (S1 Supporting Information). Thus, we opted for an extension of modern
141 coexistence theory that provides the flexibility to examine the contributions of different
142 processes to coexistence using simulations (Ellner *et al.*, 2019; Shoemaker *et al.*, 2020).

143 For each simulation, we examined coexistence outcomes across the selection param-
144 eter space of sexually antagonistic selection ($0 < S_m, S_f < 1$). To do so, we partitioned
145 the parameter space into a grid of 50×50 , which yielded 2500 pairwise combinations
146 of different w_{jm} and w_{kf} values. For each pairwise combination of w_{jm} and w_{kf} , as we
147 detail in the next sections, we iterated our model while controlling the effect size of fluc-
148 tuations in selection (σ_w) and their correlation (ρ_w), as well as fluctuations in population
149 sizes (σ_g) and their correlation (ρ_g). Then, we performed “invasion simulations” of each
150 allele invading a population, evaluated coexistence outcomes, and determined the rela-
151 tive contribution of each type of fluctuation. Finally, we calculated for each simulation
152 the proportion of the parameter space that allowed alleles to coexist.

153 We explored all of the combinations of low, intermediate, and high fluctuations in
154 fitness values and population sizes, with different extents of correlations between fluc-
155 tuations (Table 1). As a control simulation, we set $\sigma_w = 0.001$ and $\sigma_g = 0.001$, with no

correlation between fluctuations. We ran ten replicates per parameter combination, which resulted in 3780 simulations.

Timeseries

To incorporate the effects of fluctuations into our population dynamics model we generated independent timeseries of fluctuations in selection and population sizes. In the case of fluctuations in selection values, for a given value of w_{jm} and w_{kf} (i.e., a fixed point in the selection parameter space), we generated a timeseries of 500 timesteps made up of correlated fluctuations of w_{jm} and w_{kf} . We controlled the effect size of fluctuations in selection (σ_w) and its correlation (ρ_w) by using the Cholesky factorization of the variance-covariance matrix:

$$C_w = \begin{bmatrix} \sigma_w^2 & \rho_w \sigma_w^2 \\ \rho_w \sigma_w^2 & \sigma_w^2 \end{bmatrix} \quad (11)$$

We multiplied Eqn. 11 by a (2×500) matrix of random numbers from a normal distribution with mean 0 and unit variance, which yielded γ_j and γ_k . Then, we calculated the new fitness values at time $t + 1$ as $w_{jm,t+1} = w_{jm}^{\gamma_{j,t}}$ and $w_{kf,t+1} = w_{kf}^{\gamma_{k,t}}$.

Similarly, we generated a timeseries of 500 timesteps made up of correlated fluctuations in population sizes. We chose values of $N_m = 200$ and $N_f = 200$ as the initial value of population sizes throughout our simulations. We performed a Cholesky factorization of the variance-covariance matrix, controlling the effect size of fluctuations in population sizes with σ_g and their correlation with ρ_g . Similar to our previous approach, we multiplied this factorization by a random matrix of uncorrelated random variables,

which yielded γ_m and γ_f . Finally, we calculated the number of males and females in the population at time $t + 1$ as $N_{m,t+1} = N_m + \gamma_{m,t}$ and $N_{f,t+1} = N_f + \gamma_{f,t}$. Therefore, the population sizes in each timestep differed from the initial value of 200 individuals on the order of ρ_g . Note that the scales of σ_g and σ_w are different from each other. While σ_w controls the exponential change in fitness values in each timestep, σ_g controls the number of individuals added to a population in each timestep.

Finally, we performed simulations where our population dynamics model (Eqns. 1 to 9) iterated over 500 timesteps while allowing selection values and population sizes to fluctuate in each timestep. We started each simulation with the initial values of N_m and N_f described before and equal frequencies of allele j and allele k in each sex. For each timestep t in our simulations, the values of w_{jm} , w_{kf} , N_m and N_f used to calculate allele's frequencies in timestep t (e.g., Eqn. 7), corresponded to the t values calculated in each timeseries, as described previously. This approach yielded a final timeseries that captured the dynamics of sexually antagonistic alleles, with fluctuating values of selection and population sizes.

Invasion simulations

Modern coexistence theory has shown that coexistence is promoted by mechanisms that give species a population growth rate advantage over other species when they become rare (Chesson, 1982; 2003; Barabás *et al.*, 2018). Typically, one species is held at its *resident* state, as given by its steady-state abundances while the rare species is called the *invader*. In the context of alleles in a population, an allele is an *invader* when a muta-

tion occurs that introduces that allele into a population in which it is absent (e.g., if in a population with only k alleles, a random mutation made one individual carry the j allele). Within sexually antagonistic selection, each allele has two pathways of invasion, depending on whether the mutation arises in a female or in a male. If an allele's *invasion growth rate* (or the average instantaneous population growth rate when rare) is positive, it buffers it against extinction, maintaining its persistence in the population. Coexistence, and hence polymorphism, occurs when both alleles have positive invasion growth rates.

We used the timeseries that captured the dynamics of our population model as a template to perform invasion simulations of both alleles. We performed 500 independent invasion simulations, one for each timestep in our timeseries. We explored all four potential combinations of each allele invading through each pathway (e.g., allele j invading through males, and allele k invading through females, and so on). To simulate invasion, we set the density of the invading allele to one individual. For example, if allele j was invading via males, then we would set $n_{jm,i} = 1$ and $n_{jf,i} = 0$. Note that each invasion simulation was independent of the iteration that we used to generate the timeseries, therefore we denoted the initial timestep in an invasion simulation with the subscript i . We also set the resident allele, in this case k , to the corresponding value of the timeseries minus one individual, $n_{km,i} = N_{m,t} - 1$ and $n_{kf,i} = N_{f,t}$. Then, we iterated our model one timestep, $i + 1$, and calculated the logarithmic growth rate of j allele invading as:

$$r_j = \ln \left(\frac{n_{jm,i+1} + n_{jf,i+1}}{1} \right) \quad (12)$$

Correspondingly, the logarithmic growth rate of the k allele as a resident would be given by:

$$r_k = \ln \left(\frac{n_{km,i+1} + n_{kf,i+1}}{n_{km,i} + n_{kf,i}} \right) \quad (13)$$

Following the approach of Shoemaker *et al.* (2020), we treated each invasion simulation independently, and hence we performed 500 invasion simulations. We then calculated, for each allele invading via a different pathway, its mean invasion growth rate as the average of the 500 invasion growth rates. We also calculated the mean growth rate of the resident allele as the average of the 500 resident growth rates. We determined alleles to be coexisting if both of alleles had positive mean invasion growth rates, which is often referred to as the mutual invasibility criterion (Barabás *et al.*, 2018).

Functional decomposition

Our invasion simulations tell us whether or not sexually antagonistic alleles can coexist in a determined point of the selection parameter space. However, we also quantified the relative contributions of fluctuating selection and population sizes into the predicted coexistence outcome using a *functional decomposition* approach (Ellner *et al.*, 2016; 2019; Shoemaker *et al.*, 2020).

We applied the functional decomposition approach by breaking up the average growth rate of each allele into a null growth rate in the absences of fluctuations in all selected variables, a set of main effect terms that represent the effect of only one variable fluctuating, and a set of two-way interaction terms representing the effect of variables fluctuating simultaneously (Ellner *et al.*, 2019). In our simulations, this is a function of four variables:

the number of males in the population (N_m), the number of females in the population (N_f), the fitness of allele j in males (w_{jm}), and the fitness of allele k in females (w_{kf}). As an example, if only N_m and N_f were fluctuating, the growth rate of allele j when it is the invader at timestep t could be decomposed into:

$$r_{j,t}(N_m, N_f) = \mathcal{E}_j^0 + \mathcal{E}_j^{N_m} + \mathcal{E}_j^{N_f} + \mathcal{E}_j^{N_m N_f} \quad (14)$$

Where \mathcal{E}^0 is the null growth rate when N_m and N_f are set to their averages. Terms with superscripts represent the marginal effects of letting all superscripted variables vary while fixing all the other variables at their average values. For example, the term \mathcal{E}^{N_m} expresses the contribution of fluctuations in N_m when N_f is at its average, without the contribution when both variables are set to their averages :

$$\mathcal{E}_j^{N_m} = r_{j,t}(N_m, \overline{N_f}) - \mathcal{E}_j^0 \quad (15)$$

If we average Eqn. 14 across the timesteps in our simulation, we get a partition of the average population growth rate into the variance-free growth rate, the main effects of variability in N_m , the main effects of variability in N_f , and the interaction between variability in N_m and N_f

$$\bar{r}_j = \mathcal{E}_j^0 + \overline{\mathcal{E}_j^{N_m}} + \overline{\mathcal{E}_j^{N_f}} + \overline{\mathcal{E}_j^{N_m N_f}} \quad (16)$$

However, in our simulations w_{jm} and w_{kf} also fluctuated, therefore the full functional decomposition of the growth rate of allele j as an invader is found in Table 2, as well as

a brief description of the meaning of each term. The implementation and interpretation of the functional decomposition of the invasion growth rates of each allele are identical to each other. Note that Table 2 does not include three or four-way interactions (e.g., $\bar{\mathcal{E}}_j^{N_m N_f w_{jm} w_{fk}}$). This is because in our simulations, we did not allow fluctuations in selection and population sizes to be correlated, therefore their effects are solely captured by the terms in Table 2. We calculated the value of each of the terms in Table 2 by performing another set of invasion simulations as described previously, but instead of allowing all variables to fluctuate, systematically setting the required variables to their means and subtracting the corresponding \mathcal{E} values.

The functional decomposition approach further requires the *comparison* of each term, to understand if how it affects invaders and residents (i.e., the relative contribution). This is because fluctuations can promote coexistence by helping whichever allele is rare, or they can hurt whichever allele is common. Therefore, to understand the role of each type of fluctuation, it is necessary to compare how it affects invader *and* resident growth rates. In the example presented in Eqn. 16, if allele j is invading, then allele k is at its resident state and there exists an analogue decomposition of \bar{r}_k with the exact same terms as Eqn. 16. Therefore we can express the difference between contributions of fluctuations in N_m as:

$$\Delta_j^{N_m} = \bar{\mathcal{E}}_j^{N_m} - \bar{\mathcal{E}}_k^{N_m} \quad (17)$$

If $\Delta_j^{N_m}$ is positive, then fluctuations in the male population benefit allele j when it is

269 rare more than what they benefit k as a resident. If $\Delta_j^{N_m}$ is negative, then fluctuations
 270 benefit k as a resident more than j as an invader, and if it is minimal, then fluctuations
 271 have an equal effect in j and k . Therefore, for each allele invading via a different pathway,
 272 we calculated 7 Δ values, one for each one of the \mathcal{E} terms in Table 2. However, since the
 273 magnitude of each one of these values could vary considerably across the parameter space
 274 of selection, to make them comparable, we normalized each Δ value by dividing it by the
 275 square root of the sum of the squares of the 7 Δ values. For example, the normalized value
 276 of Eqn. 17 would be given by:

$$\delta_j^{N_m} = \frac{\Delta_j^{N_m}}{\sqrt{\sum_{d=1}^7 (\Delta_d)^2}} \quad (18)$$

277 This normalization bounded δ values from -1 to 1 .

278 **4 Results**

279 Our results showed that both fluctuations in selection and population sizes can substan-
 280 tially increase the expected genetic variability under sexually antagonistic selection. The
 281 average proportion of coexistence in the selection parameter space increased with the ef-
 282 fect size of fluctuations (Fig. 1). Increments in allelic coexistence were more likely when
 283 fluctuations were large, and fluctuations in population sizes were negatively correlated,
 284 while fluctuations in selection were positively correlated (Fig. 1). Importantly, our results
 285 show that environmental fluctuations can more than double the expected genetic varia-
 286 tion under sexually antagonistic selection, reaching up to ≈ 0.9 of allelic coexistence in

the selection parameter space (Fig. 1).

Fluctuations increased coexistence by allowing both alleles as invaders to have positive invasion growth rates in instances where selection would typically not allow them to (i.e., they made parts of the parameter space “flip” into coexistence). As a baseline, we show in Fig. 2A the outcome of the control simulation, which matched previous findings that without fluctuations, alleles can coexist in only ≈ 0.38 of the selection parameter space (Connallon & Hall, 2018). These “flips” occurred with both types of fluctuations and were more common with larger fluctuations and strongly correlated effects, for which we show examples in Fig. 2A . However, note that there are also parts of the parameter space where coexistence is lost compared to the control simulation, which was more likely when population sizes were fluctuating (Fig. 2A).

Alleles had positive invasion growth rates when positive contributions of fluctuations outweighed the negative contributions of fluctuations. As an example in Fig. 2B we show the functional decomposition of both alleles invading via their favored pathway in parts of the parameter space that “flipped” into coexistence and competitive exclusion (which correspond to the square and triangle in Fig. 2A). Note that each type of fluctuation made similar contributions to each allele, both when they were coexisting or experiencing competitive exclusion (Fig. 2B). However, δ_0 , which captures the effect of fluctuations set to their averages, switched between positive and negative contributions for both alleles (Fig. 2B).

The relative contribution of fluctuations in population sizes of males and females benefited alleles when alleles invaded via the fluctuating population (Fig. 2B and Fig. 3). If

alleles invaded via the opposite sex, then fluctuations contributed negatively to their invasion growth rate (Fig. 3). For example, the relative contribution of fluctuations in the male population, δ^{N_m} , was positive for both alleles when they invaded via males and negative when they invaded via females, regardless of the correlation between fluctuations (Fig. 3). The opposite pattern was shown by δ^{N_f} . The relative contribution of both populations fluctuating, $\delta^{N_m N_f}$, was positive when fluctuations were negatively correlated, it had a negligible effect when fluctuations were not correlated, and it had a negative effect when fluctuations were positively correlated (Fig. 3).

In contrast, fluctuations in selection benefited the allele that was unaffected by selection. For example, $\delta^{w_{jm}}$ which captured the relative contribution of fluctuations in selection against j in males, contributed negatively to allele j 's invasion growth rate but benefited allele k 's invasion, regardless of the correlation between fluctuations or the sex where invasion occurred (Fig. 4). The opposite pattern was shown by $\delta^{w_{kf}}$. The relative contribution of both types of selection fluctuating, was negative when fluctuations were negatively correlated, it had a negligible effect when fluctuations were not correlated, and it had a positive effect when fluctuations were positively correlated (Fig. 4).

The overall average effect of fluctuations, δ^0 , was at first glance unpredictable when fluctuations were incorporated (Fig. 2B). Without fluctuations, δ^0 captured exclusively the effect of selection and had positive values in parts of the parameter space where each allele could invade (Fig S1 Supporting Information). Fluctuations in population sizes generated stochastic changes of δ^0 across the parameter space (Fig. S1). In contrast, fluctuations in selection slightly changed the magnitude of δ^0 , which was heightened in parts

of the parameter space that were on the limit of the coexistence region (Fig. S1). When both selection and population sizes fluctuated, the value of δ_0 varied greatly across the selection parameter space (Fig. S1).

Nonetheless, changes in δ^0 can be explained by how we implemented the functional decomposition. Recall that δ_0 captured the relative contribution of fluctuations set to their mean. However, fluctuations set to their mean do not necessarily equal the mean values when there are no fluctuations. For example, we chose an initial value of 200 individuals of males and females for all of our simulations. Without fluctuations in population sizes, the mean population size for each sex was 200 individuals. In contrast, when we incorporated fluctuations in population sizes, the *realized mean* of population sizes used to calculate invasion growth rates varied from 100 to 500 individuals (Fig.S2). The realized mean of each population was correlated with the invasion growth rate of each allele, and thus the value of δ_0 (Fig.S2). For example, δ_0 tended to ‘flip’ from positive to negative compared to the control simulation if an allele invading via males was introduced to a male population with a realized mean greater than 200 individuals (Fig.S2). In contrast, δ_0 ‘flipped’ from negative to positive when an allele invading via males was introduced to a male population with a realized mean smaller than 200 individuals (Fig.S2). The opposite pattern was shown when alleles invaded via females (Fig.S3).

5 Discussion

The results of our study provide supporting evidence that environmental fluctuations can increase the expected genetic variance maintained under sexually antagonistic selec-

tion. Antagonistically selected alleles are an important component of genetic variation for many species (Foerster *et al.*, 2007; Van Doorn, 2009; Bonduriansky & Chenoweth, 2009; Innocenti & Morrow, 2010). Furthermore, as much as 20% of traits for which data is available are thought to be under sexually antagonistic selection (Morrissey, 2016). Yet, a large body of work suggests that the criteria for maintaining antagonistic genetic variation are very restrictive (i.e., we would expect the coexistence of alleles in few scenarios) (Kidwell *et al.*, 1977; Pamilo, 1979; Hedrick, 1999; Curtsinger *et al.*, 1994; Patten *et al.*, 2010). Our study shows that incorporating more realistic assumptions, such as non-constant selection and population sizes can more than double the expected genetic variation under sexually antagonistic selection (Fig. 1).

The relative contribution of fluctuations in selection

Our simulations indicate that fluctuations in selection can promote allelic coexistence in parts of the parameter space where we would typically expect selection to fix one of the alleles (Fig. 2). Fluctuations in selection had positive contributions to the invasion growth rate of the allele that was unaffected by selection and were dis-advantageous if the fluctuations directly affected the invading allele (e.g., $\delta_{w_{jm}}$ contributed positively to the growth rate of k and negatively to the growth rate of j , Fig. 4). The mechanism by which fluctuations in selection promoted coexistence can be understood as *relative non-linearity in response to selection* and arises because fluctuations in w_{jm} and w_{kf} do not affect both alleles equally, and thus each allele responds differently to them.

The term *relative non-linearity* refers to fluctuation-dependent coexistence mechanisms

that arise from competitors responding differently to limiting competitive factors (Chesson, 2000a; Ellner *et al.*, 2016; Zepeda & Martorell, 2019). Our results suggest that in the case of sexually antagonistic alleles, a non-advantageous allele can be maintained in a population if the favored allele experiences high fluctuations in selection (Fig. 4). This could be the case, for example, if traits associated with sexual dimorphism like ornaments or bright colors are also associated with higher predator rates (Bildstein *et al.*, 1989; Götmark *et al.*, 1997) or sex-biased mortality (Promislow *et al.*, 1992). However, if the non-advantageous allele is the one associated to higher fluctuations in selection, then fluctuations will likely erode genetic diversity.

The interactive effect of fluctuations in selection, $\delta_{w_{jm}, w_{kf}}$, accounts for the additional change in alleles' growth rates when both w_{jm} and w_{kf} vary, beyond the contribution of each effect varying on its own. This term only promoted allelic coexistence when fluctuations were positively correlated, while it contributed negatively to each allele's growth rate if fluctuations were negatively correlated (Fig. 4). Thus, in environments where selection on both alleles vary simultaneously, only coupled fluctuations benefit the maintenance of genetic diversity in a population. This could arise, for example, in environments where sexual selection on both sexes is stronger when climatic conditions are favorable and becomes negligible in stressful conditions (Cockburn *et al.*, 2008).

Temporal fluctuations in selection were initially thought to be of limited importance to the maintenance of polymorphism (Hedrick, 1974; 1986). However, some studies have shown that in populations with overlapping generations (Ellner & Sasaki, 1996; Sasaki & Ellner, 1995) and with sex-limited traits (Reinhold, 2000), fluctuations in selection over

time can maintain allelic coexistence due to the *storage effect*. This term refers to another coexistence mechanism that arises when fluctuations that alternate between favorable conditions to one species and those favorable to another, there exists a life-history stage for each species to survive unaffected by the unfavorable conditions, such as long-term dormancy (Chesson, 2000a; Ellner *et al.*, 2016). Similarly, Connallon & Hall (2018) found that when fluctuations in selection promote local adaptations due to life-history traits, the expected proportion of allelic coexistence in the selection parameter space can increase significantly. Our results provide further evidence that fluctuations in selection can promote the maintenance of genetic diversity, as sexual antagonism requires selection to differentially affect the alleles involved and thus promote non-linear responses to fluctuations.

The relative contribution of fluctuations in population sizes

Fluctuations in population sizes caused overall increases in the proportion of coexistence compared to the control simulation (Fig. 1). Similarly to fluctuations in selection, fluctuations in population sizes had positive contributions to the invasion growth rate of alleles due to *relative non linearity in response to population sizes*. In contrast to fluctuations in selection, fluctuations in population sizes benefited the allele that was directly affected by fluctuations (e.g., δ_{N_m} benefited the invasion of growth rate of an allele if it invaded via males Fig. 3). If an allele invaded via the non-fluctuating sex, however, fluctuations contributed negatively to its invasion growth rate (Fig. 3).

Therefore, our results suggest that a non-advantageous allele could have a positive

invasion growth rate if it invaded via a population experiencing temporal changes in its size. Temporal changes in population sizes of males and females can arise due to sex differences in movement (e.g., if males immigrate to higher quality areas (Matter & Roland, 2002)), development (e.g., females requiring more time to mature than males (Kasumovic *et al.*, 2008)), and behavior (e.g., cannibalistic mating (Elgar *et al.*, 2003)). When males and females experience different population dynamics, sexual antagonism allows alleles to differentiate in their response to fluctuations, and thus, promote allelic coexistence. The interactive effect of fluctuations in males and females, δ_{N_m, N_f} , shows that if both populations fluctuate, then negatively correlated fluctuations promote the maintenance of genetic diversity, while positively correlated fluctuations will likely impair it (Fig. 3). These insights offer an exciting avenue of research to understand if sexually selected traits are often found in populations that experience negatively correlated temporal changes in population sizes, and could help explain the high heritabilities of those traits (Reinhold, 2000).

Nonetheless, fluctuations in population sizes also caused competitive exclusion in some parts of the parameter space where we would expect selection to maintain both alleles (Fig. 2A). These “flips” into competitive exclusion were driven by changes to δ_0 compared to the control simulation when fluctuations in population sizes were incorporated (Fig. 2B). The stochastic changes in δ_0 were caused by changes to the realized mean population sizes in our simulations used to calculate the value of each allele mean invasion growth rate. Our results suggest that invasion is less likely to occur when an allele is introduced to a large population made up of predominantly the antagonistic allele (e.g.,

438). Note, however, that invasion growth rates are a function of four variables, and

439 Antagonistic selection is often ineffective in the face of genetic drift, with the impact
440 of drift being particularly pronounced at or near the parameter domain for balancing
441 selection (Connallon & Clark, 2012).

442 Sexually antagonistic alleles have a slow rate of evolutionary change Connallon &
443 Clark (2012) which also generates sensitivity to the effective size of a population. Genetic
444 drift may dominate over selection, even in populations that are quite large. There may be
445 little opportunity in small populations for stable balanced polymorphism at antagonistic
446 loci

447 **Allelic coexistence and sexual conflict**

448 Our study exclusively focused on the maintenance of polymorphism in a population.

449 **Conclusion**

450 Our study contributes to the growing body of work that shows that the criteria for main-
451 taining genetic variation under sexually antagonistic selection are overly conservative
452 (Connallon & Clark, 2012; Connallon *et al.*, 2018). Processes like recurrent mutations
453 (Radwan, 2008), genetic drift (Connallon & Clark, 2012), local adaptations in fluctuat-
454 ing environments (Connallon *et al.*, 2018), and alleles that experience seasonal changes in
455 dominance (Wittmann *et al.*, 2017) have been shown to dramatically change the levels of
456 sexually antagonistic variance in natural population. Our study

Figures and tables

Table 1: Parameters used in our simulations to control the effect size of fluctuations in population sizes (σ_g) and selection values (σ_w), as well as their respective correlations (ρ_g and ρ_w). We ran ten replicates for each one of the factorial combinations of the following parameters, which yielded a total of 3780 simulations.

Parameter	Values	Description
σ_w	0.001, 0.1, 0.3, 0.5, 0.7, 0.9	Effect size of fluctuations in fitness values
σ_g	0.001, 1, 10, 20, 30, 50	Effect size of fluctuations in population sizes
ρ_w	-0.75, 0, 0.75	Correlation between fluctuations in fitness values
ρ_g	-0.75, 0, 0.75	Correlation between fluctuation in population sizes

Table 2: Functional decomposition of the growth rate of allele j . As we exemplify in Eqn. 16, each term captures the contribution of fluctuations to an alleles' invasion growth rate.

Term	Formula	Meaning
\mathcal{E}_j^0	$\bar{r}_j(\bar{N}_m, \bar{N}_f, \bar{w}_{jm}, \bar{w}_{kf})$	Growth rate at mean population size and selection values.
$\bar{\mathcal{E}}_j^{N_m}$	$\bar{r}_j(N_m \bar{N}_f, \bar{w}_{jm}, \bar{w}_{kf}) - \mathcal{E}_j^0$	Main effect of fluctuations in N_m
$\bar{\mathcal{E}}_j^{N_f}$	$\bar{r}_j(\bar{N}_m, N_f \bar{w}_{jm}, \bar{w}_{kf}) - \mathcal{E}_j^0$	Main effect of fluctuations in N_f
$\bar{\mathcal{E}}_j^{w_{jm}}$	$\bar{r}_j(\bar{N}_m, \bar{N}_f, w_{jm}, \bar{w}_{kf}) - \mathcal{E}_j^0$	Main effect of fluctuations in w_{jm}
$\bar{\mathcal{E}}_j^{w_{kf}}$	$\bar{r}_j(\bar{N}_m, \bar{N}_f, \bar{w}_{jm}, w_{kf}) - \mathcal{E}_j^0$	Main effect of fluctuations in w_{kf}
$\bar{\mathcal{E}}_j^{N_m, N_f}$	$\bar{r}_j(N_m, N_f, \bar{w}_{jm}, \bar{w}_{kf}) - [\mathcal{E}_j^0 + \bar{\mathcal{E}}_j^{N_m} + \bar{\mathcal{E}}_j^{N_f}]$	Interaction of fluctuations in N_m and N_f
$\bar{\mathcal{E}}_j^{w_{jm}, w_{kf}}$	$\bar{r}_j(\bar{N}_m, \bar{N}_f, w_{jm}, w_{kf}) - [\mathcal{E}_j^0 + \bar{\mathcal{E}}_j^{w_{jm}} + \bar{\mathcal{E}}_j^{w_{kf}}]$	Interaction of fluctuations in w_{jm} and w_{kf}

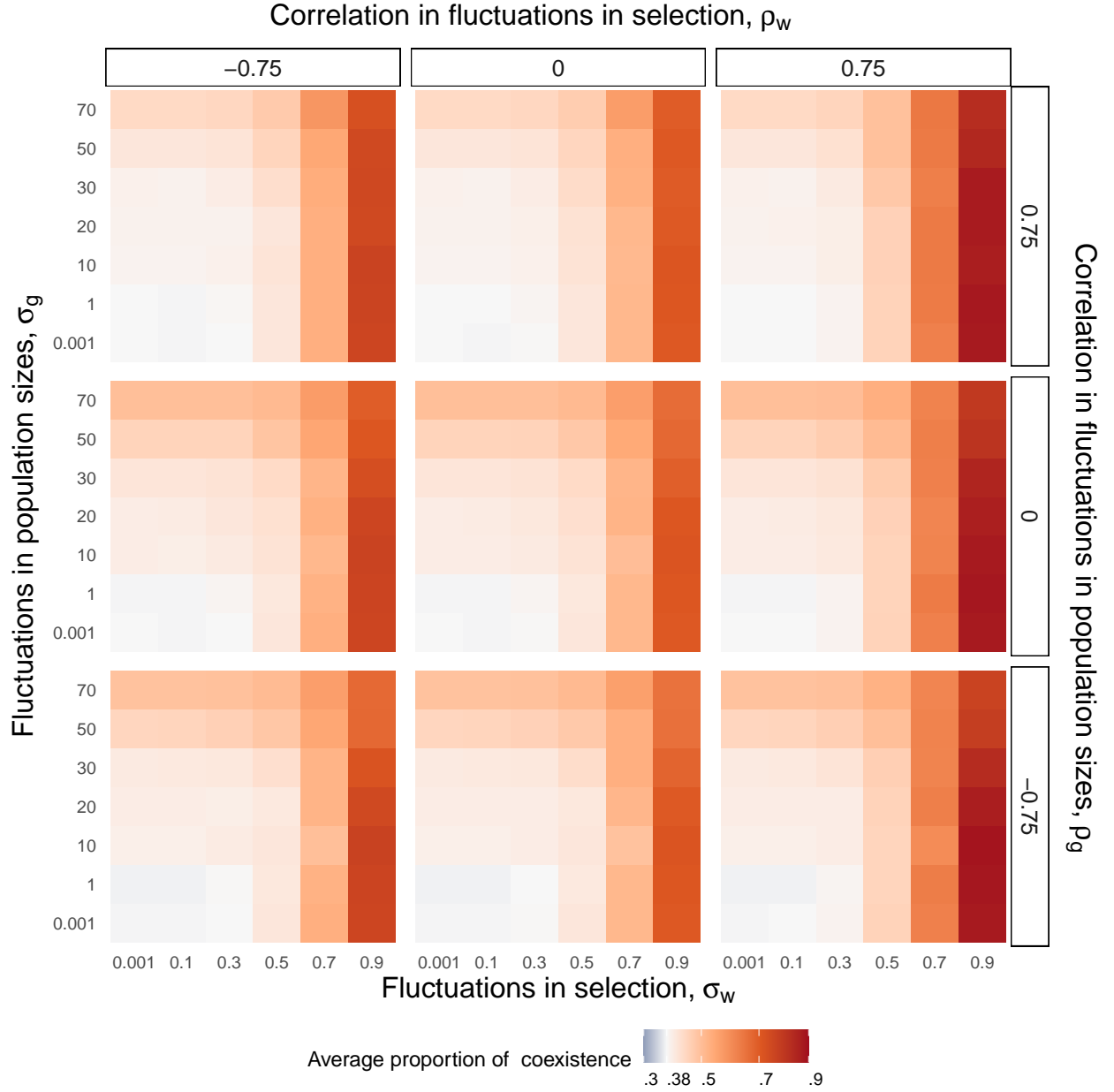


Figure 1: The average proportion of coexistence. For all parameter combinations in our simulations, we show the average proportion of coexistence across replicates and invasion types. Each panel corresponds to a different combination of correlations between fluctuations. Labels on top indicate the correlation between fluctuations in selection ρ_w , while labels on the right show the correlation in fluctuations between fluctuations in population sizes ρ_g . As a basis of comparison, we show the expected proportion of coexistence (0.38) as white in our color scheme.

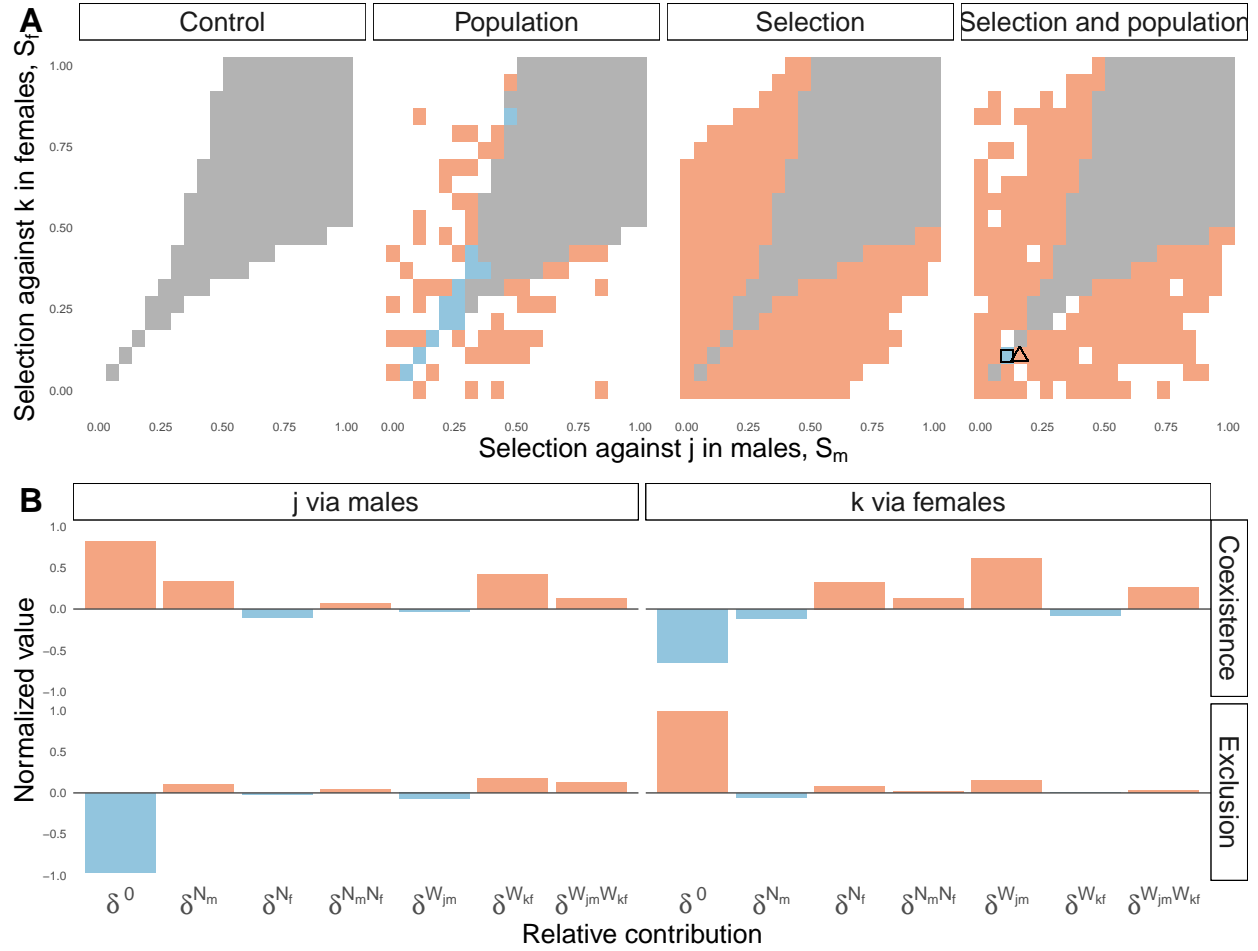


Figure 2: Coexistence outcomes and functional decomposition. In A) each panel corresponds to one replicate of a different type of simulation. All panels show the coexistence outcomes in the selection parameter space when j invaded via males and k invaded via females. As a reference, j is favored in females and k is favored in males. In the Control ($\sigma_g = 0.001$, $\rho_g = 0$, $\sigma_w = 0.001$, $\rho_w = 0$) grey areas indicate parts of the selection parameter space where alleles can coexist, while white areas indicate parts of the parameter space that correspond to competitive exclusion (following Eqn.10). In the Population ($\sigma_g = 70$, $\rho_g = -0.75$, $\sigma_w = 0.001$, $\rho_w = 0$), Selection ($\sigma_g = 0.001$, $\rho_g = 0$, $\sigma_w = 0.9$, $\rho_w = 0.75$), and Selection and population ($\sigma_g = 0.9$, $\rho_g = -0.75$, $\sigma_w = 0.9$, $\rho_w = 0.75$) panels, light red areas indicate parts of the parameter space that “flipped” into coexistence, while light blue areas show changes into competitive exclusion. We highlighted two points in the parameter space in the Selection and population panel that corresponded to changes into coexistence (triangle) and into competitive exclusion (square). In B) we show the functional decomposition of the coexistence and competitive exclusion points highlighted in A). Each panel corresponds to each allele invading via their respective pathway and shows the bar plots of the different δ values that made up the functional decomposition of each allele as an invader. Light red colors indicate positive δ values that benefited that allele as an invader more than the other allele as a resident, while light blue colors indicate negative δ values that benefited a resident more than the invader.

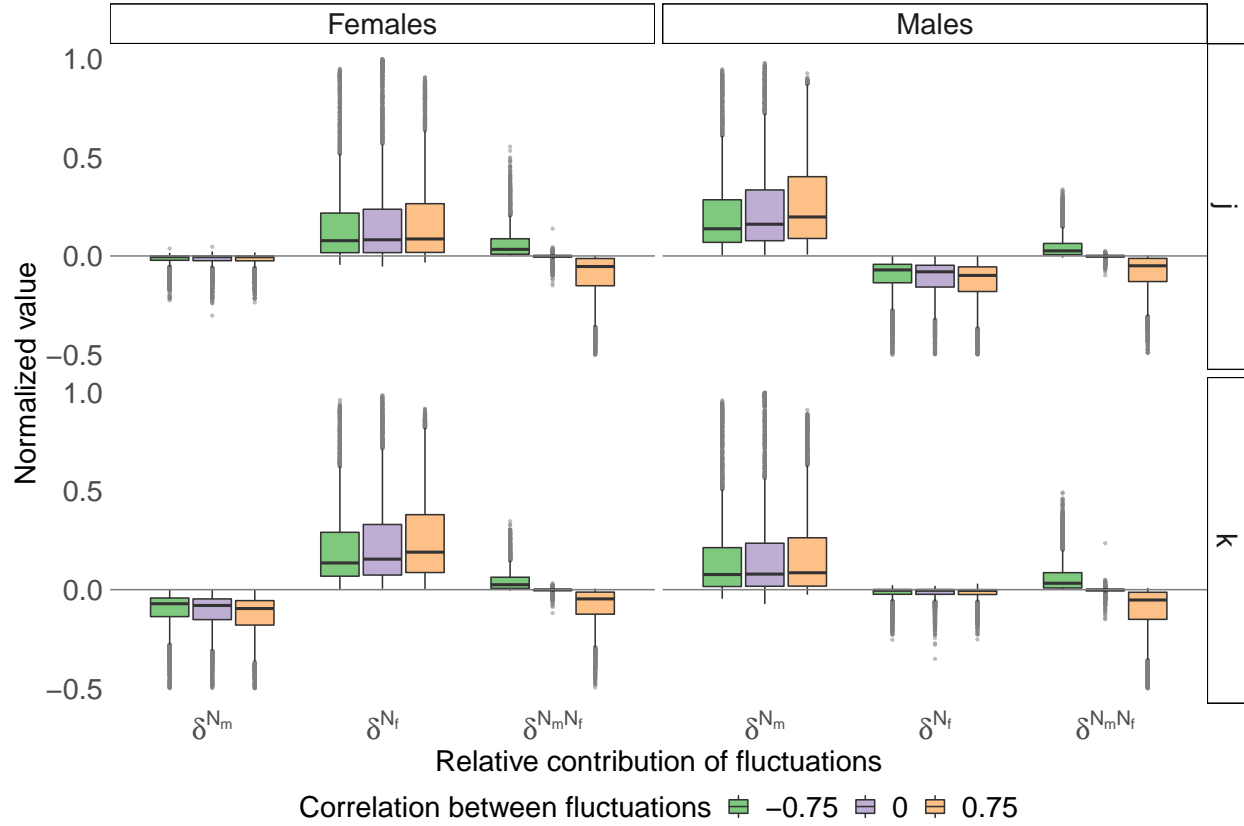


Figure 3: The relative contributions of fluctuations in population sizes. As a reference, positive δ values imply that the corresponding fluctuation benefits that allele as an invader more than the other allele as a resident, negative δ values indicate fluctuations benefit the residents more than the invader, and δ values close to zero indicate that the corresponding fluctuation has equal contributions to invaders and residents. Each panel corresponds to each allele invading via a different pathway, for which we show the boxplots of the three distinct δ values that captured the effects of fluctuations in population sizes, for all of the replicates in our simulation in which $\sigma_g = 70$. Each color corresponds to a different correlation between fluctuations in population sizes (ρ_g), as the legend indicates. Each box plot extends from the first to third quantiles of the corresponding posterior distribution of parameter values, and the line inside the the box indicates the median.

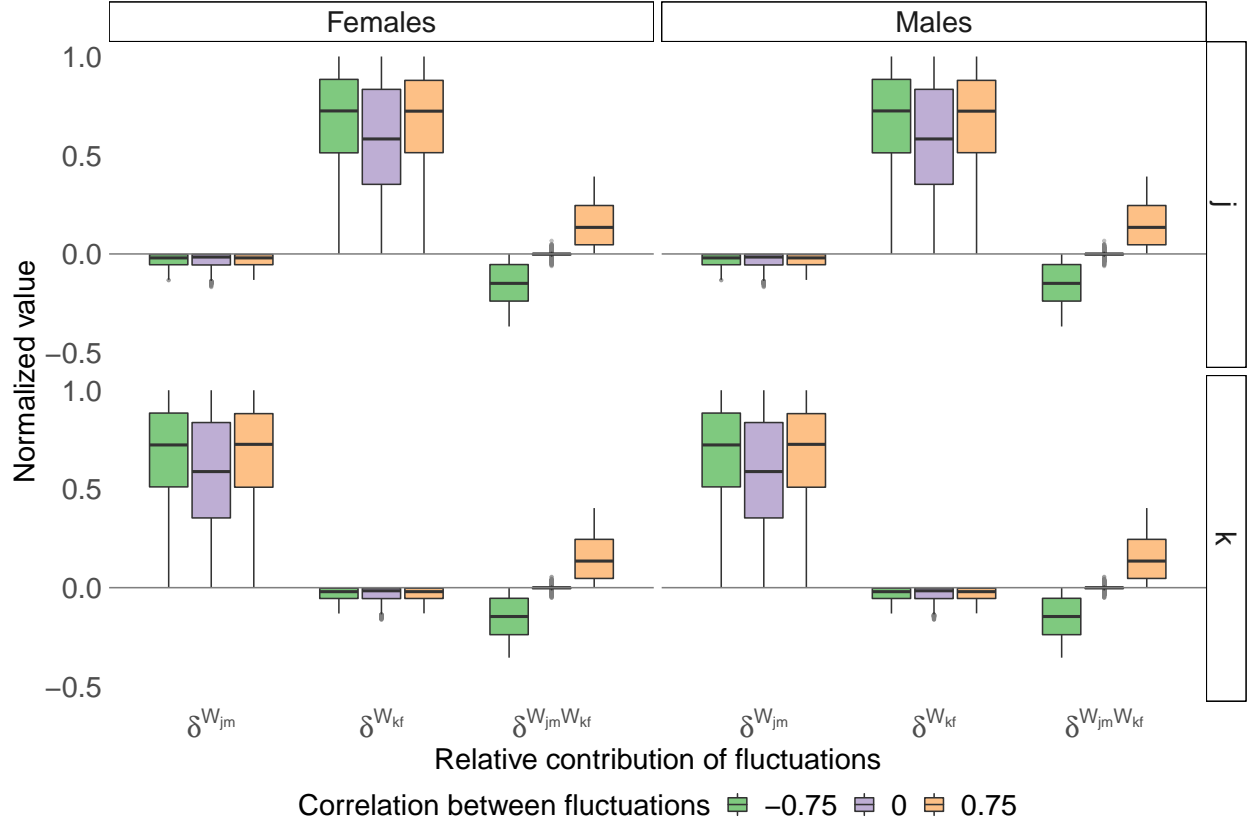


Figure 4: The relative contributions of fluctuations in selection. As a reference, positive δ values imply that the corresponding fluctuation benefits that allele as an invader more than the other allele as a resident, negative δ values indicate fluctuations benefit the residents more than the invader, and δ values close to zero indicate that the corresponding fluctuation has equal contributions to invaders and residents. Each panel corresponds to each allele invading via a different pathway, for which we show the boxplots of the three distinct δ values that captured the effects of fluctuations in population sizes, for all of the replicates in our simulation in which $\sigma_w = 0.90$. Each color corresponds to a different correlation between fluctuations in population sizes (ρ_w), as the legend indicates. Each box plot extends from the first to third quantiles of the corresponding posterior distribution of parameter values, and the line inside the the box indicates the median.

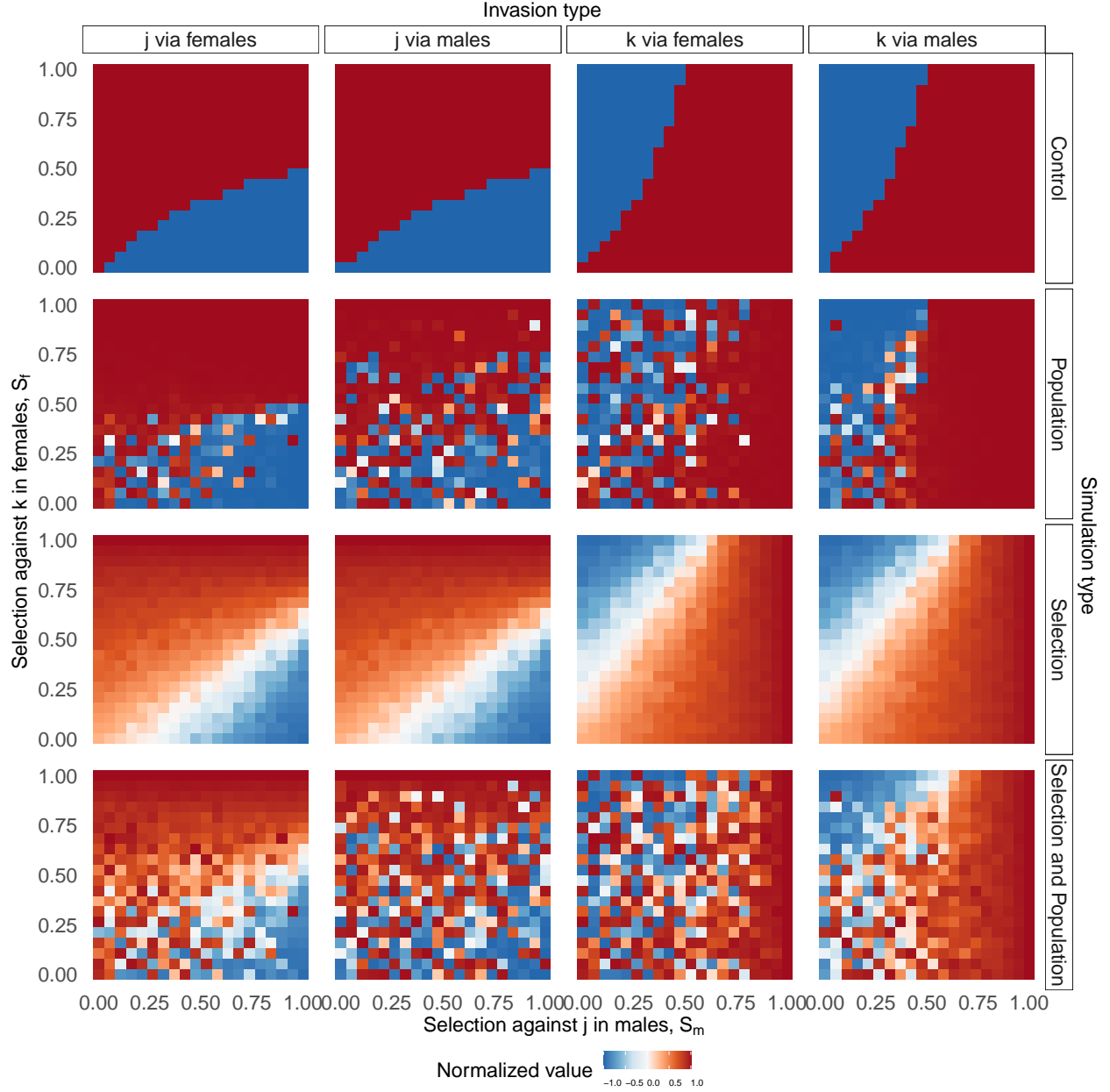


Figure 5: The normalized value of δ^0 in the selection parameter space. Each row in this figure corresponds to one replicate of a different type of simulation, while each column corresponds to a different type of invasion. In the Control simulation ($\sigma_g = 0.001$, $\rho_g = 0$, $\sigma_w = 0.001$, $\rho_w = 0$), δ^0 had a normalized value of 1 in parts of the parameter space where selection allows each allele to be fixed in a population, and a value of -1 where each allele can not be maintained in a population. The Population simulation corresponds to a replicate of a simulation where only population sizes fluctuated, without correlation between fluctuations ($\sigma_g = 70$, $\rho_g = 0$, $\sigma_w = 0.001$, $\rho_w = 0$). The Selection simulation corresponds to a replicate of a simulation where only selection fluctuated, without correlations between fluctuations ($\sigma_g = 0.001$, $\rho_g = 0$, $\sigma_w = 0.9$, $\rho_w = 0$). Finally, the Selection Population simulation corresponds to a replicate of a simulation where both selection and population sizes fluctuated, without correlations between fluctuations ($\sigma_g = 70$, $\rho_g = 0$, $\sigma_w = 0.9$, $\rho_w = 0$).

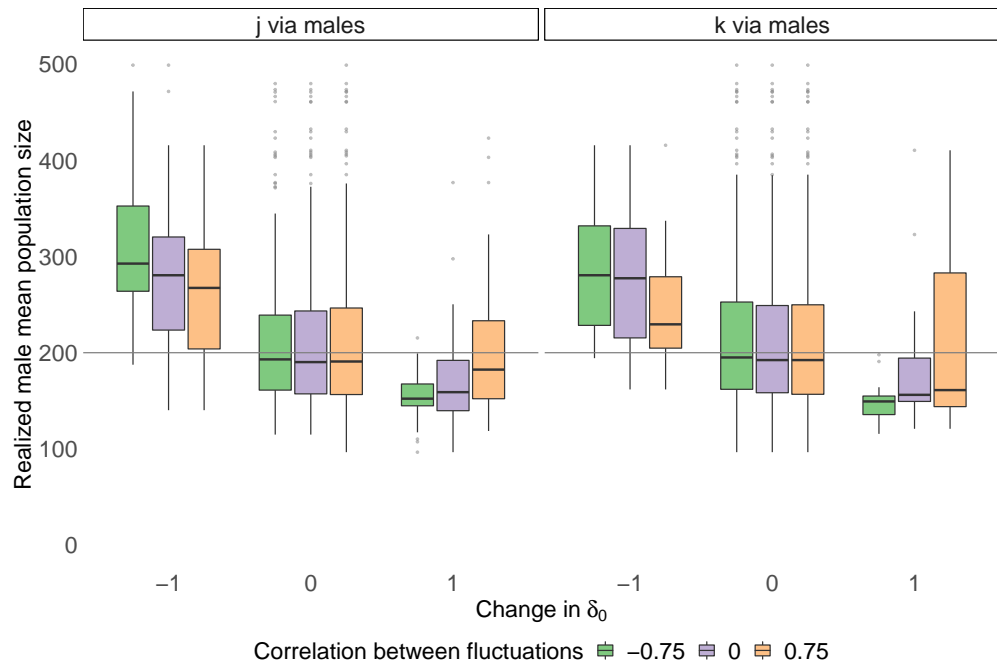


Figure 6: This is a caption

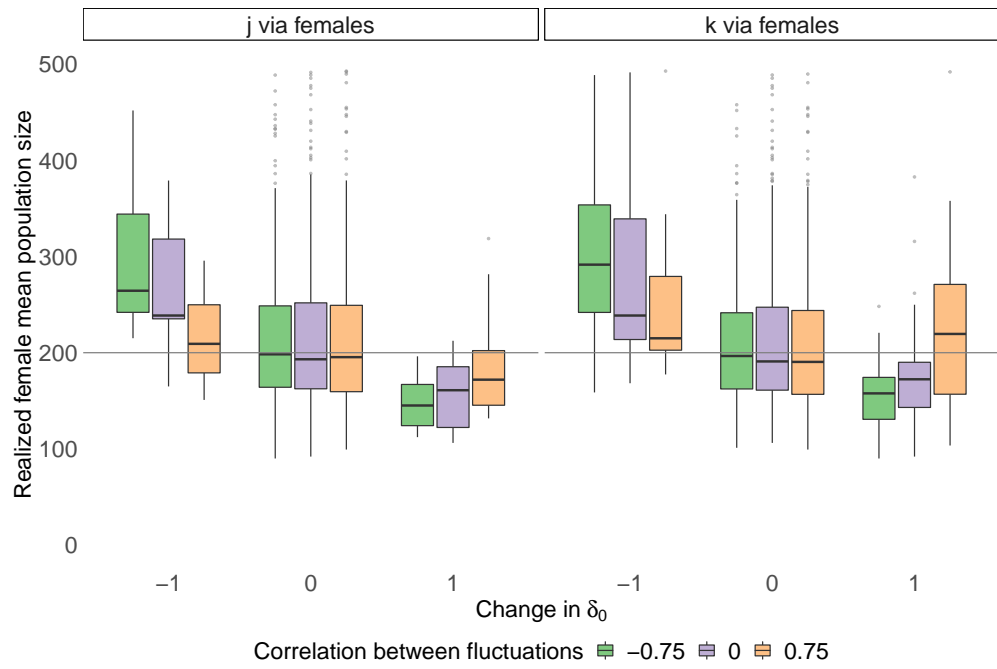


Figure 7: This is a caption

References

- Arnqvist, G. & Rowe, L. (2013). *Sexual conflict*. Princeton University Press.
- Barabás, G., D'Andrea, R. & Stump, S.M. (2018). Chesson's coexistence theory. *Ecological Monographs*, 88, 277–303.
- Bildstein, K.L., McDowell, S.G. & Brisbin, I.L. (1989). Consequences of sexual dimorphism in sand fiddler crabs, *uca pugilator*: differential vulnerability to avian predation. *Animal Behaviour*, 37, 133–139.
- Bonduriansky, R. & Chenoweth, S.F. (2009). Intralocus sexual conflict. *Trends in ecology & evolution*, 24, 280–288.
- Chesson, P. (1994). Multispecies competition in variable environments. *Theoretical population biology*, 45, 227–276.
- Chesson, P. (2000a). General theory of competitive coexistence in spatially-varying environments. *Theoretical Population Biology*, 58, 211–237.
- Chesson, P. (2000b). Mechanisms of maintenance of species diversity. *Annual review of Ecology and Systematics*, 31, 343–366.
- Chesson, P. (2003). Quantifying and testing coexistence mechanisms arising from recruitment fluctuations. *Theoretical Population Biology*, 64, 345–357.
- Chesson, P.L. (1982). The stabilizing effect of a random environment. *Journal of Mathematical Biology*, 15, 1–36.

- 477 Cockburn, A., Osmond, H.L. & Double, M.C. (2008). Swingin' in the rain: condition de-
478 pendence and sexual selection in a capricious world. *Proceedings of the Royal Society B:*
479 *Biological Sciences*, 275, 605–612.
- 480 Connallon, T. & Clark, A.G. (2012). A general population genetic framework for antag-
481 onistic selection that accounts for demography and recurrent mutation. *Genetics*, 190,
482 1477–1489.
- 483 Connallon, T. & Hall, M.D. (2018). Environmental changes and sexually antagonistic
484 selection. *eLS*, pp. 1–7.
- 485 Connallon, T., Sharma, S. & Olito, C. (2018). Evolutionary Consequences of Sex-Specific
486 Selection in Variable Environments: Four Simple Models Reveal Diverse Evolutionary
487 Outcomes. *The American Naturalist*, 193, 93–105.
- 488 Curtsinger, J.W., Service, P.M. & Prout, T. (1994). Antagonistic pleiotropy, reversal of
489 dominance, and genetic polymorphism. *The American Naturalist*, 144, 210–228.
- 490 Elgar, M.A., Bruce, M.J., De Crespigny, F.E.C., Cutler, A.R., Cutler, C.L., Gaskett, A.C.,
491 Herberstein, M.E., Ramamurthy, S. & Schneider, J.M. (2003). Male mate choice and
492 patterns of paternity in the polyandrous, sexually cannibalistic orb-web spider, *nephila*
493 *plumipes*. *Australian Journal of Zoology*, 51, 357–365.
- 494 Ellner, S. & Hairston Jr, N.G. (1994). Role of overlapping generations in maintaining
495 genetic variation in a fluctuating environment. *The American Naturalist*, 143, 403–417.

496 Ellner, S. & Sasaki, A. (1996). Patterns of genetic polymorphism maintained by fluctuating
 497 selection with overlapping generations. *theoretical population biology*, 50, 31–65.

498 Ellner, S.P., Snyder, R.E. & Adler, P.B. (2016). How to quantify the temporal storage effect
 499 using simulations instead of math. *Ecology letters*, 19, 1333–1342.

500 Ellner, S.P., Snyder, R.E., Adler, P.B. & Hooker, G. (2019). An expanded modern coexis-
 501 tence theory for empirical applications. *Ecology Letters*, 22, 3–18.

502 Foerster, K., Coulson, T., Sheldon, B.C., Pemberton, J.M., Clutton-Brock, T.H. & Kruuk,
 503 L.E. (2007). Sexually antagonistic genetic variation for fitness in red deer. *Nature*, 447,
 504 1107–1110.

505 Gavrillets, S. (2014). Is sexual conflict an “engine of speciation”? *Cold Spring Harbor*
 506 *perspectives in biology*, 6, a017723.

507 Götmark, F., Post, P., Olsson, J. & Himmelmann, D. (1997). Natural selection and sexual
 508 dimorphism: sex-biased sparrowhawk predation favours crypsis in female chaffinches.
 509 *Oikos*, pp. 540–548.

510 Hedrick, P.W. (1974). Genetic variation in a heterogeneous environment. i. temporal het-
 511 erogeneity and the absolute dominance model. *Genetics*, 78, 757–770.

512 Hedrick, P.W. (1986). Genetic polymorphism in heterogeneous environments: a decade
 513 later. *Annual review of ecology and systematics*, 17, 535–566.

514 Hedrick, P.W. (1999). Antagonistic pleiotropy and genetic polymorphism: a perspective.
 515 *Heredity*, 82, 126–133.

516 Immler, S., Arnqvist, G. & Otto, S.P. (2012). Ploidally antagonistic selection maintains
 517 stable genetic polymorphism. *Evolution: International Journal of Organic Evolution*, 66,
 518 55–65.

519 Innocenti, P. & Morrow, E.H. (2010). The sexually antagonistic genes of drosophila
 520 melanogaster. *PLoS biology*, 8, e1000335.

521 Kasumovic, M.M., Bruce, M.J., Andrade, M.C. & Herberstein, M.E. (2008). Spatial and
 522 temporal demographic variation drives within-season fluctuations in sexual selection.
 523 *Evolution: International Journal of Organic Evolution*, 62, 2316–2325.

524 Kidwell, J., Clegg, M., Stewart, F. & Prout, T. (1977). Regions of stable equilibria for
 525 models of differential selection in the two sexes under random mating. *Genetics*, 85,
 526 171–183.

527 Lande, R. (1980). Sexual dimorphism, sexual selection, and adaptation in polygenic char-
 528 acters. *Evolution*, pp. 292–305.

529 Matter, S.F. & Roland, J. (2002). An experimental examination of the effects of habitat
 530 quality on the dispersal and local abundance of the butterfly parnassius smintheus.
 531 *Ecological Entomology*, 27, 308–316.

532 Mitchell-Olds, T., Willis, J.H. & Goldstein, D.B. (2007). Which evolutionary processes
 533 influence natural genetic variation for phenotypic traits? *Nature Reviews Genetics*, 8,
 534 845–856.

Mori, E., Mazza, G. & Lovari, S. (2017). Sexual dimorphism. *Encyclopedia of Animal Cognition and Behavior* (J. Vonk, and T. Shakelford, Eds). Springer International Publishing, Switzerland, pp. 1–7.

Morrissey, M.B. (2016). Meta-analysis of magnitudes, differences and variation in evolutionary parameters. *Journal of Evolutionary Biology*, 29, 1882–1904.

Nunney, L. (2002). The effective size of annual plant populations: the interaction of a seed bank with fluctuating population size in maintaining genetic variation. *The American Naturalist*, 160, 195–204.

Pamilo, P. (1979). Genic variation at sex-linked loci: Quantification of regular selection models. *Hereditas*, 91, 129–133.

Patten, M.M., Haig, D. & Ubeda, F. (2010). Fitness variation due to sexual antagonism and linkage disequilibrium. *Evolution: International Journal of Organic Evolution*, 64, 3638–3642.

Pemberton, J., Smith, J., Coulson, T.N., Marshall, T.C., Slate, J., Paterson, S., Albon, S., Clutton-Brock, T.H. & Sneath, P.H.A. (1996). The maintenance of genetic polymorphism in small island populations: large mammals in the hebrides. *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences*, 351, 745–752.

Promislow, D.E., Montgomerie, R. & Martin, T.E. (1992). Mortality costs of sexual dimorphism in birds. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 250, 143–150.

- 555 Radwan, J. (2008). Maintenance of genetic variation in sexual ornaments: a review of the
556 mechanisms. *Genetica*, 134, 113–127.
- 557 Reinhold, K. (2000). Maintenance of a genetic polymorphism by fluctuating selection on
558 sex-limited traits. *Journal of Evolutionary Biology*, 13, 1009–1014.
- 559 Sasaki, A. & Ellner, S. (1995). The evolutionarily stable phenotype distribution in a ran-
560 dom environment. *Evolution*, 49, 337–350.
- 561 Shoemaker, L.G., Barner, A.K., Bittleston, L.S. & Teufel, A.I. (2020). Quantifying the rela-
562 tive importance of variation in predation and the environment for species coexistence.
563 *Ecology letters*, 23, 939–950.
- 564 Van Doorn, G.S. (2009). Intralocus sexual conflict. *Annals of the New York Academy of*
565 *Sciences*, 1168, 52–71.
- 566 Walsh, B. & Lynch, M. (2018). *Evolution and Selection of Quantitative Traits*. OUP Oxford.
- 567 Wittmann, M.J., Bergland, A.O., Feldman, M.W., Schmidt, P.S. & Petrov, D.A. (2017). Sea-
568 sonally fluctuating selection can maintain polymorphism at many loci via segregation
569 lift. *Proceedings of the National Academy of Sciences*, 114, E9932–E9941.
- 570 Zepeda, V. & Martorell, C. (2019). Fluctuation-independent niche differentiation and rel-
571 ative non-linearity drive coexistence in a species-rich grassland. *Ecology*, 100, e02726.