## Coexistence of alleles: insights of Modern

# Coexistence Theory into the maintenance of

## genetic diversity

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## <sub>9</sub> 1 Abstract

Sexually antagonistic selection (SAS) occurs when the selection in the traits or loci differs between the sexes. This sexual conflict offers the opportunity for maintaining poly-11 morphism in a population, but it often results in the eventual fixation of the fitter allele. However, the effects of SAS have generally been studied under strong simplifying as-13 sumptions, such as constant populations and homogeneous environments, which could considerably change the expected outcomes of SAS. Thus, in this study, we examined how fluctuations in selection and population sizes contributed to the coexistence of sexu-16 ally antagonistic alleles by adopting an ecological framework that allowed us to examine 17 evolutionary dynamics through the same lens as the coexistence of competing species. We performed simulations of alleles invading a population while allowing selection and 19 populations sizes to fluctuate over time. Then, we quantified coexistence outcomes and 20 the relative contribution of each type of fluctuation to each alleles' invasion growth rate. Our results showed that environmental fluctuations can dramatically increase the expected genetic variation under SAS. The positive contribution of fluctuations, however, 23 depended on the sex and allele where invasion occurred. This study contributes to the growing body of work that shows the importance of non-constant environments on the 25 maintenance of genetic diversity.

## 2 Introduction

The question of how genetic variation is maintained, despite the effects of selection and drift, continues to be central to the study of evolutionary biology (Walsh & Lynch, 2018).

Classical explanations include overdominance (heterozygote advantage) or frequencydependent selection, but in the modern era of genomic data, all patterns of variation that
exceed the expected variation under neutrality tend to be categorized broadly as balancing selection, regardless of the evolutionary mechanism (Mitchell-Olds *et al.*, 2007). One
of the evolutionary mechanisms coined under balancing selection is sexually antagonistic selection, which occurs when the direction of natural selection on traits or loci differs
between the sexes (Lande, 1980; Arnqvist & Rowe, 2013).

Sexually antagonistic selection can in some cases can mantain polymorphisms of otherwise dis-advantageous alleles in a population (Gavrilets, 2014), which in turn can result in phenotypically distinct sexes that express morphological, physiological, and behavioral traits to different degrees (Mori *et al.*, 2017; Connallon & Hall, 2018). However, sexually antagonistic selection can only mantain polymorphism in specific scenarios, as classical predictions show that sexual antagonism often results in the fixation of one of the alleles (Curtsinger *et al.*, 1994; Connallon *et al.*, 2018). Importantly, the effect of sexually antagonistic selection, has been generally studied under strong simplifying assumptions such as constant population sizes and homogeneous environments (e.g., Kidwell *et al.* (1977); Pamilo (1979); Immler *et al.* (2012)). Few studies have explored the effect of sexually antagonistic selection on the maintenance of polymorphism with more realistic

assumptions. Excepctions include Connallon *et al.* (2018) who found that classical predictions break down when fluctuations in the environment combined with life-history traits allow local adaptations and promote the maintenance of genetic diversity. The effect of environmental fluctuations without local adaptation, however, has not been studied in the context of sexually antagonistic selection to the best of our knowledge.

The contribution of environmental fluctuations to genetic variability remains a debated issue in evolutionary biology. Classic theoretical models predict that temporal fluctuations in environmental conditions are unlikely to maintain a genetic polymorphism (Hedrick, 1974; 1986). However, other studies have found that fluctuating selection can maintain genetic variance on sex-linked traits (Reinhold, 2000), or in populations where generations overlap (Ellner & Hairston Jr, 1994; Ellner & Sasaki, 1996). Similarly, temporal changes in population sizes have been shown to mitigate the effect of genetic drift in small populations (Pemberton *et al.*, 1996), and in annual plant systems (Nunney, 2002). Thus, both fluctuations in selection and population sizes could dramatically change the effect of sexually antagonistic selection in the maintenance of genetic diversity.

Importantly, progress requires more than just identifying if environmental fluctuations can maintain genetic diversity in a population, but to quantify how exactly they
contribute to its maintenance (Ellner *et al.*, 2016). Modern coexistence theory (modern
coexistence theory) provides a powerful conceptual framework to do so (Chesson, 2000b;
1994; Barabás *et al.*, 2018). Although its core ideas were formalized in an ecological context (Chesson, 1994; 2000a), this framework provides the necessary tools to examine the
relative contributions of fluctuations to diversity maintenance, which can also be applied

to evolutionary contexts (Ellner & Sasaki, 1996; Reinhold, 2000). From an ecological perspective, polymorphism of sexually antagonistic alleles is equivalent to the coexistence of species, and the fixation of either one of the alleles in a population is equivalent to competitive exclusion. The coexistence of alleles, thus, can be examined through the same lens as the coexistence of competing species.

Here, we seek to explicitly apply recent advances in modern coexistence theory to the question of how polymorphism is maintained under sexually antagonistic selection. We examined how fluctuations in selection values, fluctuations in population sizes, and their interactions can stabilize or hinder the coexistence of alleles. In particular, we examined i) Can fluctuations in population sizes and selection values allow sexually antagonistic alleles to coexist when differences in their fitness would typically not allow them to? and ii) What is the relative contribution of different types of fluctuations that allow two sexually antagonistic alleles to be maintained in a population? Our study provides the tools to analyze evolutionary dynamics from a novel perspective and contributes to answering long-lasting questions regarding the effect of non-constant environments on genetic diversity.

## 86 3 Methods

We first present a model that describes the evolutionary dynamics of sexually antagonistic alleles and show how changes in allele frequencies can be expressed in terms of growth rates, a necessary condition for analyses done using modern coexistence theory. We continue by simulating different scenarios of alleles invading a population, where we allowed population sizes, selection values, both, or neither to vary. Finally, we examine
the results of our simulations through a modern coexistence theory lens by calculating
the contribution of each of these fluctuations in the coexistence of alleles.

## Population dynamics of sexually antagonistic alleles

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Our model considered evolution at single, biallelic locus with frequency and density independent effects on the relative fitness of females and males. We examined the dynammics of two sexually antagonistic alleles, j and k, that affect fitness in the haploid state. We assumed allele j always has a high fitness in females ( $w_{jf}=1$ ), but variable fitness in males ( $w_{jm}<1$ ); and allele k has a high fitness in males ( $w_{km}=1$ ), but variable fitness in females ( $w_{kf}<1$ ). The selection against allele j in males is therefore  $S_m=1-w_{jm}$ , and the selection against allele k in females is  $S_f=1-w_{kf}$ .

The frequency of each allele in each sex at the beginning of a life-cycle at time t is given by:

$$p_{jm,t} = \frac{n_{jm,t}}{N_{m,t}} \tag{1}$$

$$p_{jf,t} = \frac{n_{jf,t}}{N_{f,t}} \tag{2}$$

 $p_{km,t} = \frac{N_{m,t} - n_{jm,t}}{N_{m,t}} \tag{3}$ 

$$p_{kf,t} = \frac{N_{f,t} - n_{jf,t}}{N_{f,t}} \tag{4}$$

where  $N_{m,t}$  and  $N_{t,t}$  are the numbers of males and females in a population at time t,  $n_{jf,t}$  is the number of females f with allele j, and  $n_{jm,t}$  is the number of males m with allele j at

time t, respectively.

The individuals in the population mate at random before selection occurs, and therefore the frequency of offspring with allele j after mating,  $p'_{j,t}$  can be expressed as:

$$p'_{j,t} = \frac{n_{jf}}{N_f} \frac{n_{jm}}{N_m} + \frac{1}{2} \frac{n_{jf}}{N_f} \frac{(N_m - n_{jm})}{N_m} + \frac{1}{2} \frac{(N_f - n_{jf})}{N_f} \frac{n_{jm}}{N_m},$$
 (5)

which upon rearranging and simplifying gives:

$$p'_{j,t} = \frac{(N_{m,t}n_{jf,t} + N_{f,t}n_{jm,t})}{2N_f N_m}.$$
 (6)

Selection acts upon these offspring in order to determine the allelic frequencies in females and males in the next generation, t + 1. As an example the frequency of females with allele j after selection is given by:

$$p'_{jf,t+1} = \frac{n_{jf,t+1}}{N'_{f,t+1}} = \frac{p'_j w_{jf}}{p'_j w_{jf} + (1 - p'_j) w_{kf}}$$
(7)

The logarithmic growth rate of j in females, is therefore given by the number of females with allele j after selection, divided by the original number of females carrying allele j:

$$r_{jf,t} = \ln\left(\frac{n'_{jf,t+1}}{n_{jf,t}}\right) \tag{8}$$

An equivalent expression for the per capita growth rate of allele j in males m can be obtained by exchanging f for m across the various subscripts in this expression.

Allelic coexistence in a sexual population, however, is ultimately influenced by growth and establishment of an allele across both sexes. Therefore, the full growth rate of allele j across the entire population of females *and* males is given by:

$$r_{j} = \ln \left( \frac{n'_{jf,t+1} + n'_{jm,t+1}}{n_{jf,t} + n_{jf,t}} \right) . \tag{9}$$

- An equivalent expression describes  $r_k$ , the growth rate of allele k.
- Selection mantains both alleles in the population under the condition that:

$$\frac{S_m}{1 + S_m} < S_f < \frac{S_m}{1 - S_m} \tag{10}$$

(Kidwell *et al.*, 1977; Pamilo, 1979; Connallon & Hall, 2018) Thus, the maintenance of polymorphism of sexually antagonistic alleles is solely determined by the values of  $S_m$  and  $S_f$ . Note that in our model, the values  $S_m$  and  $S_f$  are bounded from 0 to 1. Therefore the **parameter space of sexually antagonistic selection** is within the range  $0 < S_m, S_f < 1$ . Classic theoretical models predict that in constant environments, only in  $\approx 0.38$  of the selection parameter space alleles can coexist (Kidwell *et al.*, 1977; Pamilo, 1979; Connallon *et al.*, 2018). If fluctuations in population sizes or selection values have an effect on the coexistence of sexually antagonistic alleles, it would be reflected in increases or decreases of the proportion of the parameter space of selection where polymorphism is maintained.

### 135 Simulations

Typically, would require decomposing alleles' growth rates (e.g., Eqn. 9) analytically to examine the relative contributions of different types of fluctuations to their coexistence (Barabás *et al.*, 2018). Although we present an analytical approach in the Supporting Information, our general solution is not easily interpretable and soon becomes mathematically intractable (S1 Supporting Information). Thus, we opted for an extension of modern coexistence theory that provides the flexibility to examine the contributions of different processes to coexistence using simulations (Ellner *et al.*, 2019; Shoemaker *et al.*, 2020).

For each simulation, we examined coexistence outcomes across the selection param-143 eter space of sexually antagonistic selection (0  $< S_m, S_f < 1$ ). To do so, we partitioned 144 the parameter space into a grid of  $50 \times 50$ , which yielded 2500 pairwise combinations 145 of different  $w_{im}$  and  $w_{kf}$  values. For each pairwise combination of  $w_{im}$  and  $w_{kf}$ , as we detail in the next sections, we iterated our model while controlling the effect size of fluc-147 tuations in selection  $(\sigma_w)$  and their correlation  $(\rho_w)$ , as well as fluctuations in population 148 sizes  $(\sigma_g)$  and their correlation  $(\rho_g)$ . Then, we performed "invasion simulations" of each 149 allele invading a population, evaluated coexistence outcomes, and determined the rela-150 tive contribution of each type of fluctuation. Finally, we calculated for each simulation 151 the proportion of the parameter space that allowed alleles to coexist. 152

We explored all of the combinations of low, intermediate, and high fluctuations in fitness values and population sizes, with different extents of correlations between fluctuations (Table 1). As a control simulation, we set  $\sigma_w = 0.001$  and  $\sigma_g = 0.001$ , with no correlation between fluctuations. We ran ten replicates per parameter combination, which resulted in 3780 simulations.

#### 158 Timeseries

To incorporate the effects of fluctuations into our population dynamics model we generated independent timeseries of fluctuations in selection and population sizes. In the case of fluctuations in selection values, for a given value of  $w_{jm}$  and  $w_{kf}$  (i.e., a fixed point in the selection parameter space), we generated a timeseries of 500 timesteps made up of correlated fluctuations of  $w_{jm}$  and  $w_{kf}$ . We controlled the effect size of fluctuations in selection  $(\sigma_w)$  and its correlation  $(\rho_w)$  by using the Cholesky factorization of the variance-

$$C_w = \begin{bmatrix} \sigma_w^2 & \rho_w \sigma_w^2 \\ \rho_w \sigma_w^2 & \sigma_w^2 \end{bmatrix} \tag{11}$$

We multiplyed Eqn. 11 by a  $(2 \times 500)$  matrix of random numbers from a normal distribution with mean 0 and unit variance, which yielded  $\gamma_j$  and  $\gamma_k$ . Then, we calculated the new fitness values at time t+1 as  $w_{jm,t+1}=w_{jm}^{\gamma_{j,t}}$  and  $w_{kf,t+1}=w_{kf}^{\gamma_{k,t}}$ .

Similarly, we generated a timeseries of 500 timesteps made up of correlated fluctuations in population sizes. We chose values of  $N_m=200$  and  $N_f=200$  as the initial value of population sizes throughout our simulations. We performed a Cholesky factorization of the variance-covariance matrix, controlling the effect size of fluctuations in population sizes with  $\sigma_g$  and their correlation with  $\rho_g$ . Similar to our previous approach, we multiplied this factorization by a random matrix of uncorrelated random variables,

which yielded  $\gamma_m$  and  $\gamma_f$ . Finally, we calculated the number of males and females in the population at time t+1 as  $N_{m,t+1}=N_m+\gamma_{m,t}$  and  $N_{f,t+1}=N_f+\gamma_{f,t}$ . Therefore, the population sizes in each timestep differed from the inital value of 200 individuals on the order of  $\rho_g$ . Note that the scales of  $\sigma_g$  and  $\sigma_w$  are different from each other. While  $\sigma_w$  controls the exponential change in fitness values in each timestep,  $\sigma_g$  controls the number of individuals added to a population in each timestep.

Finally, we performed simulations where our population dynamics model (Eqns. 1 181 to 9) iterated over 500 timesteps while allowing selection values and population sizes 182 to fluctuate in each timestep. We started each simulation with the initial values of  $N_m$ 183 and  $N_f$  described before and equal frequencies of allele j and allele k in each sex. For each timestep t in our simulations, the values of  $w_{jm}$   $w_{kf}$ ,  $N_m$  and  $N_f$  used to calculate 185 allele's frequencies in timestep t (e.g., Eqn. 7), corresponded to the t values calculated 186 in each timeseries, as described previously. This approach yielded a final timeseries that captured the dynamics of sexually antagonistic alleles, with fluctuating values of selection 188 and population sizes. 189

#### 190 Invasion simulations

Modern coexistence theory has shown that coexistence is promoted by mechanisms that give species a population growth rate advantage over other species when they become rare (Chesson, 1982; 2003; Barabás *et al.*, 2018). Typically, one species is held at its *resident* state, as given by its steady-state abundances while the rare species is called the *invader*. In the context of alleles in a population, an allele is an *invader* when a muta-

tion occurs that introduces that allele into a population in which it is absent (e.g., if in a population with only k alleles, a random mutation made one individual carry the i al-197 lele). Within sexually antagonistic selection, each allele has two pathways of invasion, 198 depending on whether the mutation arises in a female or in a male. If an alleles' invasion growth rate (or the average instantaneous population growth rate when rare) is positive, 200 it buffers it against extinction, maintaining its persistence in the population. Coexistence, 201 and hence polymorphism, occurs when both alleles have positive invasion growth rates. 202 We used the timeseries that captured the dynamics of our population model as a tem-203 plate to perform invasion simulations of both alleles. We performed 500 independent 204 invasion simulations, one for each timestep in our timeseries. We explored all four po-205 tential combinations of each allele invading through each pathway (e.g., allele *j* invading 206 through males, and allele *k* invading through females, and so on). To simulate invasion, 207 we set the density of the invading allele to one individual. For example, if allele j was 208 invading via males, then we would set  $n_{jm,i} = 1$  and  $n_{jf,i} = 0$ . Note that each inva-209 sion simulation was independent of the iteration that we used to generate the timeseries, 210 therefore we denoted the initial timestep in an invasion simulation with the subscript i. 21 We also set the resident allele, in this case k, to the corresponding value of the timeseries minus one individual,  $n_{km,i} = N_{m,t} - 1$  and  $n_{kf,i} = N_{f,t}$ . Then, we iterated our model one 213 timestep, i + 1, and calculated the logarithmic growth rate of j allele invading as:

$$r_{j} = \ln\left(\frac{n_{jm,i+1} + n_{jf,i+1}}{1}\right) \tag{12}$$

Correspondingly, the logarithmic growth rate of the k allele as a resident would be given by:

$$r_k = \ln\left(\frac{n_{km,i+1} + n_{kf,i+1}}{n_{km,i} + n_{kf,i}}\right)$$
(13)

Following the approach of Shoemaker *et al.* (2020), we treated each invasion simulation independently, and hence we performed 500 invasion simulations. We then calculated, for each allele invading via a different pathway, its mean invasion growth rate as the average of the 500 invasion growth rates. We also calculated the mean growth rate of the resident allele as the average of the 500 resident growth rates. We determined alleles to be coexisting if both of alleles had positive mean invasion growth rates, which is often referred to as the mutual invasibility criterion (Barabás *et al.*, 2018).

#### 224 Functional decompostion

Our invasion simulations tell us whether or not sexually antagonistic alleles can coexistence outcome using a *functional decomposition* approach (Ellner *et al.*, 2016; 2019; Shoemaker *et al.*, 2020).

We applied the functional decomposition approach by breaking up the average growth rate of each allele into a null growth rate in the absences of fluctuations in all selected variables, a set of main effect terms that represent the effect of only one variable fluctuating, and a set of two-way interaction terms representing the effect of variables fluctuating simultaneously (Ellner *et al.*, 2019). In our simulations, this is a function of four variables:

the number of males in the population  $(N_m)$ , the number of females in the population  $(N_f)$ , the fitness of allele j in males  $(w_{jm})$ , and the fitness of allele k in females  $(w_{kf})$ . As an example, if only  $N_m$  and  $N_f$  were fluctuating, the growth rate of allele j when it is the invader at timestep t could be decomposed into:

$$r_{j,t}(N_m, N_f) = \mathcal{E}_j^0 + \mathcal{E}_j^{N_m} + \mathcal{E}_j^{N_f} + \mathcal{E}_j^{N_m N_f}$$
(14)

Where  $\mathcal{E}^0$  is the null growth rate when  $N_m$  and  $N_f$  are set to their averages. Terms with superscripts represent the marginal effects of letting all superscripted variables vary while fixing all the other variables at their average values. For example, the term  $\mathcal{E}^{N_m}$  expresses the contribution of fluctuations in  $N_m$  when  $N_f$  is at its average, without the contribution when both variables are set to their averages:

$$\mathcal{E}_{j}^{N_{m}} = r_{j,t}(N_{m}, \overline{N_{f}}) - \mathcal{E}_{j}^{0} \tag{15}$$

If we average Eqn. 14 across the timesteps in our simulation, we get a partition of the average population growth rate into the variance–free growth rate, the main effects of variability in  $N_m$ , the main effects of variability in  $N_f$ , and the interaction between variability in  $N_m$  and  $N_f$ 

$$\overline{r}_j = \mathcal{E}_j^0 + \overline{\mathcal{E}_j}^{N_m} + \overline{\mathcal{E}_j}^{N_f} + \overline{\mathcal{E}_j}^{N_m N_f}$$
(16)

However, in our simulations  $w_{jm}$  and  $w_{kf}$  also fluctuated, therefore the full functional decomposition of the growth rate of allele j as an invader is found in Table 2, as well as

a brief description of the meaning of each term. The implementation and interpretation of the functional decomposition of the invasion growth rates of each allele are identical 251 to each other. Note that Table 2 does not include three or four-way interactions (e.g., 252  $\overline{\mathcal{E}}_{j}^{N_{m}N_{f}w_{jm}w_{fk}}$ ). This is because in our simulations, we did not allow fluctuations in selection and population sizes to be correlated, therefore their effects are solely captured by the 254 terms in Table 2. We calculated the value of each of the terms in Table 2 by performing 255 another set of invasion simulations as described previously, but instead of allowing all 256 variables to fluctuate, systematically setting the required variables to their means and 257 subtracting the corresponding  $\mathcal{E}$  values. 258

The functional decomposition approach further requires the *comparison* of each term, 259 to understand if how it affects invaders and residents (i.e., the relative contribution). This 260 is because fluctuations can promote coexistence by helping whichever allele is rare, or 261 they can hurt whichever allele is common. Therefore, to understand the role of each 262 type of fluctuation, it is necessary to compare how it affects invader and resident growth 263 rates. In the example presented in Eqn. 16, if allele *j* is invading, then allele *k* is at it's 264 resident state and there exists an analogue decomposition of  $\bar{r}_k$  with the exact same terms 265 as Eqn. 16. Therefore we can express the difference between contributions of fluctuations 266 in  $N_m$  as: 267

$$\Delta_j^{N_m} = \overline{\mathcal{E}}_j^{N_m} - \overline{\mathcal{E}}_k^{N_m} \tag{17}$$

If  $\Delta_j^{N_m}$  is positive, then fluctuations in the male population benefit allele j when it is

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rare more than what they benefit k as a resident. If  $\Delta_j^{N_m}$  is negative, then fluctuations benefit k as a resident more than j as an invader, and if it is minimal, then fluctuations have an equal effect in j and k. Therefore, for each allele invading via a different pathway, we calculated 7  $\Delta$  values, one for each one of the  $\mathcal{E}$  terms in Table 2. However, since the magnitude of each one of these values could vary considerably across the parameter space of selection, to make them comparable, we normalized each  $\Delta$  value by dividing it by the square root of the sum of the squares of the 7  $\Delta$  values. For example, the normalized value of Eqn. 17 would be given by:

$$\delta_j^{N_m} = \frac{\Delta_j^{N_m}}{\sqrt{\sum_{d=1}^7 (\Delta_d)^2}} \tag{18}$$

This normalization bounded  $\delta$  values from -1 to 1.

## 278 4 Results

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Our results showed that both fluctuations in selection and population sizes can substantially increase the expected genetic variability under sexually antagonistic selection. The
average proportion of coexistence in the selection parameter space increased with the effect size of fluctuations when fluctuations were large enough (Fig. 1). Increments in allelic
coexistence were more likely when fluctuations were large, and fluctuations in population sizes were negatively correlated, while fluctuations in selection were positively correlated (Fig. 1). Importantly, our results show that environmental fluctuations can more
than double the expected genetic variation under sexually antagonistic selection, reaching

up to  $\approx 0.9$  of allelic coexistence in the selection parameter space (Fig. 1).

Fluctuations increased coexistence by allowing both alleles as invaders to have posi-288 tive invasion growth rates in instances where selection would typically not allow them to 289 (i.e., they made parts of the parameter space "flip" into coexistence). As a baseline, we show in Fig. 2A the outcome of the control simulation, which matched previous findings 291 that without fluctuations, alleles can coexist in only  $\approx 0.38$  of the selection parameter 292 space (Connallon & Hall, 2018). These "flips" occurred with both types of fluctuations 293 and were more common with larger fluctuations and strongly correlated effects, for which 294 we show examples in Fig. 2A. However, note that there are also parts of the parameter 295 space where coexistence is lost compared to the control simulation, which was more likely 296 when population sizes were fluctuating (Fig. 2A). 297

Alleles had positive invasion growth rates when positive contributions of fluctuations 298 outweighed the negative contributions of fluctuations. As an example in Fig. 2B we show 299 the functional decomposition of both alleles invading via their favored pathway in parts 300 of the parameter space that "flipped" into coexistence and competitive exclusion (which 301 correspond to the square and triangle in Fig. 2A). Note that each type of fluctuation made 302 similar contributions to each allele, both when they were coexisting or experiencing com-303 petitive exclusion (Fig. 2B). However,  $\delta_0$ , which captures the effect of fluctuations set 304 to their averages, switched between positive and negative contributions for both alleles 305 (Fig. 2B).

The relative contribution of fluctuations in population sizes of males and females benefited alleles when alleles invaded via the fluctuating population (Fig. 2B and Fig. 3). If alleles invaded via the opposite sex, then fluctuations contributed negatively to their invasion growth rate (Fig. 3). For example, the relative contribution of fluctuations in the male population,  $\delta^{N_m}$ , was positive for both alleles when they invaded via males and negative when they invaded via females, regardless of the correlation between fluctuations (Fig. 3). The opposite pattern was shown by  $\delta^{N_f}$ . The relative contribution of both populations fluctuating,  $\delta^{N_m N_f}$ , was positive when fluctuations were negatively correlated, it had a negligible effect when fluctuations were not correlated, and it had a negative effect when fluctuations were positively correlated (Fig. 3).

In contrast, fluctuations in selection benefited the allele that was unaffected by selection. For example,  $\delta^{w_{jm}}$  which captured the relative contribution of fluctuations in selection against j in males, contributed negatively to allele j's invasion growth rate but benefited allele k's invasion, regardless of the correlation between fluctuations or the sex where invasion occurred (Fig. 4). The opposite pattern was shown by  $\delta^{w_{kf}}$ . The relative contribution of both types of selection fluctuating, was negative when fluctuations were negatively correlated, it had a negligible effect when fluctuations were not correlated, and it had a positive effect when fluctuations were positively correlated (Fig. 4).

The overall average effect of fluctuations,  $\delta^0$ , was at first glance unpredictable when fluctuations were incorporated. Without fluctuations,  $\delta^0$  captured exclusively the effect of selection and had positive values in parts of the parameter space where each allele could invade (Fig S1 Supporting Information). Fluctuations in population sizes generated stochastic changes of  $\delta^0$  accross the parameter space and spanned a greater extent when alleles invade via their non-favored pathway (Fig S1). In contrast, fluctuations in selection slightly changed the magnitude of  $\delta^0$ , which was heightened in parts of the parameter space that were on the limit of the coexistence region (Fig S1). The stochastic changes in  $\delta^0$  when populations fluctuated, however, were correlated to the realized average population size during our simulations (Fig S2).

## 5 Discussion

The results of our study provide supporting evidence that environmental fluctuations 336 can increase the expected genetic variance maintained under sexually antagonistic selec-337 tion. Antagonistically selected alleles are an important component of genetic variation for many species (Foerster et al., 2007; Van Doorn, 2009; Bonduriansky & Chenoweth, 2009; 339 Innocenti & Morrow, 2010). Furthermore, as much as 20% of traits for which data is available are thought to be under sexually antagonistic selection (Morrissey, 2016). Yet, a large body of work suggests that the criteria for maintaining antagonistic genetic variation are 342 very restrictive (i.e., we would expect the coexistence of alleles in few scenarios) (Kid-343 well et al., 1977; Pamilo, 1979; Hedrick, 1999; Curtsinger et al., 1994; Patten et al., 2010). Our study shows that incorporating more realistic assumptions, such as non-constant se-345 lection and population sizes can more than double the expected genetic variation under 346 sexually antagonistic selection (Fig. 1).

#### The relative contribution of fluctuations in selection

Our simulations indicate that fluctuations in selection can promote allelic coexistence in parts of the parameter space where we would typically expect selection to fix one of the alleles (Fig. 2). Fluctuations in selection had positive contributions to the invasion growth rate of the allele that was unaffected by selection and were dis-advantageous if the fluctuations directly affected the invading allele (Fig. 4). How fluctuations in selection promoted coexistence can be understood as *relative non-linearity in response to selection* and arises because fluctuations in  $w_{jm}$  and  $w_{kf}$  do not affect both alleles equally, and thus each allele responds differently to fluctuations.

The term relative non-linearity refers to fluctuation-dependent coexistence mechanisms 357 that arise from competitors responding differently to limiting competitive factors (Ches-358 son, 2000a; Zepeda & Martorell, 2019). Our results suggest that in the case of sexually 359 antagonistic alleles, a non-advantageous allele can be maintained in a population if the 360 favored allele experiences high fluctuations in selection. This could be the case, for exam-36 ple, if traits associated with sexual dimorphism like ornaments or bright colors are also 362 associated with higher predator rates (Bildstein et al., 1989; Götmark et al., 1997) or sex-363 biased mortality (Promislow et al., 1992). The relative contribution of fluctuations in both 364  $w_{im}$  and  $w_{kf}$  was positive only when fluctuations were positively correlated (Fig. 4), sug-365 gesting that if the selection on both alleles is highly variable, coupled fluctuations benefit 366 the maintenance of genetic diversity in a population. 367

Temporal fluctuations in selection were initially thought to be of limited importance to the maintenance of polymorphism (Hedrick, 1974; 1986). However, some studies have shown that in populations with overlapping generations (Ellner & Sasaki, 1996; Sasaki & Ellner, 1995) and with sex-limited traits (Reinhold, 2000), fluctuations in selection over time can maintain allelic coexistence due to the *storage effect*. This term refers to another

coexistence mechanism that arises when fluctuations that alternate between favorable conditions to one species and those favorable to another, there exists a life-history stage 374 for each species to survive unaffected by the unfavorable conditions, such as long-term 375 dormancy (Chesson, 2000a; Ellner et al., 2016). Similarly, Connallon & Hall (2018) found that when fluctuations in selection promote local adaptations due to life-history traits, 377 the expected proportion of allelic coexistence in the selection parameter space can in-378 crease significantly. Our results provide further evidence that fluctuations in selection can promote the maintenance of genetic diversity, as sexual antagonism requires selec-380 tion to differentially affect the alleles involved and thus promote non-linear responses to 381 fluctuations. 382

#### 83 The relative contribution of fluctuations in population sizes

Discuss how our results show that fluctuations in population sizes benefit the allele that is invading via the fluctuating population. Put that in context in how much real populations fluctuate, and maybe highlight how an excess of genetic diversity could be explained by this mechanism (example).

Antagonistic selection is often ineffective in the face of genetic drift, with the impact of drift being particularly pronounced at or near the parameter domain for balancing selection (Connallon & Clark, 2012).

Sexually antagonistic alleles have a slow rate of evolutionary chanage Connallon & Clark (2012) which also generates sensitivity to the effective size of a population. Genetic drift my dominate over selection, even in populations that are quite large. There may be

little opportunity in small populations for stable balanced polymorphism at antagonistic loci

Many sex-limited traits involve interactions between individuals, and such traits should
be especially prone to be influenced by fluctuating population density. A male with a superior but costly coustly courtship signal might be in disadvatage under low populations
because only few females are available and the cost of the signal outweighs the advantage
of his attractiveness whereas under higher population density this male might achieve
many copulations and the benefit of attractiveness may thus surpass the cost of signal
production. (Reinhold, 2000)

#### Insights from the functional decomposition approach

In this paragraph we discuss how the functional decomposition approach showed not only the relative contributions of each type of fluctuations, but also captured unexpected stochassticity captured by  $\delta_0$ .

SAS might be resolved in extremely changing environments ??? (De Lisle et al., 2018)

### 408 Conclusion

Our study contributes to the growing body of work that shows that the criteria for maintaining genetic variation under sexually antagonistic selection are overly conservative
(Connallon & Clark, 2012; Connallon *et al.*, 2018). Processes like recurrent mutations
(Radwan, 2008), genetic drift (Connallon & Clark, 2012), local adaptations in fluctuating environments (Connallon *et al.*, 2018), and alleles that experience seasonal changes in

dominance (Wittmann et al., 2017) have been shown to dramatically change the levels of

sexually antagonistic variance in natural population. Our study contributes

## Figures and tables

Table 1: Parameters used in our simulations to control the effect size of fluctuations in population sizes ( $\sigma_g$ ) and selection values ( $\sigma_w$ ), as well as their respective correlations ( $\rho_g$  and  $\rho_w$ ). We ran ten replicates for each one of the factorial combinations of the following parameters, which yielded a total of 3780 simulations.

| Parameter  | Values                         | Description   |
|------------|--------------------------------|---|
| $\sigma_w$ | 0.001, 0.1, 0.3, 0.5, 0.7, 0.9 | Effect size of fluctuations in fitness values       |
| $\sigma_g$ | 0.001, 1, 10, 20, 30, 50       | Effect size of fluctuations in population sizes     |
| $ ho_w$    | -0.75, 0, 0.75                 | Correlation between fluctuations in fitness values  |
| $\rho_g$   | -0.75, 0, 0.75                 | Correlation between fluctuation in population sizes |

Table 2: Functional decomposition of the growth rate of allele *j*. As we exemplify in Eqn. 16, each term captures the contribution of fluctuations to an alleles' invasion growth rate.

| Term  | Formula  | Meaning   |
|---|--|---|
| $\mathcal{E}_{i}^{0}$   | $\overline{r_j}(\overline{N_m},\overline{N_f},\overline{w_{jm}},\overline{w_{kf}})$  | Growth rate at mean population size and selection values. |
| $ \frac{\overline{\mathcal{E}}_{j}^{N_{m}}}{\overline{\mathcal{E}}_{j}^{N_{f}}} $ $ \frac{\overline{\mathcal{E}}_{j}^{N_{f}}}{\overline{\mathcal{E}}_{jm}^{w_{jm}}} $ | $\overline{r}_j(N_m\overline{N_f},\overline{w_{jm}},\overline{w_{kf}})-\mathcal{E}_j^0$  | Main effect of fluctuations in $N_m$                      |
| $\overline{\mathcal{E}}_{j}^{N_{f}}$  | $\overline{r_j}(\overline{N_m},N_f,\overline{w_{jm}},\overline{w_{kf}})-\mathcal{E}_j^0$   | Main effect of fluctuations in $N_f$                      |
| $\overline{\mathcal{E}}_{j}^{w_{jm}}$   | $\overline{r_j}(\overline{N_m},\overline{N_f},w_{jm},\overline{w_{kf}})-\mathcal{E}_j^0$   | Main effect of fluctuations in $w_{jm}$                   |
| $\overline{\mathcal{E}}_{i}^{w_{kf}}$   | $\overline{r_j}(\overline{N_m},\overline{N_f},\overline{w_{jm}},w_{kf})-\mathcal{E}_j^0$   | Main effect of fluctuations in $w_{kf}$                   |
| $\overline{\mathcal{E}}_{j}^{N_{m},N_{f}}$  | $\overline{r_j}(N_m, N_f, \overline{w_{jm}}, \overline{w_{kf}}) - [\mathcal{E}_j^0 + \overline{\mathcal{E}}_j^{N_m} + \overline{\mathcal{E}}_j^{N_f}]$       | Interaction of fluctuations in $N_m$ and $N_f$            |
| $\overline{\mathcal{E}}_{j}^{w_{jm},w_{kf}}$  | $\overline{r_j}(\overline{N_m}, \overline{N_f}, w_{jm}, w_{kf}) - [\mathcal{E}_j^0 + \overline{\mathcal{E}_j^{w_{jm}}} + \overline{\mathcal{E}_j^{w_{kf}}}]$ | Interaction of fluctuations in $w_{jm}$ and $w_{kf}$      |

#### Correlation in fluctuations in selection, $\rho_w$ -0.750 0.75 70 50 30 0.75 20 Correlation in fluctuations in population sizes, $\rho_{\text{g}}$ 10 Fluctuations in population sizes, $\sigma_{g}$ 0.001 70 50 30 20 10 0.001 70 50 30 20 10 0.001 0.7 0.9 0.001 0.1 0.3 0.5 0.7 0.9 0.001 0.1 0.5 0.3 0.5 Fluctuations in selection, $\sigma_{\rm w}$ Average proportion of coexistence .3 .38 .5

Figure 1: The average proportion of coexistence. For all parameter combinations in our simulations, we show the average proportion of coexistence across replicates and invasion types. Each panel corresponds to a different combination of correlations between fluctuations. Labels on top indicate the correlation between fluctuations in selection  $\rho_w$ , while labels on the right show the correlation in fluctuations between fluctuations in population sizes  $\rho_g$ . As a basis of comparison, we show the expected proportion of coexistence (0.38) as white in our color scheme.

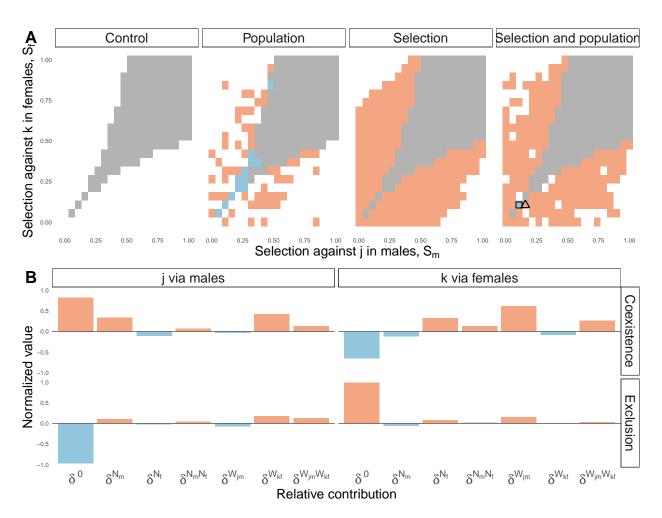


Figure 2: Coexistene outcomes and functional decomposition. In A) each panel corresponds to one replicate of a different type of simulation. All panels show the coexistence outcomes in the selection parameter space when *j* invaded via males and *k* invaded via females. As a reference, *j* is favored in females and k is favored in males. In the Control panel ( $\sigma_g = 0.001$ ,  $\rho_g = 0$ ,  $\sigma_w = 0$ 0.001,  $\rho_w = 0$ ) grey areas indicate parts of the selection parameter space where alleles can coexist, while white areas indicate parts of the parameter space that correspond to competitive exclusion (following Eqn.10). In the Population ( $\sigma_g = 70$ ,  $\rho_g = -0.75$ ,  $\sigma_w = 0.001$ ,  $\rho_w = 0$ ), Selection  $(\sigma_g = 0.001, \rho_g = 0, \sigma_w = 0.9, \rho_w = 0.75)$ , and Selection and population  $(\sigma_g = 0.9, \rho_g = -0.75, \rho_g = 0.001)$  $\sigma_w = 0.9$ ,  $\rho_w = 0.75$ ) panels, light red areas indicate parts of the parameter space that "flipped" into coexistence, while light blue areas show changes into competitive exclusion. We highlighted two points in the parameter space in the Selection and population panel that corresponded to changes into coexistence (triangle) and into competitive exlcusion (square). In B) we show the functional decomposition of the coexistence and competitive exclusion points highlighted in A). Each panel corresponds to each allele invading via their respective pathway and shows the bar plots of the different  $\delta$  values that made up the functional decomposition of each allele as an invader. Light red colors indicate positive  $\delta$  values that benefited that allele as an invader more than the other allele as a resident, while light blue colors indicate negative  $\delta$  values that benefited a resident more than the invader.

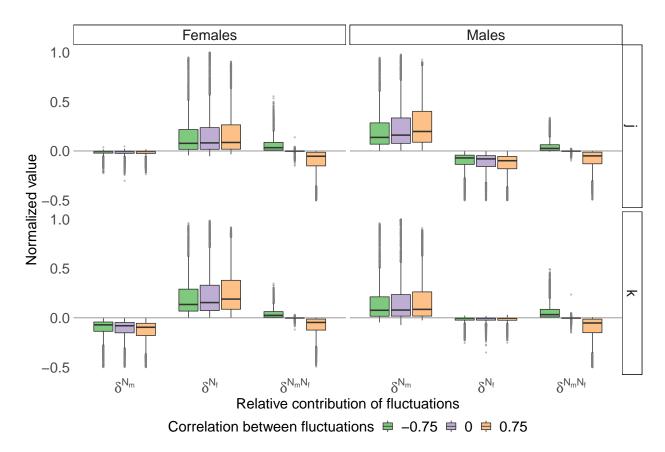


Figure 3: The relative contributions of fluctuations in population sizes. As a reference, positive  $\delta$  values imply that the corresponding fluctuation benefits that allele as an invader more than the other allele as a resident, negative  $\delta$  values indicate fluctuations benefit the residents more than the invader, and  $\delta$  values close to zero indicate that the corresponding fluctuation has equal contributions to invaders and residents. Each panel corresponds to each allele invading via a different pathway, for which we show the boxplots of the three distinct  $\delta$  values that captured the effects of fluctuations in population sizes, for all of the replicates in our simulation in which  $\sigma_g = 70$ . Each color corresponds to a different correlation between fluctuations in population sizes ( $\rho_g$ ), as the legend indicates. Each box plot extends from the first to third quantiles of the corresponding posterior distribution of parameter values, and the line inside the the box indicates the median.

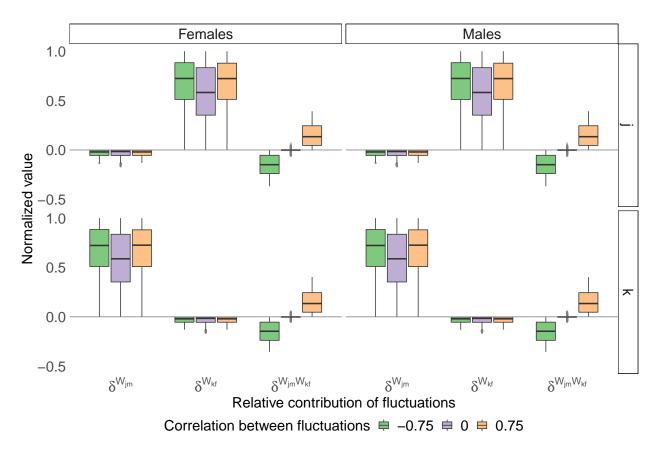


Figure 4: The relative contributions of fluctuations in selection. As a reference, positive  $\delta$  values imply that the corresponding fluctuation benefits that allele as an invader more than the other allele as a resident, negative  $\delta$  values indicate fluctuations benefit the residents more than the invader, and  $\delta$  values close to zero indicate that the corresponding fluctuation has equal contributions to invaders and residents. Each panel corresponds to each allele invading via a different pathway, for which we show the boxplots of the three distinct  $\delta$  values that captured the effects of fluctuations in population sizes, for all of the replicates in our simulation in which  $\sigma_w = 0.90$ . Each color corresponds to a different correlation between fluctuations in population sizes ( $\rho_w$ ), as the legend indicates. Each box plot extends from the first to third quantiles of the corresponding posterior distribution of parameter values, and the line inside the the box indicates the median.

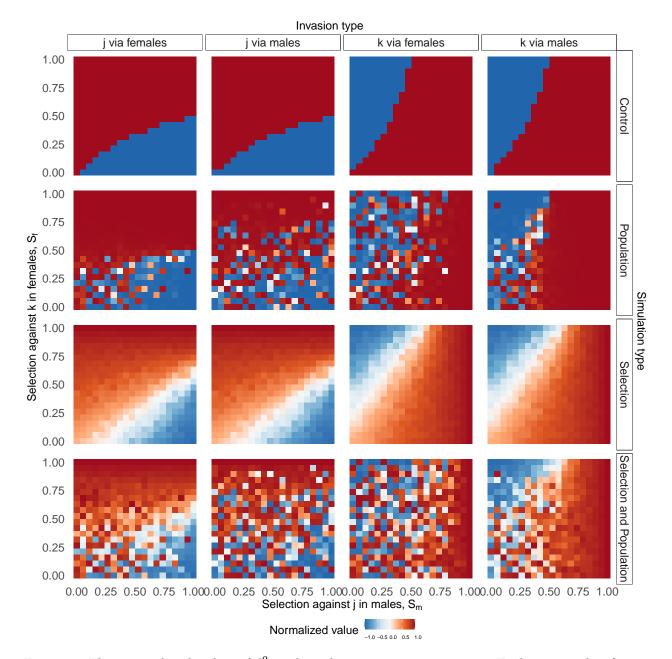


Figure 5: The normalized value of  $\delta^0$  in the selection parameter space. Each row in this figure corresponds to one replicate of a different type of simulation, while each column corresponds to a different type of invasion. In the Control simulation ( $\sigma_g = 0.001$ ,  $\rho_g = 0$ ,  $\sigma_w = 0.001$ ,  $\rho_w = 0$ ),  $\delta^0$  had a normalized value of 1 in parts of the parameter space where selection allows each allele to be fixed in a population, and a value of -1 where each allele can not be mantained in a population. The Population simulation corresponds to a replicate of a simulation where only population sizes fluctuated, without correlation between fluctuations ( $\sigma_g = 70$ ,  $\rho_g = 0$ ,  $\sigma_w = 0.001$ ,  $\rho_w = 0$ ). The Selection simulation corresponds to a replicate of a simulation where only selection fluctuated, without correlations between fluctuations ( $\sigma_g = 0.001$ ,  $\rho_g = 0$ ,  $\sigma_w = 0.9$ ,  $\rho_w = 0$ ). Finally, the Selection Population simulation corresponds to a replicate of a simulation where both selection and population sizes fluctuated, without correlations between fluctuations ( $\sigma_g = 70$ ,  $\rho_g = 0$ ,  $\sigma_w = 0.9$ ,  $\rho_w = 0$ ).

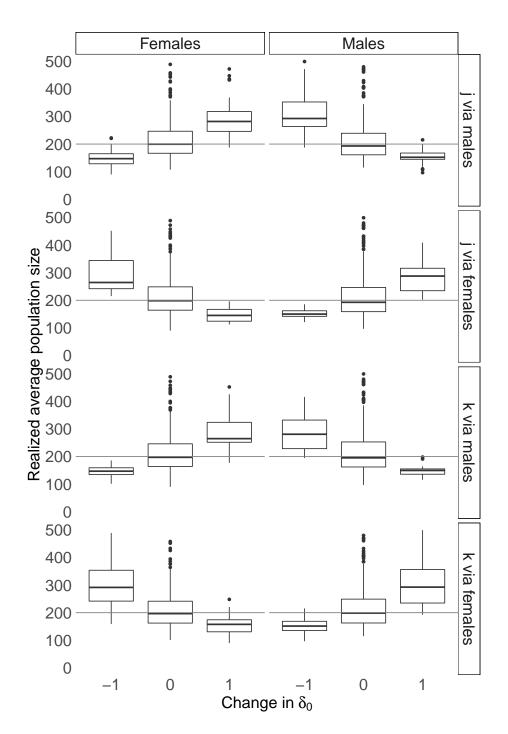


Figure 6: This is a caption

## References

- <sup>418</sup> Arnqvist, G. & Rowe, L. (2013). Sexual conflict. Princeton University Press.
- Barabás, G., D'Andrea, R. & Stump, S.M. (2018). Chesson's coexistence theory. *Ecological*
- 420 *Monographs*, 88, 277–303.
- Bildstein, K.L., McDowell, S.G. & Brisbin, I.L. (1989). Consequences of sexual dimorphism
- in sand fiddler crabs, uca pugilator: differential vulnerability to avian predation. Ani-
- *mal Behaviour*, 37, 133–139.
- Bonduriansky, R. & Chenoweth, S.F. (2009). Intralocus sexual conflict. *Trends in ecology &*
- evolution, 24, 280–288.
- <sup>426</sup> Chesson, P. (1994). Multispecies competition in variable environments. *Theoretical popula-*
- *tion biology*, 45, 227–276.
- <sup>428</sup> Chesson, P. (2000a). General theory of competitive coexistence in spatially-varying envi-
- ronments. *Theoretical Population Biology*, 58, 211–237.
- <sup>430</sup> Chesson, P. (2000b). Mechanisms of maintenance of species diversity. *Annual review of*
- Ecology and Systematics, 31, 343–366.
- 432 Chesson, P. (2003). Quantifying and testing coexistence mechanisms arising from recruit-
- ment fluctuations. *Theoretical Population Biology*, 64, 345–357.
- <sup>434</sup> Chesson, P.L. (1982). The stabilizing effect of a random environment. *Journal of Mathemat-*
- *ical Biology*, 15, 1–36.

- Connallon, T. & Clark, A.G. (2012). A general population genetic framework for antag-
- onistic selection that accounts for demography and recurrent mutation. Genetics, 190,
- 438 1477–1489.
- Connallon, T. & Hall, M.D. (2018). Environmental changes and sexually antagonistic
- selection. eLS, pp. 1–7.
- Connallon, T., Sharma, S. & Olito, C. (2018). Evolutionary Consequences of Sex-Specific
- Selection in Variable Environments: Four Simple Models Reveal Diverse Evolutionary
- Outcomes. *The American Naturalist*, 193, 93–105.
- 444 Curtsinger, J.W., Service, P.M. & Prout, T. (1994). Antagonistic pleiotropy, reversal of
- dominance, and genetic polymorphism. *The American Naturalist*, 144, 210–228.
- De Lisle, S.P., Goedert, D., Reedy, A.M. & Svensson, E.I. (2018). Climatic factors and
- species range position predict sexually antagonistic selection across taxa. *Philosophical*
- Transactions of the Royal Society B: Biological Sciences, 373, 20170415.
- Ellner, S. & Hairston Jr, N.G. (1994). Role of overlapping generations in maintaining
- genetic variation in a fluctuating environment. *The American Naturalist*, 143, 403–417.
- Ellner, S. & Sasaki, A. (1996). Patterns of genetic polymorphism maintained by fluctuating
- selection with overlapping generations. *theoretical population biology*, 50, 31–65.
- Ellner, S.P., Snyder, R.E. & Adler, P.B. (2016). How to quantify the temporal storage effect
- using simulations instead of math. *Ecology letters*, 19, 1333–1342.

- Ellner, S.P., Snyder, R.E., Adler, P.B. & Hooker, G. (2019). An expanded modern coexistence theory for empirical applications. *Ecology Letters*, 22, 3–18.
- Foerster, K., Coulson, T., Sheldon, B.C., Pemberton, J.M., Clutton-Brock, T.H. & Kruuk,
- L.E. (2007). Sexually antagonistic genetic variation for fitness in red deer. *Nature*, 447,
- 459 1107–1110.
- Gavrilets, S. (2014). Is sexual conflict an "engine of speciation"? *Cold Spring Harbor perspectives in biology*, 6, a017723.
- Götmark, F., Post, P., Olsson, J. & Himmelmann, D. (1997). Natural selection and sexual
- dimorphism: sex-biased sparrowhawk predation favours crypsis in female chaffinches.
- Oikos, pp. 540–548.
- Hedrick, P.W. (1974). Genetic variation in a heterogeneous environment. i. temporal het-
- erogeneity and the absolute dominance model. *Genetics*, 78, 757–770.
- Hedrick, P.W. (1986). Genetic polymorphism in heterogeneous environments: a decade
- later. *Annual review of ecology and systematics*, 17, 535–566.
- Hedrick, P.W. (1999). Antagonistic pleiotropy and genetic polymorphism: a perspective.
- 470 *Heredity*, 82, 126–133.
- Immler, S., Arnqvist, G. & Otto, S.P. (2012). Ploidally antagonistic selection maintains
- stable genetic polymorphism. Evolution: International Journal of Organic Evolution, 66,
- 473 55–65.

- Innocenti, P. & Morrow, E.H. (2010). The sexually antagonistic genes of drosophila melanogaster. *PLoS biology*, 8, e1000335.
- Kidwell, J., Clegg, M., Stewart, F. & Prout, T. (1977). Regions of stable equilibria for models of differential selection in the two sexes under random mating. *Genetics*, 85,
- 478 171–183.
- Lande, R. (1980). Sexual dimorphism, sexual selection, and adaptation in polygenic characters. *Evolution*, pp. 292–305.
- Mitchell-Olds, T., Willis, J.H. & Goldstein, D.B. (2007). Which evolutionary processes influence natural genetic variation for phenotypic traits? *Nature Reviews Genetics*, 8, 845–856.
- Mori, E., Mazza, G. & Lovari, S. (2017). Sexual dimorphism. *Encyclopedia of Animal Cognition and Behavior (J. Vonk, and T. Shakelford, Eds). Springer International Publishing, Switzer-land,* pp. 1–7.
- Morrissey, M.B. (2016). Meta-analysis of magnitudes, differences and variation in evolutionary parameters. *Journal of Evolutionary Biology*, 29, 1882–1904.
- Nunney, L. (2002). The effective size of annual plant populations: the interaction of a seed
  bank with fluctuating population size in maintaining genetic variation. *The American*Naturalist, 160, 195–204.
- Pamilo, P. (1979). Genic variation at sex-linked loci: Quantification of regular selection models. *Hereditas*, 91, 129–133.

- Patten, M.M., Haig, D. & Ubeda, F. (2010). Fitness variation due to sexual antagonism and linkage disequilibrium. *Evolution: International Journal of Organic Evolution*, 64, 3638–3642.
- Pemberton, J., Smith, J., Coulson, T.N., Marshall, T.C., Slate, J., Paterson, S., Albon, S.,
- <sup>498</sup> Clutton-Brock, T.H. & Sneath, P.H.A. (1996). The maintenance of genetic polymorphism
- in small island populations: large mammals in the hebrides. *Philosophical Transactions*
- of the Royal Society of London. Series B: Biological Sciences, 351, 745–752.
- Promislow, D.E., Montgomerie, R. & Martin, T.E. (1992). Mortality costs of sexual dimor-
- phism in birds. Proceedings of the Royal Society of London. Series B: Biological Sciences, 250,
- 503 143–150.
- Radwan, J. (2008). Maintenance of genetic variation in sexual ornaments: a review of the mechanisms. *Genetica*, 134, 113–127.
- Reinhold, K. (2000). Maintenance of a genetic polymorphism by fluctuating selection on sex-limited traits. *Journal of Evolutionary Biology*, 13, 1009–1014.
- Sasaki, A. & Ellner, S. (1995). The evolutionarily stable phenotype distribution in a random environment. *Evolution*, 49, 337–350.
- Shoemaker, L.G., Barner, A.K., Bittleston, L.S. & Teufel, A.I. (2020). Quantifying the rela-
- tive importance of variation in predation and the environment for species coexistence.
- 512 *Ecology letters*, 23, 939–950.

- Van Doorn, G.S. (2009). Intralocus sexual conflict. *Annals of the New York Academy of*Sciences, 1168, 52–71.
- Walsh, B. & Lynch, M. (2018). Evolution and Selection of Quantitative Traits. OUP Oxford.
- Wittmann, M.J., Bergland, A.O., Feldman, M.W., Schmidt, P.S. & Petrov, D.A. (2017). Sea-
- sonally fluctuating selection can maintain polymorphism at many loci via segregation
- lift. Proceedings of the National Academy of Sciences, 114, E9932–E9941.
- <sup>519</sup> Zepeda, V. & Martorell, C. (2019). Fluctuation-independent niche differentiation and rel-
- ative non-linearity drive coexistence in a species-rich grassland. *Ecology*, 100, e02726.