Coexistence of alleles: insights of Modern

Coexistence Theory into the maintenance of

genetic diversity

- ⁴ Alba Cervantes-Loreto¹, Michelle L. Marraffini¹, Daniel B. Stouffer¹, and
- 5 Sarah P. Flanagan¹

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- ¹Centre for Integrative Ecology, School of Biological Sciences, University of Canterbury,
- 7 Christchurch 8140, New Zealand

Words in abstract to be determined

Words in manuscript to be determined

Number of references to be determined

Number of figures to be determined

Number of tables 2

Number of text boxes 0

Corresponding author Alba Cervantes-Loreto

Phone +64 369 2880

Email alba.cervantesloreto@pg.canterbury.ac.nz

1 Introduction

The question of how genetic variation is maintained, despite the effects of selection and drift, continues to be central to the study of evolutionary biology (Walsh & Lynch, 2018). Classical explanations include overdominance (heterozygote advantage) or frequency-dependent selection, but in the modern era of genomic data, all patterns of variation that exceed the expected variation under neutrality tend to be categorized broadly as balancing selection, regardless of the evolutionary mechanism (Mitchell-Olds *et al.*, 2007). One of the evolutionary mechanisms coined under balancing selection is sexually antagonistic selection, which occurs when the direction of natural selection on traits or loci differs between the sexes (Lande, 1980; Arnqvist & Rowe, 2013).

Sexually antagonistic selection can in some cases can mantain polymorphisms of otherwise dis-advantageous alleles in a population in a population (Gavrilets, 2014), which in turn can result in phenotypically distinct sexes that express morphological, physiological, and behavioral traits to different degrees (Mori *et al.*, 2017; Connallon & Hall, 2018). However, sexually antagonistic selection can only mantain polymorphism in specific scenarios, as classical predictions show that sexual antagonism often results in the fixation of one of the alleles (Curtsinger *et al.*, 1994; Connallon *et al.*, 2018). Importantly, the effect of sexually antagonistic selection, has been generally studied under strong simplifying assumptions such as constant population sizes and homogeneous environments (e.g., Kidwell *et al.* (1977); Pamilo (1979); Immler *et al.* (2012)). Few studies have explored the effect of sexually antagonistic selection on the maintenance of polymorphism with

more realistic assumptions. Exceptions include Connallon et al. (2018) who found that classical predictions break down when fluctuations in the environment combined with 31 life-history traits allow local adaptations and promote the maintenance of genetic diversity. The effect of environmental fluctuations without local adaptation, however, has not been studied in the context of sexually antagonistic selection to the best of our knowledge. The contribution of environmental fluctuations to genetic variability remains a de-35 bated issue in evolutionary biology. Classic theoretical models predict that temporal fluctuations in environmental conditions are unlikely to maintain a genetic polymorphism 37 (Hedrick, 1974; 1986). However, other studies have found that fluctuating selection can 38 maintain genetic variance on sex-linked traits (Reinhold, 2000), or in populations where generations overlap (Ellner & Hairston Jr, 1994; Ellner & Sasaki, 1996). Similarly, tempo-40 ral changes in population sizes have been shown to mitigate the effect of genetic drift in small populations (Pemberton *et al.*, 1996), and in annual plant systems (Nunney, 2002). Thus, both fluctuations in selection and population sizes could dramatically change the effect of sexually antagonistic selection in the maintenance of genetic diversity. Importantly, progress requires more than just identifying if environmental fluctua-45 tions can maintain genetic diversity in a population, but to quantify how exactly they contribute to its maintenance (Ellner et al., 2016). Modern coexistence theory (MCT) provides a powerful conceptual framework to do so (Chesson, 2000b; 1994; Barabás et al., 2018). Although its core ideas were formalized in an ecological context (Chesson, 1994;

2000a), this framework provides the necessary tools to examine the relative contributions

of fluctuations to diversity maintenance, which can also be applied to evolutionary con-

texts (Ellner & Sasaki, 1996; Reinhold, 2000). From an ecological perspective, polymorphism of sexually antagonistic alleles is equivalent to the coexistence of species, and the fixation of either one of the alleles in a population is equivalent to competitive exclusion. The coexistence of alleles, thus, can be examined through the same lens as the coexistence

of competing species.

Here, we seek to explicitly apply recent advances in MCT to the question of how polymorphism is maintained under sexually antagonistic selection. We examined how fluctuations in selection values, fluctuations in population sizes, and their interactions can stabilize or hinder the coexistence of alleles. In particular, we examined i) Can fluctuations in
population sizes and selection values allow sexually antagonistic alleles to coexist when
differences in their fitness would typically not allow them to? and ii) What is the relative
contribution of different types of fluctuations that allow two sexually antagonistic alleles
to be maintained in a population? Our study provides the tools to analyze evolutionary
dynamics from a novel perspective and contributes to answering long-lasting questions
regarding the effect of non-constant environments on genetic diversity.

67 2 Methods

We first present a model that describes the evolutionary dynamics of sexually antagonistic alleles and show how changes in allele frequencies can be expressed in terms of growth rates, a necessary condition for analyses done using modern coexistence theory. We continue by simulating different scenarios of alleles invading a population, where we allowed population sizes, selection values, both, or neither to vary. Finally, we examine the results of our simulations through a MCT lens by calculating the contribution of each of these fluctuations in the coexistence of alleles.

Population dynamics of sexually antagonistic alleles

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Our model considered evolution at single, biallelic locus with frequency and density independent effects on the relative fitness of females and males. We examined the dynammics of two sexually antagonistic alleles, j and k, that affect fitness in the haploid state. We assumed allele j always has a high fitness in females ($w_{jf}=1$), but variable fitness in males ($w_{jm}<1$); and allele k has a high fitness in males ($w_{km}=1$), but variable fitness in females ($w_{kf}<1$). The selection against allele k in males is therefore k0 and the selection against allele k1 in females is k1 females is k2.

The frequency of each allele in each sex at the beginning of a life-cycle at time t is given by:

$$p_{jm,t} = \frac{n_{jm,t}}{N_{m,t}} \tag{1}$$

 $p_{jf,t} = \frac{n_{jf,t}}{N_{f,t}} \tag{2}$

$$p_{km,t} = \frac{N_{m,t} - n_{jm,t}}{N_{m,t}} \tag{3}$$

$$p_{kf,t} = \frac{N_{f,t} - n_{jf,t}}{N_{f,t}} \tag{4}$$

where $N_{m,t}$ and $N_{t,t}$ are the numbers of males and females in a population at time t, $n_{jf,t}$ is the number of females f with allele j, and $n_{jm,t}$ is the number of males m with allele j at time t, respectively.

The individuals in the population mate at random before selection occurs, and therefore the frequency of offspring with allele j after mating, $p'_{j,t}$ can be expressed as:

$$p'_{j,t} = \frac{n_{jf}}{N_f} \frac{n_{jm}}{N_m} + \frac{1}{2} \frac{n_{jf}}{N_f} \frac{(N_m - n_{jm})}{N_m} + \frac{1}{2} \frac{(N_f - n_{jf})}{N_f} \frac{n_{jm}}{N_m},$$
 (5)

which upon rearranging and simplifying gives:

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$$p'_{j,t} = \frac{(N_{m,t}n_{jf,t} + N_{f,t}n_{jm,t})}{2N_f N_m}.$$
 (6)

Selection acts upon these offspring in order to determine the allelic frequencies in females and males in the next generation, t + 1. As an example the frequency of females with allele j after selection is given by:

$$p'_{jf,t+1} = \frac{n_{jf,t+1}}{N'_{f,t+1}} = \frac{p'_j w_{jf}}{p'_j w_{jf} + (1 - p'_j) w_{kf}}$$
(7)

The logarithmic growth rate of j in females, is therefore given by the number of females with allele j after selection, divided by the original number of females carrying allele j:

$$r_{jf,t} = \ln\left(\frac{n'_{jf,t+1}}{n_{jf,t}}\right) \tag{8}$$

An equivalent expression for the per capita growth rate of allele j in males m can be obtained by exchanging f for m across the various subscripts in this expression.

Allelic coexistence in a sexual population, however, is ultimately influenced by growth

and establishment of an allele across both sexes. Therefore, the full growth rate of allele j across the entire population of females *and* males is given by:

$$r_{j} = \ln \left(\frac{n'_{jf,t+1} + n'_{jm,t+1}}{n_{jf,t} + n_{jf,t}} \right) . \tag{9}$$

- An equivalent expression describes r_k , the growth rate of allele k.
- Selection mantains both alleles in the population under the condition that:

$$\frac{S_m}{1 + S_m} < S_f < \frac{S_m}{1 - S_m} \tag{10}$$

(Kidwell *et al.*, 1977; Pamilo, 1979; Connallon & Hall, 2018) Thus, the maintenance of polymorphism of sexually antagonistic alleles is solely determined by the values of S_m and S_f . Note that in our model, the values S_m and S_f are bounded from 0 to 1. Therefore the **parameter space of sexually antagonistic selection** is within the range $0 < S_m, S_f < 1$. Classic theoretical models predict that in constant environments, only in ≈ 0.38 of the selection parameter space alleles can coexist (Kidwell *et al.*, 1977; Pamilo, 1979; Connallon *et al.*, 2018). If fluctuations in population sizes or selection values have an effect on the coexistence of sexually antagonistic alleles, it would be reflected in increases or decreases of the proportion of the parameter space of selection where polymorphism is maintained.

116 Simulations

Typically, MCT would require decomposing alleles' growth rates (e.g., Eqn. 9) analytically to examine the relative contributions of different types of fluctuations to their coexistence

(Barabás *et al.*, 2018). Although we present an analytical approach in the Supporting Information, our general solution is not easily interpretable and soon becomes mathematically intractable (S1 Supporting Information). Thus, we opted for an extension of MCT that provides the flexibility to examine the contributions of different processes to coexistence using simulations (Ellner *et al.*, 2019; Shoemaker *et al.*, 2020).

For each simulation, we examined coexistence outcomes across the selection param-124 eter space of sexually antagonistic selection (0 $< S_m, S_f < 1$). To do so, we partitioned the parameter space into a grid of 50×50 , which yielded 2500 pairwise combinations 126 of different w_{im} and w_{kf} values. For each pairwise combination of w_{im} and w_{kf} , as we 127 detail in the next sections, we iterated our model while controlling the effect size of fluctuations in selection (σ_w) and their correlation (ρ_w) , as well as fluctuations in population 129 sizes (σ_g) and their correlation (ρ_g) . Then, we performed "invasion simulations" of each 130 allele invading a population, evaluated coexistence outcomes, and determined the rela-131 tive contribution of each type of fluctuation. Finally, we calculated for each simulation the proportion of the parameter space that allowed alleles to coexist. 133

We explored all of the combinations of low, intermediate, and high fluctuations in fitness values and population sizes, with different extents of correlations between fluctuations (Table 1). As a control simulation, we set $\sigma_w = 0.001$ and $\sigma_g = 0.001$, with no correlation between fluctuations. We ran ten replicates per parameter combination, which resulted in 3780 simulations.

39 Timeseries

To incorporate the effects of fluctuations into our population dynamics model we generated independent timeseries of fluctuations in selection and population sizes. In the case of fluctuations in selection values, for a given value of w_{jm} and w_{kf} (i.e., a fixed point in the selection parameter space), we generated a timeseries of 500 timesteps made up of correlated fluctuations of w_{jm} and w_{kf} . We controlled the effect size of fluctuations in selection (σ_w) and its correlation (ρ_w) by using the Cholesky factorization of the variance-

$$C_w = \begin{bmatrix} \sigma_w^2 & \rho_w \sigma_w^2 \\ \rho_w \sigma_w^2 & \sigma_w^2 \end{bmatrix} \tag{11}$$

We multiplyed Eqn. 11 by a (2×500) matrix of random numbers from a normal distribution with mean 0 and unit variance, which yielded γ_j and γ_k . Then, we calculated the new fitness values at time t+1 as $w_{jm,t+1}=w_{jm}^{\gamma_{j,t}}$ and $w_{kf,t+1}=w_{kf}^{\gamma_{k,t}}$.

Similarly, we generated a timeseries of 500 timesteps made up of correlated fluctuations in population sizes. We chose values of $N_m=200$ and $N_f=200$ as the initial value of population sizes throughout our simulations. We performed a Cholesky factorization of the variance-covariance matrix, controlling the effect size of fluctuations in population sizes with σ_g and their correlation with ρ_g . Similar to our previous approach, we multiplied this factorization by a random matrix of uncorrelated random variables, which yielded γ_m and γ_f . Finally, we calculated the number of males and females in the population at time t+1 as $N_{m,t+1}=N_m+\gamma_{m,t}$ and $N_{f,t+1}=N_f+\gamma_{f,t}$. Therefore, the

population sizes in each timestep differed from the inital value of 200 individuals on the order of ρ_g . Note that the scales of σ_g and σ_w are different from each other. While σ_w controls the exponential change in fitness values in each timestep, σ_g controls the number of individuals added to a population in each timestep.

Finally, we performed simulations where our population dynamics model (Eqns. 1 to 9) iterated over 500 timesteps while allowing selection values and population sizes to fluctuate in each timestep. We started each simulation with the initial values of N_m and N_f described before and equal frequencies of allele j and allele k in each sex. For each timestep t in our simulations, the values of w_{jm} w_{kf} , N_m and N_f used to calculate allele's frequencies in timestep t (e.g., Eqn. 7), corresponded to the t values calculated in each timeseries, as described previously. This approach yielded a final timeseries that captured the dynamics of sexually antagonistic alleles, with fluctuating values of selection and population sizes.

171 Invasion simulations

Modern coexistence theory has shown that coexistence is promoted by mechanisms that give species a population growth rate advantage over other species when they become rare (Chesson, 1982; 2003; Barabás $et\ al.$, 2018). Typically, one species is held at its resident state, as given by its steady-state abundances while the rare species is called the invader. In the context of alleles in a population, an allele is an invader when a mutation occurs that introduces that allele into a population in which it is absent (e.g., if in a population with only k alleles, a random mutation made one individual carry the j al-

lele). Within sexually antagonistic selection, each allele has two pathways of invasion, depending on whether the mutation arises in a female or in a male. If an alleles' invasion 180 growth rate (or the average instantaneous population growth rate when rare) is positive, 181 it buffers it against extinction, maintaining its persistence in the population. Coexistence, and hence polymorphism, occurs when both alleles have positive invasion growth rates. 183 We used the timeseries that captured the dynamics of our population model as a tem-184 plate to perform invasion simulations of both alleles. We performed 500 independent invasion simulations, one for each timestep in our timeseries. We explored all four po-186 tential combinations of each allele invading through each pathway (e.g., allele *j* invading 187 through males, and allele k invading through females, and so on). To simulate invasion, we set the density of the invading allele to one individual. For example, if allele j was 189 invading via males, then we would set $n_{jm,i} = 1$ and $n_{jf,i} = 0$. Note that each inva-190 sion simulation was independent of the iteration that we used to generate the timeseries, therefore we denoted the initial timestep in an invasion simulation with the subscript *i*. 192 We also set the resident allele, in this case k, to the corresponding value of the timeseries 193 minus one individual, $n_{km,i} = N_{m,t} - 1$ and $n_{kf,i} = N_{f,t}$. Then, we iterated our model one timestep, i + 1, and calculated the logarithmic growth rate of j allele invading as: 195

$$r_j = \ln\left(\frac{n_{jm,i+1} + n_{jf,i+1}}{1}\right) \tag{12}$$

Correspondingly, the logarithmic growth rate of the *k* allele as a resident would be

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197 given by:

$$r_k = \ln\left(\frac{n_{km,i+1} + n_{kf,i+1}}{n_{km,i} + n_{kf,i}}\right)$$
(13)

Following the approach of Shoemaker *et al.* (2020), we treated each invasion simulation independently, and hence we performed 500 invasion simulations. We then calculated, for each allele invading via a different pathway, its mean invasion growth rate as the average of the 500 invasion growth rates. We also calculated the mean growth rate of the resident allele as the average of the 500 resident growth rates. We determined alleles to be coexisting if both of alleles had positive mean invasion growth rates, which is often referred to as the mutual invasibility criterion (Barabás *et al.*, 2018).

205 Functional decompostion

Our invasion simulations tell us whether or not sexually antagonistic alleles can coexist in a determined point of the selection parameter space. However, we also quantified
the relative contributions of fluctuating selection and population sizes into the predicted
coexistence outcome using a *functional decomposition* approach (Ellner *et al.*, 2016; 2019;
Shoemaker *et al.*, 2020).

We applied the functional decomposition approach by breaking up the average growth rate of each allele into a null growth rate in the absences of fluctuations in all selected variables, a set of main effect terms that represent the effect of only one variable fluctuating, and a set of two-way interaction terms representing the effect of variables fluctuating simultaneously (Ellner *et al.*, 2019). In our simulations, this is a function of four variables: the number of males in the population (N_m) , the number of females in the population

 (N_f) , the fitness of allele j in males (w_{jm}) , and the fitness of allele k in females (w_{kf}) . As an example, if only N_m and N_f were fluctuating, the growth rate of allele j when it is the invader at timestep t could be decomposed into:

$$r_{j,t}(N_m, N_f) = \mathcal{E}_j^0 + \mathcal{E}_j^{N_m} + \mathcal{E}_j^{N_f} + \mathcal{E}_j^{N_m N_f}$$
(14)

Where \mathcal{E}^0 is the null growth rate when N_m and N_f are set to their averages. Terms with superscripts represent the marginal effects of letting all superscripted variables vary while fixing all the other variables at their average values. For example, the term \mathcal{E}^{N_m} expresses the contribution of fluctuations in N_m when N_f is at its average, without the contribution when both variables are set to their averages:

$$\mathcal{E}_j^{N_m} = r_{j,t}(N_m, \overline{N_f}) - \mathcal{E}_j^0 \tag{15}$$

If we average Eqn. 14 across the timesteps in our simulation, we get a partition of the average population growth rate into the variance–free growth rate, the main effects of variability in N_m , the main effects of variability in N_f , and the interaction between variability in N_m and N_f

$$\overline{r}_j = \mathcal{E}_j^0 + \overline{\mathcal{E}_j}^{N_m} + \overline{\mathcal{E}_j}^{N_f} + \overline{\mathcal{E}_j}^{N_m N_f}$$
(16)

However, in our simulations w_{jm} and w_{kf} also fluctuated, therefore the full functional decomposition of the growth rate of allele j as an invader is found in Table 2, as well as a brief description of the meaning of each term. The implementation and interpretation

of the functional decomposition of the invasion growth rates of each allele are identical to each other. Note that Table 2 does not include three or four–way interactions (e.g., $\overline{\mathcal{E}}_{j}^{N_{m}N_{f}w_{jm}w_{fk}}$). This is because in our simulations, we did not allow fluctuations in selection and population sizes to be correlated, therefore their effects are solely captured by the terms in Table 2. We calculated the value of each of the terms in Table 2 by performing another set of invasion simulations as described previously, but instead of allowing all variables to fluctuate, systematically setting the required variables to their means and subtracting the corresponding \mathcal{E} values.

The functional decomposition approach further requires the *comparison* of each term, to understand if how it affects invaders and residents (i.e., the relative contribution). This is because fluctuations can promote coexistence by helping whichever allele is rare, or they can hurt whichever allele is common. Therefore, to understand the role of each type of fluctuation, it is necessary to compare how it affects invader *and* resident growth rates. In the example presented in Eqn. 16, if allele j is invading, then allele k is at it's resident state and there exists an analogue decomposition of \bar{r}_k with the exact same terms as Eqn. 16. Therefore we can express the difference between contributions of fluctuations in N_m as:

$$\Delta_j^{N_m} = \overline{\mathcal{E}}_j^{N_m} - \overline{\mathcal{E}}_k^{N_m} \tag{17}$$

If $\Delta_j^{N_m}$ is positive, then fluctuations in the male population benefit allele j when it is rare more than what they benefit k as a resident. If $\Delta_j^{N_m}$ is negative, then fluctuations

benefit k as a resident more than j as an invader, and if it is minimal, then fluctuations have an equal effect in j and k. Therefore, for each allele invading via a different pathway, we calculated 7Δ values, one for each one of the \mathcal{E} terms in Table 2. However, since the magnitude of each one of these values could vary considerably across the parameter space of selection, to make them comparable, we normalized each Δ value by dividing it by the square root of the sum of the squares of the 7Δ values. For example, the normalized value of Eqn. 17 would be given by:

$$\delta_j^{N_m} = \frac{\Delta_j^{N_m}}{\sqrt{\sum_{d=1}^7 (\Delta_d)^2}} \tag{18}$$

This normalization bounded δ values from -1 to 1.

259 3 Results

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Our results showed that both fluctuations in selection and population sizes can substantially increase the expected genetic variability under sexually antagonistic selection. The average proportion of coexistence in the selection parameter space increased with the effect size of fluctuations when fluctuations were large enough (Fig. 1). Increments in allelic coexistence were more likely when fluctuations were large, and fluctuations in population sizes were negatively correlated, while fluctuations in selection were positively correlated (Fig. 1). Importantly, our results show that environmental fluctuations can more than double the expected genetic variation under sexually antagonistic selection, reaching up to ≈ 0.9 of allelic coexistence in the selection parameter space (Fig. 1).

Fluctuations increased coexistence by allowing both alleles as invaders to have posi-269 tive invasion growth rates in instances where selection would typically not allow them to 270 (i.e., they made parts of the parameter space "flip" into coexistence). As a baseline, we 271 show in Fig. 2A the outcome of the control simulation, which matched previous findings that without fluctuations, alleles can coexist in only ≈ 0.38 of the selection parameter 273 space (Connallon & Hall, 2018). These "flips" occurred with both types of fluctuations 274 and were more common with larger fluctuations and strongly correlated effects, for which 275 we show examples in Fig. 2A . However, note that there are also parts of the parameter 276 space where coexistence is lost compared to the control simulation, which was more likely 277 when population sizes were fluctuating (Fig. 2A). 278

Alleles had positive invasion growth rates when positive contributions of fluctuations 279 outweighed the negative contributions of fluctuations. As an example in Fig. 2B we show 280 the functional decomposition of both alleles invading via their favored pathway in parts 281 of the parameter space that "flipped" into coexistence and competitive exclusion (which 282 correspond to the square and triangle in Fig. 2A). Note that each type of fluctuation made 283 similar contributions to each allele, both when they were coexisting or experiencing com-284 petitive exclusion (Fig. 2B). However, δ_0 , which captures the effect of fluctuations set 285 to their averages, switched between positive and negative contributions for both alleles 286 (Fig. 2B). 287

The relative contribution of fluctuations in population sizes of males and females benefited alleles when alleles invaded via the fluctuating population (Fig. 2B and Fig. 3). If alleles invaded via the opposite sex, then fluctuations contributed negatively to their invasion growth rate (Fig. 3). For example, the relative contribution of fluctuations in the male population, δ^{N_m} , was positive for both alleles when they invaded via males and negative when they invaded via females, regardless of the correlation between fluctuations (Fig. 3). The opposite pattern was shown by δ^{N_f} . The relative contribution of both populations fluctuating, $\delta^{N_m N_f}$, was positive when fluctuations were negatively correlated, it had a negligible effect when fluctuations were not correlated, and it had a negative effect when fluctuations were positively correlated (Fig. 3).

In contrast, fluctuations in selection benefited the allele that was unaffected by selec-298 tion. For example, $\delta^{w_{jm}}$ which captured the relative contribution of fluctuations in se-299 lection against j in males, contributed negatively to allele j's invasion growth rate but 300 benefited allele k's invasion, regardless of the correlation between fluctuations or the sex 30 where invasion occurred (Fig. 4). The opposite pattern was shown by $\delta^{w_{kf}}$. The relative 302 contribution of both types of selection fluctuating, was negative when fluctuations were 303 negatively correlated, it had a negligible effect when fluctuations were not correlated, and 304 it had a positive effect when fluctuations were positively correlated (Fig. 4). 305

The overall average effect of fluctuations, δ^0 , was at first glance unpredictable when fluctuations were incorporated. Without fluctuations, δ^0 captured exclusively the effect of selection and had positive values in parts of the parameter space where each allele could invade (Fig. 5). However, when we incorporated fluctuations, it also captured unexpected variance not captured by the terms in Table 2. Fluctuations in population sizes generated stochastic changes of δ^0 accross the parameter space and spanned a greater extent when alleles invade via their non-favored pathway (Fig. 5). Fluctuations in selection

generated δ^0 values to decrease in magnitude compared to the control simulation, which was heightened in parts of the parameter space that were on the limit of the coexistence region (Fig. 5). When we incorporated both types of fluctuations, the value of δ^0 was more likely to changed compared to the control simulation (Fig. 5).

317 4 Discussion

The results of our study provide supporting evidence that environmental fluctuations 318 can increase the expected genetic variation maintained under sexually antagonistic selec-319 tion. Antagonistically selected alleles are an important component of genetic variation for many species (Foerster et al., 2007; Van Doorn, 2009; Bonduriansky & Chenoweth, 2009; 321 Innocenti & Morrow, 2010). Furthermore, as much as 20% of traits for which data is avail-322 able are thought to be under sexually antagonistic selection (Morrissey, 2016). Yet, a large body of work suggests that the criteria for maintaining antagonistic genetic variation is 324 very restrictive (i.e., we would expect the coexistence of alleles in few scenarios) (Kid-325 well et al., 1977; Pamilo, 1979; Hedrick, 1999; Curtsinger et al., 1994; Patten et al., 2010). 326 Our study shows that incorporating more relaistic assumptions, such as non-constant se-327 lection and population sizes can more than double the expected genetic variation under 328 sexually antagonistic selection (Fig. 1).

The relative contribution of fluctuations in selection

Our results show that fluctuations in selection can promote allelic coexistence in parts of
the parameter space where we would typically expect selection to fix one of the alleles

(Fig. 2). Fluctuating selection promoted coexistence by contributing possitvely to an allele's invasion growth rate when that allele was shielded from the fluctuations (Fig. 4). Therefore, the relative contribution of both w_{jm} and w_{kf} fluctuating only promoted coexistence when fluctuations were positively correlated (Fig. 4), which is why the average proportion of coexistence was generally higher for simulations that had this correlation (Fig. 1). This

In this paragraph we highlight our results that show that fluctuations in selection help
an allele invade when it is shielded from the fluctuating selection. This could be understood as a relative non linearity to resources as (Shoemaker *et al.*, 2020) also found. Give
an example on how this could enhance coexistence when we would tipically not expect
it to, for example in instances where the allele with the highest fitness also experiences
more fluctuations, and how correlations could further enhance that.

In natural populations, fitness optima are highly variable as biotic and abiotic conditions are constantly changing (Siepielski *et al.*, 2017).

Selection may vary in space or time as are sult of variation in female preferences

Males and females often have dinstinct roles in reproduction, social interactions or the
ways they interact with the environment. These different niche and roles give rise to sexually antagonistic selection, such selection that often favors larger body syzes in females
than in males due to a significant female fecundity advantage of large size, and sex differences in selection on feeding morphology as a consequence of sex specific nutritional
requirenments.

Chippindale and collagues observed genotypes

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In this paragraph we continue by contextualizing our findings with other studies that
show that fluctuations in selection can promote polymorphism in sex-linked traits, or
when generations overlap, which could be understood as a storage effect. Also mention
(Connallon & Hall, 2018) who found that selection coupled with life history traits promote
polymorphism.

360 The effect of fluctuations in populatin sizes

Discuss how our results show that fluctuations in population sizes benefit the allele that is invading via the fluctuating population. Put that in context in how much real populations fluctuate, and maybe highlight how an excess of genetic diversity could be explained by this mechanism (example).

Antagonistic selection is often ineffective in the face of genetic drift, with the impact of drift being particularly pronounced at or near the parameter domain for balancing selection (Connallon & Clark, 2012).

Sexually antagonistic alleles have a slow rate of evolutionary chanage Connallon & Clark (2012) which also generates sensitivity to the effective size of a population. Genetic drift my dominate over selection, even in populations that are quite large. There may be little opportunity in small populations for stable balanced polymorphism at antagonistic loci

In this paragraph we contextualize how other findings have highlighted the importance of fluctuations in males and females to promote genetic diversity of sex-linked traits or by mitigating genetic drift. Mention how our findings fall into the existing understanding of non constant populations

Insights from the functional decomposition approach

In this paragraph we discuss how the functional decomposition approach showed not only the relative contributions of each type of fluctuations, but also captured unexpected stochasticity captured by δ_0 .

SAS might be resolved in extremely changing environments ??? (De Lisle *et al.*, 2018)

382 Conclusion

Our study contributes to the growing body of work that shows that the criteria for maintaining genetic variation under sexually antagonistic selection are overly conservative

(Connallon & Clark, 2012; Connallon *et al.*, 2018). Processes like recurrent mutations

(Radwan, 2008), genetic drift (Connallon & Clark, 2012), local adaptations in fluctuating
environments (Connallon *et al.*, 2018).

Figures and tables

Table 1: Parameters used in our simulations to control the effect size of fluctuations in population sizes (σ_g) and selection values (σ_w), as well as their respective correlations (ρ_g and ρ_w). We ran ten replicates for each one of the factorial combinations of the following parameters, which yielded a total of 3780 simulations.

Parameter	Values	Description
σ_w	0.001, 0.1, 0.3, 0.5, 0.7, 0.9	Effect size of fluctuations in fitness values
σ_g	0.001, 1, 10, 20, 30, 50	Effect size of fluctuations in population sizes
$ ho_w$	-0.75, 0, 0.75	Correlation between fluctuations in fitness values
ρ_g	-0.75, 0, 0.75	Correlation between fluctuation in population sizes

Table 2: Functional decomposition of the growth rate of allele *j*. As we exemplify in Eqn. 16, each term captures the contribution of fluctuations to an alleles' invasion growth rate.

Term	Formula	Meaning
\mathcal{E}_{i}^{0}	$\overline{r_j}(\overline{N_m},\overline{N_f},\overline{w_{jm}},\overline{w_{kf}})$	Growth rate at mean population size and selection values.
$rac{\mathcal{E}_{j}^{0}}{\overline{\mathcal{E}}_{j}^{Nm}}$ $rac{\overline{\mathcal{E}}_{j}^{N}}{\overline{\mathcal{E}}_{j}^{w}}$ $\overline{\mathcal{E}}_{j}^{w}$	$\overline{r}_j(N_m\overline{N_f},\overline{w_{jm}},\overline{w_{kf}})-\mathcal{E}_j^0$	Main effect of fluctuations in N_m
$\overline{\mathcal{E}}_{j}^{N_{f}}$	$\overline{r_j}(\overline{N_m},N_f,\overline{w_{jm}},\overline{w_{kf}})-\mathcal{E}_j^0$	Main effect of fluctuations in N_f
$\overline{\mathcal{E}}_{j}^{w_{jm}}$	$\overline{r_j}(\overline{N_m},\overline{N_f},w_{jm},\overline{w_{kf}})-\mathcal{E}_j^0$	Main effect of fluctuations in w_{jm}
$\overline{\mathcal{E}}_i^{w_{kf}}$	$\overline{r_j}(\overline{N_m},\overline{N_f},\overline{w_{jm}},w_{kf})-\mathcal{E}_j^0$	Main effect of fluctuations in w_{kf}
$\overline{\mathcal{E}}_{j}^{^{N_{m},N_{f}}}$	$\overline{r_j}(N_m, N_f, \overline{w_{jm}}, \overline{w_{kf}}) - [\mathcal{E}_j^0 + \overline{\mathcal{E}}_j^{N_m} + \overline{\mathcal{E}}_j^{N_f}]$	Interaction of fluctuations in N_m and N_f
$\overline{\mathcal{E}}_{j}^{'w_{jm},w_{kf}}$	$\overline{r_j}(\overline{N_m}, \overline{N_f}, w_{jm}, w_{kf}) - [\mathcal{E}_j^0 + \overline{\mathcal{E}_j^{w_{jm}}} + \overline{\mathcal{E}_j^{w_{kf}}}]$	Interaction of fluctuations in w_{jm} and w_{kf}

Correlation in fluctuations in selection, ρ_w -0.750 0.75 70 50 30 0.75 20 Correlation in fluctuations in population sizes, ρ_{g} 10 Fluctuations in population sizes, σ_{g} 0.001 70 50 30 20 10 0.001 70 50 30 20 10 0.001 0.7 0.9 0.001 0.1 0.3 0.5 0.7 0.9 0.001 0.1 0.5 0.3 0.5 Fluctuations in selection, $\sigma_{\rm w}$ Average proportion of coexistence .3 .38 .5

Figure 1: The average proportion of coexistence. For all parameter combinations in our simulations, we show the average proportion of coexistence across replicates and invasion types. Each panel corresponds to a different combination of correlations between fluctuations. Labels on top indicate the correlation between fluctuations in selection ρ_w , while labels on the right show the correlation in fluctuations between fluctuations in population sizes ρ_g . As a basis of comparison, we show the expected proportion of coexistence (0.38) as white in our color scheme.

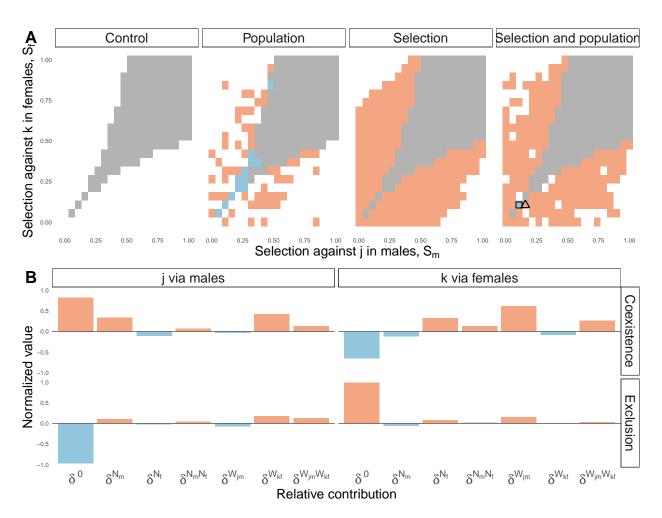


Figure 2: Coexistene outcomes and functional decomposition. In A) each panel corresponds to one replicate of a different type of simulation. All panels show the coexistence outcomes in the selection parameter space when *j* invaded via males and *k* invaded via females. As a reference, *j* is favored in females and k is favored in males. In the Control panel ($\sigma_g = 0.001$, $\rho_g = 0$, $\sigma_w = 0$ 0.001, $\rho_w = 0$) grey areas indicate parts of the selection parameter space where alleles can coexist, while white areas indicate parts of the parameter space that correspond to competitive exclusion (following Eqn.10). In the Population ($\sigma_g = 70$, $\rho_g = -0.75$, $\sigma_w = 0.001$, $\rho_w = 0$), Selection $(\sigma_g = 0.001, \rho_g = 0, \sigma_w = 0.9, \rho_w = 0.75)$, and Selection and population $(\sigma_g = 0.9, \rho_g = -0.75, \rho_g = 0.001)$ $\sigma_w = 0.9$, $\rho_w = 0.75$) panels, light red areas indicate parts of the parameter space that "flipped" into coexistence, while light blue areas show changes into competitive exclusion. We highlighted two points in the parameter space in the Selection and population panel that corresponded to changes into coexistence (triangle) and into competitive exlcusion (square). In B) we show the functional decomposition of the coexistence and competitive exclusion points highlighted in A). Each panel corresponds to each allele invading via their respective pathway and shows the bar plots of the different δ values that made up the functional decomposition of each allele as an invader. Light red colors indicate positive δ values that benefited that allele as an invader more than the other allele as a resident, while light blue colors indicate negative δ values that benefited a resident more than the invader.

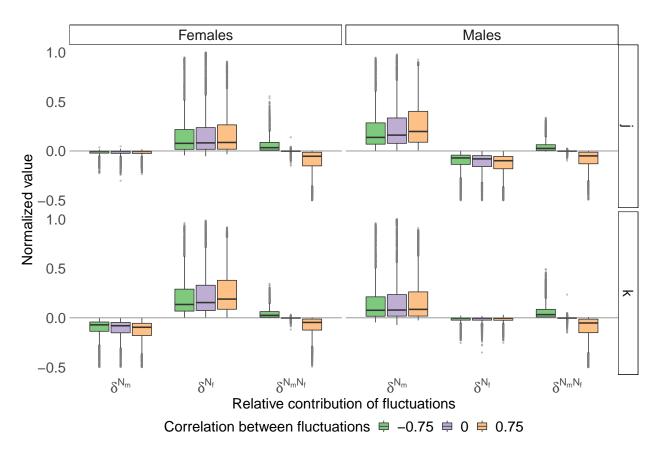


Figure 3: The relative contributions of fluctuations in population sizes. As a reference, positive δ values imply that the corresponding fluctuation benefits that allele as an invader more than the other allele as a resident, negative δ values indicate fluctuations benefit the residents more than the invader, and δ values close to zero indicate that the corresponding fluctuation has equal contributions to invaders and residents. Each panel corresponds to each allele invading via a different pathway, for which we show the boxplots of the three distinct δ values that captured the effects of fluctuations in population sizes, for all of the replicates in our simulation in which $\sigma_g = 70$. Each color corresponds to a different correlation between fluctuations in population sizes (ρ_g), as the legend indicates. Each box plot extends from the first to third quantiles of the corresponding posterior distribution of parameter values, and the line inside the the box indicates the median.

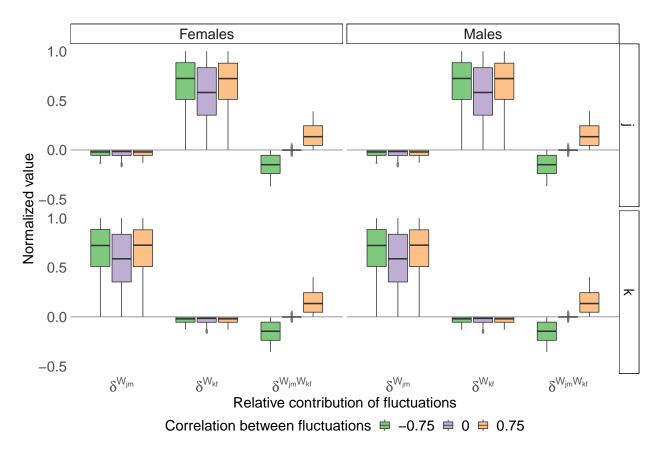


Figure 4: The relative contributions of fluctuations in selection. As a reference, positive δ values imply that the corresponding fluctuation benefits that allele as an invader more than the other allele as a resident, negative δ values indicate fluctuations benefit the residents more than the invader, and δ values close to zero indicate that the corresponding fluctuation has equal contributions to invaders and residents. Each panel corresponds to each allele invading via a different pathway, for which we show the boxplots of the three distinct δ values that captured the effects of fluctuations in population sizes, for all of the replicates in our simulation in which $\sigma_w = 0.90$. Each color corresponds to a different correlation between fluctuations in population sizes (ρ_w), as the legend indicates. Each box plot extends from the first to third quantiles of the corresponding posterior distribution of parameter values, and the line inside the the box indicates the median.

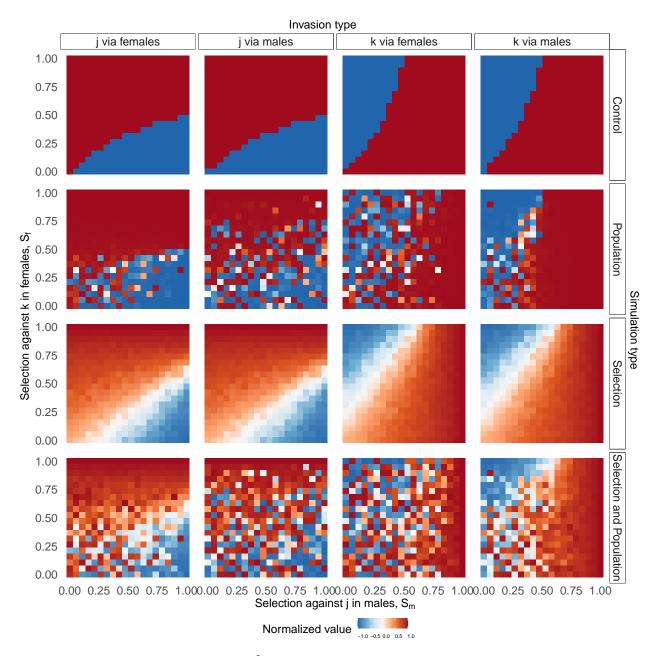


Figure 5: The normalized value of δ^0 in the selection parameter space. Each row in this figure corresponds to one replicate of a different type of simulation, while each column corresponds to a different type of invasion. In the Control simulation ($\sigma_g = 0.001$, $\rho_g = 0$, $\sigma_w = 0.001$, $\rho_w = 0$), δ^0 had a normalized value of 1 in parts of the parameter space where selection allows each allele to be fixed in a population, and a value of -1 where each allele can not be mantained in a population. The Population simulation corresponds to a replicate of a simulation where only population sizes fluctuated, without correlation between fluctuations ($\sigma_g = 70$, $\rho_g = 0$, $\sigma_w = 0.001$, $\rho_w = 0$). The Selection simulation corresponds to a replicate of a simulation where only selection fluctuated, without correlations between fluctuations ($\sigma_g = 0.001$, $\rho_g = 0$, $\sigma_w = 0.9$, $\rho_w = 0$). Finally, the Selection Population simulation corresponds to a replicate of a simulation where both selection and population sizes fluctuated, without correlations between fluctuations ($\sigma_g = 70$, $\rho_g = 0$, $\sigma_w = 0.9$, $\rho_w = 0$).

References

- ³⁹⁰ Arnqvist, G. & Rowe, L. (2013). Sexual conflict. Princeton University Press.
- Barabás, G., D'Andrea, R. & Stump, S.M. (2018). Chesson's coexistence theory. *Ecological Monographs*, 88, 277–303.
- Bonduriansky, R. & Chenoweth, S.F. (2009). Intralocus sexual conflict. *Trends in ecology & evolution*, 24, 280–288.
- Chesson, P. (1994). Multispecies competition in variable environments. *Theoretical popula-*tion biology, 45, 227–276.
- ³⁹⁷ Chesson, P. (2000a). General theory of competitive coexistence in spatially-varying environments. *Theoretical Population Biology*, 58, 211–237.
- Chesson, P. (2000b). Mechanisms of maintenance of species diversity. *Annual review of Ecology and Systematics*, 31, 343–366.
- Chesson, P. (2003). Quantifying and testing coexistence mechanisms arising from recruitment fluctuations. *Theoretical Population Biology*, 64, 345–357.
- Chesson, P.L. (1982). The stabilizing effect of a random environment. *Journal of Mathemat- ical Biology*, 15, 1–36.
- Connallon, T. & Clark, A.G. (2012). A general population genetic framework for antagonistic selection that accounts for demography and recurrent mutation. *Genetics*, 190, 1477–1489.

- Connallon, T. & Hall, M.D. (2018). Environmental changes and sexually antagonistic selection. *eLS*, pp. 1–7.
- 410 Connallon, T., Sharma, S. & Olito, C. (2018). Evolutionary Consequences of Sex-Specific
- Selection in Variable Environments: Four Simple Models Reveal Diverse Evolutionary
- Outcomes. *The American Naturalist*, 193, 93–105.
- Curtsinger, J.W., Service, P.M. & Prout, T. (1994). Antagonistic pleiotropy, reversal of dominance, and genetic polymorphism. *The American Naturalist*, 144, 210–228.
- De Lisle, S.P., Goedert, D., Reedy, A.M. & Svensson, E.I. (2018). Climatic factors and species range position predict sexually antagonistic selection across taxa. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 373, 20170415.
- Ellner, S. & Hairston Jr, N.G. (1994). Role of overlapping generations in maintaining genetic variation in a fluctuating environment. *The American Naturalist*, 143, 403–417.
- Ellner, S. & Sasaki, A. (1996). Patterns of genetic polymorphism maintained by fluctuating selection with overlapping generations. *theoretical population biology*, 50, 31–65.
- Ellner, S.P., Snyder, R.E. & Adler, P.B. (2016). How to quantify the temporal storage effect using simulations instead of math. *Ecology letters*, 19, 1333–1342.
- Ellner, S.P., Snyder, R.E., Adler, P.B. & Hooker, G. (2019). An expanded modern coexistence theory for empirical applications. *Ecology Letters*, 22, 3–18.
- Foerster, K., Coulson, T., Sheldon, B.C., Pemberton, J.M., Clutton-Brock, T.H. & Kruuk,

- L.E. (2007). Sexually antagonistic genetic variation for fitness in red deer. *Nature*, 447,
- 428 1107–1110.
- Gavrilets, S. (2014). Is sexual conflict an "engine of speciation"? Cold Spring Harbor
- perspectives in biology, 6, a017723.
- Hedrick, P.W. (1974). Genetic variation in a heterogeneous environment. i. temporal het-
- erogeneity and the absolute dominance model. *Genetics*, 78, 757–770.
- Hedrick, P.W. (1986). Genetic polymorphism in heterogeneous environments: a decade
- later. Annual review of ecology and systematics, 17, 535–566.
- Hedrick, P.W. (1999). Antagonistic pleiotropy and genetic polymorphism: a perspective.
- 436 *Heredity*, 82, 126–133.
- Immler, S., Arnqvist, G. & Otto, S.P. (2012). Ploidally antagonistic selection maintains
- stable genetic polymorphism. Evolution: International Journal of Organic Evolution, 66,
- 439 55–65.
- Innocenti, P. & Morrow, E.H. (2010). The sexually antagonistic genes of drosophila
- melanogaster. *PLoS biology*, 8, e1000335.
- 442 Kidwell, J., Clegg, M., Stewart, F. & Prout, T. (1977). Regions of stable equilibria for
- models of differential selection in the two sexes under random mating. *Genetics*, 85,
- 444 171–183.
- Lande, R. (1980). Sexual dimorphism, sexual selection, and adaptation in polygenic char-
- acters. *Evolution*, pp. 292–305.

- Mitchell-Olds, T., Willis, J.H. & Goldstein, D.B. (2007). Which evolutionary processes influence natural genetic variation for phenotypic traits? *Nature Reviews Genetics*, 8,
- 449 845-856.
- 450 Mori, E., Mazza, G. & Lovari, S. (2017). Sexual dimorphism. Encyclopedia of Animal Cogni-
- tion and Behavior (J. Vonk, and T. Shakelford, Eds). Springer International Publishing, Switzer-
- 452 *land*, pp. 1–7.
- 453 Morrissey, M.B. (2016). Meta-analysis of magnitudes, differences and variation in evolu-
- tionary parameters. *Journal of Evolutionary Biology*, 29, 1882–1904.
- Nunney, L. (2002). The effective size of annual plant populations: the interaction of a seed
- bank with fluctuating population size in maintaining genetic variation. *The American*
- 457 Naturalist, 160, 195–204.
- Pamilo, P. (1979). Genic variation at sex-linked loci: Quantification of regular selection
- models. *Hereditas*, 91, 129–133.
- Patten, M.M., Haig, D. & Ubeda, F. (2010). Fitness variation due to sexual antagonism
- and linkage disequilibrium. Evolution: International Journal of Organic Evolution, 64,
- 462 3638–3642.
- Pemberton, J., Smith, J., Coulson, T.N., Marshall, T.C., Slate, J., Paterson, S., Albon, S.,
- ⁴⁶⁴ Clutton-Brock, T.H. & Sneath, P.H.A. (1996). The maintenance of genetic polymorphism
- in small island populations: large mammals in the hebrides. *Philosophical Transactions*
- of the Royal Society of London. Series B: Biological Sciences, 351, 745–752.

- Radwan, J. (2008). Maintenance of genetic variation in sexual ornaments: a review of the mechanisms. *Genetica*, 134, 113–127.
- Reinhold, K. (2000). Maintenance of a genetic polymorphism by fluctuating selection on sex-limited traits. *Journal of Evolutionary Biology*, 13, 1009–1014.
- Shoemaker, L.G., Barner, A.K., Bittleston, L.S. & Teufel, A.I. (2020). Quantifying the rela-
- tive importance of variation in predation and the environment for species coexistence.
- 473 *Ecology letters*, 23, 939–950.
- Siepielski, A.M., Morrissey, M.B., Buoro, M., Carlson, S.M., Caruso, C.M., Clegg, S.M.,
- Coulson, T., DiBattista, J., Gotanda, K.M., Francis, C.D. et al. (2017). Precipitation drives
- global variation in natural selection. *Science*, 355, 959–962.
- Van Doorn, G.S. (2009). Intralocus sexual conflict. *Annals of the New York Academy of*Sciences, 1168, 52–71.
- Walsh, B. & Lynch, M. (2018). Evolution and Selection of Quantitative Traits. OUP Oxford.