

BDA - Assignment 6

Anonymous

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Import some useful libraries

```
library(aaltobda)
library(rstan)
```

```
## Loading required package: StanHeaders
```

```
## Loading required package: ggplot2
```

```
## rstan (Version 2.21.2, GitRev: 2e1f913d3ca3)
```

```
## For execution on a local, multicore CPU with excess RAM we recommend calling
```

```
## options(mc.cores = parallel::detectCores()).
```

```
## To avoid recompilation of unchanged Stan programs, we recommend calling
```

```
## rstan_options(auto_write = TRUE)
```

```
## Do not specify '-march=native' in 'LOCAL_CPPFLAGS' or a Makevars file
```

```
library(bayesplot)
```

```
## This is bayesplot version 1.7.2
```

```
## - Online documentation and vignettes at mc-stan.org/bayesplot
```

```
## - bayesplot theme set to bayesplot::theme_default()
```

```
## * Does _not_ affect other ggplot2 plots
```

```
## * See ?bayesplot_theme_set for details on theme setting
```

```
# rstan options
```

```
options(mc.cores = parallel::detectCores())
```

```
rstan_options(auto_write = TRUE)
```

```
# Import the data
```

```
data("bioassay")
```

Generalized linear model: Bioassay with Stan

Exercise 1

In this exercise, we model a dose-response relation for the bioassay data using the Stan probabilistic programming language. We assume a Gaussian prior distribution characterized by

$$\begin{bmatrix} \alpha \\ \beta \end{bmatrix} \sim N(\boldsymbol{\mu}_0, \boldsymbol{\Sigma}_0), \text{ where } \boldsymbol{\mu}_0 = \begin{bmatrix} 0 \\ 10 \end{bmatrix} \text{ and } \boldsymbol{\Sigma}_0 = \begin{bmatrix} 2^2 & 10 \\ 10 & 10^2 \end{bmatrix}.$$

The model is built in Stan as follows

```
code_bioassay <- "bioassay_model.stan"
writeLines(readLines(code_bioassay))

## // The input data is a vector 'x' of length 'N'.
## data {
##   int<lower=0> N;      // total number of rows
##   vector[N] x;        // dose level
##   int<lower=0> n[N];   // number of animals
##   int y[N];           // number of deaths (positive outcomes)
##
##   // parameters of prior distribution
##   vector[2] mu;
##   matrix[2, 2] Sigma;
##
## }
##
## // The parameters accepted by the model. Our model
## // accepts the 2D parameter 'theta'.
## parameters {
##   vector[2] theta;
## }
##
## transformed parameters {
##   vector[N] rel;
##   rel = theta[1] + theta[2]*x; // dose-response relation
## }
##
## // The model to be estimated.
## model {
##   theta ~ multi_normal(mu, Sigma); // prior
##   y ~ binomial_logit(n, rel);      // likelihood
## }
```

Now, we can prepare the data in R with

```
bioassay_data <- list(N = length(bioassay$x),
                      x = bioassay$x,
                      n = bioassay$n,
                      y = bioassay$y,
                      mu = c(0, 10),
                      Sigma = matrix(data = c(4, 10, 10, 100), nrow = 2, ncol = 2))
```

Then, we can get a fit with the following R command and sample from the posterior distribution.

```

max_iters <- 2000
n_chains <- 5
fit <- stan(file = 'bioassay_model.stan', data = bioassay_data, chains = n_chains,
            iter = max_iters, warmup = floor(max_iters/2))

```

Exercise 2

In order to check the convergence of the chains, we can use `stan::monitor` to compute \hat{R} and other convergence analytics. \hat{R} is an indicator of the between- and within-chain variance of the estimates. If the between- or within-chain estimates are generally not agreeing, we will get a larger value. A larger value thus tells that we should continue with further simulations to improve our inference of the target distribution. If the value of \hat{R} is less than 1.05, it is considered safe to use the sample.

```

monitor(fit)

## Inference for the input samples (5 chains: each with iter = 2000; warmup = 0):
##
##           Q5  Q50  Q95 Mean  SD  Rhat Bulk_ESS Tail_ESS
## theta[1] -0.4  0.9  2.6  1.0 0.9    1    1644    1830
## theta[2]  4.0  9.9 19.0 10.5 4.6    1    1629    1870
## rel[1]   -14.4 -7.6 -3.2 -8.0 3.5    1    1911    2080
## rel[2]    -4.2 -2.0 -0.6 -2.2 1.1    1    3047    2587
## rel[3]    -0.8  0.4  1.8  0.5 0.8    1    1941    2000
## rel[4]     2.9  8.2 15.9  8.6 4.0    1    1478    1930
## lp__     -9.1 -6.8 -6.1 -7.1 1.0    1    1806    2282
##
## For each parameter, Bulk_ESS and Tail_ESS are crude measures of
## effective sample size for bulk and tail quantities respectively (an ESS > 100
## per chain is considered good), and Rhat is the potential scale reduction
## factor on rank normalized split chains (at convergence, Rhat <= 1.05).

```

As we can see, α (i.e. `theta[1]`) and β (i.e. `theta[2]`) both have an $\hat{R}_\alpha = \hat{R}_\beta = 1$. Both results are below 1.05, which means that we can accept the sample as the chains have converged.

Exercise 3

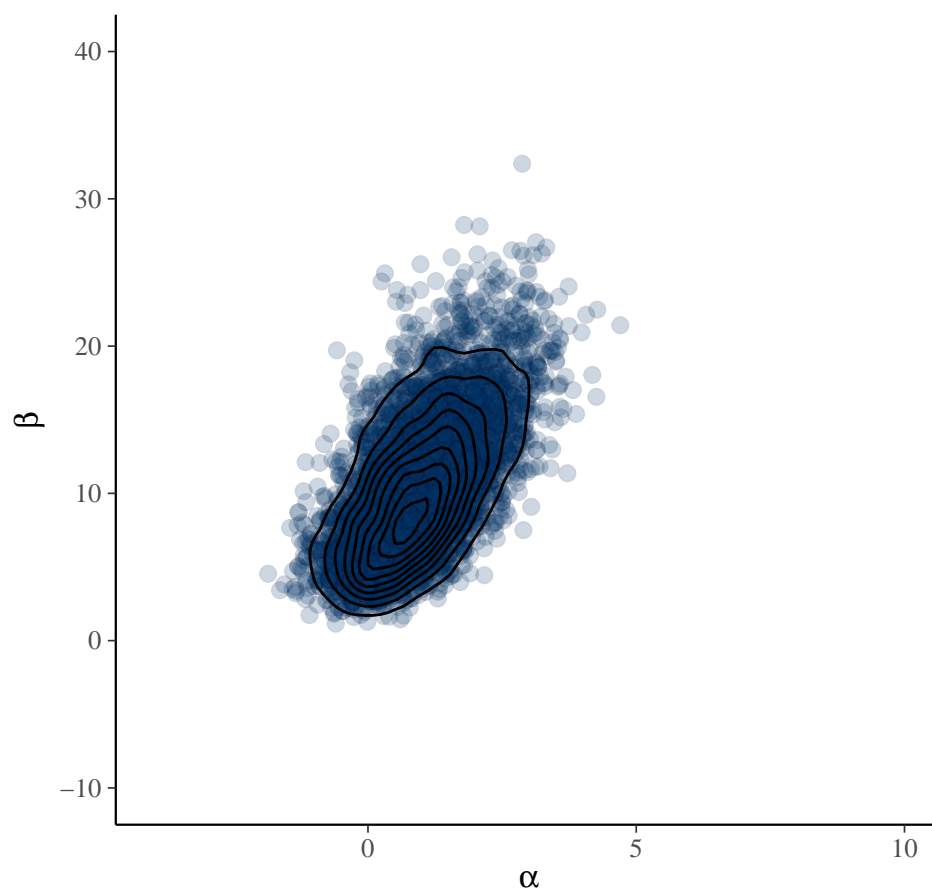
Here, we plot the draws from α and β in a scatter plot to compare the results with Figure 3.3b in BDA3. The x- and y-axis limits are adjusted to capture the same grid as in the book figure.

```

p <- mcmc_scatter(fit, pars = c('theta[1]', 'theta[2]'), alpha = 0.2)
(p + labs(title = "Scatter plot of alpha and beta draws",
           x = expression(alpha),
           y = expression(beta)) + stat_density_2d(color = "black", size = .5)
  + xlim(-4, 10) + ylim(-10, 40))

```

Scatter plot of alpha and beta draws



All in all, the results look quite similar with some slight difference due to the different choice of prior (uniform prior compared to Gaussian).

Exercise 4

This exercise has been written and compiled in Windows using R and the package `Rstan`. I had some installation issues in the beginning since I did not follow the instructions of installing the `Rstan` from source as I had already used it in the previous assignment. The problems disappeared when I followed the instructions. I thought about using Aalto's jupyter when I run into problems, but I resolved those problems locally in the end.

The hardest part about the assignment was getting the hang of Stan syntax and how the model is defined. I believe this will get easier with practice.