# **Dataset Exploration!**

This notebook takes an initial look at the datasets downloaded. It identifies techniques to view the features of each image, as well as to generate videos to understand the movement and change in forms of the cells over time.

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This Jupyter Notebook aims at exploring the datasets for the project, and finding ways to represent the data in an easy way

## **Imports**

```
In [1]: from os import getcwd, walk, mkdir, stat, remove
    from os import sep # used later on, in a function, to print directory contents
    from os.path import exists, basename, join

from shutil import move, copy

import matplotlib.pyplot as plt
import cv2
```

# dataset File Heirarchy

This section of the notebook focusses on understanding the file heirarchy of the datasets

Let us start with some imports:

The file path used by author should be: "C:\Users\G5\UKZN\COMP700"

```
In [3]: desired_directory = "..\\..\\Comp700_DataSets"
    current_directory = getcwd()
    print( exists(current_directory + "\\" + desired_directory) )
True
```

The output should be true as the file path we are interested in is:

```
Documents
|_____Comp700_DataSets
```

```
_____ ...
_____Github
_____Comp700
_____ ...
```

There are many files inside this dataset folder, so let us get the first 2 'branches' to show the user

```
In [4]: path = walk(current directory + "\\" + desired directory)
       count = 0
       for root, dirs, files in path:
          print(dirs)
           print("Length is:", len(dirs), "\n")
           if (count == 1):
               break
           count += 1
       ['Extracted', 'OriginalZipped']
       Length is: 2
       ['BF-C2DL-HSC', 'BF-C2DL-HSC (1)', 'BF-C2DL-MuSC', 'BF-C2DL-MuSC (1)', 'DIC-C2DH-HeLa',
       'DIC-C2DH-HeLa (1)', 'Fluo-C2DL-Huh7', 'Fluo-C2DL-Huh7 (1)', 'Fluo-C2DL-MSC', 'Fluo-C2DL
       -MSC (1)', 'Fluo-N2DH-GOWT1', 'Fluo-N2DH-GOWT1 (1)', 'Fluo-N2DH-SIM+', 'Fluo-N2DH-SIM+
       (1)', 'Fluo-N2DL-HeLa', 'Fluo-N2DL-HeLa (1)', 'PhC-C2DH-U373', 'PhC-C2DH-U373 (1)', 'PhC
       -C2DL-PSC', 'PhC-C2DL-PSC (1)']
       Length is: 20
       I.e. :
           Documents
             ____
               ____Comp700_DataSets
                         Extracted
                            BF-C2DL-HSC
                               \_BF-C2DL-HSC (1)
                          OriginalZipped
                           BF-C2DL-HSC.zip
                            _____BF-C2DL-HSC (1).zip
                            _____
             Github
```

Zipped files are contained in the directory called *OriginalZipped* 

A detailed breakdown of all directories can be shown with following function, adapted from stack overflow:

```
In [5]: # from os import sep
# Author: dhobbs on Mar 15, 2012 at 21:29,
```

```
# https:\\\\stackoverflow.com\\questions\\9727673\\list-directory-tree-structure-in-pyth
def list files(startpath):
    for root, dirs, files in walk(startpath):
        level = root.replace(startpath, '').count(sep)
        indent = ' ' * 4 * (level)
        print('{}{}\\'.format(indent, basename(root)))
        subindent = ' ' * 4 * (level + 1)
        for f in files:
            # 3 lines of code added to improve readibility
            if (".tif" in f):
                print('{}{}'.format(subindent, "..."))
            print('{}{}'.format(subindent, f))
path = (current directory + "\\" + desired directory)
list files (path)
Comp700 DataSets\
   Extracted\
       BF-C2DL-HSC\
            BF-C2DL-HSC\
                01\
                01 GT\
                    SEG\
                    TRA\
                       man track.txt
                01 ST\
                    SEG\
                       . . .
                02\
                02 GT\
                    SEG\
                        . . .
                    TRA\
                        man track.txt
                02 ST\
                    SEG\
        BF-C2DL-HSC (1) \
            BF-C2DL-HSC (1)\
                01\
                02\
        BF-C2DL-MuSC\
            BF-C2DL-MuSC\
                01\
                01 GT\
                    SEG\
                    TRA\
                       man track.txt
                01 ST\
                    SEG\
                02\
                02 GT\
```

```
SEG\
           TRA\
           man_track.txt
       02 ST\
          SEG\
          . . .
BF-C2DL-MuSC (1) \
  BF-C2DL-MuSC (1) \
      01\
          . . .
       02\
DIC-C2DH-HeLa\
   DIC-C2DH-HeLa\
     01\
       01 GT\
         SEG\
          ...
           TRA\
           man_track.txt
       01 ST\
        SEG\
           . . .
       02\
       02 GT\
          SEG\
          TRA\
           man_track.txt
       02 ST\
          SEG\
DIC-C2DH-HeLa (1) \
   DIC-C2DH-HeLa (1) \
      01\
       02\
Fluo-C2DL-Huh7\
   Fluo-C2DL-Huh7\
      01\
       01 GT\
          SEG\
           TRA\
           man_track.txt
       02\
          . . .
       02 GT\
          SEG\
           . . .
           TRA\
           man_track.txt
Fluo-C2DL-Huh7 (1) \
   Fluo-C2DL-Huh7 (1) \
      01\
```

```
02\
Fluo-C2DL-MSC\
   Fluo-C2DL-MSC\
       01\
       01 GT\
          SEG\
          TRA\
           man_track.txt
              . . .
       01 ST\
          SEG\
           ...
       02\
       02 GT\
          SEG\
          TRA\
             man_track.txt
       02 ST\
          SEG\
Fluo-C2DL-MSC (1) \
  Fluo-C2DL-MSC (1) \
       01\
          . . .
       02\
Fluo-N2DH-GOWT1\
   Fluo-N2DH-GOWT1\
       01\
       01 GT\
          SEG\
          TRA\
           man_track.txt
       01_ST\
          SEG\
           ...
       02\
          . . .
       02 GT\
          SEG\
            . . .
           TRA\
            man track.txt
       02_ST\
          SEG\
Fluo-N2DH-GOWT1 (1) \
  Fluo-N2DH-GOWT1 (1)
       01\
       02\
Fluo-N2DH-SIM+\
   Fluo-N2DH-SIM+\
      01\
```

```
01 GT\
          SEG\
           TRA\
           man_track.txt
             . . .
       02\
          . . .
       02 GT\
          SEG\
          TRA\
            man track.txt
Fluo-N2DH-SIM+ (1) \
   Fluo-N2DH-SIM+ (1) \
      01\
       02\
Fluo-N2DL-HeLa\
   Fluo-N2DL-HeLa\
       01\
       01 GT\
          SEG\
              . . .
          TRA\
            man_track.txt
       01 ST\
         SEG\
           ...
       02\
       02 GT\
          SEG\
           TRA\
           man_track.txt
              . . .
       02 ST\
          SEG\
Fluo-N2DL-HeLa (1) \
   Fluo-N2DL-HeLa (1) \
      01\
          . . .
       02\
PhC-C2DH-U373\
   PhC-C2DH-U373\
       01\
       01 GT\
         SEG\
            . . .
            man track.txt
       01_ST\
          SEG\
              . . .
       02\
```

02 GT\

```
SEG\
                TRA\
                  man track.txt
            02 ST\
                SEG\
                   . . .
    PhC-C2DH-U373 (1) \
       PhC-C2DH-U373 (1)\
           01\
            02\
    PhC-C2DL-PSC\
       PhC-C2DL-PSC\
           01\
            01 GT\
               SEG\
                 . . .
                TRA\
                  man track.txt
            01 ST\
               SEG\
                   . . .
            02\
            02 GT\
                SEG\
                TRA\
                man track.txt
            02 ST\
               SEG\
    PhC-C2DL-PSC (1) \
       PhC-C2DL-PSC (1) \
           01\
            02\
OriginalZipped\
   BF-C2DL-HSC (1).zip
   BF-C2DL-HSC.zip
   BF-C2DL-MuSC(1).zip
   BF-C2DL-MuSC.zip
   DIC-C2DH-HeLa(1).zip
   DIC-C2DH-HeLa.zip
   Fluo-C2DL-Huh7(1).zip
   Fluo-C2DL-Huh7.zip
   Fluo-C2DL-MSC(1).zip
   Fluo-C2DL-MSC.zip
   Fluo-N2DH-GOWT1(1).zip
   Fluo-N2DH-GOWT1.zip
   Fluo-N2DH-SIM+(1).zip
   Fluo-N2DH-SIM+.zip
   Fluo-N2DL-HeLa(1).zip
   Fluo-N2DL-HeLa.zip
   PhC-C2DH-U373(1).zip
   PhC-C2DH-U373.zip
    PhC-C2DL-PSC(1).zip
    PhC-C2DL-PSC.zip
```

The ellipses above indicate that images are contained in that location.

In order to view every folder's collection of images, we need to travel individually into each folder and look at the images contained there.

Before we go any further, it is important to acknowledge the structure of the directories.

There are 20 datasets contained here. Every folder that has the suffix (1) represents the CHALLENGE dataset, provided by the Cell Tracking Challenge.

Inside the TRAINING datasets, additional folders appear, either as shown on the left OR as shown on the right:

01	01
01_GT	01_GT
01_ST	02
02	02_GT
02_GT	
02 ST	

If the folder contains the suffix "\_GT" of "\_ST", then the image contents have the naming convention "man\_segXXXX.tif" or "man\_trackXXXX.tif", otherwise the images have the naming convention "tXXXX.tif", where XXXX represents the number of the image in that folder

The author suspects that "man\_seg" represents a manually segmented cell, whereas "man\_track" represents a manually tracked cell. The other files are the "Petri Dish" files, showing the complete landscape of the cells under the microscope

Now, windows photo viewer (and the extension in VS Code) show the "man\_" images as pure black images.... This is not helpful or useful. We need to identify how to see the details here!

# Image recognition

In the section below, the notebook will explore ways to extract the valuable information from these seemingly black images

To verify, let us bring 3 images to the local directory and try open them using OpenCV

```
In [7]: # get 3 images and save to local folder

path = walk(desired_directory)
```

```
# hard coded to generate a local copy
locations = [
            "Extracted\\BF-C2DL-HSC\\BF-C2DL-HSC\\01",
            "Extracted\\BF-C2DL-HSC\\BF-C2DL-HSC\\01 GT\\SEG",
            "Extracted\\BF-C2DL-HSC\\BF-C2DL-HSC\\01 GT\\TRA"
flags = ["t0058.tif", "man seg0058.tif", "man track0058.tif"]
flag count = 0
for root, dirs, files in path:
    # print(files)
    # terminate
   if (flag count > 2):
        break
    for file in files:
        # terminate
        if (flag count > 2):
           break
        elif (flags[flag count] == file):
            # print("Yerp")
            copy( join( join(desired directory, locations[flag count]) , file) , sample
            flag count += 1
           break
```

However, If we use Matplotlib to view a sample of each of the TIF images, we see the following:

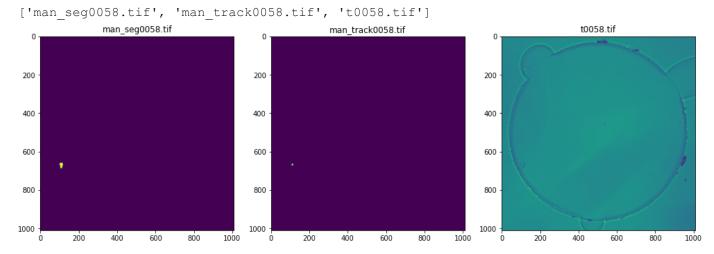
```
In [8]: path = walk(current_directory + "\\" + sample_directory)

i = 1
for root, dirs, files in path:
    print(files)
    fig = plt.figure(figsize=(14, 8))

for item in files:
    location = (current_directory + "\\" + sample_directory + "\\" + item)
    img = plt.imread(location)

    fig.add_subplot(1, 3, i)
    plt.title(item)
    plt.imshow(img)
    i += 1

plt.tight_layout()
plt.show()
```



Here is the same, as grayscale:

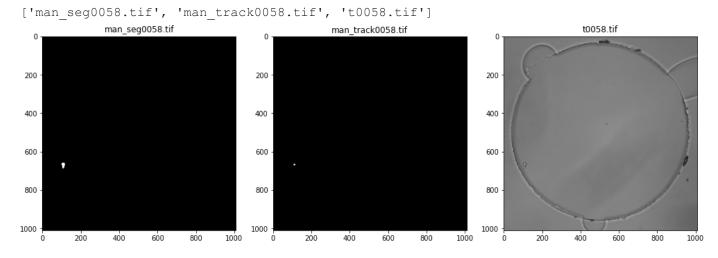
```
In [9]: path = walk(current_directory + "\\" + sample_directory)

i = 1
for root, dirs, files in path:
    print(files)
    fig = plt.figure(figsize=(14, 8))

for item in files:
        location = (current_directory + "\\" + sample_directory + "\\" + item)
        img = plt.imread(location)

        fig.add_subplot(1, 3, i)
        plt.title(item)
        plt.imshow(img, cmap="gray")
        i += 1

plt.tight_layout()
plt.show()
```

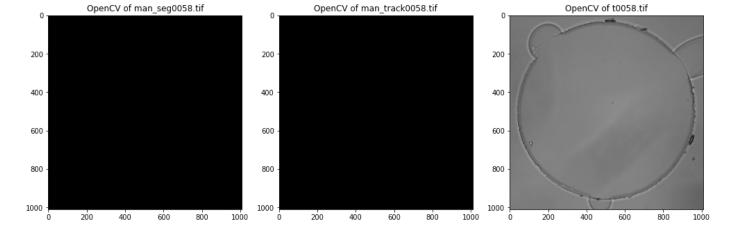


As we can see from the pop-up window - the details in the Opencv window are not being shown... They ARE being shown in the Matplotlib.Pyplot package.

This is not desirable, as Opencv has some useful tools we'd like to use for data visualization. So we need to find a way to get the features shown from Matplotlib, and preserve them for future use with Opencv

Let us see if we can mix and match:

```
path = walk(current directory + "\\" + sample directory)
In [10]:
         i = 1
         for root, dirs, files in path:
            print(files)
             fig = plt.figure(figsize=(14, 8))
             for item in files:
                 location = (current directory + "\\" + sample directory + "\\" + item)
                 img = cv2.imread(location, cv2.IMREAD GRAYSCALE)
                 fig.add subplot(1, 3, i)
                 plt.title("OpenCV of " + item)
                 plt.imshow(img, cmap="gray")
                 i += 1
         plt.tight layout()
         plt.show()
         ['man seg0058.tif', 'man track0058.tif', 't0058.tif']
```



NO! We cannot mix and match. What about saving a file using Matplotlib and then opening it using Opency?

```
destination directory 1 = "002 DataSetExploration"
In [11]:
         destination directory 2 = "MatplotlibSavedImages"
         destination directory = destination directory 1 + "\\" + destination directory 2
         try:
             mkdir( join( join(current directory, destination directory 1) , destination director
         except FileExistsError:
            pass
         except:
             print("Unknown Error Encountered...")
In [12]: path = walk(current_directory + "\\" + sample directory)
         \# i = 1
         for root, dirs, files in path:
             print(files)
             # fig = plt.figure(figsize=(14, 8))
             for item in files:
                 location = (current directory + "\\" + sample directory + "\\" + item)
                 img = plt.imread(location)
                 name = destination directory + "\\" + "Matplotlib " + item[ : -4] + ".jpg"
                 plt.imsave(name, img, cmap="gray")
         path = walk(current directory + "\\" + destination directory)
         for root, dirs, files in path:
            print(files)
             for item in files:
                 location = (current directory + "\\" + destination directory + "\\" + item)
                 img = cv2.imread(location, cv2.IMREAD GRAYSCALE)
                 (x, y) = img.shape
                 break
         imgSmall = cv2.resize(img, (x // 2, y // 2))
         cv2.imshow(item, imgSmall)
         cv2.waitKey(0)
         ['man seg0058.tif', 'man track0058.tif', 't0058.tif']
         ['Matplotlib man seg0058.jpg', 'Matplotlib man track0058.jpg', 'Matplotlib t0058.jpg']
         -1
```

YES! We can do that. So that is one option for how we can address the inconsitent black images. However, this option changes the quality from TIFF to JPG...

Out[12]:

Another option is to attempt to read the images in using matplotlib.pyplot and then write them using Opencv

Let us try to do that now:

```
destination directory 1 = "002 DataSetExploration"
In [13]:
         destination directory 2 = "OpencySavedImages"
         destination directory = destination directory 1 + "\\" + destination directory 2
         try:
            mkdir(join(current directory, destination directory 1), destination director
        except FileExistsError:
            pass
        except:
            print("Unknown Error Encountered...")
In [14]: path = walk(current directory + "\\" + sample directory)
         \# i = 1
         for root, dirs, files in path:
            print(files)
            # fig = plt.figure(figsize=(14, 8))
            for item in files:
                location = (current directory + "\\" + sample directory + "\\" + item)
                img = plt.imread(location)
                name = destination_directory + "\\" + "Opencv " + item
                cv2.imwrite(name, img)
        path = walk(current directory + "\\" + destination directory)
         for root, dirs, files in path:
            print(files)
            for item in files:
                 location = (current directory + "\\" + destination directory + "\\" + item)
                img = cv2.imread(location, cv2.IMREAD GRAYSCALE)
                (x, y) = img.shape
                break
         imgSmall = cv2.resize(img, (x // 2, y // 2))
         cv2.imshow(item, imgSmall)
         cv2.waitKey(0)
         ['man seg0058.tif', 'man track0058.tif', 't0058.tif']
         ['Opencv_man_seg0058.tif', 'Opencv man track0058.tif', 'Opencv t0058.tif']
        -1
Out[14]:
```

Unfortunately, that does not work... So our best bet is to read them in and save them using Matplotlib. This is not ideal, as we may lose a bit of quality converting from TIF to JPG

We desire to read in the image using Matplotlib, and write it to disk as a grayscale TIF. Though this does not appear possible, as Matplotlib does not support TIFF extensions. Let's try create Grayscale TIFF's from the sample pictures?

```
In [15]: destination_directory_1 = "002_DataSetExploration"
    destination_directory_2 = "GrayscaleTiffs"
    destination_directory = destination_directory_1 + "\\" + destination_directory_2

try:
    mkdir( join( join(current_directory, destination_directory_1) , destination_director
except FileExistsError:
```

```
pass
except:
    print("Unknown Error Encountered...")

In [16]: path = walk(current_directory + "\\" + sample_directory)

# i = 1
for root, dirs, files in path:
    print(files)
    # fig = plt.figure(figsize=(14, 8))

for item in files:
    location = (current_directory + "\\" + sample_directory + "\\" + item)
    img = cv2.imread(location, cv2.IMREAD_GRAYSCALE)
    name = destination directory + "\\" + "Grayscale" + item
```

```
['man seg0058.tif', 'man track0058.tif', 't0058.tif']
```

Unfortunately, no difference! Looks like our best bet is to convert it to JPG when we need to... Let's move on

We can use a function to calculate the size of a file like this:

cv2.imwrite(name, img)

```
In [17]: # from os import stat

bytes = stat("002_DataSetExploration\\GrayscaleTiffs\\Grayscale_man_seg0058.tif").st_siz
print("File is", bytes / 1024, "kb's")

File is 19.185546875 kb's
```

If we compare the Sample Images file size to that of our other files, we recognize the following:

```
In [18]: places array = ["002 DataSetExploration\\SampleImages", "002 DataSetExploration\\Graysca
                        "002 DataSetExploration\\MatplotlibSavedImages", "002 DataSetExploration
        spacing length = len("002 DataSetExploration\\MatplotlibSavedImages") # used to improve
        for i in range(len(places array)):
            location = current directory + "\\" + places array[i]
            path = walk(location)
            for root, dirs, files in path:
                print(places array[i], " " * ( spacing length - len(places array[i]) ), end="\t"
                for item in files:
                   bytes = stat(location + "\\" + item).st size
                   bytes = bytes / 1024
                   print( str(bytes)[ : 6], "kb's", end="\t") # keep first 5 sig figs (ignore f
                print()
        002 DataSetExploration\SampleImages
                                                      38.419 kb's 38.281 kb's 509.12 k
        b's
        002 DataSetExploration\GrayscaleTiffs
                                                     19.185 kb's 19.185 kb's 414.22 k
        002 DataSetExploration\MatplotlibSavedImages
                                                                     16.738 kb's
                                                                                    42.908 k
                                                     16.892 kb's
        002 DataSetExploration\OpencvSavedImages
                                                     38.277 kb's
                                                                    38.201 kb's
                                                                                    414.22 k
        b's
```

From the results above, we can see that the Grayscale TIFFs we created result in a loss of information for the manually segmented and tracked images, but only a small loss for the main image.

The file sizes for MatplotlibSavedImages follows a similar pattern, however the original Petri Dish image has a large loss in quality...

OpencvSavedImages should be virtually identical to the original pictures, as Opencv merely read them in and saved them, as is.

From the above, we can infer that a loss in quality in the manually segmented and tracked images is acceptable, however we would like to try preserve the data in the Petri dish image.

Before we conclude this, let us look at the Pillow package to identify if we can see the details in the manually segmented and tracked images

```
In [19]: # Imports PIL module
from PIL import Image

im = Image.open("002_DataSetExploration\\SampleImages\\man_seg0058.tif")
im.show()
```

Unfortunately not... We have no choice but to convert each image into a grayscale image through Matplotlib.Pyplot, in order to see the details for the manually segmented and tracked images. One option we have is to leave the Petri Dish images as Tiff, and convert the rest to JPG... that solution may be ideal as we preserve the most amount of information for our segmentation software.

#### dataset Cell Variation

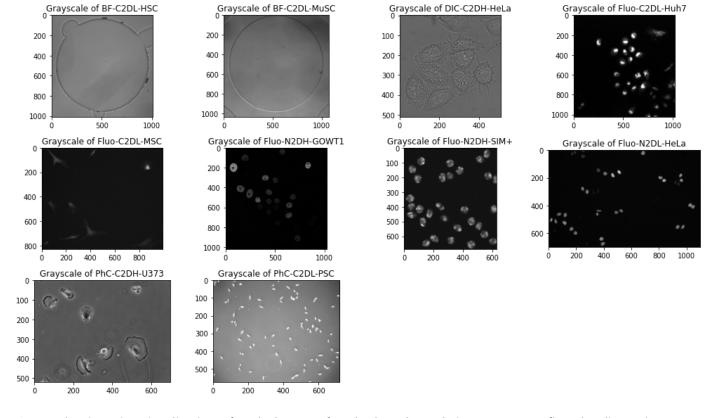
This section of the notebook focusses on sampling one Perti Dish image from each folder and placing them together, to see the different cells present

```
In [20]:
         We only need to show every OTHER folder, as each dataset has a
         training and challenge set. So out of 20 files, we need to show 10
         First things first, let us create an array of the directory locations
         data sets = "..\\..\\Comp700 DataSets"
        path = walk(current directory + "\\" + data sets)
         directory array = [] # contains the main folders
         i = 1
         for root, dirs, files in path:
            if (i == 2):
                directory array = dirs
                break
            i += 1
        print(directory array)
        print("\nStarting matplotlib\n")
        path = walk(current directory + "\\" + data sets) # reset path
         fig = plt.figure(figsize=(14, 8))
```

```
i = -1
temp = -1
counter = 1 # used for matplotlib subplots
for root, dirs, files in path:
    # print(dirs)
    for item in files:
        # only execute for first picture in directory
        if ("t0000.tif" == item) or ("t000.tif" == item):
            i += 1
            # skips folder "02" in datasets
            if (i % 2 == 1):
                break
            # print(i)
            temp = i // 2
            # skip Challenge datasets
            if ("(1)" in directory array[temp]):
                break
            location = ( current directory + "\\" + data sets + "\\Extracted\\" + direct
                        "\\" + directory array[temp] + "\\01\\" + item)
            # print(location)
            img = plt.imread(location)
            fig.add subplot(3, 4, counter)
            plt.title("Grayscale of " + directory array[temp])
            plt.imshow(img, cmap="gray")
            counter += 1
            break
        else:
            break
plt.tight layout()
# save file for future use
plt.savefig("002 DataSetExploration\\Visualization Of Cells.jpg")
plt.show()
# save file for future use
# plt.savefig("Visualization Of Cells.jpg")
['BF-C2DL-HSC', 'BF-C2DL-HSC (1)', 'BF-C2DL-MuSC', 'BF-C2DL-MuSC (1)', 'DIC-C2DH-HeLa',
'DIC-C2DH-HeLa (1)', 'Fluo-C2DL-Huh7', 'Fluo-C2DL-Huh7 (1)', 'Fluo-C2DL-MSC', 'Fluo-C2DL
```

['BF-C2DL-HSC', 'BF-C2DL-HSC (1)', 'BF-C2DL-MuSC', 'BF-C2DL-MuSC (1)', 'DIC-C2DH-HeLa', 'DIC-C2DH-HeLa (1)', 'Fluo-C2DL-Huh7', 'Fluo-C2DL-Huh7 (1)', 'Fluo-C2DL-MSC', 'Fluo-C2DL-MSC (1)', 'Fluo-N2DH-GOWT1', 'Fluo-N2DH-GOWT1 (1)', 'Fluo-N2DH-SIM+', 'Fluo-N2DH-SIM+ (1)', 'Fluo-N2DL-HeLa', 'Fluo-N2DL-HeLa (1)', 'PhC-C2DH-U373', 'PhC-C2DH-U373 (1)', 'PhC-C2DL-PSC', 'PhC-C2DL-PSC (1)']

Starting matplotlib



Great! That is a nice visualization of each dataset. If we look at the website, we can confirm the dimensions of each microscope used:

dataset Name	Pixel Size (Microns)	Time Step (Min)
BF-C2DL-HSC	0.645 X 0.645	5
BF-C2DL-MuSC	0.645 X 0.645	5
DIC-C2DH-HeLa	0.19 x 0.19	10
Fluo-C2DL-Huh7	0.65 x 0.65	15
Fluo-C2DL-MSC	0.3 x 0.3	20
Fluo-N2DH-GOWT1	0.240 x 0.240	5
Fluo-N2DH-SIM+	0.125 x 0.125	29
Fluo-N2DL-HeLa	0.645 x 0.645	30
PhC-C2DH-U373	0.65 x 0.65	15
PhC-C2DL-PSC	1.6 x 1.6	10

## dataset Videos Variables

This section of the notebook will generate videos for each of the datasets

We desire to have a short video showing the movement of the cells over time, which we can refer to when we want to understand the features of the data.

We will need to use OpenCV to generate the videos, depending on the length of data present. For example, folders containing 1000 images may run for more time than those with 300 images.

Each video (for Training and Challenge) will be saved in a folder, outside of Github, for ease of reference. This is because the combined file size can be ~200MB, so it is same in the same directory as the dataset images

First, let us create an a few arrays for the information we need:

```
In [21]: # a list of the directories - will be used to name the videos
        print(directory array)
         ['BF-C2DL-HSC', 'BF-C2DL-HSC (1)', 'BF-C2DL-MuSC', 'BF-C2DL-MuSC (1)', 'DIC-C2DH-HeLa',
         'DIC-C2DH-HeLa (1)', 'Fluo-C2DL-Huh7', 'Fluo-C2DL-Huh7 (1)', 'Fluo-C2DL-MSC', 'Fluo-C2DL
        -MSC (1)', 'Fluo-N2DH-GOWT1', 'Fluo-N2DH-GOWT1 (1)', 'Fluo-N2DH-SIM+', 'Fluo-N2DH-SIM+
         (1)', 'Fluo-N2DL-HeLa', 'Fluo-N2DL-HeLa (1)', 'PhC-C2DH-U373', 'PhC-C2DH-U373 (1)', 'PhC
        -C2DL-PSC', 'PhC-C2DL-PSC (1)']
In [22]: | # First, generate a list of the locations for each folder of Petri Dish images
        path = walk(current directory + "\\" + data sets) # reset path
        location array = []
         # used to cycle between 2 folders, present in each directory
         sub directory choice = ["\01\", "\02\"]
         i = 0 # will grow from 0 to 39
         index = 0 # we need to lie in [0, 19]
         for root, dirs, files in path:
            # print(dirs)
            for item in files:
                 if ("t0000.tif" == item) or ("t000.tif" == item):
                     index = i // 2
                     location = ( current directory + "\\" + data sets + "\\Extracted\\" + direct
                                 "\\" + directory array[index] + sub directory choice[i % 2])
                     i += 1
                     # print(location)
                     location array.append(location)
        print(location array)
```

['c:\\Users\\G5\\Documents\\GitHub\\COMP700\\..\\Comp700 DataSets\\Extracted\\BF-C2D L-HSC\\BF-C2DL-HSC\\01\\', 'c:\\Users\\G5\\Documents\\GitHub\\COMP700\\..\\..\\Comp700 D \\COMP700\\..\\.\\Comp700 DataSets\\Extracted\\BF-C2DL-HSC (1)\\BF-C2DL-HSC (1)\\01\\', c:\\Users\\G5\\Documents\\GitHub\\COMP700\\..\\..\\Comp700 DataSets\\Extracted\\BF-C2DL -HSC (1) \\BF-C2DL-HSC (1) \\02\\', 'c:\\Users\\G5\\Documents\\GitHub\\COMP700\\..\\..\\Co  $\label{thm:condition} $$ mp700 \ DataSets\Extracted\BF-C2DL-MuSC\BF-C2DL-MuSC\01\\', 'c:\Users\G5\Documents $$$ \\GitHub\\COMP700\\..\\.\\Comp700 DataSets\\Extracted\\BF-C2DL-MuSC\\BF-C2DL-MuSC\\02  $\ 'c:\Users\G5\Documents\GitHub\COMP700\...\Comp700\ DataSets\Extracted\BF$ -C2DL-MuSC (1) \\BF-C2DL-MuSC (1) \\01\\', 'c:\\Users\\G5\\Documents\\GitHub\\COMP700 \\..\\Comp700 DataSets\\Extracted\\BF-C2DL-MuSC (1)\\BF-C2DL-MuSC (1)\\02\\', 'c:\\U sers\\G5\\Documents\\GitHub\\COMP700\\..\\Comp700 DataSets\\Extracted\\DIC-C2DH-HeLa \\DIC-C2DH-HeLa\\01\\', 'c:\\Users\\G5\\Documents\\GitHub\\COMP700\\..\\..\\Comp700 Data Sets\\Extracted\\DIC-C2DH-HeLa\\DIC-C2DH-HeLa\\02\\', 'c:\\Users\\G5\\Documents\\GitHub \\COMP700\\..\\.\\Comp700 DataSets\\Extracted\\DIC-C2DH-HeLa (1)\\DIC-C2DH-HeLa (1)\\01 \\', 'c:\\Users\\G5\\Documents\\GitHub\\COMP700\\..\\.\\Comp700 DataSets\\Extracted\\DI \\..\\Comp700 DataSets\\Extracted\\Fluo-C2DL-Huh7\\Fluo-C2DL-Huh7\\01\\', 'c:\\Users \\G5\\Documents\\GitHub\\COMP700\\..\\..\\Comp700 DataSets\\Extracted\\Fluo-C2DL-Huh7\\F  $\label{locality} $$ 1uo-C2DL-Huh7\02\', 'c:\Users\G5\Documents\GitHub\COMP700\...\Comp700 DataSe $$$  $ts\Times C2DL-Huh7 (1)\Times C2DL-Huh7 (1)\V01\V', 'c:\Users\V65\Documents$ \\GitHub\\COMP700\\..\\.\\Comp700 DataSets\\Extracted\\Fluo-C2DL-Huh7 (1)\\Fluo-C2DL-Hu h7 (1)\\02\\', 'c:\\Users\\G5\\Documents\\GitHub\\COMP700\\..\\..\\Comp700 DataSets\\Ext

 $\verb|racted|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL-MSC|\fluo-C2DL$ \\..\\Comp700 DataSets\\Extracted\\Fluo-C2DL-MSC\\Fluo-C2DL-MSC\\02\\', 'c:\\Users \\G5\\Documents\\GitHub\\COMP700\\..\\..\\Comp700 DataSets\\Extracted\\Fluo-C2DL-MSC (1) \\Fluo-C2DL-MSC (1)\\01\\', 'c:\\Users\\G5\\Documents\\GitHub\\COMP700\\..\\..\\Comp700 nts\\GitHub\\COMP700\\..\\Comp700 DataSets\\Extracted\\Fluo-N2DH-GOWT1\\Fluo-N2DH-GO  $\label{thm:linear} $$ T1\ol\', 'c:\Users\G5\Documents\GitHub\COMP700\...\Comp700 DataSets\Extrac Paracle Par$ ted\\Fluo-N2DH-GOWT1\\Fluo-N2DH-GOWT1\\02\\', 'c:\\Users\\G5\\Documents\\GitHub\\COMP700 c:\\Users\\G5\\Documents\\GitHub\\COMP700\\..\\..\\Comp700 DataSets\\Extracted\\Fluo-N2 DH-GOWT1 (1)\\Fluo-N2DH-GOWT1 (1)\\02\\', 'c:\\Users\\G5\\Documents\\GitHub\\COMP700 \\G5\\Documents\\GitHub\\COMP700\\..\\.\\Comp700 DataSets\\Extracted\\Fluo-N2DH-SIM+\\F luo-N2DH-SIM+\\02\\', 'c:\\Users\\G5\\Documents\\GitHub\\COMP700\\..\\..\\Comp700 DataSe  $ts\xspace{$ts\xspace{0.5}\xspace{0.5}} ts\xspace{0.5}\xspace{0.5$ \\GitHub\\COMP700\\..\\.\\Comp700 DataSets\\Extracted\\Fluo-N2DH-SIM+ (1)\\Fluo-N2DH-SI racted\\Fluo-N2DL-HeLa\\Fluo-N2DL-HeLa\\01\\', 'c:\\Users\\G5\\Documents\\GitHub\\COMP70 0\\..\\Comp700 DataSets\\Extracted\\Fluo-N2DL-HeLa\\Fluo-N2DL-HeLa\\02\\', 'c:\\User s\\G5\\Documents\\GitHub\\COMP700\\...\\Comp700 DataSets\\Extracted\\Fluo-N2DL-HeLa (1)  $\label{eq:comp700}...\$  c:\\Users\\G5\\Documents\\GitHub\\COMP700\\...\\Comp 700 DataSets\\Extracted\\Fluo-N2DL-HeLa (1)\\Fluo-N2DL-HeLa (1)\\02\\', 'c:\\Users\\G5 \\Documents\\GitHub\\COMP700\\..\\Comp700 DataSets\\Extracted\\PhC-C2DH-U373\\PhC-C2 DH-U373\\01\\', 'c:\\Users\\G5\\Documents\\GitHub\\COMP700\\..\\..\\Comp700 DataSets\\Ex tracted\\PhC-C2DH-U373\\PhC-C2DH-U373\\02\\', 'c:\\Users\\G5\\Documents\\GitHub\\COMP700 \\..\\Comp700 DataSets\\Extracted\\PhC-C2DH-U373 (1)\\PhC-C2DH-U373 (1)\\01\\', c:\\Users\\G5\\Documents\\GitHub\\COMP700\\..\\Comp700 DataSets\\Extracted\\PhC-C2D H-U373 (1)\\PhC-C2DH-U373 (1)\\02\\', 'c:\\Users\\G5\\Documents\\GitHub\\COMP700 \\Documents\\GitHub\\COMP700\\..\\Comp700 DataSets\\Extracted\\PhC-C2DL-PSC\\PhC-C2D  $L-PSC\\02\\', 'c:\\05\\Documents\\GitHub\\COMP700\\...\\Comp700 DataSets\\Extr$ acted\\PhC-C2DL-PSC (1)\\PhC-C2DL-PSC (1)\\01\\', 'c:\\Users\\G5\\Documents\\GitHub\\COM P700\\..\\Comp700 DataSets\\Extracted\\PhC-C2DL-PSC (1)\\PhC-C2DL-PSC (1)\\02\\']

The cell below verifies that all cells have exactly 1 set of dimensions across the folder. It may take a few minutes to run

```
In [23]: # # Now, use the location array to determine if each dataset has consistent image sizes
         # path = walk(current directory + "\\" + data sets) # reset path
         # temp array = []
         \# i = -1 \# will grow from 0 to 39
         # for root, dirs, files in path:
         #
             # print(files)
               for item in files:
                   if ("man " not in item) and (".zip" not in item):
         #
                        # update on first element only
         #
                       if ("t0000.tif" == item) or ("t000.tif" == item):
                           i += 1
         #
                        # print(location array[i] + item, exists(location array[i] + item))
         #
                       # break
         #
                       img = cv2.imread( (location array[i] + item), cv2.IMREAD GRAYSCALE)
         #
                       (x, y) = img.shape
         #
                       # only keep distinct sizes
         #
                       if ([x, y] \text{ not in temp array}):
                           temp array.append([x, y])
                   # skip "man " images
                   else:
```

```
# break

# if (len(temp_array) != 0):
# print(temp_array, end="\t")

# temp_array = []
```

From the test above, we can see that the images are already consistent in their dimensions, however the dimensions vary per dataset. We can use this knowledge to just find the dimensions of the first image in each folder, which will be used further on

```
In [24]: image size array = []
         path = walk(current directory + "\\" + data sets) # reset path
         i = -1 \# will grow from 0 to 39
         for root, dirs, files in path:
             for item in files:
                 if ("man " not in item) and (".zip" not in item):
                     # update on first element only
                     if ("t0000.tif" == item) or ("t000.tif" == item):
                         i += 1
                     img = cv2.imread( (location array[i] + item), cv2.IMREAD GRAYSCALE)
                     (x, y) = img.shape
                     image size array.append([x, y])
                     break
                 # skip "man " images
                 else:
                     break
         print(image size array)
```

[[1010, 1010], [1010, 1010], [1010, 1010], [1010, 1010], [1010, 1010], [1036, 1070], [1036, 1070], [1036, 1070], [1036, 1070], [512, 512], [512, 512], [512, 512], [512, 512], [1024, 1024], [1024, 1024], [1024, 1024], [832, 992], [782, 1200], [832, 992], [782, 1200], [832, 992], [782, 1200], [1024, 1024], [1024, 1024], [1024, 1024], [1024, 1024], [690, 628], [773, 739], [718, 660], [790, 664], [700, 1100], [700, 1100], [700, 1100], [700, 1100], [520, 696], [520, 696], [520, 696], [576, 720], [576, 720], [576, 720]]

OKAY! We now have useful arrays we can use to generate our videos: location\_array is used to find the position of the images, and image\_size\_array is used to find the dimensions of the folder. Next, we need to determine the quantity of pictures in each folder, as that will become our frame rate:

```
count = 0 # reset
print(quantity_images_per_folder)

[1764, 1764, 1763, 1763, 1375, 1376, 1376, 1375, 84, 84, 115, 115, 30, 30, 30, 30, 48, 4
8, 48, 48, 92, 92, 92, 65, 150, 110, 138, 92, 92, 92, 92, 115, 115, 115, 115, 300, 3
00, 300, 300]
```

# dataset Colour options

This section of the notebook explores if changing the colour scheme of the image reveals the details in the simulated images

Before going any further, recall that OpenCV is unable to show the details on a black screen when reading the image in as Colour:

```
sample directory 1 = "002 DataSetExploration"
In [26]:
         sample directory 2 = "SimulatedSamples"
         sample_directory = sample_directory_1 + "\\" + sample directory 2
         try:
            mkdir( join( join(current directory, sample directory 1) , sample directory 2) )
         except FileExistsError:
            pass
         except:
            print("Unknown Error Encountered...")
In [27]: # copy 1 image to desired folder
         location = "Extracted\\Fluo-N2DH-SIM+\\Fluo-N2DH-SIM+\\01"
         file = "t000.tif"
         copy( join( join( join(current directory, data sets) , location) , file) , sample direct
         '002 DataSetExploration\\SimulatedSamples\\t000.tif'
Out[27]:
In [29]:
         # BGR
         path = walk(current directory + "\\002 DataSetExploration\\SimulatedSamples")
         img = cv2.imread( current directory + "\\002 DataSetExploration\\SimulatedSamples\\t000.
         cv2.imshow("BGR", img)
         cv2.waitKey(0)
Out[29]:
```

We recognize that OpenCV uses the BGR colour order, so perhaps inverting the colours is better?

```
In [30]: # RGB
path = walk(current_directory + "\\002_DataSetExploration\\SimulatedSamples")

img = cv2.imread( current_directory + "\\002_DataSetExploration\\SimulatedSamples\\t000.
img = img[ ... , ::-1 ] # invert array order, because based on Numpy array
cv2.imshow("RGB", img)
cv2.waitKey(0)
Out[30]:
```

Still nothing... What about different combinations of the colour channels?

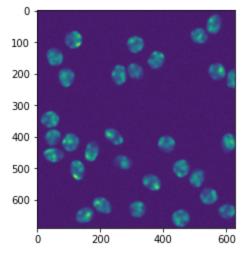
```
In [31]: # RBG
        path = walk(current directory + "\\002 DataSetExploration\\SimulatedSamples")
        bgr img = cv2.imread( current directory + "\\002 DataSetExploration\\SimulatedSamples\\t
        b, g, r = cv2.split(bgr img)
        rbg img = cv2.merge([r, b, g])
        cv2.imshow("RBG", rbg img)
         cv2.waitKey(0)
Out[31]:
        # GBR
In [32]:
        path = walk(current directory + "\\002 DataSetExploration\\SimulatedSamples")
        bgr img = cv2.imread( current directory + "\\002 DataSetExploration\\SimulatedSamples\\t
        b, g, r = cv2.split(bgr img)
        gbr img = cv2.merge([g, b, r])
        cv2.imshow("GBR", gbr img)
        cv2.waitKey(0)
Out[32]:
In [33]:
         # GRB
        path = walk(current directory + "\\002 DataSetExploration\\SimulatedSamples")
        bgr img = cv2.imread( current directory + "\\002 DataSetExploration\\SimulatedSamples\\t
        b, g, r = cv2.split(bgr img)
        grb img = cv2.merge([g, r, b])
         cv2.imshow("GRB", grb img)
         cv2.waitKey(0)
        -1
Out[33]:
         # BRG
In [34]:
         path = walk(current directory + "\\002 DataSetExploration\\SimulatedSamples")
        bgr img = cv2.imread( current directory + "\\002 DataSetExploration\\SimulatedSamples\\t
        b, g, r = cv2.split(bgr img)
        brg img = cv2.merge([b, r, g])
        cv2.imshow("BRG", brg img)
        cv2.waitKey(0)
        -1
Out[34]:
```

None of those combinations produce the hidden data... so OpenCV is not accessing the information present. NOW, Matploylib.pyplot is a different story:

If we read in the data and plot it via plt we see the following:

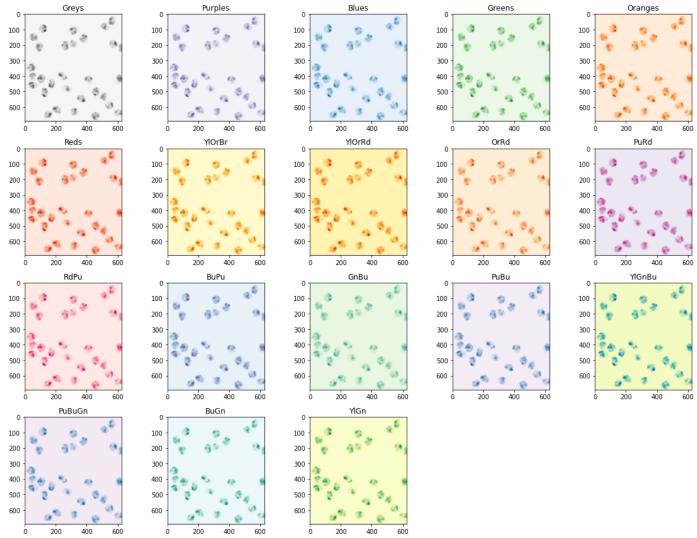
```
In [35]: path = walk(current_directory + "\\002_DataSetExploration\\SimulatedSamples")
    img = plt.imread( current_directory + "\\002_DataSetExploration\\SimulatedSamples\\t000.
    plt.imshow(img)

Out[35]: <matplotlib.image.AxesImage at 0x2d2b1106440>
```

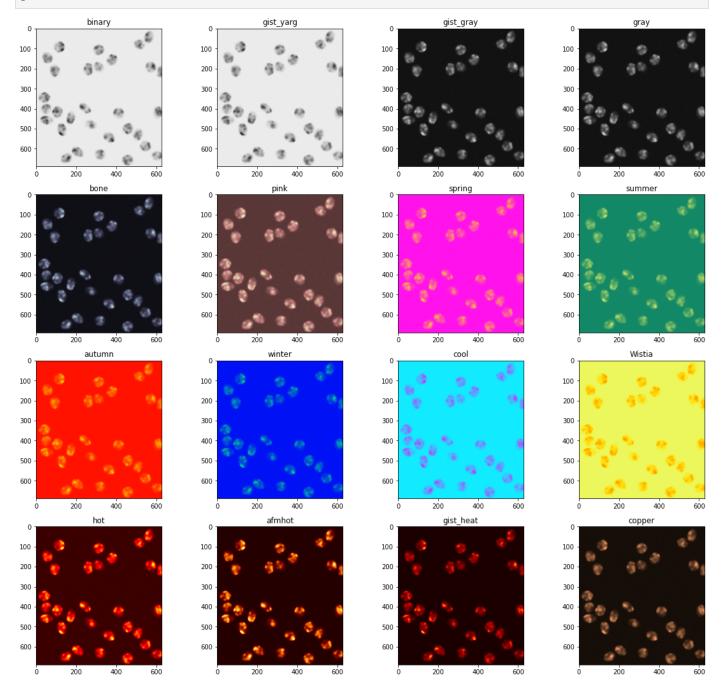


So the colour scheme is off... The default colour scheme for Pyplot is a Sequential colour 'Viridis'. Let us explore the other choices:

```
# Perceptually Uniform Sequential Colourmaps
In [36]:
          path = walk(current directory + "\\002 DataSetExploration\\SimulatedSamples")
          rgb_img = plt.imread( current_directory + "\\002_DataSetExploration\\SimulatedSamples\\t
          colour choices = ["viridis", "plasma", "inferno", "magma", "cividis"]
          fig = plt.figure(figsize=(14, 8))
          for i in range(5):
               fig.add subplot(2, 3, i+1)
               plt.title(colour choices[i])
               plt.imshow(rgb img, cmap=colour choices[i])
          plt.tight layout()
          plt.show()
                        viridis
                                                                                                    inferno
                                                              plasma
          100
                                                100
                                                                                       100
          200
                                                200
                                                                                       200
          300
                                                300
                                                                                       300
          400
                                                400
                                                                                       400
          500
                                                500
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          600
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                        300 400
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                                                              300 400
                100
                    200
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                                                   ò
                                                      100
                                                                      500
                                                                           600
                                                                                            100
                                                                                                     300
                                                                                                         400
                                                                                                             500
             Ò
                                                                                         Ò
                                                                                                 200
                                                              cividis
                       magma
           0
                                                  0
          100
                                                100
          200
                                                200
          300
                                                300
                                                400
          400
          500
                                                500
          600
                                                600
                100
                    200
                        300
                            400
                                                          200
                                                               300
                                                                  400
                                                                       500
```



```
plt.tight_layout()
plt.show()
```



The sequential colours are pretty, but not realistic... Let us explore the other options:

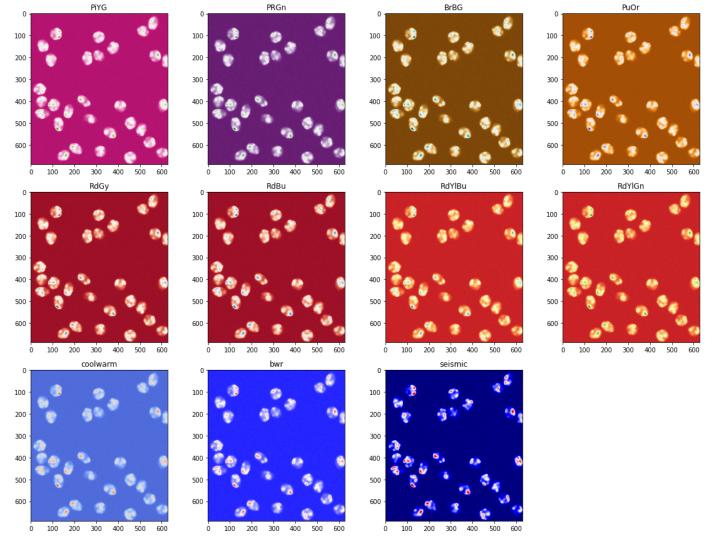
```
In [39]: # Diverging
   path = walk(current_directory + "\\002_DataSetExploration\\SimulatedSamples")

rgb_img = plt.imread( current_directory + "\\002_DataSetExploration\\SimulatedSamples\\t
colour_choices = ["PiYG", "PRGn", "BrBG", "PuOr", "RdGy", "RdBu", "RdYlBu", "RdYlGn", "c

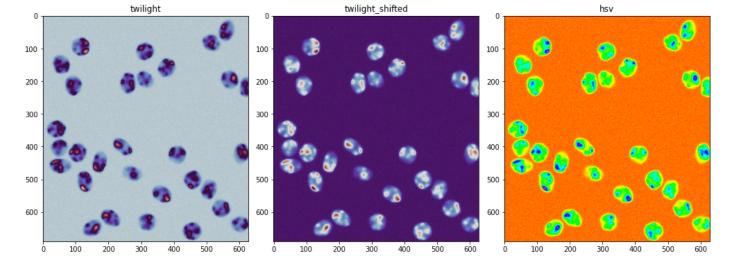
fig = plt.figure(figsize=(16, 12))

for i in range(len(colour_choices)):
    fig.add_subplot(3, 4, i+1)
    plt.title(colour_choices[i])
    plt.timshow(rgb_img, cmap=colour_choices[i])

plt.tight_layout()
plt.show()
```



```
In [40]: # Cyclic
  path = walk(current_directory + "\\002_DataSetExploration\\SimulatedSamples")
  rgb_img = plt.imread( current_directory + "\\002_DataSetExploration\\SimulatedSamples\\t colour_choices = ["twilight", "twilight_shifted", "hsv"]
  fig = plt.figure(figsize=(14, 8))
  for i in range(len(colour_choices)):
     fig.add_subplot(1, 3, i+1)
     plt.title(colour_choices[i])
     plt.imshow(rgb_img, cmap=colour_choices[i])
  plt.tight_layout()
  plt.show()
```

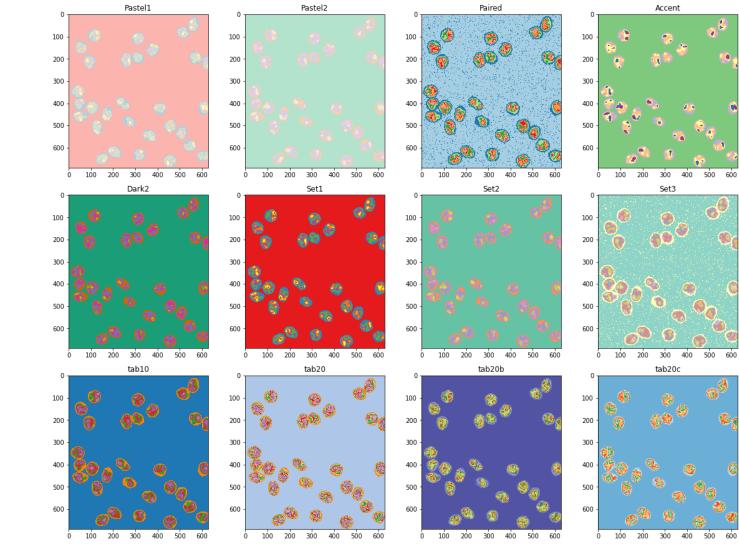


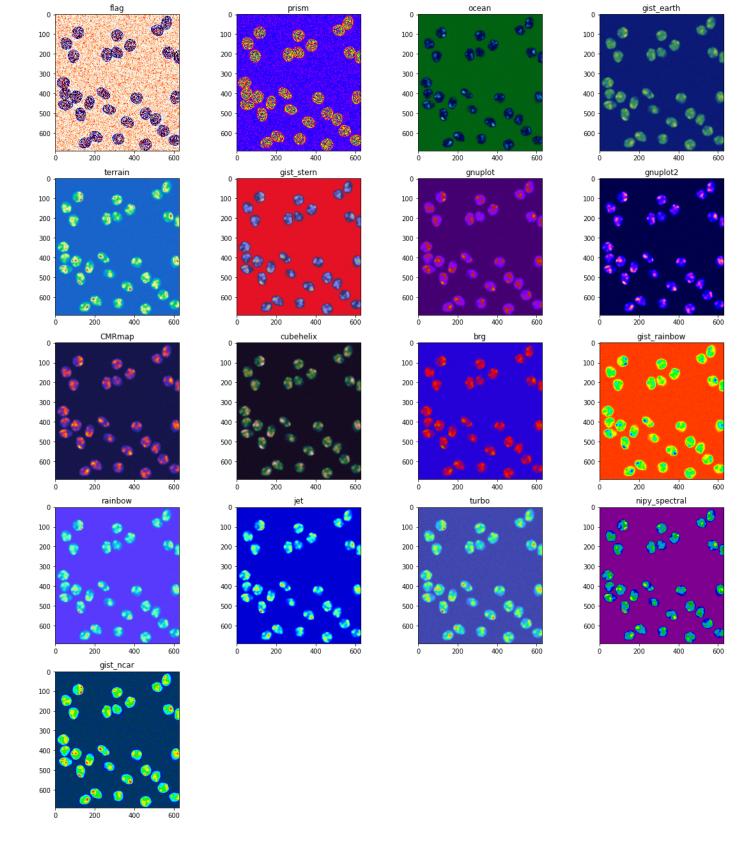
```
In [41]: # Qualitative
    path = walk(current_directory + "\\002_DataSetExploration\\SimulatedSamples")

    rgb_img = plt.imread( current_directory + "\\002_DataSetExploration\\SimulatedSamples\\t
    colour_choices = ["Pastel1", "Pastel2", "Paired", "Accent", "Dark2", "Set1", "Set2", "Se

fig = plt.figure(figsize=(16, 16))

for i in range(len(colour_choices)):
    fig.add_subplot(4, 4, i+1)
    plt.title(colour_choices[i])
    plt.tight_layout()
    plt.show()
```



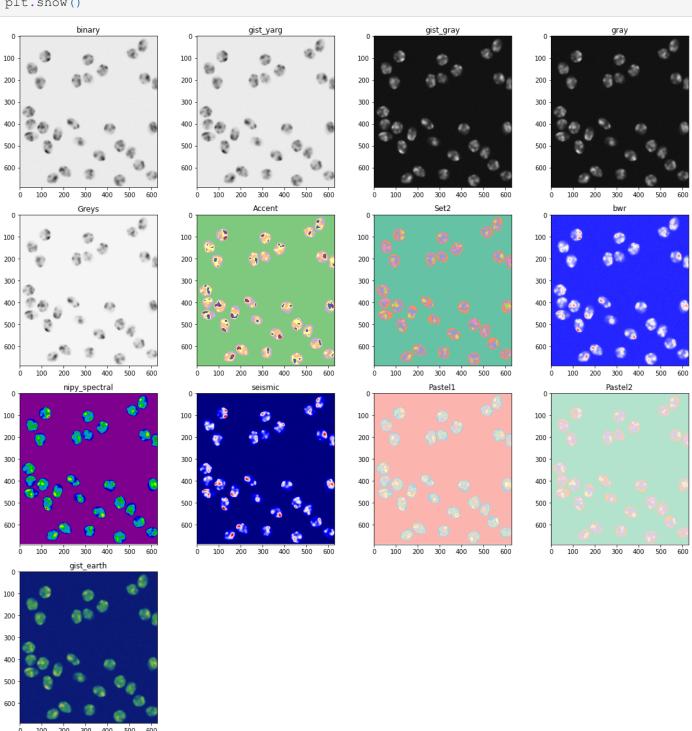


From the above pictures, we can see a lot of options, some of which capture more information than others! 'binary', 'gist\_yarg', 'gist\_gray', 'gray', 'Greys' are the obvious ones we can use.

However, some promising colours may include: 'Accent', 'Set2', 'bwr', 'nipy\_spectral', 'seismic', 'pastel1', 'pastel2', 'gist\_earth'

These combined choices are shown here:

```
rgb img = plt.imread( current directory + "\\002 DataSetExploration\\SimulatedSamples\\t
colour choices = ['binary', 'gist yarg', 'gist gray', 'gray', 'Greys', 'Accent', 'Set2',
                      'nipy spectral', 'seismic', 'Pastel1', 'Pastel2', 'gist earth']
fig = plt.figure(figsize=(16, 16))
for i in range(len(colour choices)):
    fig.add subplot(4, 4, i+1)
    plt.title(colour choices[i])
    plt.imshow(rgb img, cmap=colour choices[i])
plt.tight layout()
plt.show()
                                                            gist_gray
                                                                                      gray
100
                         100
200
                         200
                                                  200
                                                                           200
300
                         300
                                                  300
                                                                           300
                                                  400
600
                         600
                                                  600
                                                                           600
```



In another notebook, we can compare the colour choices for the collection of datasets - but for this notebook, we will use *gray* for the simulated colour images, for consistency

#### dataset Video Generation

This part of the notebook generates the videos we will use to understand the data

That seems to fix it! Let's try use that with the simulated folder of images

Upon closer experimentation, using the quantity of images in a directory as a frame rate results in a video that is super duper short in length! Instead, the author used a frame rate of 10, in order to more clearly see the journey of the cells.

Below is the code to generate the 40 videos showing the data. It may take 15 minutes to run

```
# only progress if files don't exist
In [44]:
         desired folder = "..\\..\\Comp700 VideosOfDataSets Colour"
         makeVideos = False
         if (exists(current directory + "\\" + desired folder)):
             # Now, go to directory and verify all is there
            path = walk(current directory + "\\" + desired folder)
            count = 0
            for root, dirs, files in path:
                 for item in files:
                    count += 1
             if (count == 40):
                print("All Videos exist already!")
                print("Not all Videos exist")
                makeVideos = True
         else:
            makeVideos = True
         if (makeVideos):
            path = walk(current directory + "\\" + data sets) # reset path
             sub directory choice = ["01", "02"]
             output video = cv2.VideoWriter()
            frames per second = 10
            petri dish images = False
             # Generates Colour Videos
             for root, dirs, files in path:
                 for item in files:
                     if ("man " not in item) and (".zip" not in item):
                         # update on first element only
                         if ("t0000.tif" == item) or ("t000.tif" == item):
                             petri dish images = True
                             i += 1
                             index = i // 2 # used for output video as 2 copies for each director
                             size = (image size array[i][1], image size array[i][0] ) # notice or
                             fileName = "Color " + directory array[index] + " " + sub directory c
                             output video = cv2.VideoWriter(
                                 fileName,
                                 cv2.VideoWriter fourcc(*'DIVX'),
                                 frames per second,
```

```
size
            )
        # used to update the pictures for the Simulated Videos
        if ("+" in directory array[index]):
            img = plt.imread( location array[i] + item ) # grayscale
            plt.imsave("temp.jpg", img, cmap='gray')
            img = cv2.imread( "temp.jpg")
        else:
            # Colour Video
            img = cv2.imread( (location array[i] + item) )
        output video.write(img)
    else:
        petri_dish images = False
        break
if (petri dish images):
   cv2.destroyAllWindows()
    output video.release()
    print("Video finished for ", fileName, sep="")
    petri dish images = False # update incase next iteration containes empty arr
```

All Videos exist already!

Now, remove the final temporary picture

```
In [45]: # from os import remove
if (exists("temp.jpg")):
    remove("temp.jpg")
```

ouput\_video.release() saves the contents of the files into the current directory, so the next block of code moves them to our a seperate folder

```
In [46]: # from os.path import join
         # from shutil import move # moves and replaces files
         # only progress if files don't exist
         desired folder = "..\\..\\Comp700 VideosOfDataSets Colour"
         if (exists(current directory + "\\" + desired folder)):
            print("Videos already exist!")
         else:
             try:
                 mkdir(desired folder)
             except FileExistsError:
                 pass
             except:
                 print("Unknown Error Encountered...")
             path = walk(current directory)
             # count = 0
             for root, dirs, files in path:
                 for item in files:
                     if (".mp4" in item):
                         # count += 1
                         origin path = current directory
                         new destination = current directory + "\\" + desired folder
                         move(join(current directory, item), join(new destination, item)) # shoul
             # Now, go to directory and verify all is there
             path = walk(current directory + "\\" + desired folder)
```

```
count = 0
for root, dirs, files in path:
    for item in files:
        count += 1

if (count == 40):
    print("All Videos Moved Successfully!")

else:
    print("Not all Videos Moves Successfully")
```

Videos already exist!

The simulated videos have a bit of noise. This probably comes from the Conversion between plt and cv2. To try rememdy this in the future, let us explore if we can create grayscale videos:

Here is the same cell block as above, but generates Grayscale Videos

```
# only progress if files don't exist
In [47]:
         desired folder = "..\\..\\Comp700 VideosOfDataSets Grayscale"
        makeVideos = False
         if (exists(current directory + "\\" + desired folder)):
             # Now, go to directory and verify all is there
            path = walk(current directory + "\\" + desired folder)
            count = 0
            for root, dirs, files in path:
                for item in files:
                     count += 1
            if (count == 40):
                print("All Videos already exist!")
             else:
                print("Not all Videos exist")
                makeVideos = True
         else:
            makeVideos = True
         if (makeVideos):
            path = walk(current directory + "\\" + data sets) # reset path
            sub_directory_choice = ["01", "02"]
             output video = cv2. VideoWriter() # needed for compiler to process loop below, update
            frames per second = 10
            petri dish images = False
             # Grayscale Videos
             for root, dirs, files in path:
                 for item in files:
                     if ("man " not in item) and (".zip" not in item):
                         # update on first element only
                         if ("t0000.tif" == item) or ("t000.tif" == item):
                             petri dish images = True
                             index = i // 2 # used for output video as 2 copies for each director
                             size = (image size array[i][1], image size array[i][0] ) # notice or
                             fileName = "Grayscale " + directory array[index] + " " + sub directo
                             # isColor below is what we use to create grayscale videos!
                             output video = cv2.VideoWriter(
                                 fileName,
```

```
cv2.VideoWriter fourcc(*'DIVX'),
                frames per second,
                size,
                isColor=False
        # used to update the pictures for the Simulated Videos
        # write to disk as grayscale, then read in again
        if ("+" in directory array[index]):
            img = plt.imread( location array[i] + item ) # grayscale
            plt.imsave("temp.jpg", img, cmap="gray")
            img = cv2.imread( "temp.jpg", cv2.IMREAD GRAYSCALE)
        else:
            img = cv2.imread( (location array[i] + item), cv2.IMREAD GRAYSCALE)
        output video.write(img)
    else:
       petri dish images = False
if (petri dish images):
   cv2.destroyAllWindows()
    output video.release()
    print("Video finished for ", fileName, sep="")
   petri dish images = False # update incase next iteration containes empty arr
```

All Videos already exist!

Now, let us move to a grayscale video folder:

```
In [48]: # from os.path import join
         # from shutil import move # moves and replaces files
         # only progress if files don't exist
         desired folder = "...\\...\Comp700 VideosOfDataSets Grayscale"
         if (exists(current directory + "\\" + desired folder)):
             print("Videos already exist!")
         else:
             try:
                 mkdir(desired folder)
             except FileExistsError:
                pass
             except:
                 print("Unknown Error Encountered...")
            path = walk(current directory)
             \# count = 0
             for root, dirs, files in path:
                for item in files:
                     if (".mp4" in item):
                         # count += 1
                         origin path = current directory
                         new destination = current directory + "\\" + desired folder
                         move(join(current directory, item), join(new destination, item)) # shoul
             # Now, go to directory and verify all is there
             path = walk(current directory + "\\" + desired_folder)
             count = 0
             for root, dirs, files in path:
```

Videos already exist!

Now, remove the final temporary picture

```
In [49]: # from os import remove
if (exists("temp.jpg")):
    remove("temp.jpg")
```

The grayscale videos successfully generate and the noise present in the simulated videos goes away! Fantastic! We may need to incorporate these findings later on

#### **Conclusions**

This section of the notebook summarises the findings so far

The dataset has 2 folders, one contains the original zipped data, and the other the extracted data.

The extracted data is broken up into 2 sets: Training Data, and Challenge Data. The Challenge Data is indicated with a (1) in the folder name.

The distinction between the Training Data and the Challenge Data is that the Training Data contains additional folders of manual segmentation and manual tracking of the images. This will be useful later on when training the models.

To assist with understanding of the data, the author has created a Matplotlib plot showing 10 samples of the images from the 10 data sets (Training Data)

The author has also generated Videos of each folder, in both colour and grayscale, to show the movement of the cells over time.

Much later on, we will create these files in a similar way in order to show tracking and segmentation!

The most significant findings from the work is: The pictures may need to be read in via Matplotlib, and saved onto disk in order for OpenCV to register the information in the image.

The author is not entirely sure why this is necessary, but it may be the case that the images are saved in a format that OpenCV struggles to process. Whatever the reason is though, one of the processing tips may be to do a bulk save of all images via Matplotlib first!

Thank You!