

# Mice Data Analysis

## Importing the Data

```
fetus_full <- read.csv("data/fetus-cleaned.csv", header=T)

fetus <- fetus_full %>%
  filter(Fetus_genotype != "resorp") %>%
  na.omit() # One empty row. Not sure why...
```

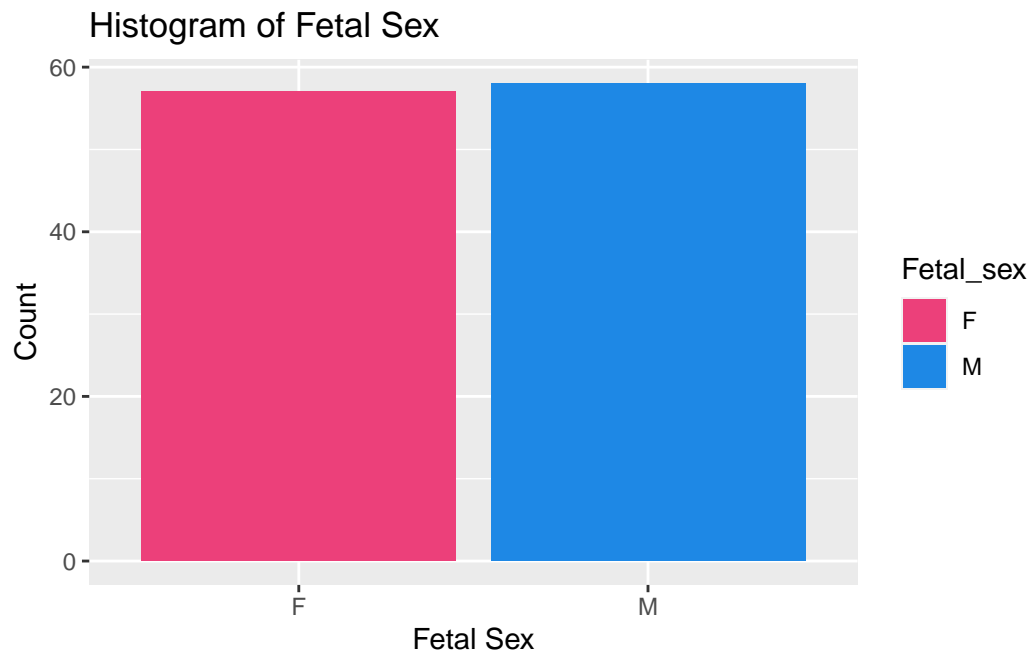
We drop all resorp fetuses. This is because they do not provide information. Of course, the data are not missing at random in this case. However, we can conduct an entirely separate analysis of resorp vs living fetuses later on if that's required.

## Assessing Normality of Features and Dependent Variables

First we assess the proportion of male v. female fetuses to make sure our data are not skewed.

```
# Define colors for each sex
color_female <- "#ec407a" # pink
color_male <- "#1e88e5"   # blue

ggplot(fetus, aes(x = Fetal_sex, fill = Fetal_sex)) +
  geom_bar() +
  scale_fill_manual(values = c(color_female, color_male)) +
  xlab("Fetal Sex") +
  ylab("Count") +
  ggtitle("Histogram of Fetal Sex")
```



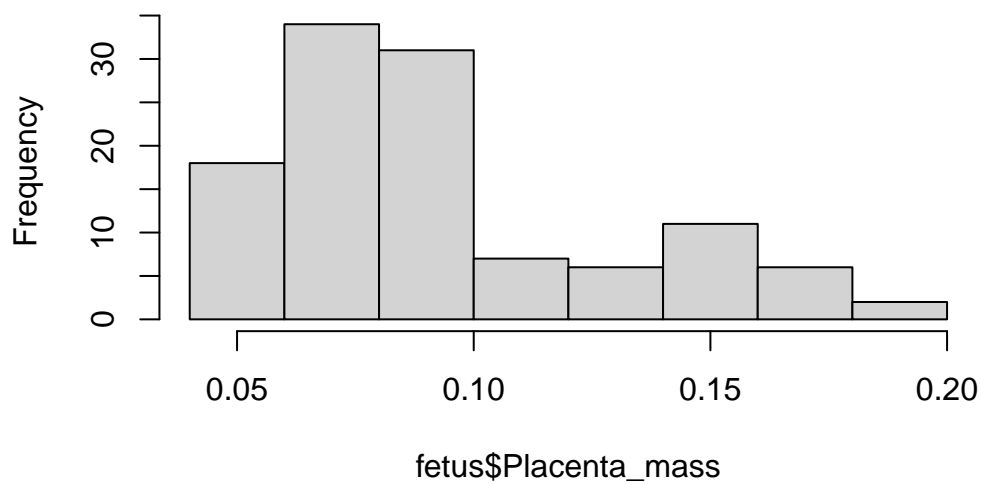
```
mean(fetus$isFemale)
```

```
[1] 0.4956522
```

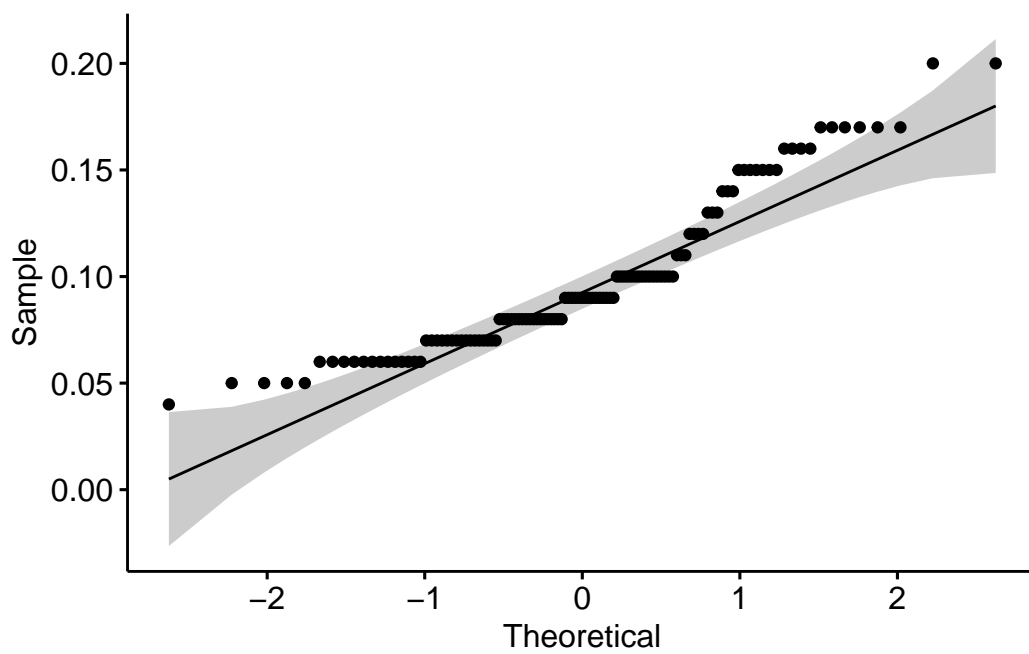
Approximately 50% of surviving fetuses are female, so no worries about composition of the data.

```
hist(fetus$Placenta_mass)
```

**Histogram of fetus\$Placenta\_mass**



```
ggqqplot(fetus$Placenta_mass)
```



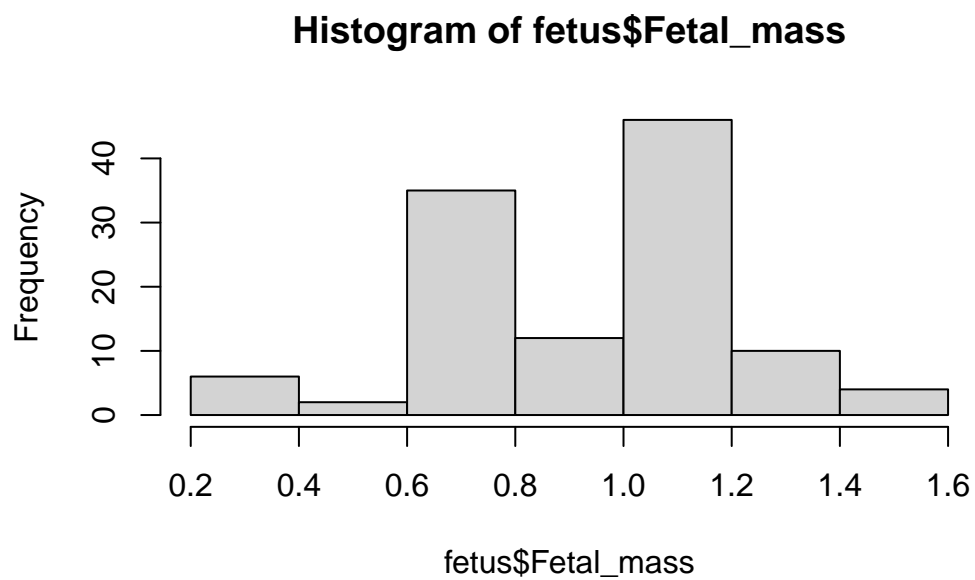
```
shapiro.test(fetus$Placenta_mass)
```

Shapiro-Wilk normality test

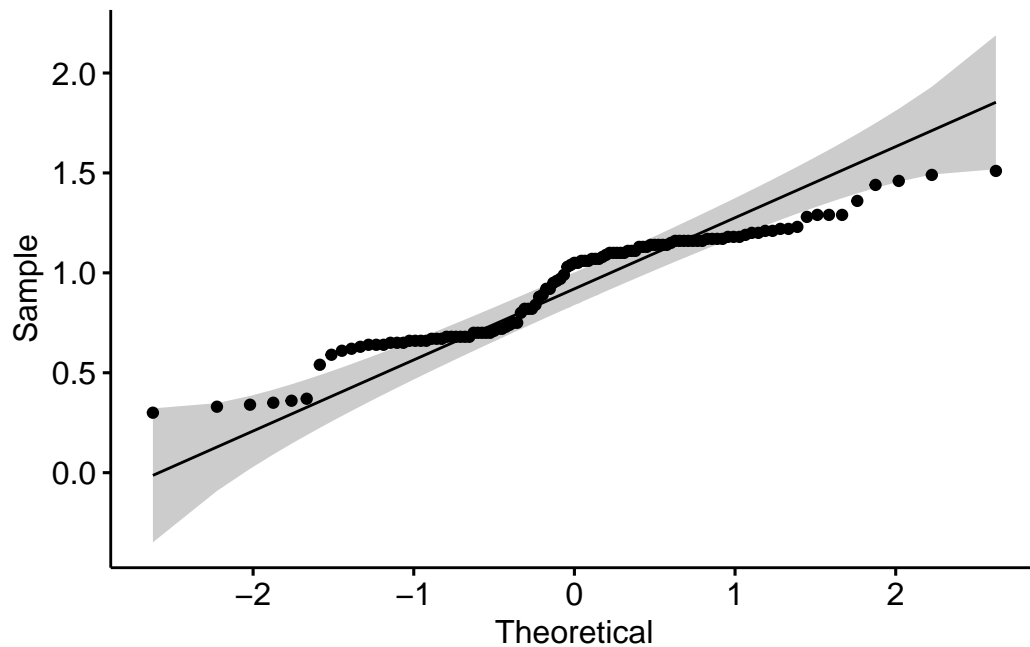
```
data: fetus$Placenta_mass  
W = 0.90156, p-value = 3.798e-07
```

Placenta mass is not normally distributed (right-skewed)

```
hist(fetus$Fetal_mass)
```



```
ggqqplot(fetus$Fetal_mass)
```



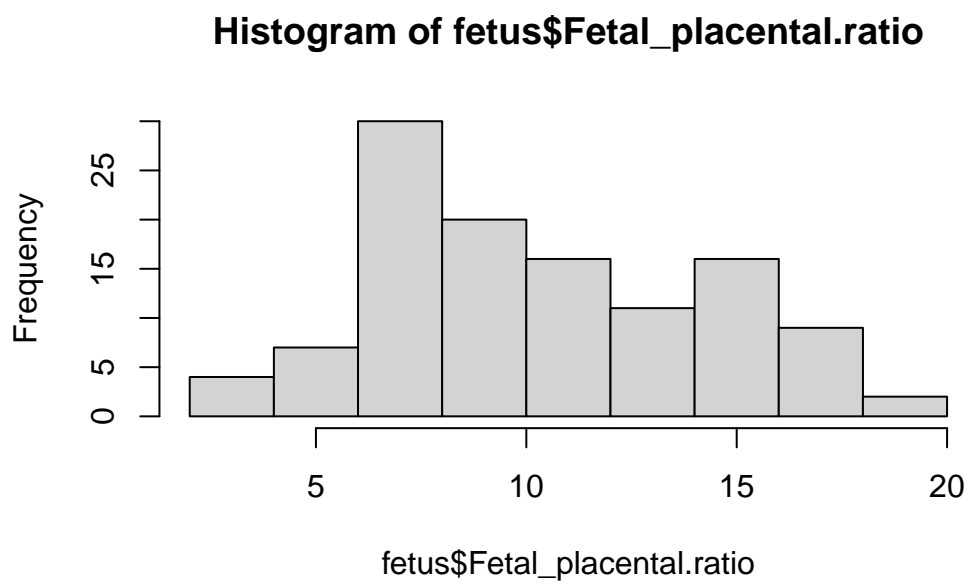
```
shapiro.test(fetus$Fetal_mass)
```

Shapiro-Wilk normality test

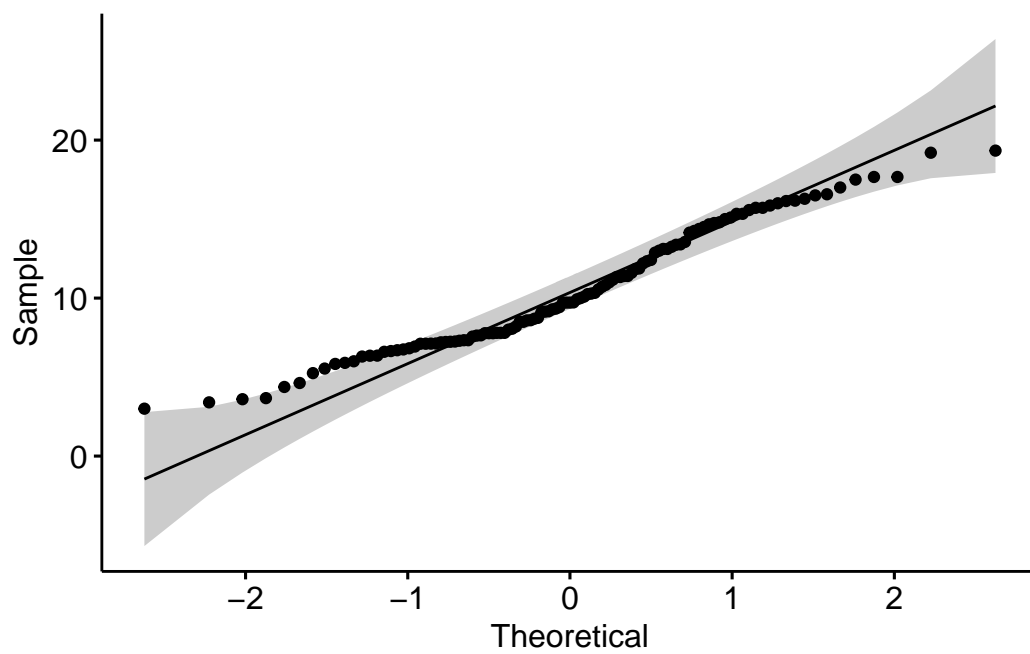
```
data: fetus$Fetal_mass  
W = 0.93942, p-value = 5.661e-05
```

Fetal mass not normally distributed (bimodal). Perhaps bimodality has to do with the sex of the fetus? Worth investigating because, if not, could be related to the genotype.

```
hist(fetus$Fetal_placental_ratio)
```



```
ggqqplot(fetus$Fetal_placental.ratio)
```



```
shapiro.test(fetus$Fetal_placental_ratio)
```

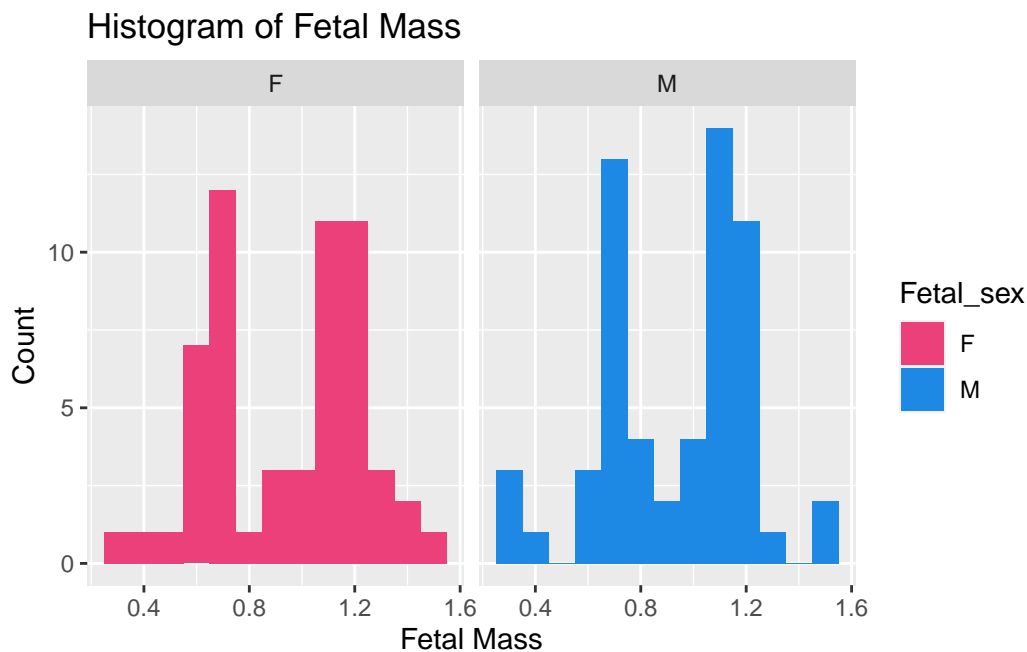
Shapiro-Wilk normality test

```
data: fetus$Fetal_placental_ratio  
W = 0.96431, p-value = 0.003709
```

Fetal/placental ratio is not normally distributed (right skew). This makes sense since it's a transformation that is not a sum of two non-normal random variables.

Let's return to assessing fetal mass. Maybe it is related to gender?

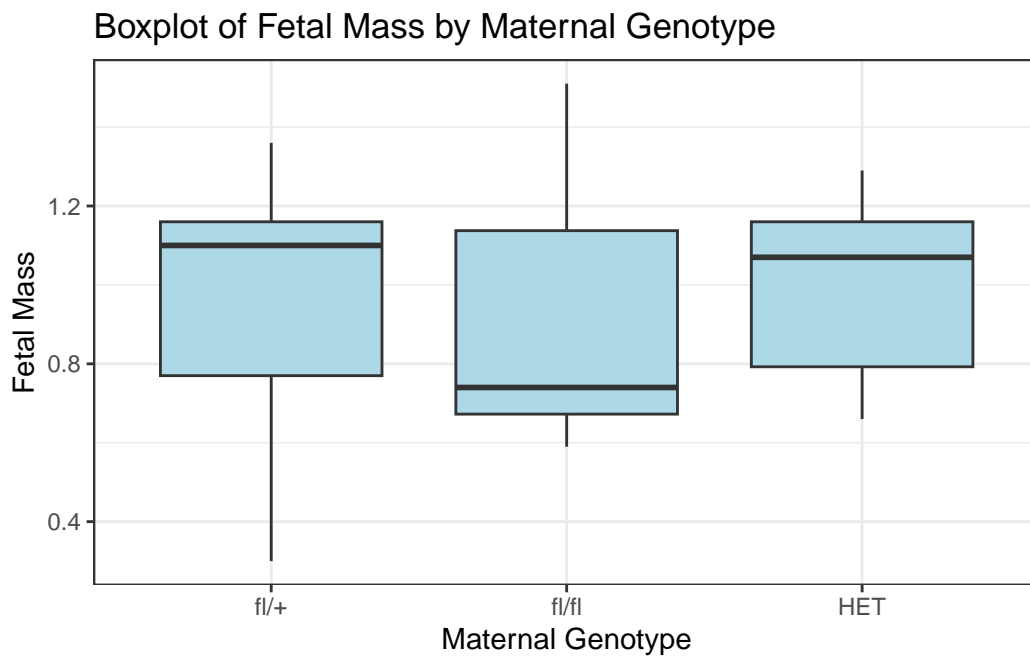
```
# Plot side by side histograms of fetal mass faceted by fetal sex with different colors  
ggplot(fetus, aes(x = Fetal_mass, fill = Fetal_sex)) +  
  geom_histogram(binwidth = 0.1) +  
  scale_fill_manual(values = c(color_female, color_male)) +  
  xlab("Fetal Mass") +  
  ylab("Count") +  
  ggtitle("Histogram of Fetal Mass") +  
  facet_wrap(~ Fetal_sex, ncol = 2)
```



This did not help the bimodality. There is more going on here. Maybe it is related to the genotypes?

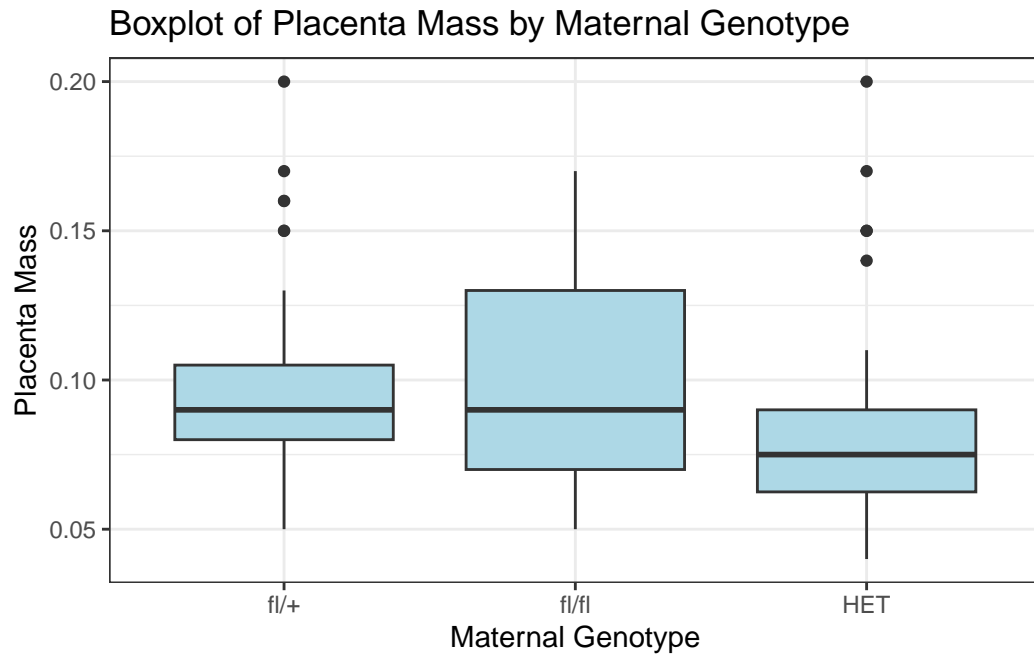
Now we visually assess the relationship between our variables of interest and the genotypes using boxplots.

```
# Boxplot of Fetal_mass by Maternal_genotype
ggplot(fetus, aes(x = Maternal_genotype, y = Fetal_mass)) +
  geom_boxplot(fill = "lightblue") +
  xlab("Maternal Genotype") +
  ylab("Fetal Mass") +
  ggtitle("Boxplot of Fetal Mass by Maternal Genotype") +
  theme_bw()
```

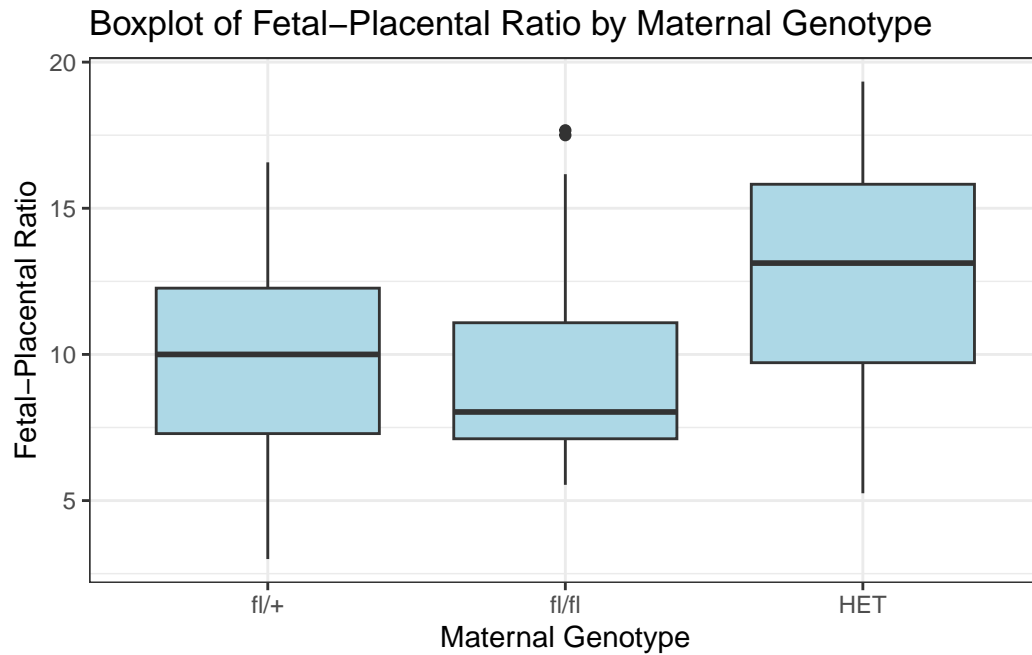


```
# Boxplot of Placenta_mass by Maternal_genotype
ggplot(fetus, aes(x = Maternal_genotype, y = Placenta_mass)) +
  geom_boxplot(fill = "lightblue") +
  xlab("Maternal Genotype") +
  ylab("Placenta Mass") +
  ggtitle("Boxplot of Placenta Mass by Maternal Genotype") +
  theme_bw()
```

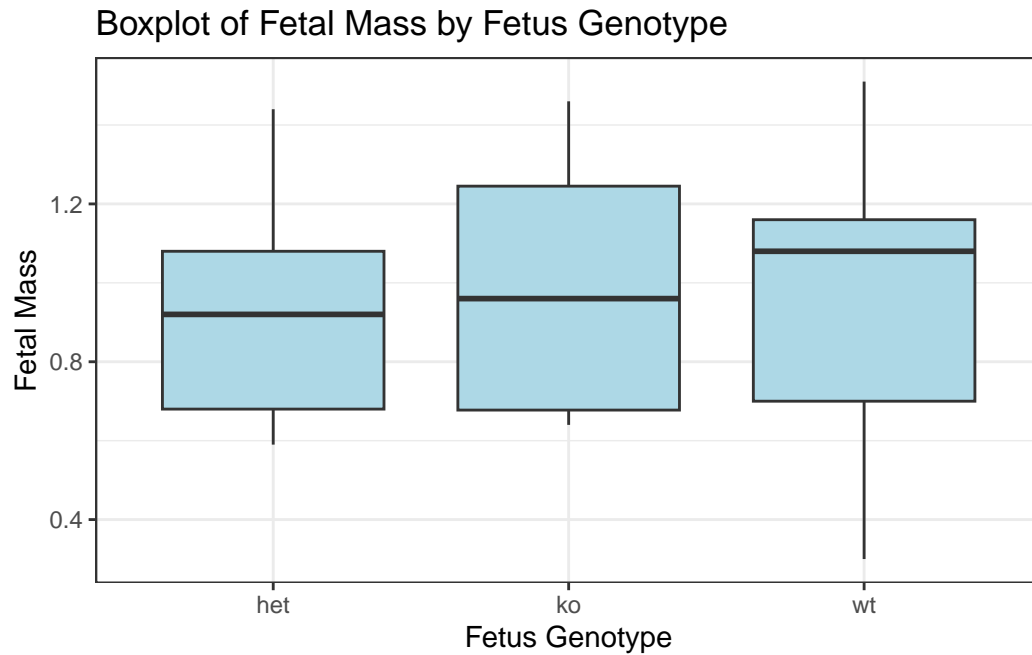




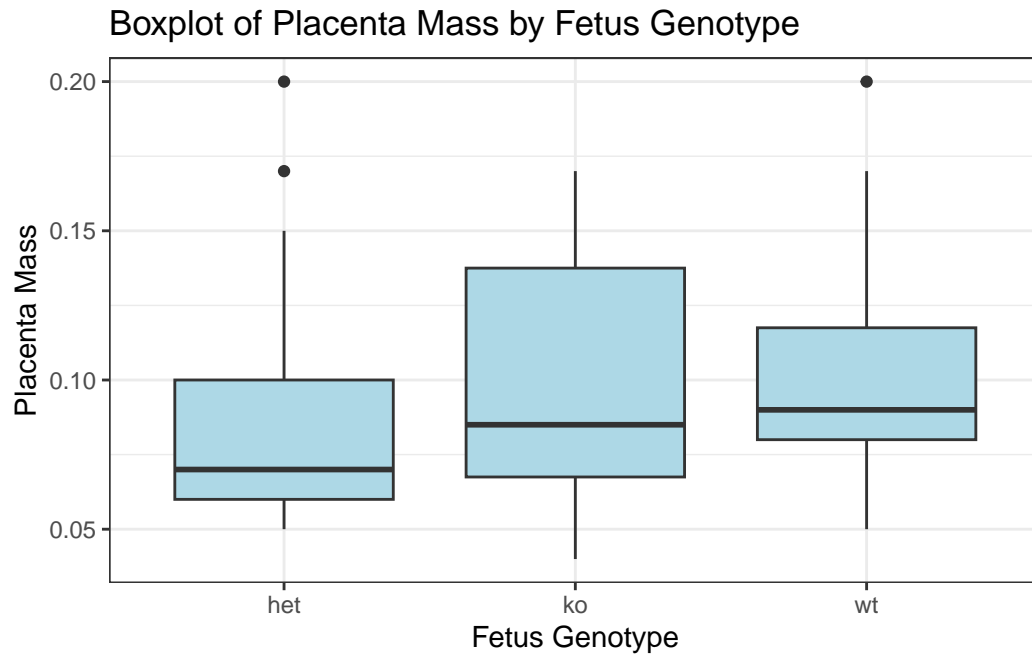
```
# Boxplot of Fetal_placental.ratio by Maternal_genotype
ggplot(fetus, aes(x = Maternal_genotype, y = Fetal_placental.ratio)) +
  geom_boxplot(fill = "lightblue") +
  xlab("Maternal Genotype") +
  ylab("Fetal-Placental Ratio") +
  ggtitle("Boxplot of Fetal-Placental Ratio by Maternal Genotype") +
  theme_bw()
```



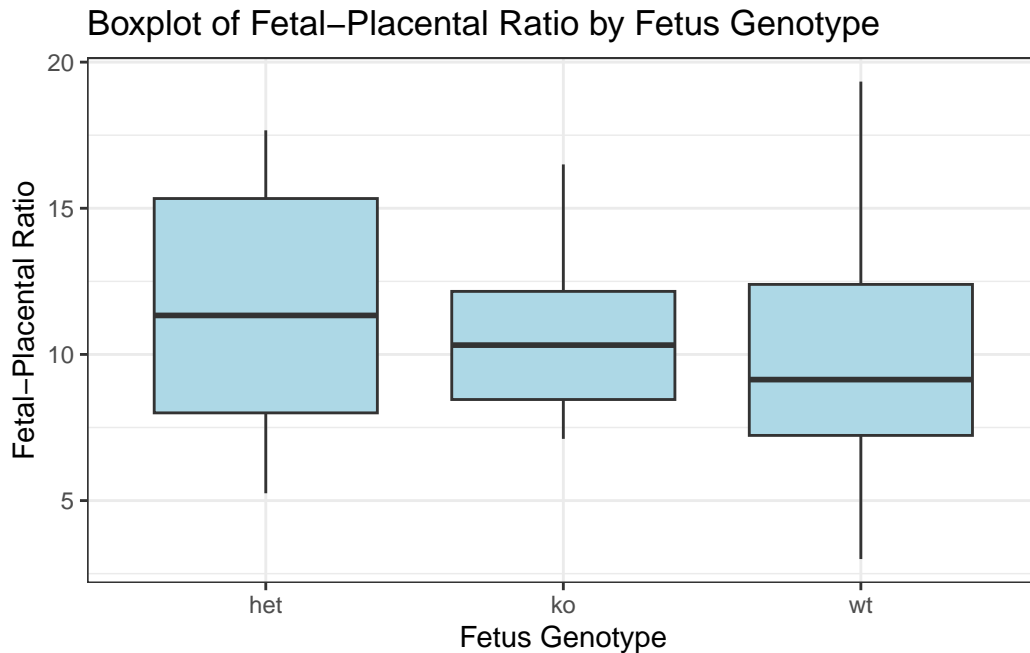
```
# Boxplot of Fetal_mass by Fetus_genotype
ggplot(fetus, aes(x = Fetus_genotype, y = Fetal_mass)) +
  geom_boxplot(fill = "lightblue") +
  xlab("Fetus Genotype") +
  ylab("Fetal Mass") +
  ggtitle("Boxplot of Fetal Mass by Fetus Genotype") +
  theme_bw()
```



```
# Boxplot of Placenta_mass by Fetus_genotype
ggplot(fetus, aes(x = Fetus_genotype, y = Placenta_mass)) +
  geom_boxplot(fill = "lightblue") +
  xlab("Fetus Genotype") +
  ylab("Placenta Mass") +
  ggtitle("Boxplot of Placenta Mass by Fetus Genotype") +
  theme_bw()
```



```
# Boxplot of Fetal_placental.ratio by Fetus_genotype
ggplot(fetus, aes(x = Fetus_genotype, y = Fetal_placental.ratio)) +
  geom_boxplot(fill = "lightblue") +
  xlab("Fetus Genotype") +
  ylab("Fetal-Placental Ratio") +
  ggtitle("Boxplot of Fetal-Placental Ratio by Fetus Genotype") +
  theme_bw()
```



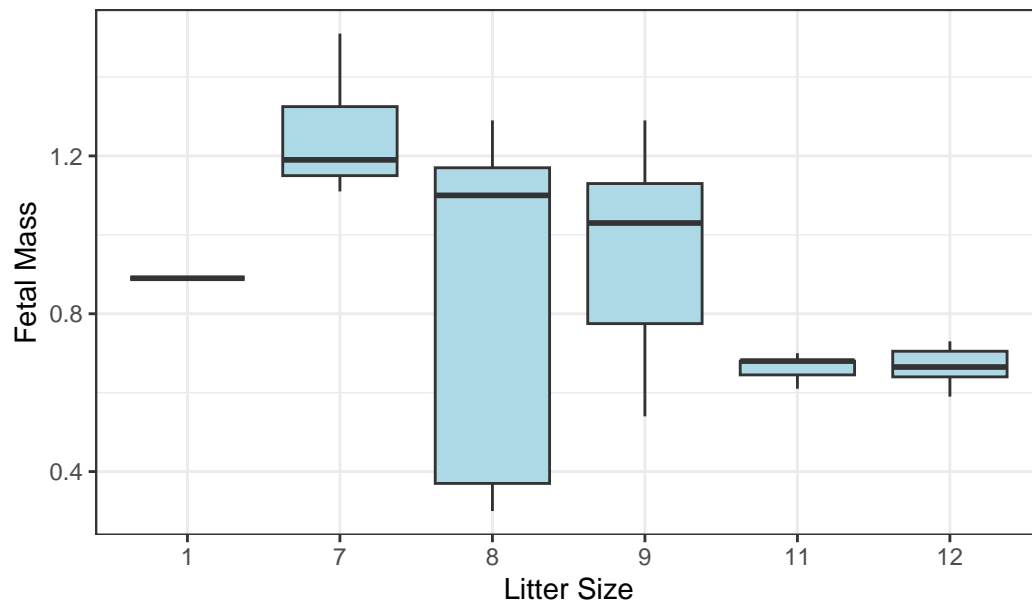
In all cases, it looks like the maternal/fetal genotype has no impact on our dependent variables. In all honesty, it's likely that masses have much more to do with the mass of the mother/father rats since it is genetic. It also might have to do with the day of conception. Presumably, if a fetus is a few days older, it will weigh more on average. I have limited subject expertise though, so I am not sure.

This could be assessed by linking the mother dataset to the fetus dataset, but there is no joining variable (dam id missing from mother dataset?)

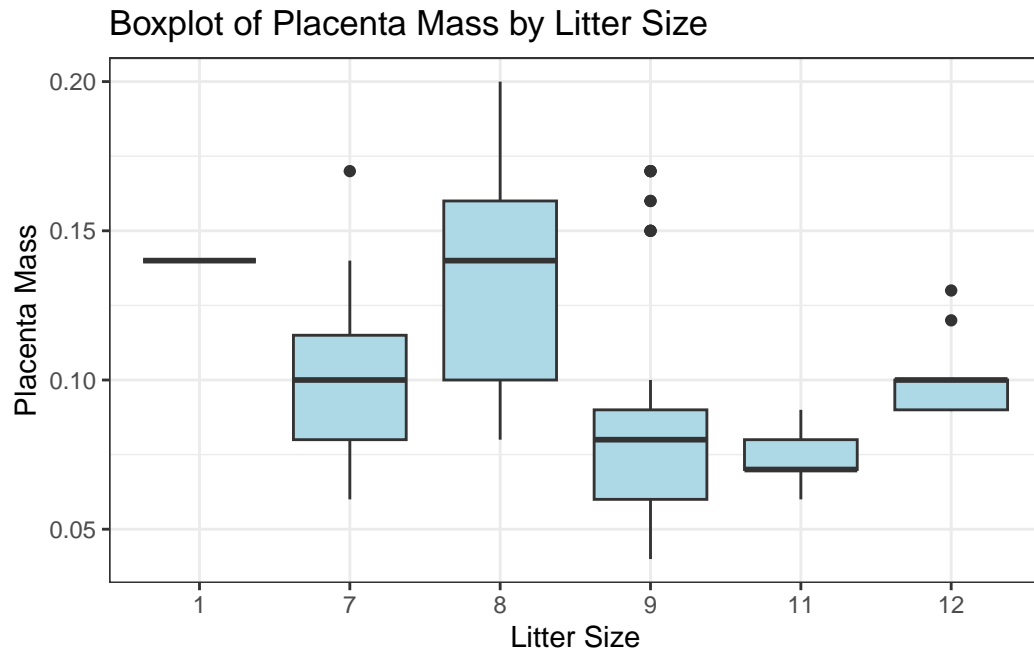
Let's look at the last variables of interest for modeling, Litter\_size and Fetal\_sex.

```
# Boxplot of Fetal_mass by Litter_size
ggplot(fetus, aes(x = as.factor(Litter_size), y = Fetal_mass)) +
  geom_boxplot(fill = "lightblue") +
  xlab("Litter Size") +
  ylab("Fetal Mass") +
  ggtitle("Boxplot of Fetal Mass by Litter Size") +
  theme_bw()
```

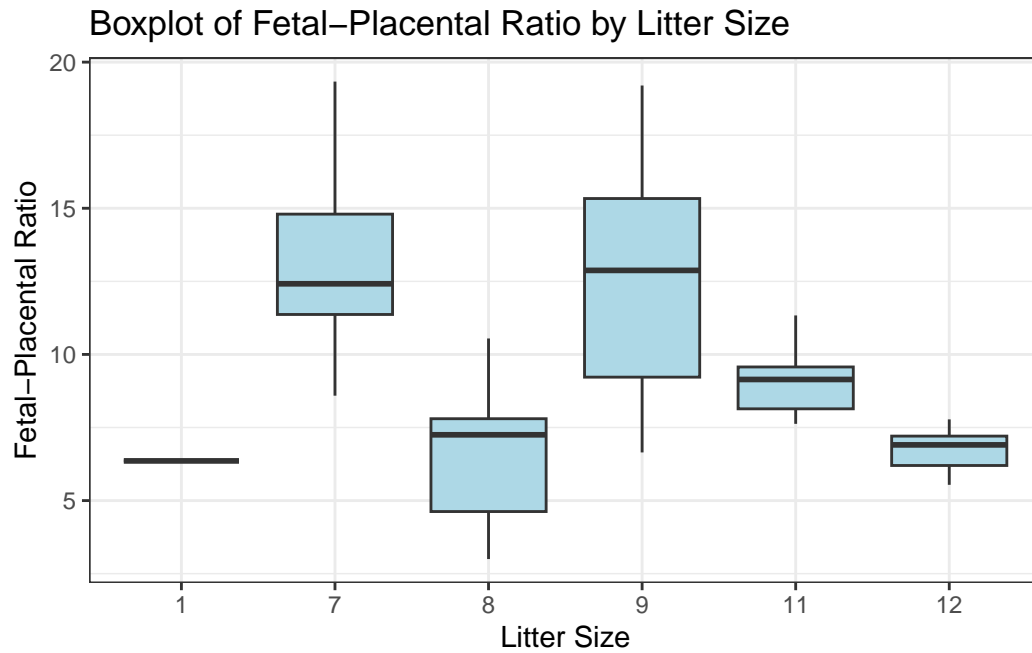
Boxplot of Fetal Mass by Litter Size



```
# Boxplot of Placenta_mass by Litter_size
ggplot(fetus, aes(x = as.factor(Litter_size), y = Placenta_mass)) +
  geom_boxplot(fill = "lightblue") +
  xlab("Litter Size") +
  ylab("Placenta Mass") +
  ggtitle("Boxplot of Placenta Mass by Litter Size") +
  theme_bw()
```

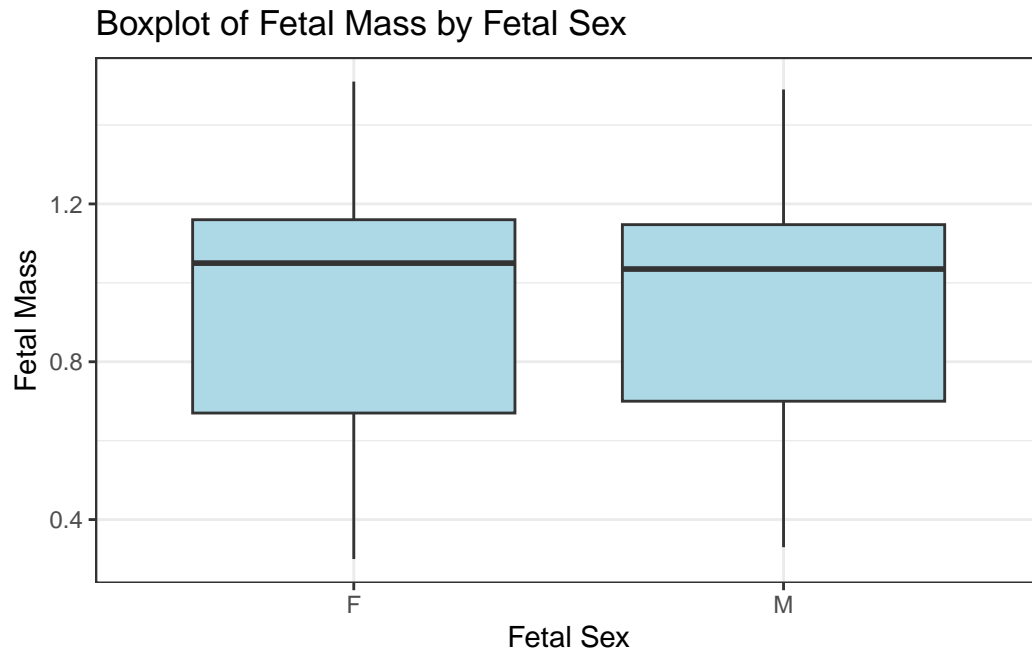


```
# Boxplot of Fetal_placental_ratio by Litter_size
ggplot(fetus, aes(x = as.factor(Litter_size), y = Fetal_placental_ratio)) +
  geom_boxplot(fill = "lightblue") +
  xlab("Litter Size") +
  ylab("Fetal-Placental Ratio") +
  ggtitle("Boxplot of Fetal-Placental Ratio by Litter Size") +
  theme_bw()
```

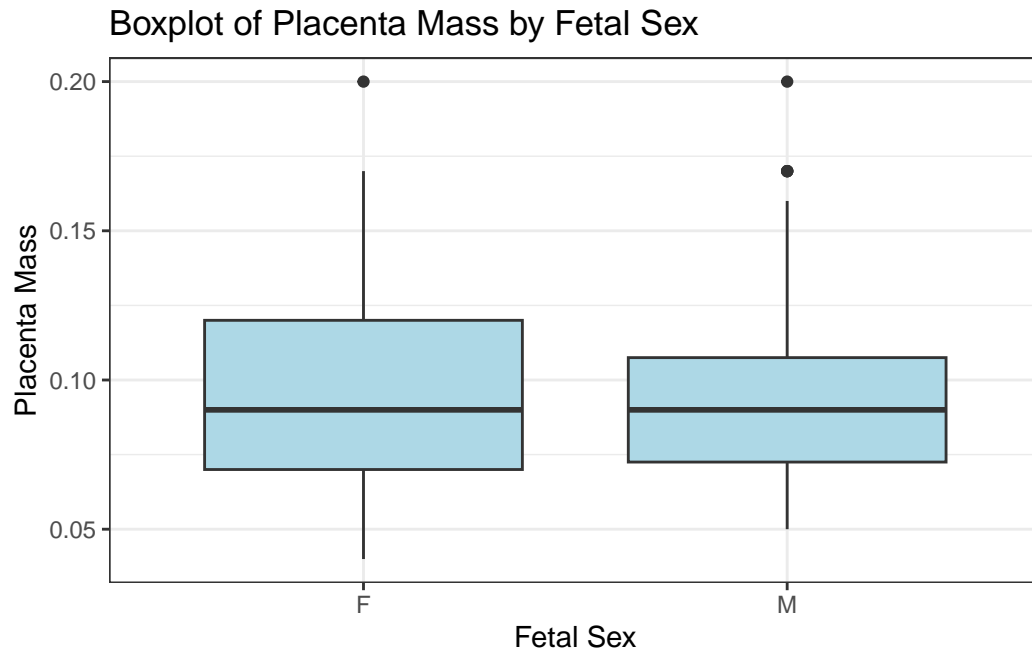


```
# Boxplot of Fetal_mass by Fetal_sex
ggplot(fetus, aes(x = Fetal_sex, y = Fetal_mass)) +
  geom_boxplot(fill = "lightblue") +
  xlab("Fetal Sex") +
  ylab("Fetal Mass") +
  ggtitle("Boxplot of Fetal Mass by Fetal Sex") +
  theme_bw()
```

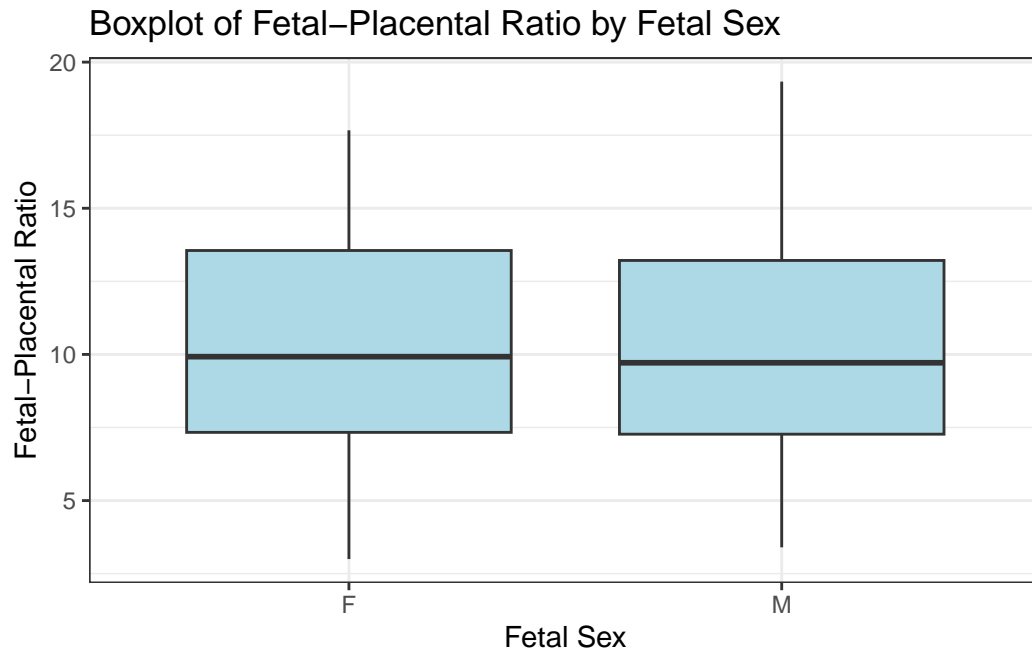




```
# Boxplot of Placenta_mass by Fetal_sex
ggplot(fetus, aes(x = Fetal_sex, y = Placenta_mass)) +
  geom_boxplot(fill = "lightblue") +
  xlab("Fetal Sex") +
  ylab("Placenta Mass") +
  ggtitle("Boxplot of Placenta Mass by Fetal Sex") +
  theme_bw()
```



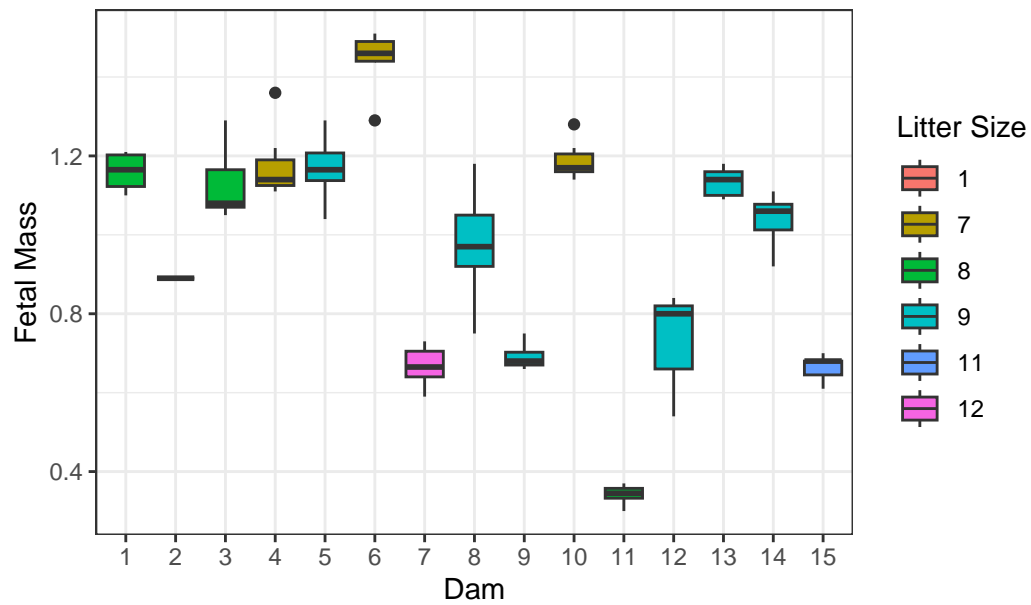
```
# Boxplot of Fetal_placental_ratio by Fetal_sex
ggplot(fetus, aes(x = Fetal_sex, y = Fetal_placental_ratio)) +
  geom_boxplot(fill = "lightblue") +
  xlab("Fetal Sex") +
  ylab("Fetal-Placental Ratio") +
  ggtitle("Boxplot of Fetal-Placental Ratio by Fetal Sex") +
  theme_bw()
```



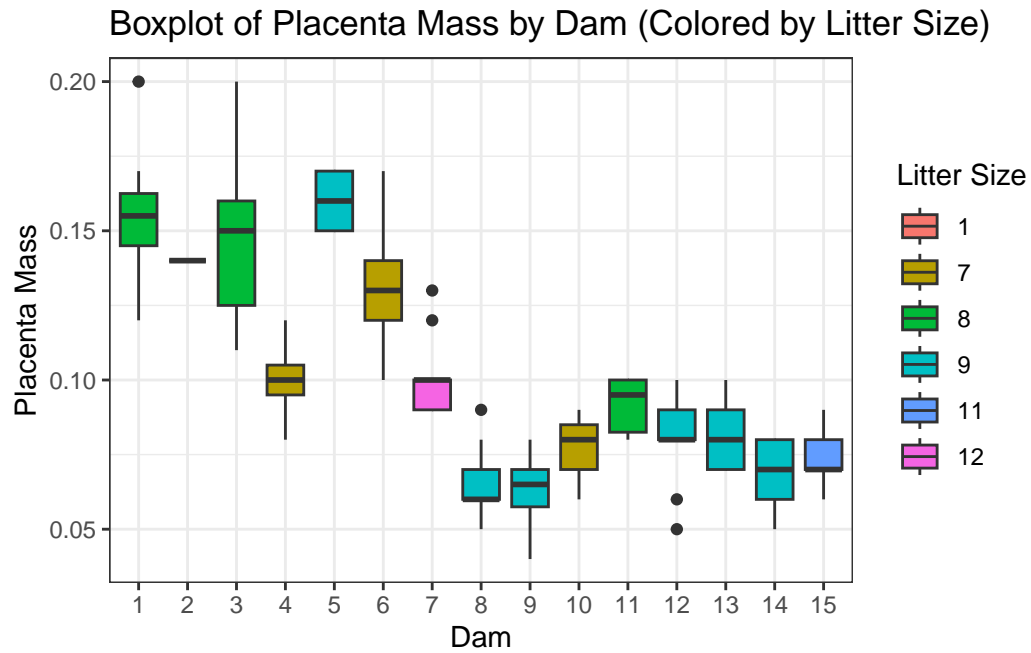
Litter size could have an effect. But, what is far more likely, is that litter size captures the variance of Dam.

```
# Boxplot of Fetal_mass by Dam with box color by Litter_size
ggplot(fetus, aes(x = as.factor(Dam), y = Fetal_mass, fill = as.factor(Litter_size))) +
  geom_boxplot() +
  xlab("Dam") +
  ylab("Fetal Mass") +
  labs(fill = "Litter Size") +
  ggtitle("Boxplot of Fetal Mass by Dam (Colored by Litter Size)") +
  theme_bw()
```

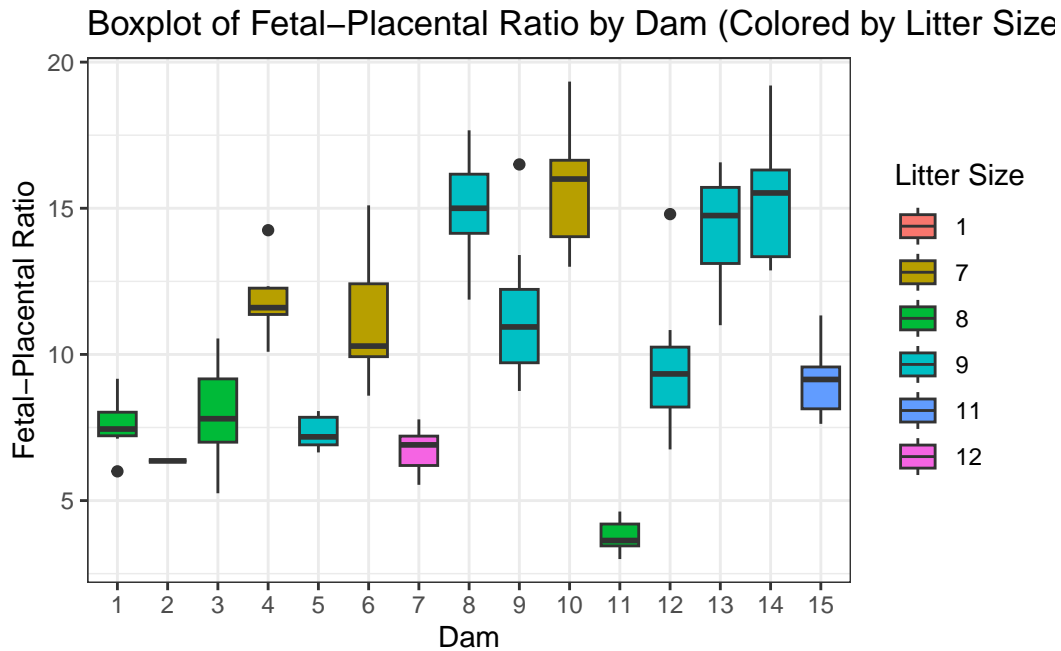
Boxplot of Fetal Mass by Dam (Colored by Litter Size)



```
# Boxplot of Placenta_mass by Dam with box color by Litter_size
ggplot(fetus, aes(x = as.factor(Dam), y = Placenta_mass, fill = as.factor(Litter_size))) +
  geom_boxplot() +
  xlab("Dam") +
  ylab("Placenta Mass") +
  labs(fill = "Litter Size") +
  ggtitle("Boxplot of Placenta Mass by Dam (Colored by Litter Size)") +
  theme_bw()
```



```
# Boxplot of Fetal_placental_ratio by Dam with box color by Litter_size
ggplot(fetus, aes(x = as.factor(Dam), y = Fetal_placental_ratio, fill = as.factor(Litter_size))) +
  geom_boxplot() +
  xlab("Dam") +
  ylab("Fetal-Placental Ratio") +
  labs(fill = "Litter Size") +
  ggtitle("Boxplot of Fetal-Placental Ratio by Dam (Colored by Litter Size)") +
  theme_bw()
```



While these plots are a bit cluttered, what they do show is that there is little correlation between litter size and any of our variables of interest. It also shows that these variables are highly dependent on Dam. Babies from the same mother looks similar (hence why the boxes are not tall and have short tails on average).

### Modeling of the Dependent Variables

Dam is obviously the most important factor here. If we were to train a regression without it, we'd certainly get awful results. In fact, we can see this here. We only do this for Fetal\_mass.

```
model <- lm(Fetal_mass ~ Litter_size + isFemale + Maternal_genotype + Fetus_genotype, data = fetus)
summary(model)
```

Call:

```
lm(formula = Fetal_mass ~ Litter_size + isFemale + Maternal_genotype + Fetus_genotype, data = fetus)
```

Residuals:

```
Min      1Q  Median      3Q      Max
```

-0.90129 -0.10817 0.02673 0.15308 0.31921

Coefficients:

	Estimate	Std. Error	t value	Pr(> t )
(Intercept)	1.727050	0.151470	11.402	< 2e-16 ***
Litter_size	-0.098878	0.015497	-6.380	4.51e-09 ***
isFemale	-0.011653	0.045971	-0.253	0.8004
Maternal_genotypefl/fl	0.142784	0.065461	2.181	0.0313 *
Maternal_genotypeHET	0.086907	0.066400	1.309	0.1934
Fetus_genotypeko	0.002518	0.083843	0.030	0.9761
Fetus_genotypewt	0.031989	0.060187	0.531	0.5962

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 0.2392 on 108 degrees of freedom

Multiple R-squared: 0.2931, Adjusted R-squared: 0.2539

F-statistic: 7.465 on 6 and 108 DF, p-value: 1.055e-06

Notice that nothing except for the intercept and Litter\_size (which before we proposed might just indirectly be capturing the variance of Dam ID) are significant. With Bayesian model selection, we can confirm this is the case.

```
y<-fetus$Fetal_mass
fetus_encoded <- fetus %>%
  mutate(
    Maternal_genotype = as.factor(Maternal_genotype),
    Fetus_genotype = as.factor(Fetus_genotype)
  )

# Create the design matrix X
X <- model.matrix(Fetal_mass ~ isFemale + Litter_size + Maternal_genotype + Fetus_genotype

lmratio.gprior<-function(z0,z1,y,X,g=dim(X)[1],nu0=1,
                          s200=mean( lm(y~-1+X[,z0==1])$res^2),
                          s201=mean( lm(y~-1+X[,z1==1])$res^2) )
{
  n<-dim(X)[1]

  X0<-X[,z0==1]
  X1<-X[,z1==1]

  H0<- (g/(g+1)) * X0%*%solve(t(X0)%*%X0)%*%t(X0)
```

```

SS0<- t(y)%*%( diag(1,nrow=n) - H0 ) %*%y
p0<-sum(z0==1)

H1<- (g/(g+1)) * X1%%solve(t(X1)%*%X1)%*%t(X1)
SS1<- t(y)%*%( diag(1,nrow=n) - H1 ) %*%y
p1<-sum(z1==1)

-.5*(p1-p0)*log( 2*pi*(1+g)) +
.5*nu0*log(s201/s200) + .5*(nu0+n)*log( (nu0*s200+SS0)/(nu0+s201+SS1) )
}

lpy.X<-function(y,X,g=length(y),nu0=1,s20=try(summary(lm(y~-1+X))$sigma^2,silent=TRUE))
{
  n<-dim(X)[1] ; p<-dim(X)[2]
  if(p==0) { s20<-mean(y^2) }
  H0<-0 ; if(p>0) { H0<- (g/(g+1)) * X%%solve(t(X)%*%X)%*%t(X) }
  SS0<- t(y)%*%( diag(1,nrow=n) - H0 ) %*%y

  -.5*n*log(2*pi) +lgamma(.5*(nu0+n)) - lgamma(.5*nu0) - .5*p*log(1+g) +
  .5*nu0*log(.5*nu0*s20) - .5*(nu0+n)*log(.5*(nu0*s20+SS0))
}

#### Bayesian model selection
p<-dim(X)[2]
S<-1000
z<-rep(1,p)
Z<-matrix(NA,S,p)
lpy.c<-lpy.X(y,X[,z==1,drop=FALSE])
for(s in 1:S)
{
  for(j in sample(1:p))
  {
    zp<-z ; zp[j]<-1-zp[j]
    lpy.p<-lpy.X(y,X[,zp==1,drop=FALSE])
    r<- (lpy.p - lpy.c)*(-1)^(zp[j]==0)
    z[j]<-rbinom(1,1,1/(1+exp(-r)))
    if(z[j]==zp[j]) {lpy.c<-lpy.p}
  }
  Z[s,]<-z
}

```



```
means <- colMeans(Z)
matrix(means, nrow = 1, ncol = ncol(Z), dimnames = list(NULL, colnames(X)))
```

```
      (Intercept) isFemale Litter_size Maternal_genotypefl/fl
[1,]           1    0.079           1           0.3
      Maternal_genotypeHET Fetus_genotypeko Fetus_genotypewt
[1,]           0.106           0.085           0.084
```

Only the intercept and Litter Size are probably features. We will thus switch over to a mixed model to see if there is any value to the genotypes.

```
fetal_mass_model<-lmer(Fetal_mass ~ Litter_size + isFemale*Fetus_genotype + (1|Dam) + (1|Maternal_genotype))
```

boundary (singular) fit: see help('isSingular')

```
summary(fetal_mass_model)
```

```
Linear mixed model fit by REML. t-tests use Satterthwaite's method [
lmerModLmerTest]
Formula: Fetal_mass ~ Litter_size + isFemale * Fetus_genotype + (1 | Dam) +
      (1 | Maternal_genotype)
Data: fetus
```

REML criterion at convergence: -189.1

```
Scaled residuals:
      Min       1Q   Median       3Q      Max
-2.81303 -0.48478 -0.05347  0.56638  2.72874
```

```
Random effects:
 Groups          Name      Variance Std.Dev.
 Dam              (Intercept) 0.082115 0.28656
 Maternal_genotype (Intercept) 0.000000 0.00000
 Residual                        0.004689 0.06848
Number of obs: 115, groups:  Dam, 15; Maternal_genotype, 3
```

```
Fixed effects:
              Estimate Std. Error    df t value Pr(>|t|)
(Intercept)    1.16725    0.27595 13.97137   4.230 0.000844 ***
```

Litter_size	-0.03086	0.03217	13.65742	-0.959	0.354159
isFemale	0.03554	0.02693	94.99296	1.320	0.190007
Fetus_genotypeko	0.03493	0.03413	95.14271	1.023	0.308707
Fetus_genotypewt	0.06588	0.02709	95.43547	2.432	0.016887 *
isFemale:Fetus_genotypeko	-0.01959	0.05207	95.06774	-0.376	0.707546
isFemale:Fetus_genotypewt	-0.05356	0.03322	95.20069	-1.612	0.110204

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

Correlation of Fixed Effects:

	(Intr)	Lttr_s	isFeml	Fts_gntypk	Fts_gntypw	isFml:Fts_gntypk
Litter_size	-0.960					
isFemale	-0.061	0.000				
Fets_gntypk	-0.051	0.002	0.508			
Fts_gntypwt	-0.077	0.006	0.638	0.487		
isFml:Fts_gntypk	0.025	0.002	-0.502	-0.602	-0.252	
isFml:Fts_gntypw	0.040	0.013	-0.838	-0.421	-0.741	0.396

optimizer (nloptwrap) convergence code: 0 (OK)  
boundary (singular) fit: see help('isSingular')

```
placenta_mass_model<-lmer(Placenta_mass ~ Litter_size + isFemale*Fetus_genotype + (1|Dam)
```

boundary (singular) fit: see help('isSingular')

```
summary(placenta_mass_model)
```

Linear mixed model fit by REML. t-tests use Satterthwaite's method [  
lmerModLmerTest]

Formula: Placenta\_mass ~ Litter\_size + isFemale \* Fetus\_genotype + (1 |  
Dam) + (1 | Maternal\_genotype)  
Data: fetus

REML criterion at convergence: -522.4

Scaled residuals:

Min	1Q	Median	3Q	Max
-2.29513	-0.58310	-0.01651	0.52652	3.05477

Random effects:

Groups	Name	Variance	Std.Dev.
--------	------	----------	----------

```

Dam (Intercept) 0.0011418 0.03379
Maternal_genotype (Intercept) 0.0000000 0.00000
Residual 0.0002511 0.01584
Number of obs: 115, groups: Dam, 15; Maternal_genotype, 3

```

Fixed effects:

	Estimate	Std. Error	df	t value	Pr(> t )
(Intercept)	0.149157	0.034623	16.993231	4.308	0.000477 ***
Litter_size	-0.005834	0.003981	15.749549	-1.466	0.162446
isFemale	0.005232	0.006227	95.410430	0.840	0.402842
Fetus_genotypeko	0.010757	0.007885	95.954771	1.364	0.175679
Fetus_genotypewt	0.001410	0.006244	97.004093	0.226	0.821820
isFemale:Fetus_genotypeko	-0.016641	0.012034	95.687772	-1.383	0.169936
isFemale:Fetus_genotypewt	-0.009003	0.007670	96.161517	-1.174	0.243413

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

Correlation of Fixed Effects:

	(Intr)	Lttr_s	isFeml	Fts_gntypk	Fts_gntypw	isFml:Fts_gntypk
Litter_size	-0.956					
isFemale	-0.113	0.000				
Fets_gntypk	-0.096	0.005	0.508			
Fts_gntypwt	-0.144	0.011	0.639	0.489		
isFml:Fts_gntypk	0.046	0.004	-0.502	-0.603	-0.255	
isFml:Fts_gntypw	0.076	0.023	-0.838	-0.421	-0.741	0.397

optimizer (nloptwrap) convergence code: 0 (OK)  
boundary (singular) fit: see help('isSingular')

```

fpratio_model<-lmer(Fetal_placental.ratio ~ Litter_size + isFemale*Fetus_genotype + (1|Dam)
summary(fpratio_model)

```

Linear mixed model fit by REML. t-tests use Satterthwaite's method [  
lmerModLmerTest]

Formula: Fetal\_placental.ratio ~ Litter\_size + isFemale \* Fetus\_genotype +  
(1 | Dam) + (1 | Maternal\_genotype)  
Data: fetus

REML criterion at convergence: 489.6

Scaled residuals:

Min	1Q	Median	3Q	Max
-----	----	--------	----	-----

-1.9672 -0.5152 -0.1121 0.4116 3.0492

Random effects:

Groups	Name	Variance	Std.Dev.
Dam	(Intercept)	15.0830	3.8837
Maternal_genotype	(Intercept)	0.1468	0.3832
Residual		2.8984	1.7025

Number of obs: 115, groups: Dam, 15; Maternal\_genotype, 3

Fixed effects:

	Estimate	Std. Error	df	t value	Pr(> t )
(Intercept)	8.71167	3.94680	15.49467	2.207	0.0428 *
Litter_size	0.14759	0.45407	12.39835	0.325	0.7506
isFemale	0.15960	0.66913	95.17382	0.239	0.8120
Fetus_genotypeko	-1.15735	0.84746	95.65656	-1.366	0.1752
Fetus_genotypewt	0.47085	0.67146	96.45555	0.701	0.4848
isFemale:Fetus_genotypeko	2.45928	1.29330	95.41909	1.902	0.0602 .
isFemale:Fetus_genotypewt	-0.06081	0.82446	95.84057	-0.074	0.9414

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

Correlation of Fixed Effects:

	(Intr)	Lttr_s	isFeml	Fts_gntypk	Fts_gntypw	isFml:Fts_gntypk
Litter_size	-0.955					
isFemale	-0.107	0.000				
Fets_gntypk	-0.090	0.005	0.508			
Fts_gntypwt	-0.135	0.011	0.639	0.489		
isFml:Fts_gntypk	0.043	0.003	-0.502	-0.603	-0.254	
isFml:Fts_gntypw	0.071	0.022	-0.838	-0.421	-0.741	0.397

We've noticed that a fetus of genotype WT might have, on average, higher fetal mass. We investigate this claim on 4 mothers with the largest litter sizes.

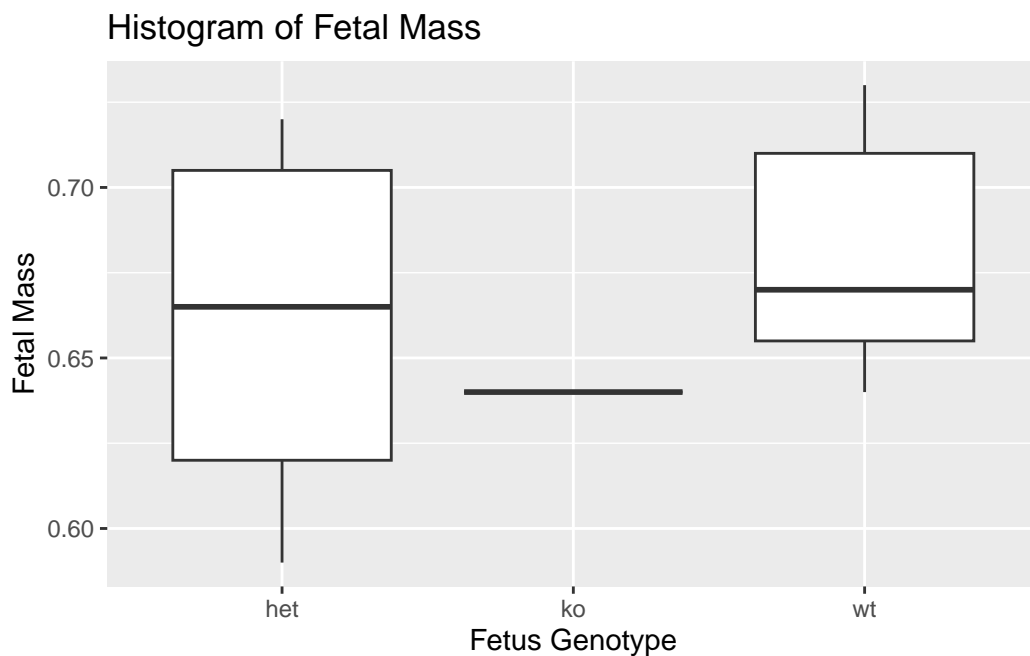
filtered\_fetus

```
# Filter the dataset to include observations from the four Dams with the largest litter si
dam_values <- unique(fetus %>%
  group_by(Dam) %>%
  arrange(desc(Litter_size)) %>%
  top_n(4, Dam) %>%
  pull(Dam)
)[1:4]
```

```
# Filter the dataset to include observations where Dam is one of the specified values
dam1 <- fetus %>%
  filter(Dam == dam_values[1])
dam2 <- fetus %>%
  filter(Dam == dam_values[2])
dam3 <- fetus %>%
  filter(Dam == dam_values[3])
dam4 <- fetus %>%
  filter(Dam == dam_values[4])
```

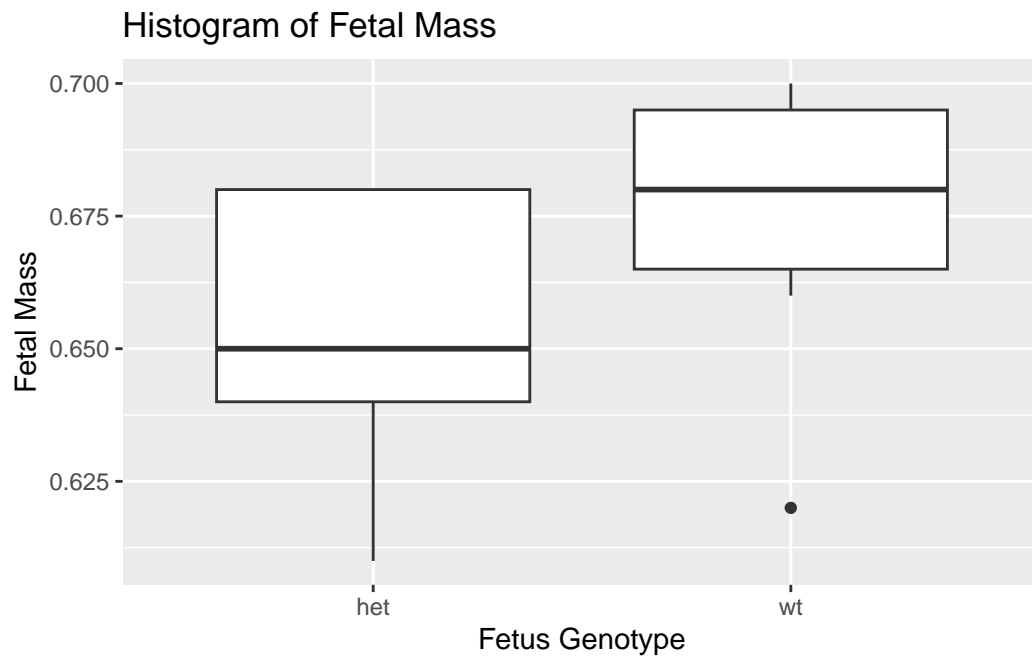
The plots below show the actual values.

```
ggplot(dam1, aes(y = Fetal_mass, x = Fetus_genotype)) +
  geom_boxplot() +
  ylab("Fetal Mass") +
  xlab("Fetus Genotype") +
  ggtitle("Histogram of Fetal Mass")
```

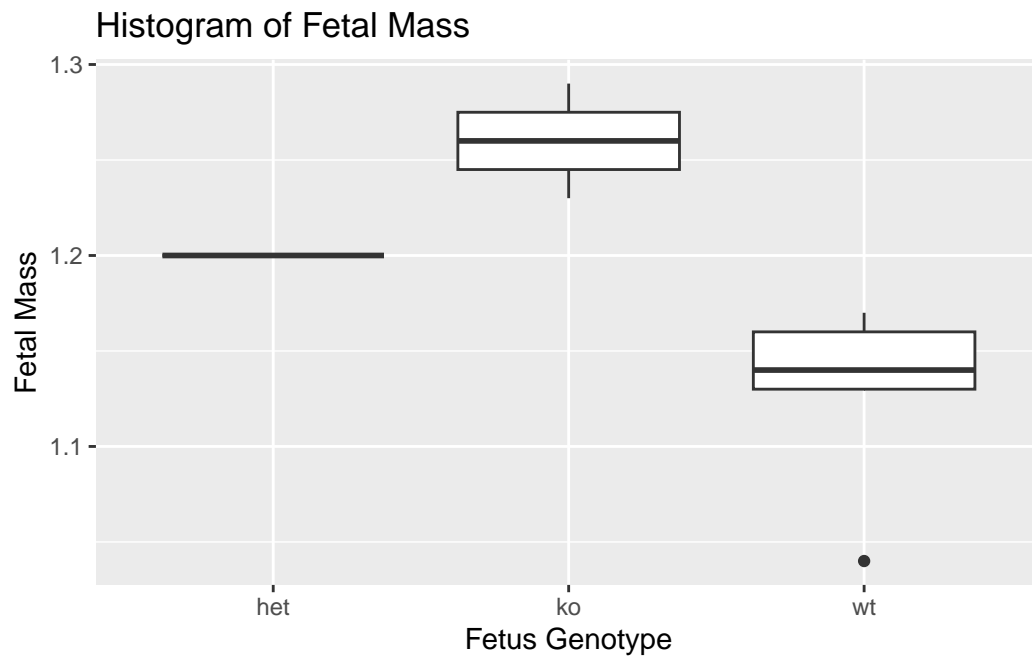


```
ggplot(dam2, aes(y = Fetal_mass, x = Fetus_genotype)) +
  geom_boxplot() +
  ylab("Fetal Mass") +
```

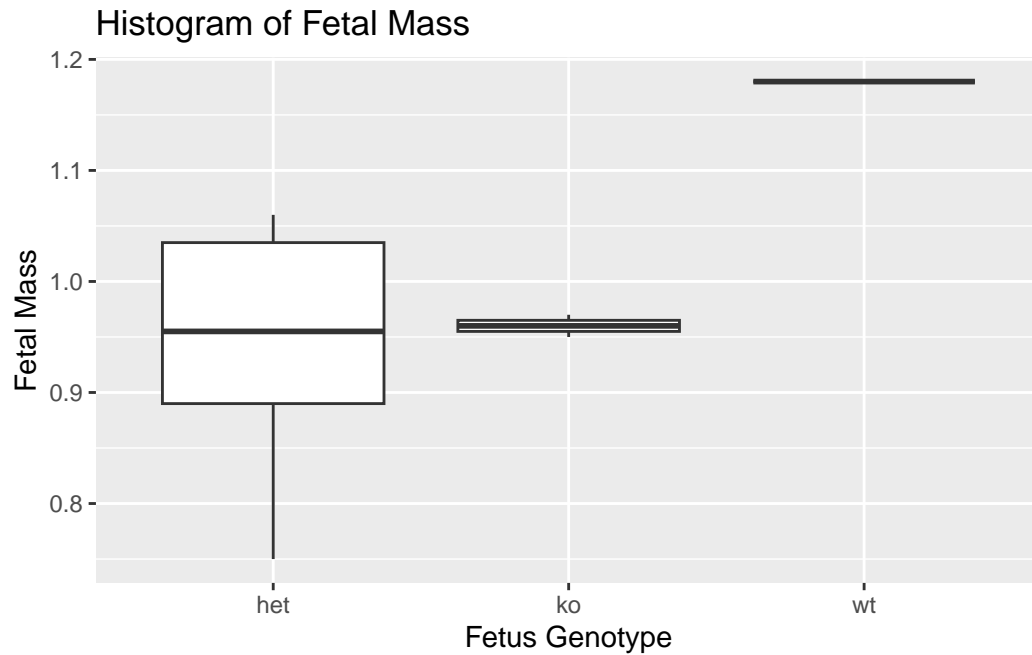
```
xlab("Fetus Genotype") +  
ggtitle("Histogram of Fetal Mass")
```



```
ggplot(dam3, aes(y = Fetal_mass, x = Fetus_genotype)) +  
  geom_boxplot() +  
  ylab("Fetal Mass") +  
  xlab("Fetus Genotype") +  
  ggtitle("Histogram of Fetal Mass")
```



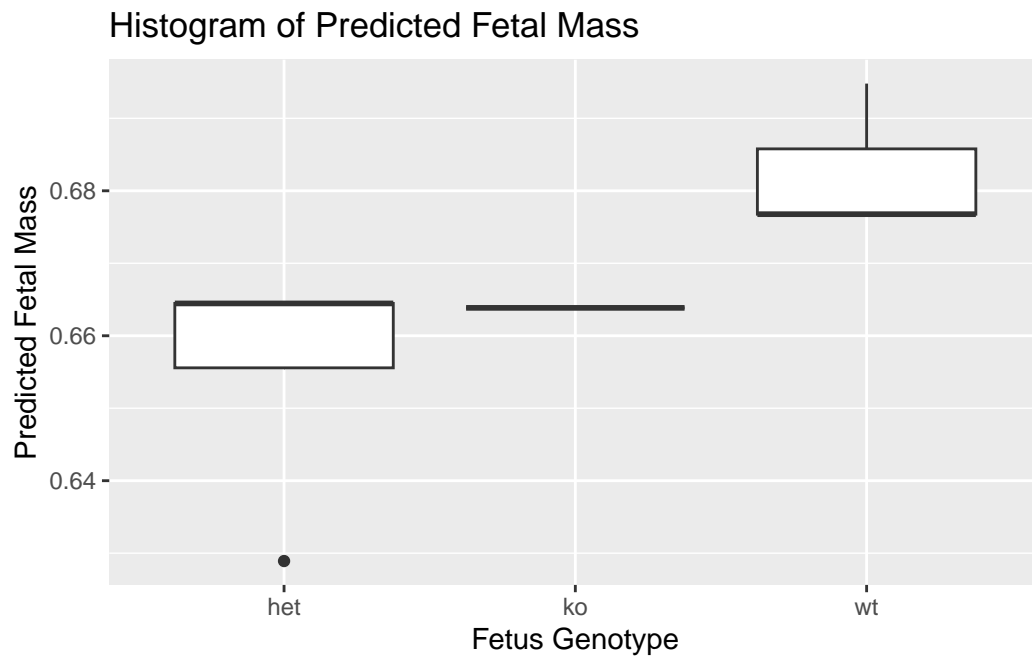
```
ggplot(dam4, aes(y = Fetal_mass, x = Fetus_genotype)) +  
  geom_boxplot() +  
  ylab("Fetal Mass") +  
  xlab("Fetus Genotype") +  
  ggtitle("Histogram of Fetal Mass")
```



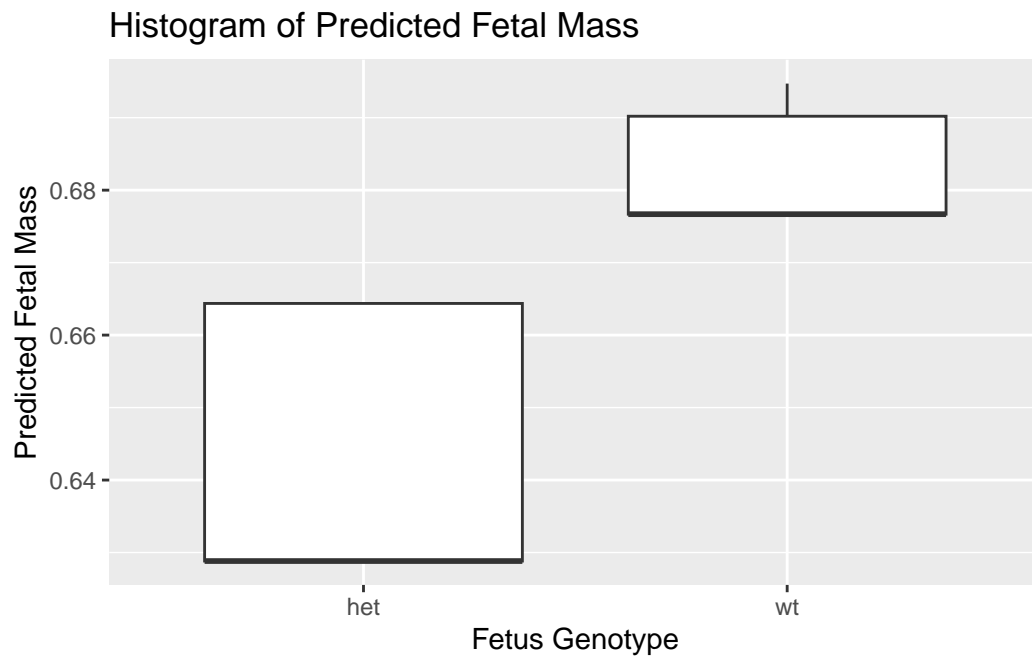
```
dam1$predicted = predict(fetal_mass_model, newdata=dam1)
dam2$predicted = predict(fetal_mass_model, newdata=dam2)
dam3$predicted = predict(fetal_mass_model, newdata=dam3)
dam4$predicted = predict(fetal_mass_model, newdata=dam4)

ggplot(dam1, aes(y = predicted, x = Fetus_genotype)) +
  geom_boxplot() +
  ylab("Predicted Fetal Mass") +
  xlab("Fetus Genotype") +
  ggtitle("Histogram of Predicted Fetal Mass")
```

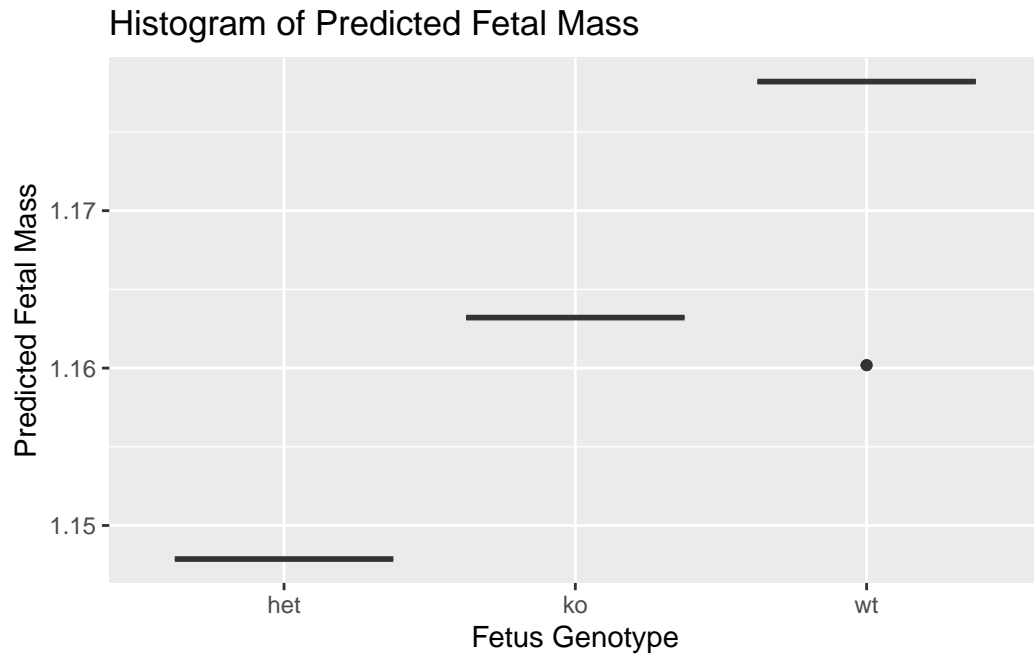




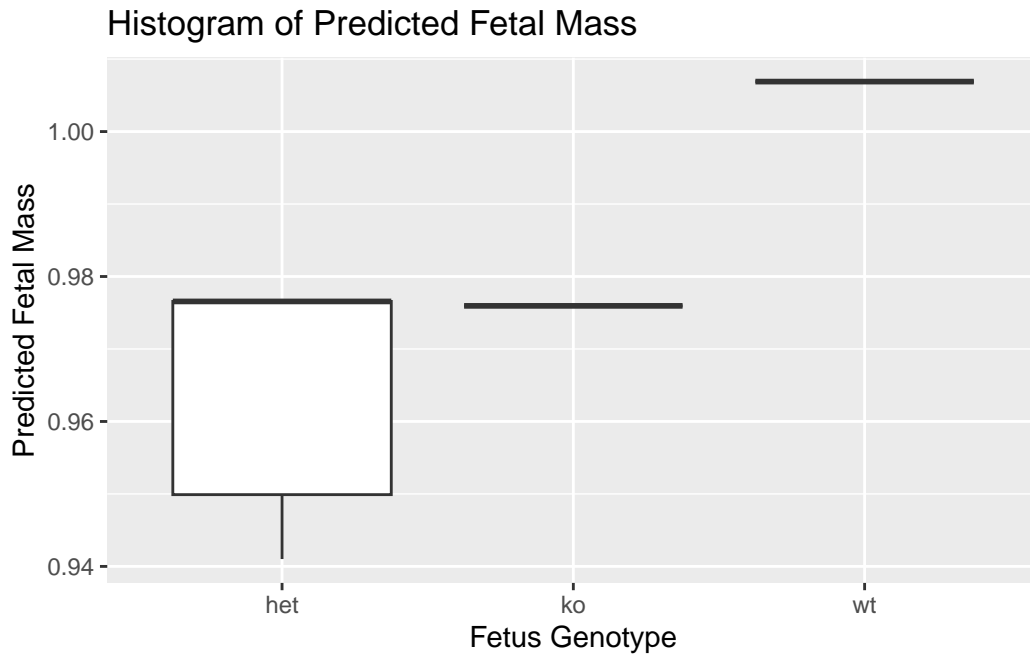
```
ggplot(dam2, aes(y = predicted, x = Fetus_genotype)) +  
  geom_boxplot() +  
  ylab("Predicted Fetal Mass") +  
  xlab("Fetus Genotype") +  
  ggtitle("Histogram of Predicted Fetal Mass")
```



```
ggplot(dam3, aes(y = predicted, x = Fetus_genotype)) +  
  geom_boxplot() +  
  ylab("Predicted Fetal Mass") +  
  xlab("Fetus Genotype") +  
  ggtitle("Histogram of Predicted Fetal Mass")
```



```
ggplot(dam4, aes(y = predicted, x = Fetus_genotype)) +  
  geom_boxplot() +  
  ylab("Predicted Fetal Mass") +  
  xlab("Fetus Genotype") +  
  ggtitle("Histogram of Predicted Fetal Mass")
```

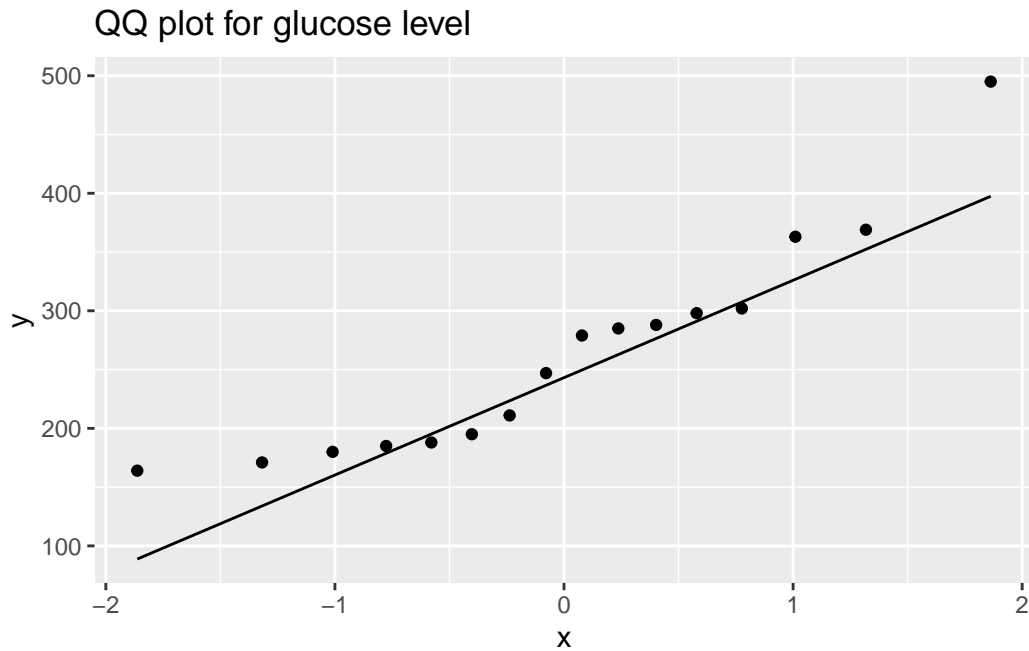


## Mother Data

```
mother <- read.csv("data/mother-data.csv", header=T)
```

Now we'd like to assess the differences in mean blood glucose levels between maternal genotypes. For each time period, we will check if there is normality and constant variance, then conduct an ANCOVA analysis to see if the genotype makes a difference.

```
# Checking visually
ggplot(mother, aes(sample=gluc_60)) +
  stat_qq() +
  stat_qq_line() +
  ggtitle("QQ plot for glucose level")
```



```
# Checking mathematically
shapiro.test(mother$gluc_60)
```

Shapiro-Wilk normality test

```
data:  mother$gluc_60
W = 0.88957, p-value = 0.05483
```

Normality is satisfied.

```
mother %>%
  group_by(Maternal_genotype) %>%
  summarise(n = n(), mean = mean(gluc_60), sd = sd(gluc_60))
```

```
# A tibble: 3 x 4
  Maternal_genotype      n  mean    sd
  <chr>             <int> <dbl> <dbl>
1 fl/+               5  223.  60.7
2 fl/fl              7  327.  95.3
3 HET                 4  204.  34.5
```

Constant variance is satisfied ( $95/34 \approx 2.8$  and sample size is small, so this is fine).

```
# First, construct a model including covariates
model <- lm(gluc_60 ~ Maternal_genotype + num_fetus + percent_body_weight_gain, data = mot

# Then perform ANCOVA using the model
anova(model)
```

#### Analysis of Variance Table

Response: gluc\_60

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Maternal_genotype	2	50758	25379.1	6.2132	0.01564 *
num_fetus	1	27732	27732.4	6.7894	0.02445 *
percent_body_weight_gain	1	187	186.8	0.0457	0.83456
Residuals	11	44932	4084.7		

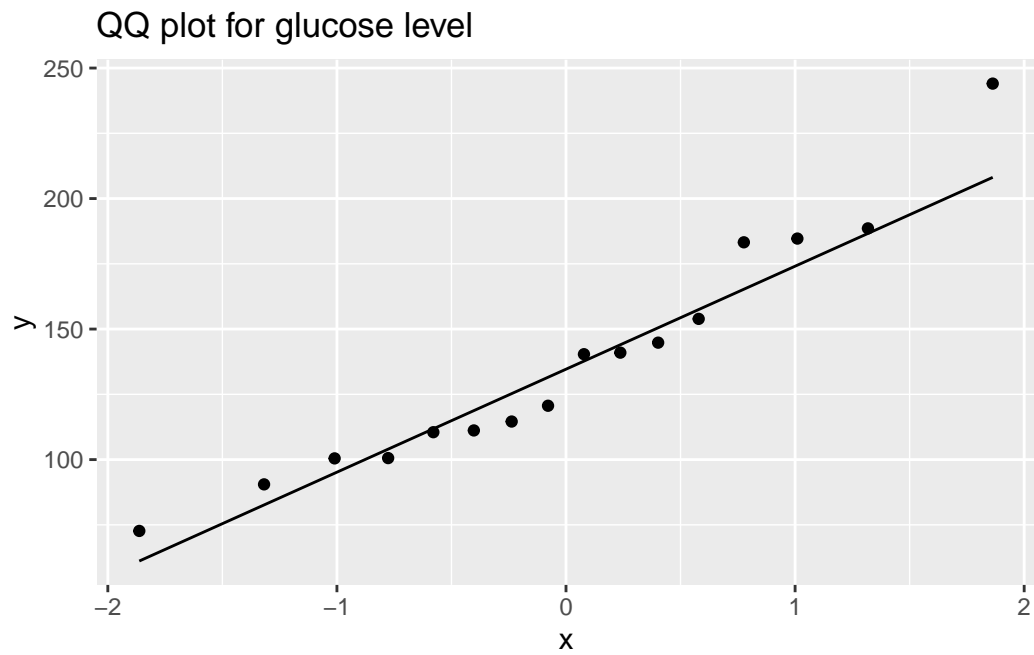
---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

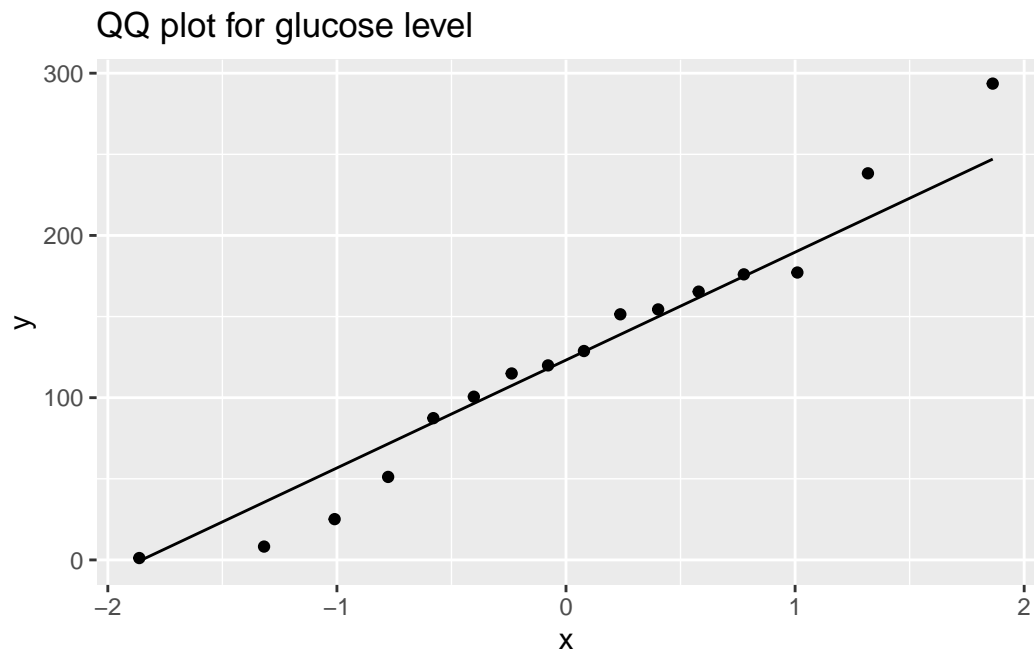
We find that only at 60 minutes are we 95% confident that there is a difference in average blood glucose level.

Now we repeat this for % change in blood glucose levels

```
# Checking visually
ggplot(mother, aes(sample=chng_base_15)) +
  stat_qq() +
  stat_qq_line() +
  ggtitle("QQ plot for glucose level")
```



```
ggplot(mother, aes(sample=chng_base_30)) +  
  stat_qq() +  
  stat_qq_line() +  
  ggtitle("QQ plot for glucose level")
```



```
ggplot(mother, aes(sample=chng_base_60)) +  
  stat_qq() +  
  stat_qq_line() +  
  ggtitle("QQ plot for glucose level")
```





```
# Checking mathematically  
shapiro.test(mother$chng_base_15)
```

Shapiro-Wilk normality test

```
data:  mother$chng_base_15  
W = 0.93838, p-value = 0.3297
```

```
shapiro.test(mother$chng_base_30)
```

Shapiro-Wilk normality test

```
data:  mother$chng_base_30  
W = 0.96662, p-value = 0.7814
```

```
shapiro.test(mother$chng_base_60)
```

Shapiro-Wilk normality test

```
data: mother$chng_base_60
W = 0.88822, p-value = 0.0522
```

Normality is satisfied.

```
mother %>%
  group_by(Maternal_genotype) %>%
  summarise(n = n(), mean = mean(chng_base_15), sd = sd(chng_base_15))
```

```
# A tibble: 3 x 4
  Maternal_genotype      n  mean    sd
  <chr>             <int> <dbl> <dbl>
1 fl/+              5  116.  42.7
2 fl/fl             7  163.  46.8
3 HET                4  121.  23.2
```

```
mother %>%
  group_by(Maternal_genotype) %>%
  summarise(n = n(), mean = mean(chng_base_30), sd = sd(chng_base_30))
```

```
# A tibble: 3 x 4
  Maternal_genotype      n  mean    sd
  <chr>             <int> <dbl> <dbl>
1 fl/+              5  85.1  70.1
2 fl/fl             7 186.   59.7
3 HET                4  67.0  51.7
```

```
mother %>%
  group_by(Maternal_genotype) %>%
  summarise(n = n(), mean = mean(chng_base_60), sd = sd(chng_base_60))
```

```
# A tibble: 3 x 4
  Maternal_genotype      n  mean    sd
  <chr>             <int> <dbl> <dbl>
1 fl/+              5  33.9  49.3
2 fl/fl             7 131.   59.8
3 HET                4  16.3  29.8
```

Constant variance is satisfied.

```
# First, construct a model including covariates
model_15_30 <- lm(chng_15_30 ~ Maternal_genotype + num_fetus + percent_body_weight_gain, d
model_base_30 <- lm(chng_base_30 ~ Maternal_genotype + num_fetus + percent_body_weight_gai
model_base_60 <- lm(chng_base_60 ~ Maternal_genotype + num_fetus + percent_body_weight_gai

# Then perform ANCOVA using the model
anova(model_15_30)
```

#### Analysis of Variance Table

Response: chng\_15\_30

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Maternal_genotype	2	3565.2	1782.60	5.9329	0.01787 *
num_fetus	1	153.8	153.83	0.5120	0.48918
percent_body_weight_gain	1	1.0	0.99	0.0033	0.95521
Residuals	11	3305.0	300.46		

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

```
anova(model_base_30)
```

#### Analysis of Variance Table

Response: chng\_base\_30

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Maternal_genotype	2	47244	23621.9	5.4670	0.02247 *
num_fetus	1	38	37.9	0.0088	0.92709
percent_body_weight_gain	1	1499	1499.2	0.3470	0.56773
Residuals	11	47529	4320.8		

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

```
anova(model_base_60)
```

#### Analysis of Variance Table

Response: chng\_base\_60

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Maternal_genotype	2	43659	21829.3	9.7512	0.003663 **
num_fetus	1	7821	7821.0	3.4937	0.088443 .
percent_body_weight_gain	1	1396	1396.3	0.6237	0.446353
Residuals	11	24625	2238.6		

---

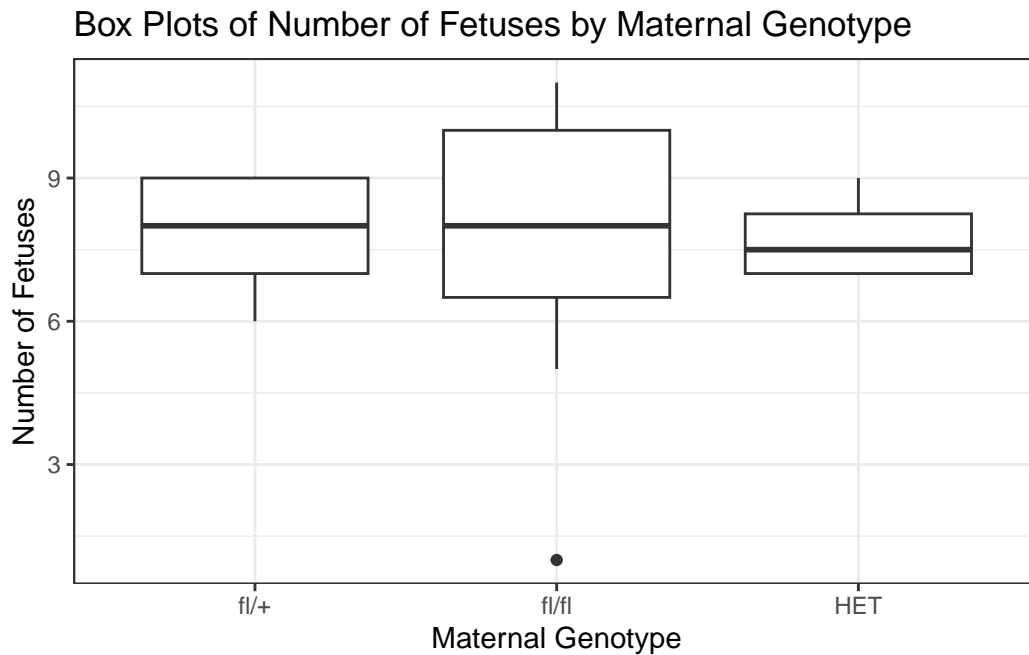
Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

15-30, base-30, base-60 all have significant p-values to suggest a difference among maternal genotype groups.

Now we are interested in assessing the impact of Maternal Genotype on the # of fetuses, # of absorptions, body weight gain, and % body weight gain.

We first do this visually and with linear models.

```
# Num Fetuses
ggplot(data = mother, aes(x = Maternal_genotype, y = num_fetus)) +
  geom_boxplot() +
  xlab("Maternal Genotype") +
  ylab("Number of Fetuses") +
  ggtitle("Box Plots of Number of Fetuses by Maternal Genotype") +
  theme_bw()
```



```
model <- lm(num_fetus ~ Maternal_genotype, data=mother)

summary(model)
```

Call:

```
lm(formula = num_fetus ~ Maternal_genotype, data = mother)
```

Residuals:

Min	1Q	Median	3Q	Max
-6.5714	-0.7625	0.3393	1.2125	3.4286

Coefficients:

	Estimate	Std. Error	t value	Pr(> t )
(Intercept)	7.8000	1.1453	6.810	1.24e-05 ***
Maternal_genotypefl/fl	-0.2286	1.4996	-0.152	0.881
Maternal_genotypeHET	-0.0500	1.7180	-0.029	0.977

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

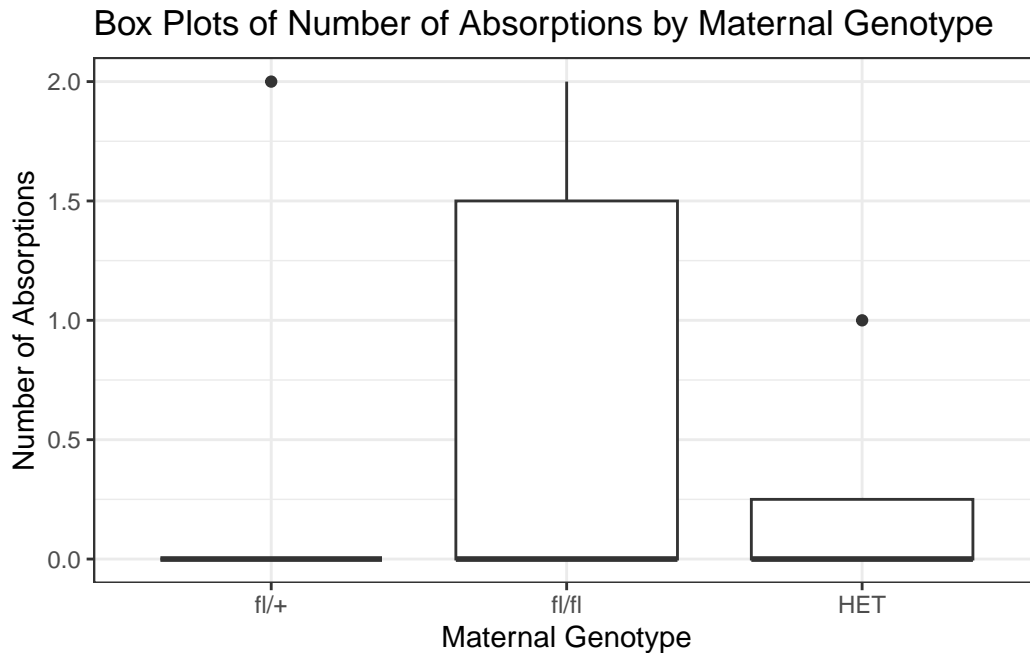
Residual standard error: 2.561 on 13 degrees of freedom

Multiple R-squared: 0.002027, Adjusted R-squared: -0.1515

F-statistic: 0.0132 on 2 and 13 DF, p-value: 0.9869

No impact of maternal genotype on the # of fetus

```
# Num Absorptions
ggplot(data = mother, aes(x = Maternal_genotype, y = absorptions)) +
  geom_boxplot() +
  xlab("Maternal Genotype") +
  ylab("Number of Absorptions") +
  ggtitle("Box Plots of Number of Absorptions by Maternal Genotype") +
  theme_bw()
```



```
# Use poisson regression for count data
model <- glm(absorptions ~ Maternal_genotype, data=mother, family="poisson")

summary(model)
```

Call:

```
glm(formula = absorptions ~ Maternal_genotype, family = "poisson",
    data = mother)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-1.1952	-0.9696	-0.8008	0.5210	1.7994

Coefficients:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	-0.9163	0.7071	-1.296	0.195
Maternal_genotypefl/fl	0.5798	0.8367	0.693	0.488
Maternal_genotypeHET	-0.4700	1.2247	-0.384	0.701

(Dispersion parameter for poisson family taken to be 1)

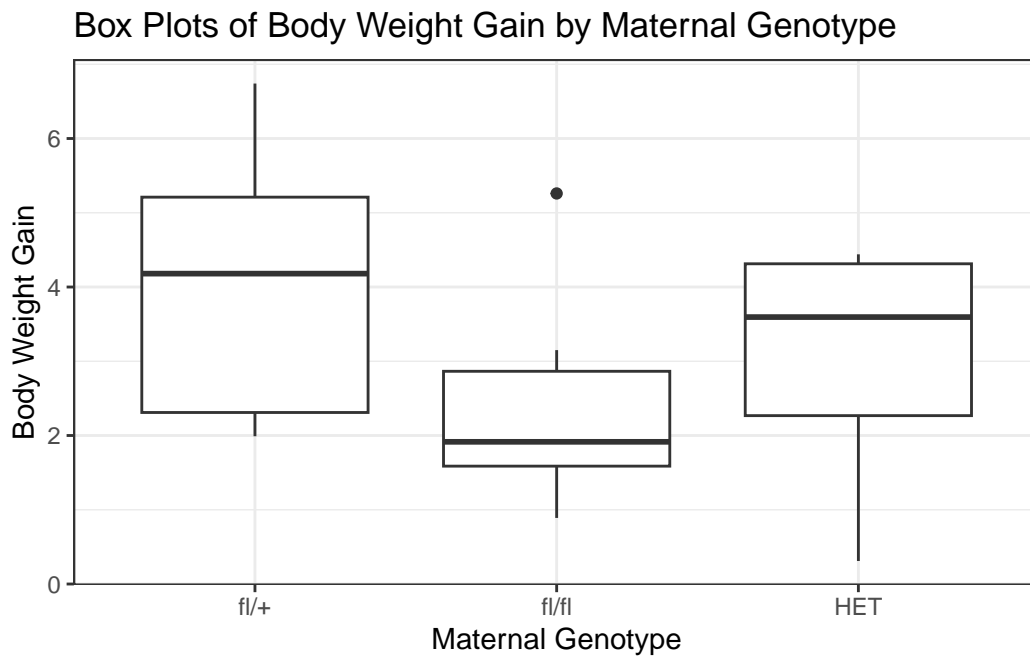
Null deviance: 19.408 on 15 degrees of freedom  
Residual deviance: 18.120 on 13 degrees of freedom  
AIC: 35.961

Number of Fisher Scoring iterations: 6

No attributable effect of maternal genotype on number of absorptions, almost meaningless analysis though since there's very little data

```
# Body weight gain
ggplot(data = mother, aes(x = Maternal_genotype, y = body_weight_gain)) +
  geom_boxplot() +
  xlab("Maternal Genotype") +
  ylab("Body Weight Gain") +
  ggtitle("Box Plots of Body Weight Gain by Maternal Genotype") +
  theme_bw()
```

Warning: Removed 1 rows containing non-finite values (``stat_boxplot()``).



```
model <- lm(body_weight_gain ~ Maternal_genotype, data=mother)

summary(model)
```

Call:

```
lm(formula = body_weight_gain ~ Maternal_genotype, data = mother)
```

Residuals:

Min	1Q	Median	3Q	Max
-2.675	-1.240	-0.065	1.204	2.820

Coefficients:

	Estimate	Std. Error	t value	Pr(> t )
(Intercept)	4.0860	0.8071	5.063	0.000279 ***
Maternal_genotypefl/fl	-1.6460	1.0928	-1.506	0.157879
Maternal_genotypeHET	-1.1010	1.2106	-0.909	0.381027

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 1.805 on 12 degrees of freedom

(1 observation deleted due to missingness)

Multiple R-squared: 0.1612, Adjusted R-squared: 0.02138

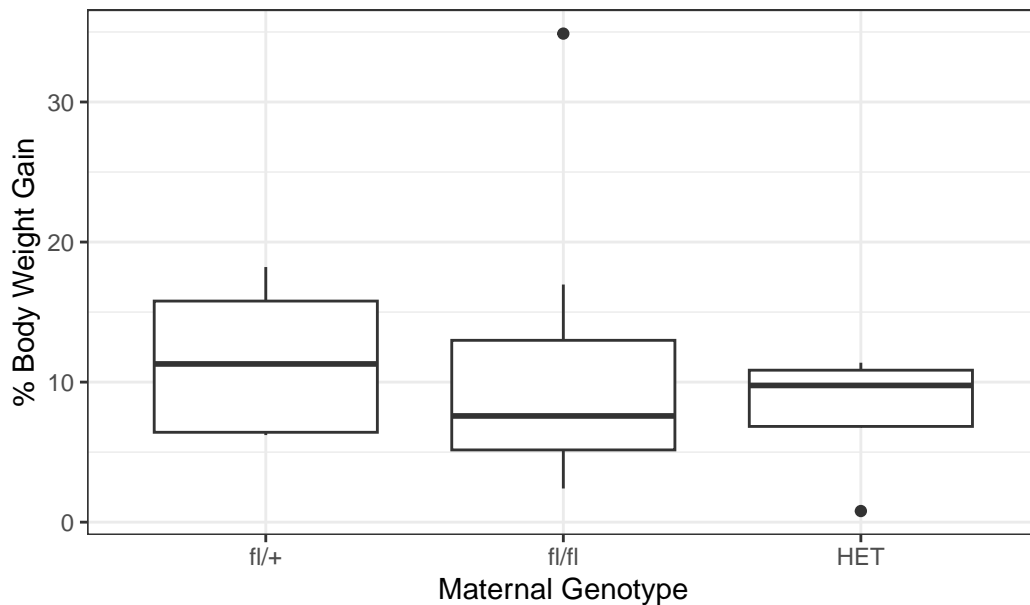
F-statistic: 1.153 on 2 and 12 DF, p-value: 0.3483

No significant effect of maternal genotype on body weight gain.

```
# % body weight gain
ggplot(data = mother, aes(x = Maternal_genotype, y = percent_body_weight_gain)) +
  geom_boxplot() +
  xlab("Maternal Genotype") +
  ylab("% Body Weight Gain") +
  ggtitle("Box Plots of % Body Weight Gain by Maternal Genotype") +
  theme_bw()
```



Box Plots of % Body Weight Gain by Maternal Genotype



```
model <- lm(percent_body_weight_gain ~ Maternal_genotype, data=mother)

summary(model)
```

Call:

```
lm(formula = percent_body_weight_gain ~ Maternal_genotype, data = mother)
```

Residuals:

Min	1Q	Median	3Q	Max
-9.189	-5.489	-1.442	3.644	23.290

Coefficients:

	Estimate	Std. Error	t value	Pr(> t )
(Intercept)	11.587362	3.823964	3.030	0.00966 **
Maternal_genotypefl/fl	0.006755	5.006744	0.001	0.99894
Maternal_genotypeHET	-3.661619	5.735946	-0.638	0.53432

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 8.551 on 13 degrees of freedom

Multiple R-squared: 0.04068, Adjusted R-squared: -0.1069

F-statistic: 0.2757 on 2 and 13 DF, p-value: 0.7634

No significant effect of % bw gain.