

Week 2: Sequence alignment & search

Substitution, BLAST

- Substitution matrix
 - Construction
 - Properties
- BLAST
 - Statistics of similarity search

Substitution matrix to measure similarity in sequence alignments



Dr. Margaret Dayhoff

Applying math & computational techniques to the sequencing of proteins and nucleic acids.

- 1965: First collection of protein seqs.
- Single-letter code for amino acids.
- 1966: 'Evolutionary trees'.
- **1978: First AA similarity-scoring matrix.**
- 1980: Launched the Protein Information Resource, the first online database system that could be accessed by telephone line.

Substitution matrix: A collection of scores for aligning nucleotides or amino acids with one another.

- The scores represent the relative ease with which one nucleotide or amino acid may mutate into or substitute for another.
- Purely statistical, nothing directly to do with structure/biochemistry.

[illegible]

Substitution matrix to measure similarity in sequence alignments

Substitution matrix: Each score is a log-odds score equal to the logarithm of the ratio of the likelihoods of two hypotheses: i) the residues can substitute for one another, or ii) not.

$$s(a,b) = \frac{1}{\lambda} \log \frac{p_{ab}}{f_a f_b}$$

- p_{ab} : likelihood of these two residues being correlated because they're homologous.
 - p_{ab} are the target frequencies: the probability that we expect to observe residues a and b aligned in homologous sequence alignments.
- $f_a f_b$: likelihood of these two residues being uncorrelated and unrelated, occurring independently.
 - f_a and f_b are background frequencies: the probabilities that we expect to observe amino acids a and b on average in any protein sequence.
- λ : a scaling factor, usually set to something that helps round off all the terms in the score matrix to sensible integers.

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- λ : a scaling factor

Assuming that each aligned residue pair is statistically independent of the others (biologically dubious, but mathematically convenient):

- The score of an alignment ("**alignment score**") = sum of individual log-odds scores for each aligned residue pair.

Substitution matrix to measure similarity in sequence alignments

BLOSUM (BLOcks SUBstitution Matrix) for protein sequence alignment.

- Scan the BLOCKS database for very conserved regions of protein families (w/o gaps in the alignment).
- Count the relative frequencies of AA and their substitution probabilities.
- Calculate a log-odds score for each of the 210 possible substitution pairs of the 20 standard amino acids.

Ala	4																			
Arg	-1	5																		
Asn	-2	0	6																	
Asp	-2	-2	1	6																
Cys	0	-3	-3	-3	9															
Gln	-1	1	0	0	-3	5														
Glu	-1	0	0	2	-4	2	5													
Gly	0	-2	0	-1	-3	-2	-2	6												
His	-2	0	1	-1	-3	0	0	-2	8											
Ile	-1	-3	-3	-3	-1	-3	-3	-4	-3	4										
Leu	-1	-2	-3	-4	-1	-2	-3	-4	-3	2	4									
Lys	-1	2	0	-1	-3	1	1	-2	-1	-3	-2	5								
Met	-1	-1	-2	-3	-1	0	-2	-3	-2	1	2	-1	5							
Phe	-2	-3	-3	-3	-2	-3	-3	-3	-1	0	0	-3	0	6						
Pro	-1	-2	-2	-1	-3	-1	-1	-2	-2	-3	-3	-1	-2	-4	7					
Ser	1	-1	1	0	-1	0	0	0	-1	-2	-2	0	-1	-2	-1	4				
Thr	0	-1	0	-1	-1	-1	-1	-2	-2	-1	-1	-1	-1	-2	-1	1	5			
Trp	-3	-3	-4	-4	-2	-2	-3	-2	-2	-3	-2	-3	-1	1	-4	-3	-2	11		
Tyr	-2	-2	-2	-3	-2	-1	-2	-3	2	-1	-1	-2	-1	3	-3	-2	-2	2	7	
Val	0	-3	-3	-3	-1	-2	-2	-3	-3	3	1	-2	1	-1	-2	-2	0	-3	-1	4
	Ala	Arg	Asn	Asp	Cys	Gln	Glu	Gly	His	Ile	Leu	Lys	Met	Phe	Pro	Ser	Thr	Trp	Tyr	Val

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Ile	-1	-3	-3	-3	-1	-3	-3	-4	-3	4										
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	Ala	Arg	Asn	Asp	Cys	Gln	Glu	Gly	His	Ile	Leu	Lys	Met	Phe	Pro	Ser	Thr	Trp	Tyr	Val

- The rarer the amino acid is, the more surprising it would be to see two of them align together by chance.
- L/L more common than WW:
 - $p_{LL} = 0.0371$, $p_{WW} = 0.0065$
- W is a much rarer amino acid:
 - $f_L = 0.099$, $f_W = 0.013$.

Check with
 $\lambda = 0.347$.

Substitution matrix to measure similarity in sequence alignments

BLOSUM (BLOcks SUBstitution Matrix) for protein sequence alignment.

- A/L pairs are slightly more frequent in homologous alignments than K/E pairs:
 - $p_{AL} = 0.0044$, $p_{KE} = 0.0041$.
- But, A and L are more common amino acids:
 - $f_A = 0.074$, $f_L = 0.099$, $f_K = 0.058$, $f_E = 0.054$.

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Pro	-1	-2	-2	-1	-3	-1	-1	-2	-2	-3	-3	-1	-2	-4	7					
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BLOSUM_r: the matrix built from blocks with less than **r**% of similarity.

- E.g., BLOSUM62: built using sequences with less than 62% similarity (sequences with $\geq 62\%$ identity were clustered)
 - BLOSUM62 among the best for detecting most weak protein similarities.
 - Default matrix for protein BLAST.

Substitution matrix to measure similarity in sequence alignments

Substitution
matrix for DNA

A				
T				
G				
C				
	A	T	G	C

Making-up an arbitrary matrix by fixing the p_{ab} values \rightarrow directly describes what homologous alignments are expected to look like.

- The resulting score matrix is optimal for detecting alignments that match these target frequencies.

Say, the matrix should be optimized for finding 88% identity alignments.

- Assume that all mismatches are equiprobable, and composition of both alignments and background sequences is uniform at 25% for each nucleotide ($f_a, f_b = 0.25$ for all a, b). Then,
 - Four identities: $p_{aa} = 0.22$
 - 12 types of mismatch: $p_{ab} = 0.01$.
- If we set $\lambda = 1$, this gives +1.26 for a match and -1.83 for a mismatch.
- Setting $\lambda = 0.25$ and round off: we have a new scoring system of +4/-7.

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Substitution matrix to measure similarity in sequence alignments

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matrix for DNA

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G				
C				
	A	T	G	C

Given a scoring matrix, we can back calculate target frequencies if two conditions are met:

$$s(a,b) = \frac{1}{\lambda} \log \frac{p_{ab}}{f_a f_b}$$

1. It must have at least one positive score, and
2. The expected score for random sequence alignments must be negative.

True for most score matrices:

- These properties are necessary to make local sequence alignment algorithms like BLAST and Smith-Waterman work.
- Both conditions are met by definition for matrices derived as log-odds scores, except for the useless case of $p_{ab} = f_a f_b$ for all a, b .

Examples:

- FASTA & WU-BLASTN: arbitrary +5/−4 scoring system; Optimal for detecting alignments that are 65% identical.
- NCBI BLASTN: +1/−2 scoring system; Optimal for detecting alignments that are 95% identical.

How do we scale this up to search an entire sequence database?

Given a query sequence, and a large set of target sequences (millions), which target sequences (if any) are related to the query?

- Individual alignments need not be perfect: Once initial matches are found, they can fine-tune them later.
- Must be very fast.

Exploit the nature of the problem (most sequences will be unrelated to the query):

- If any match with % identity ≤ 90 is going to be rejected, can ignore sequences which don't have a stretch of 10 nucleotides in a row.
- Pre-screen sequences for common long stretches.
- Pre-process the database offline and index k-mers.

TITLE

CITED BY

YEAR


Basic local alignment search tool

SF Altschul, W Gish, W Miller, EW Myers, DJ Lipman

Journal of molecular biology 215 (3), 403-410

136003 *

1990


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Wed, 15 Nov 2017 16:00:00 EST

More BLAST news...

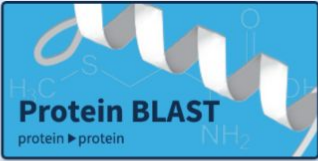
Web BLAST



Nucleotide BLAST
nucleotide ► nucleotide

blastx
translated nucleotide ► protein

tblastn
protein ► translated nucleotide



Protein BLAST
protein ► protein

BLAST Genomes

Human Mouse Rat Microbes

<https://www.ncbi.nlm.nih.gov/BLAST/>

Upcoming project deadline: Project topic due on Jan 29

- Briefly describe a project idea:
 - Title
 - Project advisor (if someone outside class)
 - 250-word abstract addressing the following 4 Qs:
 - What is the problem?
 - How is it addressed currently & what are the limitations?
 - What is your approach to addressing it & why is likely to be successful?
 - If successful, why does it matter (what is the impact)?
- I will post a link to a google form for submission.