

Class10: Structural Bioinformatics pt.1

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What is in the PDB database

The main repository of biomolecular structure info is the PDB < www.rcsb.org >.

Let's see what this database contains:

```
url <- "Data Export Summary.csv"
stats <- read.csv(url, row.names = 1)
stats
```

	X-ray	EM	NMR	Multiple.methods	Neutron	Other
Protein (only)	163,468	13,582	12,390	204	74	32
Protein/Oligosaccharide	9,437	2,287	34	8	2	0
Protein/NA	8,482	4,181	286	7	0	0
Nucleic acid (only)	2,800	132	1,488	14	3	1
Other	164	9	33	0	0	0
Oligosaccharide (only)	11	0	6	1	0	4
Total						
Protein (only)	189,750					
Protein/Oligosaccharide	11,768					
Protein/NA	12,956					
Nucleic acid (only)	4,438					
Other	206					
Oligosaccharide (only)	22					

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

The function `as.numeric` can turn strings into numbers

```
as.numeric(c(10,"1000"))
```

```
[1] 10 1000
```

```
as.numeric(stats$X.ray)
```

Warning: NAs introduced by coercion

```
[1] NA NA NA NA 164 11
```

How to get rid of the commas?

```
sum(as.numeric(sub(",", "", stats$X.ray)))
```

```
[1] 184362
```

Turning it into a function:

```
sumcomma <- function(x) {  
  sum(as.numeric(gsub(",", "", x)))  
}
```

```
n.total <- sumcomma(stats$Total)  
n.total
```

```
[1] 219140
```

```
apply(stats, 2, sumcomma)
```

X.ray	EM	NMR	Multiple.methods
184362	20191	14237	234
Neutron	Other	Total	
79	37	219140	

```
apply(stats, 2, sumcomma) / n.total
```

X.ray	EM	NMR	Multiple.methods
0.8412978005	0.0921374464	0.0649676006	0.0010678105
Neutron	Other	Total	
0.0003605001	0.0001688418	1.0000000000	

X.ray: $0.8412978005 * 100 = 84.12978005 \%$ EM: $0.0921374464 * 100 = 9.21374464 \%$

Q2: What proportion of structures in the PDB are protein?

```
189750/n.total * 100
```

```
[1] 86.58848
```

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

248805733 - 219140

```
219140/248805733 * 100
```

```
[1] 0.08807675
```

Visualizing the HIV-1 protease structure

Mol* (“molstar”) is a web-based molecular viewer <https://molstar.org/viewer/>





Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

The two H atoms are smaller than the resolution can catch, so it's not on the image.

Working with the bio3d package

```
library(bio3d)
```

```
pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
head(pdb$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10

2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40

	segid	elemsy	charge
1	<NA>	N	<NA>
2	<NA>	C	<NA>
3	<NA>	C	<NA>
4	<NA>	O	<NA>
5	<NA>	C	<NA>
6	<NA>	C	<NA>

```
pdbseq(pdb)[25]
```

```
25
"D"
```

Predicting functional motions of a single structure

We can do a bioinformatics prediction

```
pdb <- read.pdb("6s36")
```

Note: Accessing on-line PDB file

PDB has ALT records, taking A only, rm.alt=TRUE

```
pdb
```

```
Call: read.pdb(file = "6s36")
```

```
Total Models#: 1
```

```
Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)
```

```
Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
```

```
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

Non-protein/nucleic Atoms#: 244 (residues: 244)
Non-protein/nucleic resid values: [CL (3), HOH (238), MG (2), NA (1)]

Protein sequence:

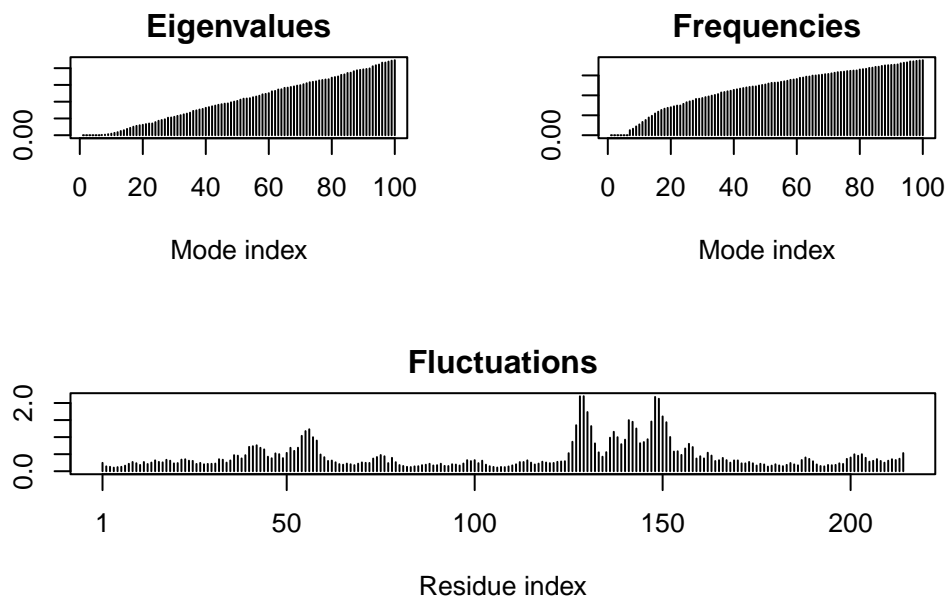
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLRAAVKSGSELGKQAKDIMDAGKLV
DELVIALVKERIAQEDCRNGFLLDGFPRTPQADAMKEAGINVDYVLEFDVPDELIVDKI
VGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG

+ attr: atom, xyz, seqres, helix, sheet,
calpha, remark, call

```
m <- nma(pdb)
```

Building Hessian... Done in 0.013 seconds.
Diagonalizing Hessian... Done in 0.259 seconds.

```
plot(m)
```



```
mktrj(m, file="adk_m7.pdb")
```