

Module 9: Variant Calling and Annotation (Part 2)

Key Concepts

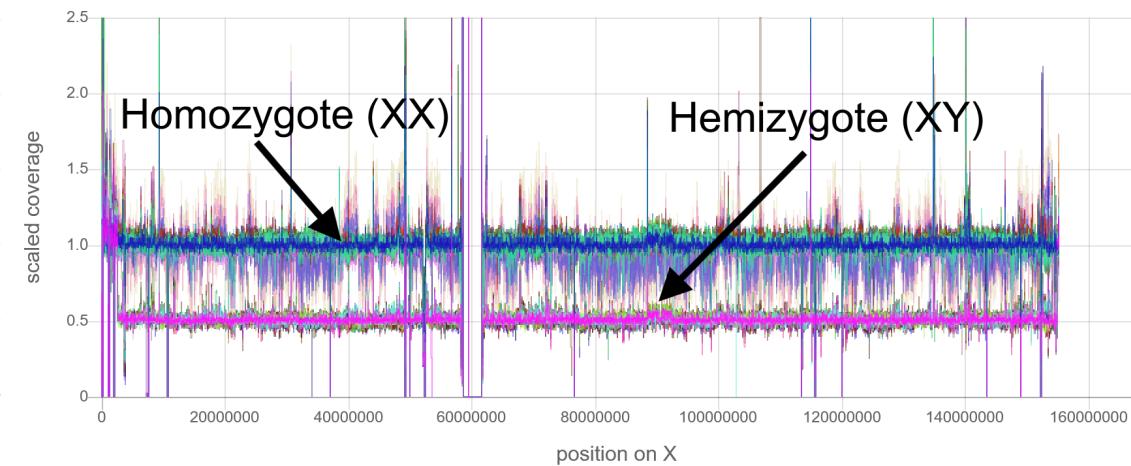
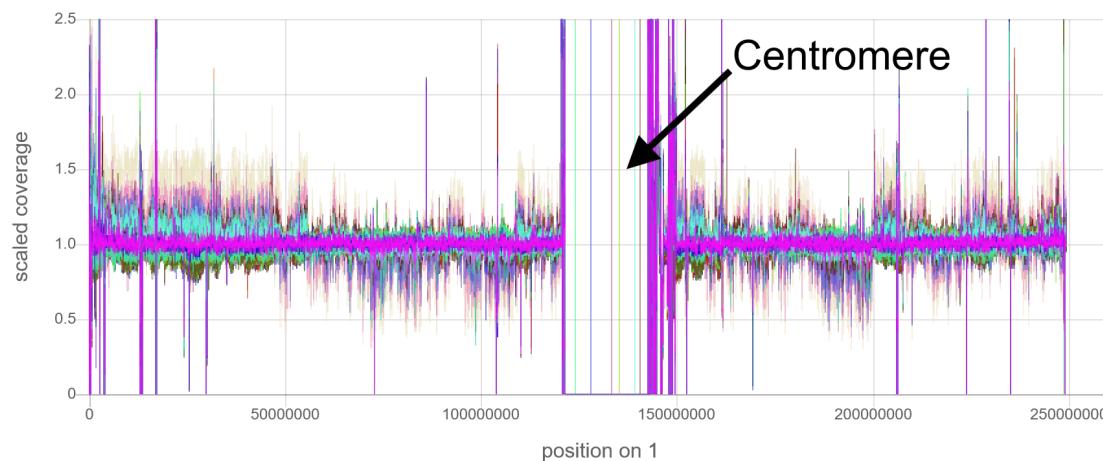
- Variant Calling
- Variant Annotation
- Types of input and output files
- Databases for clinical variants

Variant Calling: Structural Variants

indexcov

Tool: Indexcov ¹

- Quickly estimate coverage from a **WGS** bam or cram index.
- A long stretch with values of 1.5 would be a heterozygous duplication.



Variant Calling: Structural Variants



Tool: Indexcov¹

| | Description |
|-------------------------|---|
| Purpose | Quickly estimates genome-wide coverage using BAM file index data. |
| Usage | Identifies regions with abnormal coverage levels, flagging potential large deletions, duplications, or aneuploidies. |
| Key Feature | Provides a fast QC overview of structural changes without full-scale variant calling. |
| Assumptions/Limitations | Assumes that index data accurately reflects read depth; cannot detect balanced rearrangements or provide detailed breakpoint information. |

Variant Calling: Structural Variants

Tool: Manta²

| | Description |
|-------------|---|
| Purpose | Detects a variety of SVs, including deletions, insertions, inversions, and translocations. |
| Usage | <ul style="list-style-type: none">- Joint analysis of small sets of diploid individuals (where 'small' means family-scale <10 samples)- Subtractive analysis of a matched tumor/normal sample pair- Analysis of an individual tumor sample |
| Key Feature | High sensitivity and specificity; suitable for clinical diagnostics and research. |

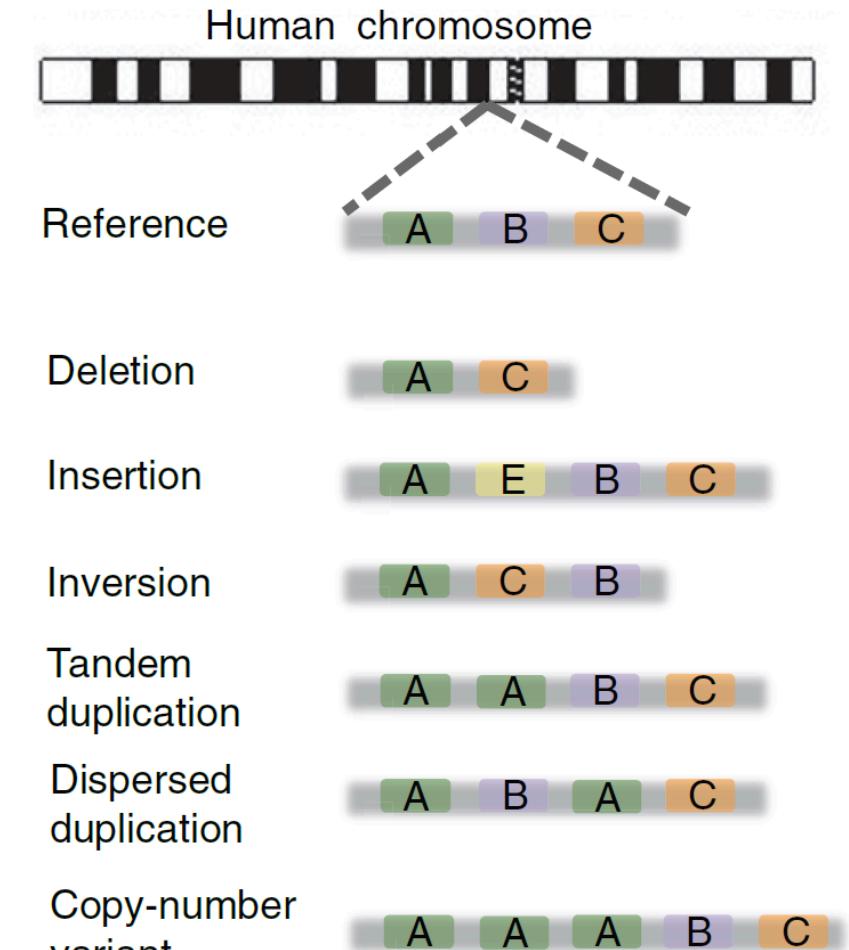
Variant Calling: Structural Variants

Tool: Manta²

Assumptions/Limitations: Cannot detect

- small inversions (<200bp),
- fully-assembled large insertions > 2 x read-pair fragment size,
- dispersed duplications

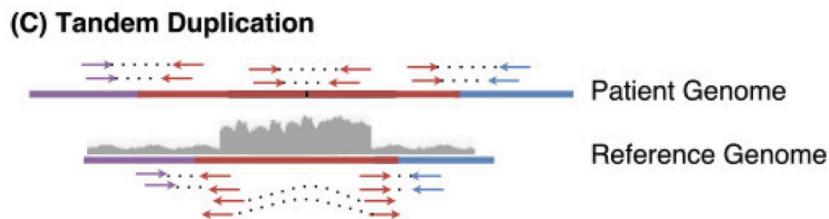
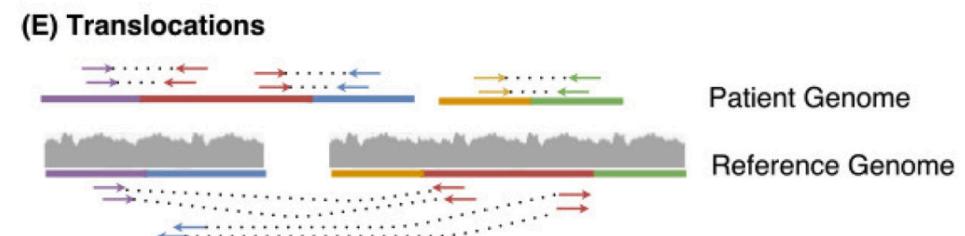
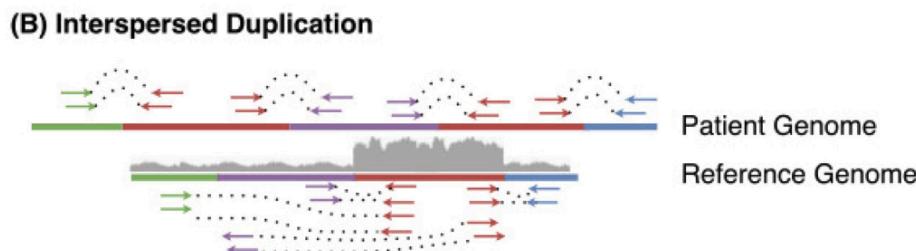
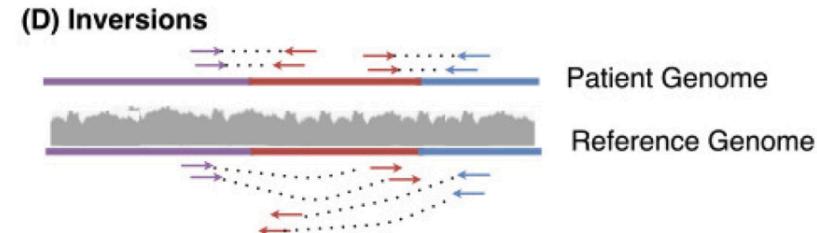
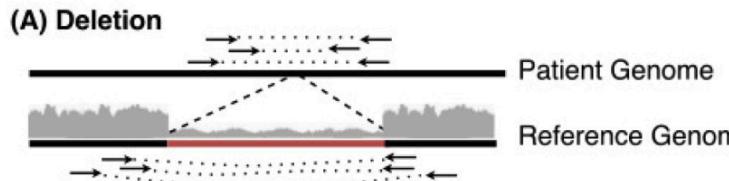
Does not support mate-pair libraries from public data sets



Variant Calling: Structural Variants

Tool: TIDDIT³

- Detects many structural variants



Variant Calling: Structural Variants

Tool: TIDDIT³

| | Description |
|-------------------------|---|
| Usage | <ul style="list-style-type: none">- Uses discordant pairs and split reads to detect the genomic location of structural variants- Uses the read depth information for classification and quality assessment of the variants |
| Key Feature | <ul style="list-style-type: none">- Distributed with a database functionality SVDB (Structural Variant DataBase) to create structural variant frequency databases- Uses SVDB to call and evaluate rare disease causing structural variants |
| Assumptions/Limitations | Does not perform well on small variants but performs really well on large variants , especially balanced variants (translocations) |

Variant Calling: Structural Variants

Tool: TIDDIT³

Table 2. Sensitivity and precision of the structural variant callers on a simulated dataset consisting of 6000 variants of each variant type. The variants were simulated using SVsim and Simseq.

| Caller | SV detection on simulated data | | | | |
|----------|--------------------------------|-----------|----------------|-----------|--|
| | Sensitivity | Precision | Sensitivity | Precision | |
| | Deletions | | Duplications | | |
| TIDDIT | 0.96 | 0.99 | 0.96 | 0.99 | |
| CNVnator | 0.9 | 0.92 | 0.86 | 0.91 | |
| Delly | 0.94 | 1 | 0.95 | 1 | |
| Fermikit | 0.41 | 1 | 0.33 | 1 | |
| Lumpy | 0.95 | 0.97 | 0.95 | 1 | |
| Manta | 0.95 | 1 | 0.95 | 1 | |
| | Inversions | | Translocations | | |
| TIDDIT | 0.97 | 0.99 | 0.92 | 0.93 | |
| Delly | 0.94 | 1 | 0.87 | 0.95 | |
| Fermikit | 0.35 | 1 | 0.26 | 0.99 | |
| Lumpy | 0.5 | 1 | 0.87 | 0.9 | |
| Manta | 0.95 | 1 | 0.88 | 0.95 | |

Table 4. CPU hour consumption of the structural variant callers. Each caller except Fermikit was run on a single core of a Intel Xeon E5-2660 CPU. Fermikit was run on 16 CPU cores. The CPU hour consumption of the Simseq data is reported as the median time consumption across the four Simseq samples.

| CPU hour consumption on SV calling | | | |
|------------------------------------|---------|-------|--------|
| Caller | NA12878 | HG002 | Simseq |
| TIDDIT | 2 | 1 | 1 |
| CNVnator | 2 | 1 | 1 |
| Delly | 30 | 15 | 7 |
| Fermikit | 640 | 120 | 15 |
| Lumpy | 45 | 2 | 7 |
| Manta | 3 | NA | 1 |

Variant Calling: Copy Number Variation (CNV)

Tool: ASCAT

| | Description |
|-------------------------|---|
| Purpose | Allele-specific copy number analysis of tumors. |
| How | Uses SNP allele frequencies to estimate copy number changes while accounting for tumor purity and ploidy in complex tumor genomes. |
| Key Feature | Provides precise CNV calls in heterogeneous tumor samples. |
| Assumptions/Limitations | <ul style="list-style-type: none">- Assumes availability of high-quality SNP data and matched normal samples- May be less reliable with low tumor purity or highly rearranged genomes. |

Variant Calling: Copy Number Variation (CNV)

Tool: CNVKit

| | Description |
|-------------------------|---|
| Purpose | Detects copy number variations from targeted sequencing data (exomes, gene panels). |
| How | Combines on-target and off-target read data to create copy number profiles and visualizations, with comprehensive normalization and segmentation. |
| Key Feature | Optimized for clinical sequencing data with user-friendly visual outputs. |
| Assumptions/Limitations | <ul style="list-style-type: none">- Assumes that off-target reads provide sufficient coverage- May be sensitive to capture biases and not as robust when applied to whole-genome data without modifications. |

Variant Calling: Copy Number Variation (CNV)

Tool: Control-FREEC

| | Description |
|-------------------------|--|
| Purpose | Detects CNVs and allelic imbalances from whole-genome or exome sequencing data. |
| How | Normalizes read depth (accounting for GC-content) and segments the genome. |
| Key Feature | You can use this with or without a matched normal sample |
| Assumptions/Limitations | <ul style="list-style-type: none">- Assumes that read depth variations correlate with copy number changes- May struggle in regions with extreme GC content or |

Variant Calling: Microsatellite Instability

Tool: MSIsensorPro

| | Description |
|-------------------------|---|
| Purpose | Detects microsatellite instability (MSI) by comparing microsatellite regions between tumor and normal samples. |
| How | Analyzes repeat length distributions at defined microsatellite loci to compute an MSI score that reflects the level of instability. |
| Key Feature | Provides a sensitive and specific measure of MSI status |
| Assumptions/Limitations | <ul style="list-style-type: none">- Assumes sufficient coverage at microsatellite loci and availability of matched normal data (or a reliable reference)- Sensitivity may be impacted by sequencing quality. |

Variant Annotation Tools

| | snpEff | VEP (Variant Effect Predictor) | bcftools |
|----------|--|---|---|
| Purpose | Predicts variant effects (e.g., missense, nonsense, frameshift) on genes. | Provides comprehensive variant annotations using Ensembl data. | Primarily a variant processing tool that also offers basic annotation capabilities. |
| Features | Uses pre-built databases for many organisms; highly configurable for custom annotations. | Annotates variants with gene, transcript, and regulatory information; integrates population frequency data and supports custom plugins. | Can add custom INFO tags and combine external annotations to VCF files. |
| Usage | Ideal for quick, high-throughput annotation of SNPs and small indels. | Suitable when extensive, detailed annotation is required and integration with Ensembl resources is desired. | Useful for post-calling processing and integrating annotations from other tools into your workflow. |

Variant Calling: File Types

PON (Panel of Normals)

- Aggregated file (often in VCF format) created from multiple normal samples
- Used in somatic variant calling to filter out false positives by comparing tumor data against a baseline.
- Improves specificity by removing systematic errors.
- **Limitations:** Quality depends on the number and quality of normal samples; may not capture all artifacts.

Variant Calling and Annotation: File Types

| Property | VCF (Variant Call Format) | MAF (Mutation Annotation Format) |
|-----------------------|---|--|
| Description | Text-based format for storing variant calls. | Tab-delimited format, commonly used in cancer genomics. |
| Key Fields / Features | CHROM, POS, ID, REF, ALT, QUAL, FILTER, INFO, FORMAT. | Contains curated annotations (e.g., gene name, variant classification, sample-specific details). |
| Usage | Widely used for both germline and somatic variant calls. | Ideal for generating human-readable, standardized reports of somatic mutations. |
| Strengths | Extensible with custom annotations; standard format in many pipelines. | Focused on detailed mutation annotation; facilitates downstream interpretation. |
| Limitations | Can become large with extensive annotations; requires proper indexing for efficient querying. | Less common for germline variants; may need conversion from VCF. |

File Types in Variant Calling and Annotation

- **Alignment Files (BAM/CRAM):** For mapping reads and ensuring quality alignments.
 - BAM is standard but large; CRAM offers better compression.
- **Variant Files (VCF/MAF):** For storing and interpreting variant calls.
 - VCF and MAF provide variant-level annotations with MAF offering more cancer-specific details.
- **PON:** For filtering out recurrent artifacts, especially in somatic pipelines.

Variant Databases

| gnomAD | |
|-------------|---|
| Description | Aggregates large-scale exome and genome sequencing data from diverse populations. |
| Purpose | Provides allele frequency data to distinguish common variants from rare variants. |
| Usage | Used for filtering variants based on population frequency; supports studies in population genetics and variant prioritization. |
| Key Points | <ul style="list-style-type: none">- Extensive dataset with rigorous quality filters.- Continuously updated with diverse sample representation. |
| Limitations | <ul style="list-style-type: none">- Lacks detailed clinical annotation.- May not capture all sub-population specific variants. |

Variant Databases

| | ClinVar |
|-------------|---|
| Description | A public archive aggregating information about the relationships between human variants and phenotypes. |
| Purpose | Provides clinical significance and supporting evidence for genetic variants. |
| Usage | Essential for clinical interpretation; used to assess variant pathogenicity and inform diagnostics. |
| Key Points | <ul style="list-style-type: none">- Curated submissions from clinical laboratories and research groups.- Integrates multiple clinical assertions and evidence. |
| Limitations | <ul style="list-style-type: none">- May contain conflicting interpretations.- Focused primarily on clinically relevant variants. |

Variant Databases

| dbGaP (Database of Genotypes and Phenotypes) | |
|--|--|
| Description | A controlled-access repository for genotype and phenotype data from a wide array of studies. |
| Purpose | Facilitates research by providing access to raw and processed genomic and phenotypic data. |
| Usage | Used for large-scale genomic studies, variant-disease association research, and validation of genetic findings. |
| Key Points | <ul style="list-style-type: none">- Contains data from diverse study cohorts.- Access is governed by ethical and legal restrictions. |
| Limitations | <ul style="list-style-type: none">- Data access is restricted and may require approval.- May require additional processing and harmonization. |