The genetic architecture of target-site resistance to pyrethroid insecticides in the African malaria vectors *Anopheles gambiae* and *Anopheles coluzzii*

Chris S. Clarkson, Alistair Miles, …, and the *Anopheles gambiae* 1000 Genomes Consortium

**Abstract**

Resistance to pyrethroid insecticides is a major concern for malaria vector control, because these are the only compounds currently approved for use in insecticide treated bed-nets (ITNs) and are also widely used for indoor residual spraying (IRS). Pyrethroids target the voltage-gated sodium channel (VGSC), an essential component of the mosquito nervous system, but mutations in the *Vgsc* gene can disrupt the activity of these insecticides, inducing a “knock-down resistance” (*kdr*) phenotype. Here we use Illumina sequence data from phase 1 of the *Anopheles gambiae* 1000 Genomes Project (Ag1000G) to provide a comprehensive account of genetic variation at the *Vgsc* locus in mosquito populations from 8 African countries. In addition to three known variants that alter the protein-coding sequence of the gene, we describe 19 previously unknown non-synonymous variants at appreciable frequency in one or more populations. We report a highly significant enrichment for non-synonymous variation occurring in linkage with the known L995F resistance allele, indicating the evolution of multiple secondary variants that may either enhance or compensate for the L995F phenotype. We also describe a possible resistance variant I1527T, which occurs in linkage with two non-synonymous variants in codon 402. We use an analysis of haplotype sharing on the flanks of the gene to refine our understanding of the origins and geographical spread of resistance variants within the gene. We characterise 11 distinct lineages, each of which carries one or more resistance alleles and appears to be undergoing a rapid and recent expansion in one or more populations. We provide preliminary evidence that the most successful and widespread resistance lineage (F1) originates in West Africa and has subsequently spread to countries in Central and Southern Africa. We also reconstruct a putative ancestral haplotype for each lineage, and analyse patterns of recombination to show that lineages are unrelated and thus represent independent outbreaks of resistance. Our data demonstrate that the molecular basis of pyrethroid resistance in African malaria vectors is more complex than previously appreciated, and provide a foundation for the development of new genetic tools to predict resistance phenotype and track the further spread of resistance.

# Introduction

It has been estimated that 663 million cases of malaria were averted in Africa between 2000 and 2015 due to public health interventions, of which 68% were prevented through the use of insecticide-treated bed-nets (ITNs) and @@N% through indoor residual spraying (IRS) of insecticides @@REF. However, over this same period, insecticide resistance has become increasingly prevalent in malaria vector populations @@REF. Four chemical classes of insecticides – organophosphates, carbamates, pyrethroids and organochlorines – are licensed for use in public health, but only pyrethroids are approved by the World Health Organisation (WHO) for use in ITNs. With pyrethroids also commonly used for IRS as well as in agriculture, there is a strong pressure on mosquito populations to evolve molecular mechanisms of pyrethroid resistance. There is evidence that pyrethroid resistance has a direct impact on the effectiveness of ITNs (@@REF) and IRS (@@REF), although assessing the impact is difficult and has been hampered by the fact that pyrethroid resistance is now so pervasive that it is nearly impossible to find fully susceptible mosquito populations to use as controls (@@REF). Nevertheless, the position of the WHO remains that insecticide resistance poses a grave threat to the future of malaria control in Africa (@@REF GPIRM). Improvements are needed in our ability to monitor resistance, and gaps must be filled in our knowledge of the molecular mechanisms of resistance.

The voltage-gated sodium channel (VGSC) is the physiological target of pyrethroids, and of the organochlorine DDT. The VGSC protein is integral to the insect nervous system, enabling transmission of nerve impulses. Both pyrethroids and DDT have a similar mode of action, binding to sites within the protein channel and causing it to remain open, causing paralysis (“knock-down”) and then death (@@REFs). However, amino acid substitutions at key positions within the channel can alter this interaction between the channel and the insecticide molecule, and thereby substantially increase the dosage of insecticide required to induce knock-down (@@REFs). If this tolerance exceeds the dosage present in ITNs or on indoor surfaces following IRS, then these interventions may be rendered ineffective. In the African malaria vectors *Anopheles gambiae* and *Anopheles coluzzii*, three substitutions have been found in natural populations and shown to induce pyrethroid and DDT resistance. Two of these substitutions occur in codon 995[[1]](#footnote-1), with the Leucine→Phenylalanine (L995F) substitution prevalent in West and Central Africa, and the Leucine→Serine (L995S) substitution found in Central and East Africa (@@REFs). A third variant N1570Y has been found in association with L995F in West and Central Africa and shown to further increase resistance above L995F alone (@@REF).

Target-site resistance to pyrethroids and DDT has also been studied in other insect species that experience insecticide pressure, including other disease vectors as well as domestic and crop pests (@@REF). Because of its essential function, the VGSC protein is highly conserved across these species, and knowledge gained from one species is relevant to another. Many resistance-associated variants have been described in these other species, and thus there are many possible amino acid substitutions that could induce a resistance phenotype in malaria vectors. Some of these variants are within the channel and thus may directly interfere with insecticide binding, however others are in regions of the protein that are internal to the cell and thus must alter the resistance phenotype indirectly. In the absence of any insecticide pressure, it has been shown that substitutions within the channel cause a reduction in fitness, and therefore substitutions within internal domains may provide compensatory changes to the gating dynamics of the channel (@@REF). Although only three variants have been found in *An. gambiae* or *An. coluzzii*, most previous studies have performed targeted sequencing of small regions within the gene, and there has been no comprehensive survey of variation across the entire gene in multiple populations.

@@TODO para on spread and origins

1. Codon numbering is given relative to transcript @@TODO as defined in the AgamP4.@@N gene annotations. A mapping of codon numbering from *An. gambiae* @@TODO transcript to *Musca domestica* @@TODO transcript is given in Table 1. [↑](#footnote-ref-1)