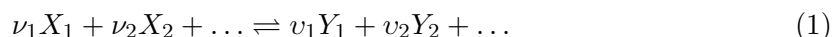


Note on Using Dissipation Tracking in MEDYAN

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To use the dissipation tracking feature to produce thermodynamically consistent results, some care needs to be taken when parameterizing the system. To understand how this parameterization has been done previously, users are encouraged to read the papers “Quantifying Dissipation in Actomyosin Networks” by Carlos Floyd, Garegin Papoian, and Christopher Jarzynski in *Interface Focus* (<https://doi.org/10.1098/rsfs.2018.0078>), and the Supplementary Material within, as well as “A Discrete Approximation to Gibbs Free Energy of Chemical Reactions is Needed for Accurately Calculating Entropy Production in Mesoscopic Simulations” by the same authors (<https://arxiv.org/abs/1901.10520v1>). This parameterization process consists mainly of choosing values for ΔG^0 to use in the input file. This process consists first of determining a good value based on the literature expressions, and next converting it into a form that will produce unbiased results.

For a reaction of the general form



we define X_i as the reactant species, Y_i as the product species, and ν_i and v_j as their stoichiometric coefficients, and where the rate constant is k_+ to the right and k_- to the left. The “stoichiometric difference” is defined as

$$\sigma = \sum_{j \in P} v_j - \sum_{i \in R} \nu_i \quad (2)$$

where P is the set of products and R is the set of reactants. We further define the conversion factor between the copy number of species i , N_i , and its concentration C_i ,

$$\Theta = N_{\text{Av}} V \quad (3)$$

where N_{Av} is Avogadro’s number, V is the volume of the compartment where the reaction occurs, and we have $N_i = C_i \Theta$. The formula for change in Gibbs free energy during this reaction used in MEDYAN is

$$\Delta G = \Delta G^0 - \sigma k_B T \log \Theta - \sigma k_B T + k_B T \log \tilde{Q} \quad (4)$$

where

$$\tilde{Q} = \prod_{i \in R} \frac{(N_i - \nu_i)^{N_i - \nu_i}}{N_i^{N_i}} \prod_{j \in P} \frac{(N_j + v_j)^{N_j + v_j}}{N_j^{N_j}}. \quad (5)$$

We refer the reader to the “A Discrete Approximation is Needed...” paper mentioned above for a derivation of this expression.

In MEDYAN the last term in Equation 4 is computed based on the instantaneous compartment concentrations of the reaction species when the reaction occurs, and the first three terms on the right hand side are provided as a constant user input called `DELGZERO`, thus

$$\text{DELGZERO} = \Delta G^0 - \sigma k_B T \log \Theta - \sigma k_B T. \quad (6)$$

So, once the user determines ΔG^0 based on their parameterization (which is discussed next), this value should be used to set the `DELGZERO` parameter in chemistry input value based on the value of σ and Θ for the reaction.

For balanced reactions, in which the number of products is equal to the number of reactants, $\sigma = 0$ and $\text{DELGZERO} = \Delta G^0$. Polymerization and linker binding reactions are assumed to be of the form $\sigma = -1$, whereas depolymerization and unbinding reactions have $\sigma = 1$. For reaction in which ATP, ADP, or Pi are implicitly involved as products or reactants (for instance a nucleotide exchange reaction converting ADP-bound G-actin to ATP-bound G-actin), the assumed constant concentrations of these molecules should be incorporated into the value of `DELGZERO`. For greater detail on these concerns, please consult the Supplementary Material in “Quantifying Dissipation in Actomyosin Networks.”

To determine values of ΔG^0 for reversible reactions, the formula

$$\Delta G^0 = k_B T \ln K_{\text{eq}} = k_B T \ln \frac{k_-}{k_+}. \quad (7)$$

may be used. For effectively irreversible reactions (i.e. when is k_- too small to be measured), then ΔG^0 is often reported directly. When no value for ΔG^0 can be found in the literature, then indirect methods of determining it for a certain reaction may be used. The method entails constructing closed loops of reactions such that traversing the loops results in no net change in the copy numbers of any species. For such a loop the net $\Delta G^0 = 0$, which provides a constraint on the unknown value in terms of the known ones. An example of this method is illustrated in the Supplementary Material of “Quantifying Dissipation in Actomyosin Networks.”

An additional issue to be aware of is that the myosin motor step size for actomyosin networks should be set as close to physiological value ($\sim 6 \text{ nm}$ for NMIIA) as possible to prevent the motors from always taking large steps and increasing their stretching energy more than the chemical energy of the ATP molecules they consumed. The binding sites per cylinder which determines the motor step size is simultaneously constrained by the cross-linker binding distance ($\sim 30 \text{ nm}$ for α -actinin). This need motivates the `DIFBINDING` feature, which allows the motors and cross-linkers to have different a number of binding sites per cylinder. This feature should be used to produce reasonable results, however it currently is only supported using the original neighborlist implementation, not the SIMD or hybrid neighborlist.