

FML Assignment_4 CLUSTERING

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Statement : Directions - An equities analyst is studying the pharmaceutical industry and would like your help in exploring and understanding the financial data collected by her firm. Her main objective is to understand the structure of the pharmaceutical industry using some basic financial measures. Financial data gathered on 21 firms in the pharmaceutical industry are available in the file Pharmaceuticals.csv Download Pharmaceuticals.csv. For each firm, the following variables are recorded:

Market capitalization (in billions of dollars) Beta Price/earnings ratio Return on equity Return on assets Asset turnover Leverage Estimated revenue growth Net profit margin Median recommendation (across major brokerages) Location of firm's headquarters Stock exchange on which the firm is listed Use cluster analysis to explore and analyze the given dataset as follows:

Use only the numerical variables (1 to 9) to cluster the 21 firms. Justify the various choices made in conducting the cluster analysis, such as weights for different variables, the specific clustering algorithm(s) used, the number of clusters formed, and so on. Interpret the clusters with respect to the numerical variables used in forming the clusters. Is there a pattern in the clusters with respect to the numerical variables (10 to 12)? (those not used in forming the clusters) Provide an appropriate name for each cluster using any or all of the variables in the dataset.

Summary : As per my observation The provided statement describes a situation in which a financial analyst for stocks is examining information from 21 pharmaceutical companies. The goal is to use numerical variable cluster analysis to comprehend the structure of the pharmaceutical industry. Market capitalization, beta, price/earnings ratio, return on equity, return on assets, asset turnover, leverage, projected revenue growth, and net profit margin are just a few of the financial metrics included in the data. So in this R I mentioned various library tools and formulated to get plot diagrams. Since each

explanation mentioned on the codes respectively

```
# Let's add the required library
```

```
library(class)
library(caret)
```

```
## Loading required package: ggplot2
```

```
## Warning: package 'ggplot2' was built under R version 4.3.2
```

```
## Loading required package: lattice
```

```
library(e1071)
library(dbscan)
```

```
## Warning: package 'dbscan' was built under R version 4.3.2
```

```
##
## Attaching package: 'dbscan'
```

```
## The following object is masked from 'package:stats':
##
##   as.dendrogram
```

```
library(tidyverse)
```

```
## — Attaching core tidyverse packages ————— tidyverse 2.0.0 —
## ✓ dplyr      1.1.3      ✓ readr      2.1.4
## ✓ forcats    1.0.0      ✓ stringr    1.5.0
## ✓ lubridate  1.9.2      ✓ tibble     3.2.1
## ✓ purrr      1.0.2      ✓ tidyr      1.3.0
```

```
## — Conflicts ————— tidyverse_conflicts() —
## ✗ dplyr::filter() masks stats::filter()
## ✗ dplyr::lag()     masks stats::lag()
## ✗ purrr::lift()    masks caret::lift()
## i Use the conflicted package (<http://conflicted.r-lib.org/>) to force all conflicts to be
come errors
```

```
library(ISLR)
library(factoextra)
```

```
## Warning: package 'factoextra' was built under R version 4.3.2
```

```
## Welcome! Want to learn more? See two factoextra-related books at https://goo.gl/ve3WBa
```

```
library(fpc)
```

```
## Warning: package 'fpc' was built under R version 4.3.2
```

```
##  
## Attaching package: 'fpc'  
##  
## The following object is masked from 'package:dbscan':  
##  
##      dbscan
```

Each library serves a specific purpose and provides functions and tools that can be used for various tasks in data analysis, machine learning, and statistics.

Let import the CSV file from the directory

```
# Import CSV file  
pharma.data <- read.csv("C:/Users/Vishal/Downloads/Pharmaceuticals.csv")  
dim(pharma.data)
```

```
## [1] 21 14
```

```
t(t(names(pharma.data)))# The 't' function creates a transpose of the dataframe
```

```
##      [,1]  
## [1,] "Symbol"  
## [2,] "Name"  
## [3,] "Market_Cap"  
## [4,] "Beta"  
## [5,] "PE_Ratio"  
## [6,] "ROE"  
## [7,] "ROA"  
## [8,] "Asset_Turnover"  
## [9,] "Leverage"  
## [10,] "Rev_Growth"  
## [11,] "Net_Profit_Margin"  
## [12,] "Median_Recommendation"  
## [13,] "Location"  
## [14,] "Exchange"
```

Interpretation : Checking the dimensions of the data frame gives an idea of the number of observations (rows) and variables (columns). Transposing the column names might be done to display them in a different format or orientation

Dropping the columns which not required for clustering

```
pharma.data <- pharma.data[ , -c(1,2,12,13,14)]# 1 and 5 are the indexes for columns ID and ZIP
dim(pharma.data)
```

```
## [1] 21 9
```

```
summary(pharma.data)
```

```
##      Market_Cap      Beta      PE_Ratio      ROE
## Min.   : 0.41   Min.   :0.1800   Min.    : 3.60   Min.    : 3.9
## 1st Qu.: 6.30   1st Qu.:0.3500   1st Qu.:18.90   1st Qu.:14.9
## Median :48.19   Median :0.4600   Median :21.50   Median :22.6
## Mean   :57.65   Mean   :0.5257   Mean    :25.46   Mean    :25.8
## 3rd Qu.:73.84   3rd Qu.:0.6500   3rd Qu.:27.90   3rd Qu.:31.0
## Max.   :199.47   Max.    :1.1100   Max.    :82.50   Max.    :62.9
##      ROA      Asset_Turnover      Leverage      Rev_Growth
## Min.   : 1.40   Min.    :0.3   Min.    :0.0000   Min.    :-3.17
## 1st Qu.: 5.70   1st Qu.:0.6   1st Qu.:0.1600   1st Qu.: 6.38
## Median :11.20   Median :0.6   Median :0.3400   Median : 9.37
## Mean   :10.51   Mean    :0.7   Mean    :0.5857   Mean   :13.37
## 3rd Qu.:15.00   3rd Qu.:0.9   3rd Qu.:0.6000   3rd Qu.:21.87
## Max.   :20.30   Max.    :1.1   Max.    :3.5100   Max.    :34.21
## Net_Profit_Margin
## Min.   : 2.6
## 1st Qu.:11.2
## Median :16.1
## Mean   :15.7
## 3rd Qu.:21.1
## Max.   :25.5
```

```
t(t(names(pharma.data)))
```

```
##      [,1]
## [1,] "Market_Cap"
## [2,] "Beta"
## [3,] "PE_Ratio"
## [4,] "ROE"
## [5,] "ROA"
## [6,] "Asset_Turnover"
## [7,] "Leverage"
## [8,] "Rev_Growth"
## [9,] "Net_Profit_Margin"
```

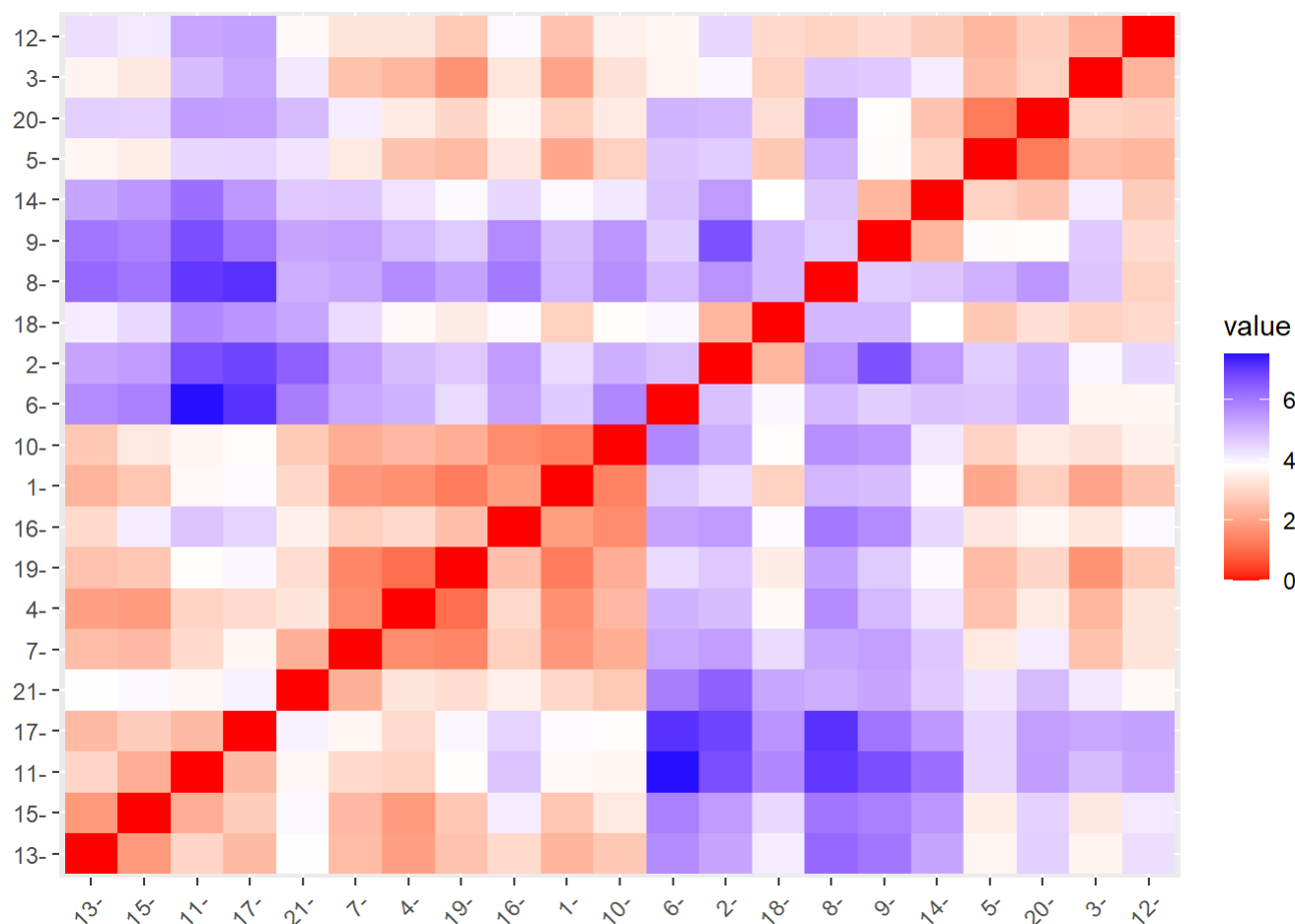
Interpretation : The removal of columns with indexes 1, 2, 12, 13, and 14 may be due to the fact that the information in those columns is duplicated, unnecessary for the analysis, or determined to be unrelated to the work at hand. A brief description of the distribution and central tendency of the remaining variables in the altered data frame can be obtained by computing summary statistics. This can be useful in comprehending the data's properties.

#kmeans

```
pharma.data1 <- scale(pharma.data)
head(pharma.data1)
```

```
##      Market_Cap      Beta  PE_Ratio      ROE      ROA Asset_Turnover
## [1,]  0.1840960 -0.80125356 -0.04671323  0.04009035  0.2416121    0.0000000
## [2,] -0.8544181 -0.45070513  3.49706911 -0.85483986 -0.9422871    0.9225312
## [3,] -0.8762600 -0.25595600 -0.29195768 -0.72225761 -0.5100700    0.9225312
## [4,]  0.1702742 -0.02225704 -0.24290879  0.10638147  0.9181259    0.9225312
## [5,] -0.1790256 -0.80125356 -0.32874435 -0.26484883 -0.5664461   -0.4612656
## [6,] -0.6953818  2.27578267  0.14948233 -1.45146000 -1.7127612   -0.4612656
##      Leverage Rev_Growth Net_Profit_Margin
## [1,] -0.2120979 -0.5277675      0.06168225
## [2,]  0.0182843 -0.3811391     -1.55366706
## [3,] -0.4040831 -0.5721181     -0.68503583
## [4,] -0.7496565  0.1474473      0.35122600
## [5,] -0.3144900  1.2163867     -0.42597037
## [6,] -0.7496565 -1.4971443     -1.99560225
```

```
distance <- get_dist(pharma.data1)
fviz_dist(distance)
```



Interpretation : # Interpretation : The pharmacy's columns are standardized by applying the scale function. data frame of data. Standardization entails scaling by the standard deviation and centering the variables by deducting the mean. The code probably belongs to an exploratory study or is used as a warm-up for a clustering task, where knowing the separations between data points are crucial. The particular needs of the study or algorithm being utilized to determine which standardization and distance calculation needs to apply.

Consider k=3

```
set.seed(159)
k <- 3
k3 <- kmeans(pharma.data1, centers = k, nstart=21)
k3$centers
```

```
## Market_Cap      Beta  PE_Ratio      ROE      ROA Asset_Turnover
## 1  0.6733825 -0.3586419 -0.2763512  0.6565978  0.8344159    0.4612656
## 2 -0.8261772  0.4775991 -0.3696184 -0.5631589 -0.8514589   -0.9994088
## 3 -0.6125361  0.2698666  1.3143935 -0.9609057 -1.0174553    0.2306328
##      Leverage Rev_Growth Net_Profit_Margin
## 1 -0.3331068 -0.2902163      0.6823310
## 2  0.8502201  0.9158889      -0.3319956
## 3 -0.3592866 -0.5757385      -1.3784169
```

```
k3$size
```

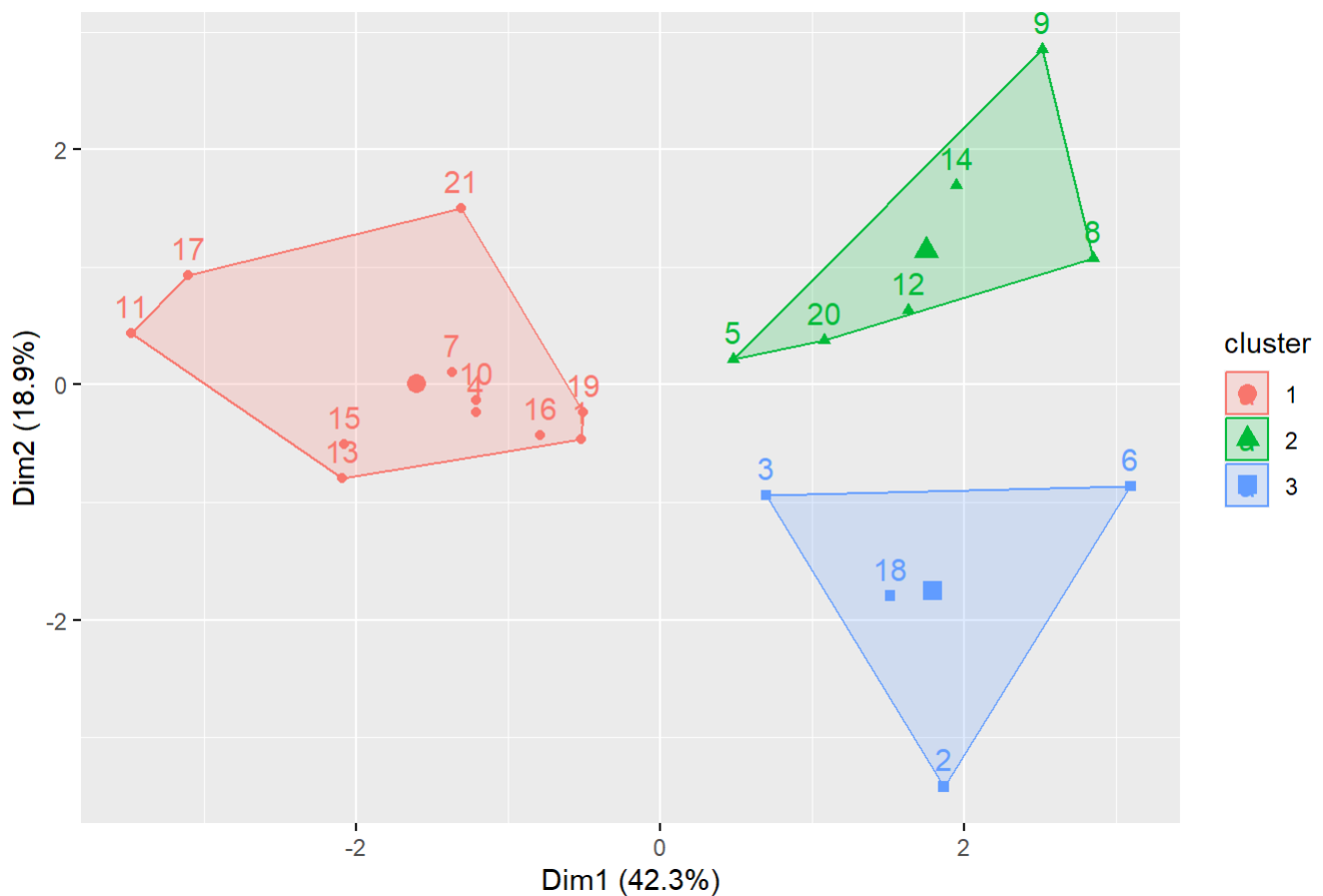
```
## [1] 11 6 4
```

```
k3$cluster
```

```
## [1] 1 3 3 1 2 3 1 2 2 1 1 2 1 2 1 1 1 3 1 2 1
```

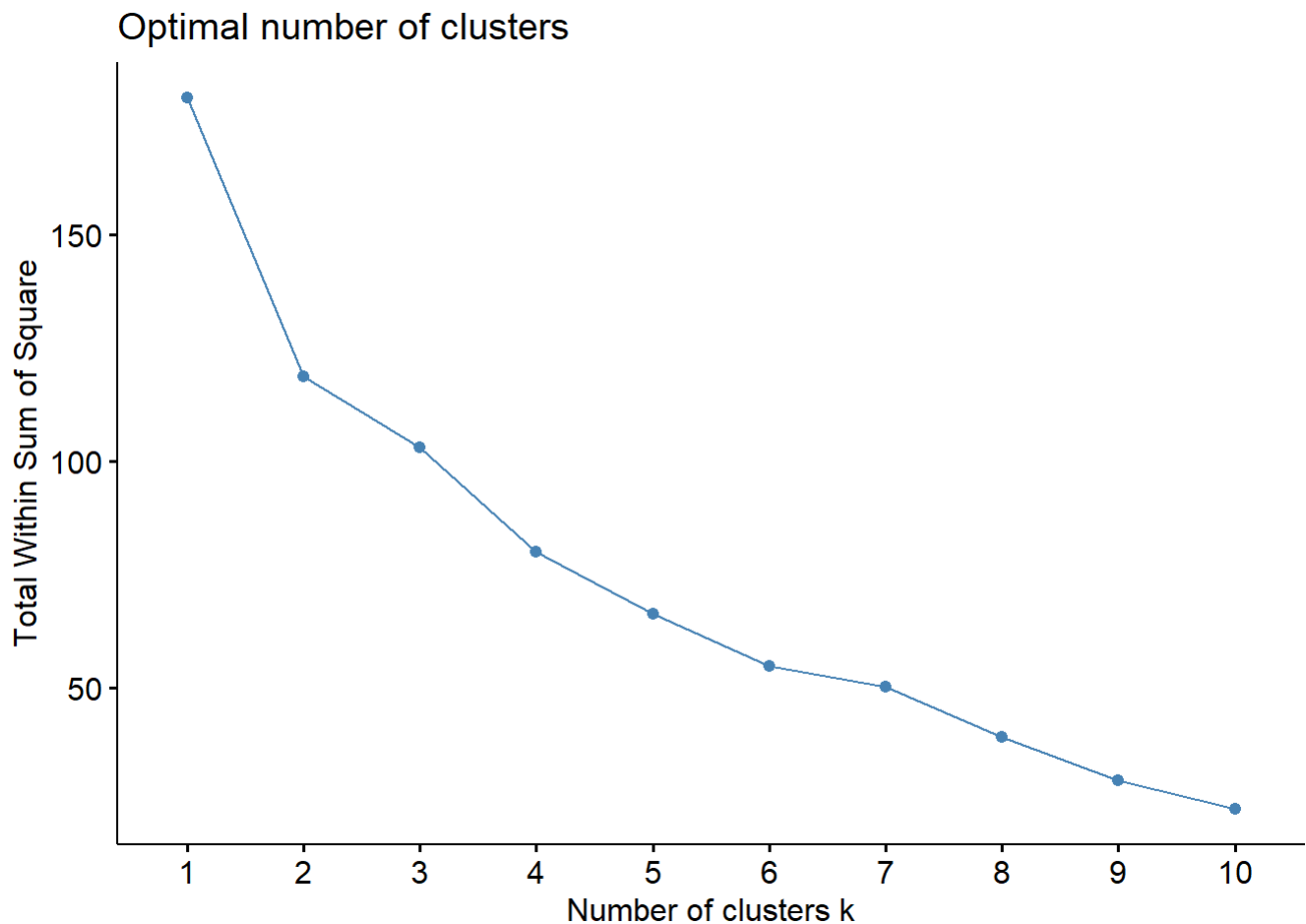
```
fviz_cluster(k3, pharma.data1)
```

Cluster plot



Interpretation : Using three clusters, the code applies k-means clustering to the standardized pharmaceutical data. The information about the cluster centers, sizes, and assignments is extracted and shown in the following lines. In order to examine the detected clusters in the data graphically, a visualization is created at the end. The assumption or previous knowledge that the data may be efficiently categorized into three different clusters is the basis for the selection of three clusters ($k = 3$).

```
fviz_nbclust(pharma.data1, kmeans, method = "wss")
```



Interpretation : The plot usually looks like an elbow, and the "elbow" represents a good candidate for the optimal number of clusters. This is the point where the rate of decline in the within-cluster sum of squares slows down, indicating that performance decreases as the within-cluster variance decreases as you increase the number of clusters. Looking at this plot, you can visually identify the point where adding more clusters improves the reduction of within-cluster variability. "elbow" is often considered a reasonable choice for the optimal number of clusters.

DBSCAN

```
library(dbSCAN)
d <- read.csv("C:/Users/Vishal/Downloads/Pharmaceuticals.csv")
```

Interpretation : It may include performing density-based cluster analysis of pharmaceutical data. DBSCAN is a clustering algorithm that groups data points that are close to each other in terms of density and is effective in identifying clusters of arbitrary shape.

```
data1 <- d[ , -c(1,2,12,13,14)] # subsetting the data depend on the goals of the analysis
data1
```


##	Market_Cap	Beta	PE_Ratio	ROE	ROA	Asset_Turnover	Leverage	Rev_Growth
## 1	68.44	0.32	24.7	26.4	11.8	0.7	0.42	7.54
## 2	7.58	0.41	82.5	12.9	5.5	0.9	0.60	9.16
## 3	6.30	0.46	20.7	14.9	7.8	0.9	0.27	7.05
## 4	67.63	0.52	21.5	27.4	15.4	0.9	0.00	15.00
## 5	47.16	0.32	20.1	21.8	7.5	0.6	0.34	26.81
## 6	16.90	1.11	27.9	3.9	1.4	0.6	0.00	-3.17
## 7	51.33	0.50	13.9	34.8	15.1	0.9	0.57	2.70
## 8	0.41	0.85	26.0	24.1	4.3	0.6	3.51	6.38
## 9	0.78	1.08	3.6	15.1	5.1	0.3	1.07	34.21
## 10	73.84	0.18	27.9	31.0	13.5	0.6	0.53	6.21
## 11	122.11	0.35	18.0	62.9	20.3	1.0	0.34	21.87
## 12	2.60	0.65	19.9	21.4	6.8	0.6	1.45	13.99
## 13	173.93	0.46	28.4	28.6	16.3	0.9	0.10	9.37
## 14	1.20	0.75	28.6	11.2	5.4	0.3	0.93	30.37
## 15	132.56	0.46	18.9	40.6	15.0	1.1	0.28	17.35
## 16	96.65	0.19	21.6	17.9	11.2	0.5	0.06	-2.69
## 17	199.47	0.65	23.6	45.6	19.2	0.8	0.16	25.54
## 18	56.24	0.40	56.5	13.5	5.7	0.6	0.35	15.00
## 19	34.10	0.51	18.9	22.6	13.3	0.8	0.00	8.56
## 20	3.26	0.24	18.4	10.2	6.8	0.5	0.20	29.18
## 21	48.19	0.63	13.1	54.9	13.4	0.6	1.12	0.36

##	Net_Profit_Margin
## 1	16.1
## 2	5.5
## 3	11.2
## 4	18.0
## 5	12.9
## 6	2.6
## 7	20.6
## 8	7.5
## 9	13.3
## 10	23.4
## 11	21.1
## 12	11.0
## 13	17.9
## 14	21.3
## 15	14.1
## 16	22.4
## 17	25.2
## 18	7.3
## 19	17.6
## 20	15.1
## 21	25.5

```
# These all the characteristics of the dataset
```

```
set.seed(12)
db <- dbscan::dbscan(data1, eps = 25, MinPts = 2) #perform clustering
```

```
## Warning in dbscan::dbscan(data1, eps = 25, MinPts = 2): converting argument
## MinPts (fpc) to minPts (dbscan)!
```

```
print(db) #print cluster details
```

```
## DBSCAN clustering for 21 objects.
## Parameters: eps = 25, minPts = 2
## Using euclidean distances and borderpoints = TRUE
## The clustering contains 2 cluster(s) and 7 noise points.
##
## 0 1 2
## 7 7 7
##
## Available fields: cluster, eps, minPts, dist, borderPoints
```

```
#install.packages("fpc")
library('factoextra')
library('fpc')
df <- data1[, 1:9]

set.seed(123)
db <- fpc::dbscan(data1, eps = 35, MinPts = 1) # DBSCAN using fpc package

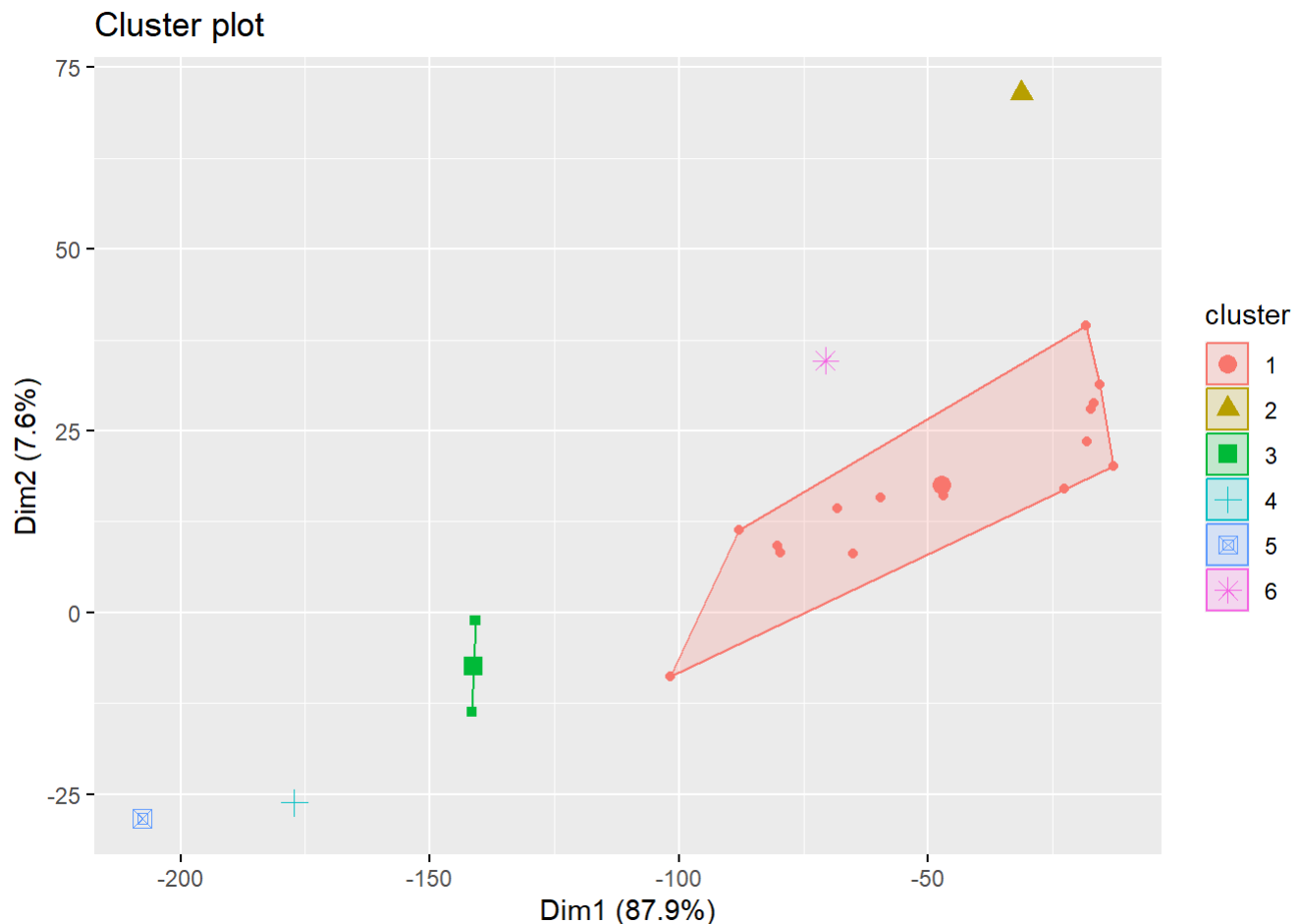
print(db) # show clusters' details
```

```
## dbscan Pts=21 MinPts=1 eps=35
##      1 2 3 4 5 6
## seed 15 1 2 1 1 1
## total 15 1 2 1 1 1
```

To identify dense regions or clusters within the data, and the specific parameters (eps and MinPts) are set based on the characteristics of the data or the analysis goals. The factoextra library can later be used to visualize clustering results. The print(db) statement is used to view details of the clusters detected by the algorithm, such as the number of clusters and the points assigned to each cluster. This step helps to understand the results of cluster analysis.

```
fviz_cluster(db, data1, stand = FALSE, frame = FALSE, geom = "point") # Alternative way to depict plot
```

```
## Warning: argument frame is deprecated; please use ellipse instead.
```



Visualizing clusters can be beneficial for interpreting the results of a clustering algorithm like DBSCAN. It helps to understand the spatial distribution of data points within clusters and assess the effectiveness of the clustering algorithm in grouping similar data points together.

#Let do Hierarchical Clustering

```
A <- read.csv("C:/Users/Vishal/Downloads/Pharmaceuticals.csv")
Sorted.data <- A[, -c(1,2,12,13,14)]
```

Let Compute Euclidean distance

```
# (to compute other distance measures, change the value in method = )
d <- dist(Sorted.data, method = "euclidean")
d.norm <- dist(Sorted.data[,c(4,8)], method = "euclidean")
```

Euclidean distance is often used in clustering algorithms to measure the dissimilarity between data points. It forms the basis for many clustering methods, including hierarchical clustering and k-means clustering.

Table 15.4

```
# normalize input variables
filt.data <- sapply(Sorted.data, scale)

# add row names: utilities
row.names(filt.data) <- row.names(Sorted.data)

# compute normalized distance based on variables ROE and Rev_Growth
d.norm <- dist(filt.data[,c(4,8)], method = "euclidean")

# The plot usually has an elbow-like shape, and the "elbow point" is a good place to look for
the ideal number of clusters. This is the point at which the within-cluster sum of squares de
clines more slowly, indicating that the number of clusters increases with diminishing returns
in terms of reducing the variance within clusters. This plot makes it easy to see when the imp
rovement in reducing within-cluster variability decreases with the number of clusters added.
Many people believe that the "elbow" number of clusters corresponds to a reasonable number of
clusters, which is the ideal number.
```

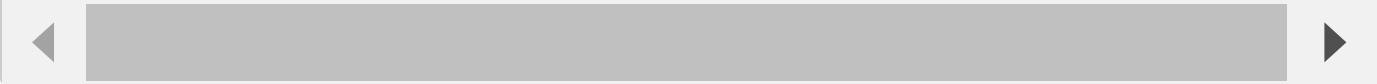
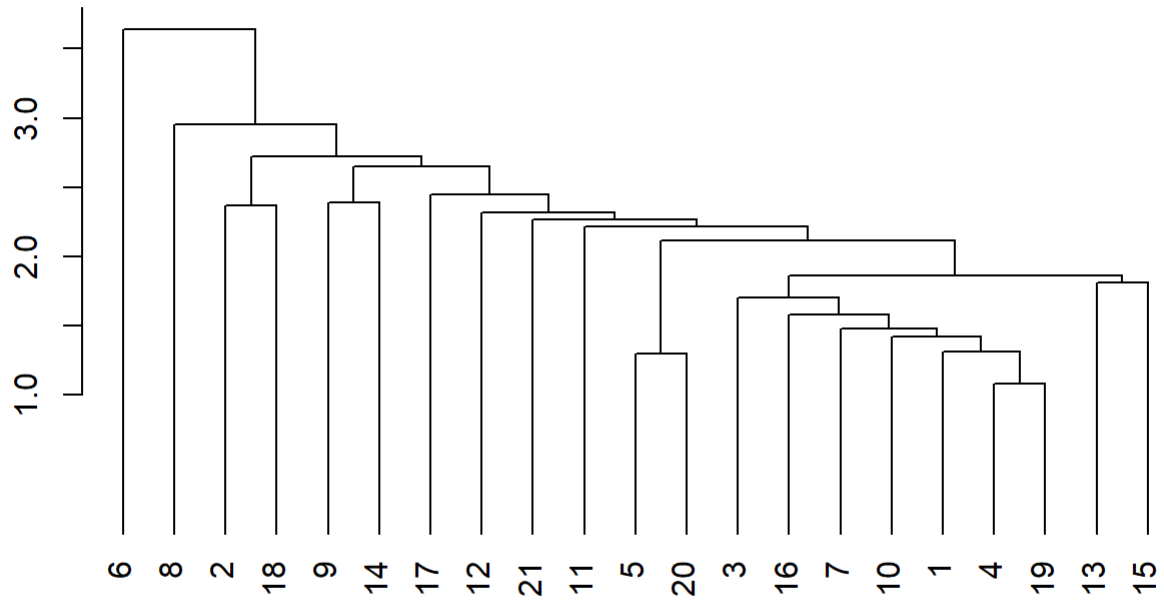


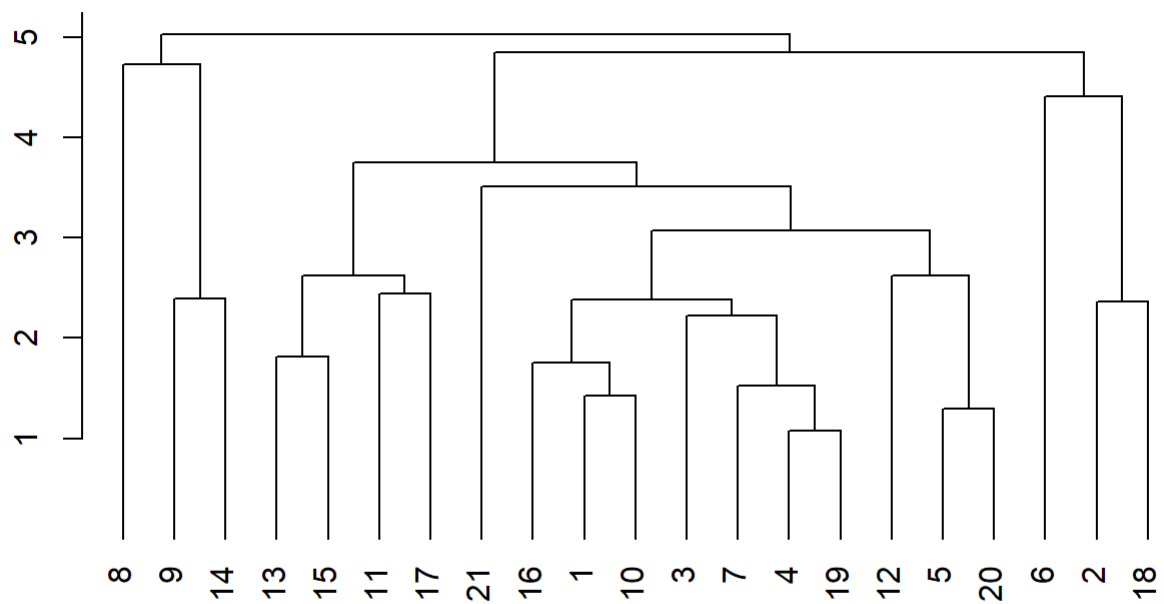
Figure 15.3

```
# compute normalized distance based on all 8 variables
d.norm <- dist(filt.data, method = "euclidean")

# in hclust() set argument method =
# to "ward.D", "single", "complete", "average", "median", or "centroid"
hc1 <- hclust(d.norm, method = "single")
plot(hc1, hang = -1, ann = FALSE)
```



```
hc2 <- hclust(d.norm, method = "average")
plot(hc2, hang = -1, ann = FALSE)
```



Interpretation : Data is arranged using hierarchical clustering into a hierarchy of nested clusters, and the formation of clusters is influenced by the linkage method selected. Various linkage techniques capture different facets of the data structure. The dendrogram that results from employing the plot function to visualize the hierarchical clustering structures illustrates how the observations are arranged into clusters according to varying degrees of similarity. It is common practice to investigate various linkage methods in order to obtain insights into the underlying clustering patterns of the data, as the choice of linkage method can impact the shape and structure of the resulting dendrogram.

Table 15.6

```
memb <- cutree(hc1, k = 3)
memb
```

```
##  1  2  3  4  5  6  7  8  9 10 11 12 13 14 15 16 17 18 19 20 21
##  1  1  1  1  1  2  1  3  1  1  1  1  1  1  1  1  1  1  1  1  1
```

```
memb <- cutree(hc2, k = 3)
memb
```

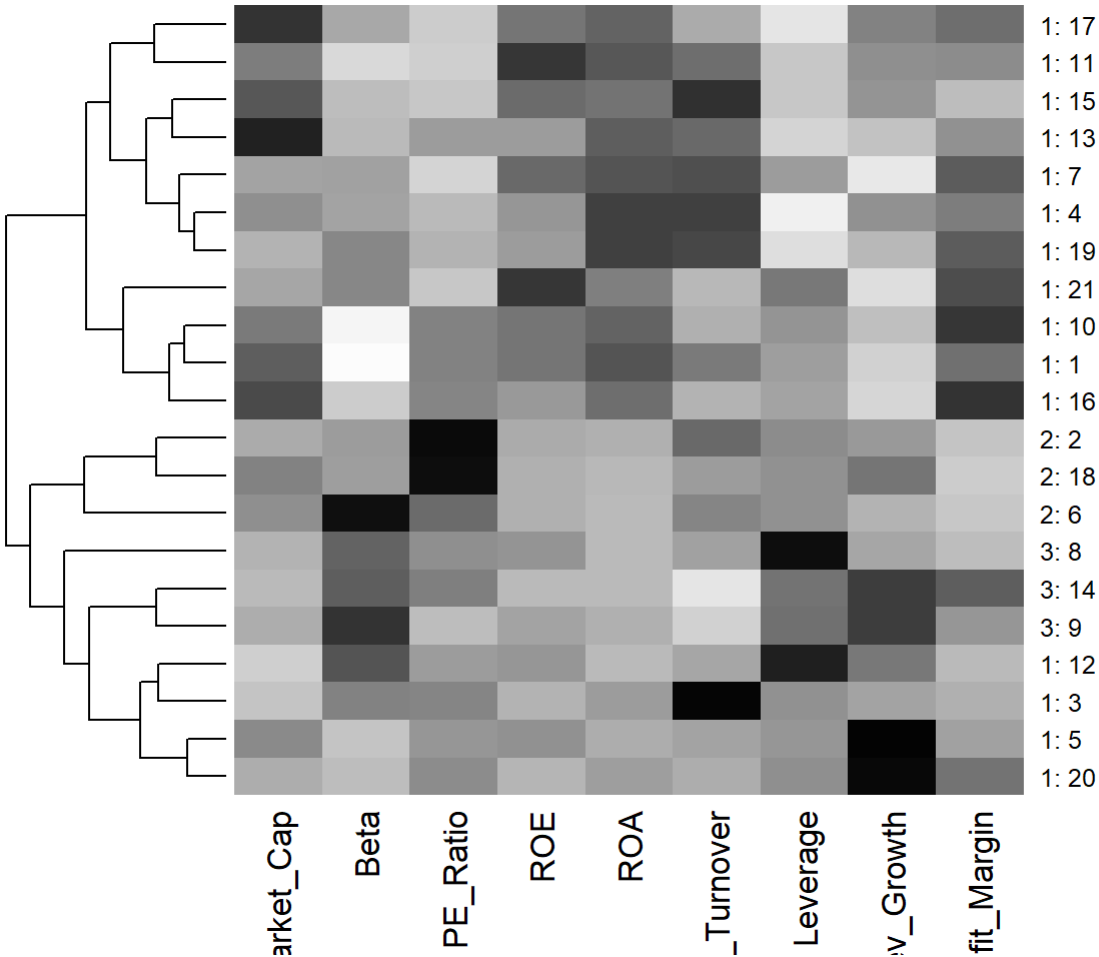
```
##  1  2  3  4  5  6  7  8  9 10 11 12 13 14 15 16 17 18 19 20 21
##  1  2  1  1  1  2  1  3  3  1  1  1  1  3  1  1  1  2  1  1  1
```

Figure 15.4

```
# set labels as cluster membership and utility name
row.names(filt.data) <- paste(memb, ": ", row.names(Sorted.data), sep = "")

# plot heatmap
# rev() reverses the color mapping to large = dark
heatmap(as.matrix(filt.data), Colv = NA, hclustfun = hclust,
        col=rev(paste("grey",1:99,sep="")))

```



Interpretation : In order to highlight the patterns and connections both within and between clusters, the code attempts to graphically depict the data's structure. This can be especially helpful for preliminary data analysis and learning more about the traits of various clusters.