

RESEARCH ARTICLE

Wild and laboratory exposure to cues of predation risk increases relative brain mass in male guppies

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Abstract

1. There is considerable diversity in brain size within and among species, and substantial dispute over the causes, consequences and importance of this variation. Comparative and developmental studies are essential in addressing this controversy.
2. Predation pressure has been proposed as a major force shaping brain, behaviour and life history. The Trinidadian guppy, *Poecilia reticulata*, shows dramatic variation in predation pressure across populations. We compared the brain mass of guppies from high and low predation populations collected in the wild. Male but not female guppies exposed to high predation possessed heavier brains for their body size compared to fish from low predation populations.
3. The brain is a plastic organ, so it is possible that the population differences we observed were partly due to developmental responses rather than evolved differences. In a follow-up study, we raised guppies under cues of predation risk or in a control condition. Male guppies exposed to predator cues early in life had heavier brains relative to their body size than control males, while females showed no significant effect of treatment.
4. Collectively our results suggest that male guppies exposed to predation invest more in neural tissue, and that these differences are at least partly driven by plastic responses.

KEYWORDSbrain size, development, plasticity, *Poecilia reticulata*, sex differences

1 | INTRODUCTION

Brains vary considerably in volume and organization both within and between species (Gonda, Herczeg, & Merilä, 2013; Striedter, 2005; de Winter & Oxnard, 2001). Such variation is often understood as the result of a trade-off between costs and benefits, balancing, for example, the energetic or developmental costs of brain enlargement against proposed benefits such as increased efficacy of perception, cognition or motor skills (Barton, 1998; Benson-Amram, Dantzer, Stricker, Swanson, & Holekamp, 2016; MacLean et al., 2014). Identifying the factors that shape brain evolution and development furthers our understanding of these costs and benefits (Sherry, 2006).

Predation poses a major challenge for many species (Edmunds, 1974; Lima & Dill, 1990) and may have a substantial influence on

brain evolution (van der Bijl & Kolm, 2016). Animals faced with frequent predation threats may be selected for greater investment in neural tissues that help them to sense, integrate or act upon information from the environment to evade predators (Gonda, Valimaki, Herczeg, & Merila, 2012). For example, birds with larger brains have shorter flight initiation distances, potentially reflecting superior predator monitoring abilities (Møller & Erritzøe, 2014). Larger-brained bird species have lower adult mortality (Sol, Szekely, Liker, & Lefebvre, 2007) and have reduced depredation of their nests (Öst & Jaatinen, 2015). Mammalian predators capture smaller-brained prey more often than expected by their abundance (Shultz & Dunbar, 2006), and the presence of predators is associated with larger brains in mammalian prey species (Jerison, 1973). In fish, prey species tend to have larger relative brain sizes than their predators and there is a

positive association between the brain sizes of predators and prey (Kondoh, 2010). By contrast, Walsh, Broyles, Beston, and Munch (2016) found that in the Trinidad killifish, *Rivulus hartii*, males from high-predation populations had smaller brains than those from low-predation populations. The authors speculated that killifish with fewer predators might be selected for larger brains because of the greater competition for food and mates in these populations. Similarly, a recent study on three-spine stickleback, *Gasterosteus aculeatus*, found that experimental exposure to predators selected for fish with smaller rather than larger brains (Samuk, Xue, & Rennison, 2018). Collectively, these results illustrate that the drivers of brain size variation are complex, and the effect of predation on relative brain size may depend on multiple interacting ecological and social pressures (Dunbar & Shultz, 2017).

The majority of studies that examine the evolution of brain size have made use of cross-species comparisons; however, these analyses can be complicated by phylogenetic relationships and unaccounted for ecological or life-history factors (Harris, O'Connell, & Hofmann, 2016; Healy & Rowe, 2007; Logan et al., 2018). Intraspecific studies across populations are valuable as they can partially control for some of the potentially confounding variables that inherently complicate the interpretation of interspecies comparisons (Gonda et al., 2012; Logan et al., 2018). Leveraging natural variation in ecological conditions among populations represents a powerful approach to the study of brain evolution (Walsh et al., 2016).

While the brain is shaped by evolution, it is also a highly malleable organ and phenotypic plasticity may also play a key role in generating individual variation in brain size (Gonda et al., 2013; Healy & Rowe, 2007). For example, environmental complexity during early life increases relative brain size in rodents (Diamond et al., 1966; Rosenzweig & Bennett, 1969), insects (Heisenberg, Heusipp, & Wanke, 1995) and fish (DePasquale, Neuberger, Hirrlinger, & Braithwaite, 2016; Gonda, Herczeg, & Merilä, 2011), while low oxygen during development decreases relative brain size in fish (Chapman, Albert, & Galis, 2008).

The Trinidadian guppy, *Poecilia reticulata*, is a small live-bearing freshwater fish that experiences pronounced interpopulation variation in predation threat (Magurran, 2005) and thus provides a valuable system to study how predation shapes the brain. Throughout Trinidad, guppies have repeatedly colonized independent river reaches above natural waterfall barriers, where aquatic predators are scarce, while simultaneously living below the same barriers where abundant aquatic predators impose substantial mortality (Magurran, 1998). As a result, there has been repeated parallel evolution of distinct behavioural, morphological and life-history traits among guppy populations that are heavily depredated compared to those that are relatively free from predation pressure (Magurran, 2005).

A recent series of papers has examined the effect of artificially selecting guppies for large or small relative brain mass, finding that increased investment in brain tissue can provide antipredator benefits, but also carry costs. Female guppies artificially selected for larger brains exhibited greater survival under predation and altered predator responses compared to small-brained individuals (van der

Bijl, Thyselius, Kotrschal, & Kolm, 2015; Kotrschal, Buechel, et al., 2015). However, larger-brained female guppies also had smaller guts, produced fewer offspring and had reduced innate immune responses, suggesting a trade-off between neural investment and other fitness-relevant parameters (Kotrschal, Corral-Lopez, Szidat, & Kolm, 2015; Kotrschal, Kolm, & Penn, 2016; Kotrschal et al., 2013). If antipredator advantages were sufficient to overcome the costs of maintaining a larger brain, then we would expect that guppies from high-predation populations would consistently have larger brains for their body size than guppies from low-predation environments. Indeed, female guppies under greater threat from predatory prawns have larger relative brain sizes than do females under lesser threat from these predators (Kotrschal, Deacon, Magurran, & Kolm, 2017). Artificial selection on brain size in guppies has consistently revealed differing effects in males and females, suggesting that sex may be a key modulator of the relationship between brain size and performance in this species (e.g., van der Bijl et al., 2015; Kotrschal, Rogell, Maklakov, & Kolm, 2012; Kotrschal et al., 2013; Kotrschal, Buechel, et al., 2015), and therefore, it is important to examine both males and females.

Guppies also show plasticity in brain size; for example, guppies raised in the laboratory have smaller brains than fish born in the wild (Burns & Rodd, 2008; Burns, Saravanan, & Rodd, 2009; Eifert et al., 2015). Furthermore, guppy males that cohabitated with females have larger brains than those that lived with only males (Kotrschal et al., 2012). If guppies can adjust their investment in neural tissue to local conditions during development, it is possible that plastic responses to cues of predation risk may at least partially explain any observed population differences in brain size. To understand the expression of a quantitative phenotypic trait, evolutionary studies on interpopulation differences in trait expression should be combined with studies of phenotypic plasticity (Gonda et al., 2013).

Our study aimed to help illuminate the importance of predation in shaping within-species variation in brain mass and to elucidate the potential role of plasticity in generating these differences. Specifically, we had two objectives: First, we aimed to determine whether there are differences in relative brain mass between wild guppies collected from high- and low-predation populations. We predicted that guppies from high-predation populations would have relatively heavier brains. Second, we aimed to determine whether guppies show brain mass plasticity in response to cues of predation risk during development. We conducted a laboratory experiment in which guppies were exposed to multisensory cues of predation risk or a control condition during the first 45 days of life. We predicted that guppies exposed to cues of predation risk would show increased relative brain mass.

2 | MATERIALS AND METHODS

2.1 | Field collections

In March 2016, we collected 151 adult guppies (79 males and 72 females) from four sites, one high-predation site and one low-predation site in each of two rivers (Aripo and Marianne) in the northern

TABLE 1 Collection site and sample sizes for wild-caught fish. Site names and predation regime classifications are based on Gotanda et al. (2013)

Site	UTM coordinates (x, y)	Predation regime	n males	n females
Aripo 1	693,188, 1,181,605	Low	15	15
Aripo 2	694,231, 1,177,709	High	27	20
Marianne 10	686,711, 1,191,358	Low	21	26
Marianne 14	684,934, 1,191,469	High	16	11

Note. UTM, Universal Transverse Mercator.

mountain range of Trinidad (Table 1). These rivers belong to independent drainages, and therefore, they are subject to a distinct suite of biotic and abiotic conditions (Gotanda et al., 2013). Assignment of predation regime followed previous studies at these sites (Gotanda et al., 2013) and was based on the presence or absence of dangerous fish predators (e.g., cichlids such as *Crenicichla* sp. and *Aequidens pulcher* in the Aripo River, and eleotrids such as *Eleotris pisonis* and *Gobiomorus dormitor* in the Marianne River; Magurran, 2005; Reznick, Callahan, & Llauredo, 1996), which is consistent across years (Schwartz & Hendry, 2010). Guppies were collected from each site using butterfly nets and then were transported to the William Beebe Research Station near Arima, Trinidad. Each fish was euthanized with an overdose of tricaine methanesulfonate (Finquel MS-222; Argent Chemical Laboratories, USA) buffered to a neutral pH with NaHCO_3 . We measured each fish for standard length (SL; from the tip of the snout to the caudal peduncle) and then dissected out the brains using a portable stereomicroscope (Ken-a-vision VisionScope 2) at 10 \times magnification. Care was taken to sever the spinal cord and optic nerves at a consistent position on each brain. We placed the brains in RNAlater (Sigma-Aldrich) and incubated them for 24 h at room temperature before transferring them to -20°C . We transported the samples back to McGill University (Montreal, Canada) where we removed them from RNAlater and gently dabbed them dry. Blind to the population of origin, we weighed each whole brain to the nearest 0.1 mg using an analytic laboratory balance (Mettler-Toledo ME104E). Because all brains were treated identically, any storage effects on brain mass should affect all samples similarly. Following measurement, the brains were used in another study.

2.2 | Developmental experiment

We exposed developing guppies to cues of predation threat during the first 45 days of life. The parental generation were guppies from a laboratory-reared population, descended from a mixture of fish captured in high-predation sites in the Aripo and Quare rivers of northern Trinidad in 2009 and 2010. Parental fish were housed in mixed-sex groups of ~ 10 adults in 18-L aquaria. We maintained the water at $26 \pm 1^\circ\text{C}$ and fed the fish ad libitum daily on a mixture of dried prepared tropical fish flakes (TetraMin, Tetra, Germany) and rehydrated decapsulated brine shrimp eggs (Brine Shrimp Direct, Inc., Ogden, Utah, USA). Lights were on from 07:00 to 19:00 hr, with a 30-min dawn/dusk period. To collect fry for the experiment, we moved groups of 10–12 visibly gravid females into separate aquaria, which we checked daily for newborn fry. We mixed fry born to

different females and randomly assigned them to one of two treatments: exposure to cues of predation risk or a control condition. Fry in both treatments were held at densities of 30 individuals per 18-L aquarium during the treatment period. We had three replicate aquaria in each experimental condition (six aquaria total). The experimental aquaria contained 1 cm of white coral sand and were furnished with an artificial plant to serve as a refuge. Water and light conditions were the same as for the parental generation, but the developing fry were fed twice daily.

Five days per week during the 45-day treatment period, the fish in the predator cue condition were visually exposed to a sympatric cichlid fish predator (*Crenicichla* sp.) living in an adjacent aquarium by removing an opaque barrier between them for 5 min. Concurrent with the visual exposure, we infused 5 ml of water previously collected from aquaria housing live *Crenicichla* that had recently been fed freshly euthanized guppies (following Brown, Paige, & Godin, 2000). Guppies respond to the odour of damaged conspecifics and predator dietary cues with antipredator responses (Brown & Godin, 1999). On four of the five weekly cue exposure days, we also added 5 ml of odour cue harvested from the skin and muscle tissue of adult guppies in addition to the predator housing water. To collect this cue, we euthanized adult guppies of both sexes by briefly immersing them in an ice water bath and then swiftly decapitating them (Matthews & Varga, 2012). We then homogenized skin and muscle tissues with dH_2O , filtered the solution with cotton floss and diluted it with dH_2O until we obtained a concentration of 0.1 cm^2 of tissue per ml of cue (following Brown & Godin, 1999). We exposed the guppies in the control condition to the sight and housing water of a nonpredatory suckermouth catfish (*Pterygoplichthys* sp.) that had been fed blanched spinach leaves. In lieu of the damaged conspecific cue, the control fish received blank dH_2O . We exposed the guppies to these heterospecific fish stimuli at a randomly chosen time (between 10:00 and 16:00 hr) on each exposure day.

After 45 days, we ceased all heterospecific stimuli exposures. On day 50, we reduced the housing density of the experimental fish by splitting each group into three separate 18-L aquaria with ~ 10 individuals of mixed sex in each, resulting in a total of 18 housing aquaria, nine per treatment. We also reduced the feeding frequency to once per day to match the standard adult husbandry protocols in our laboratory. The experimental fish were held in these conditions until approximately 300 days of age, during which time behavioural and hormonal measures were taken for other studies (Chouinard-Thuly, Reddon, Leris, Earley, & Reader, 2018; Leris, 2016). We then euthanized 73 individuals (22

predator-exposed males, 27 control males, 11 predator-exposed females and 13 control females), by briefly immersing them in an ice water bath and then swiftly decapitating them. We then dissected out their brains using a stereomicroscope (Leica EZ4W) at 10× magnification. Care was taken to sever the spinal cord and optic nerves at a consistent position on each brain. We weighed the fresh brains to the nearest 0.1 mg on an analytic laboratory balance (Mettler-Toledo ME104E). Brain mass and body size values were taken blind to treatment. Following measurement, the brains were used in another study.

2.3 | Analysis

We used linear models to investigate the relationship between brain mass and body size with exposure to predation both naturally in the field and in our developmental experiment. To account for the allometric relationship between brain mass and body size (Brandstätter & Kotschal, 2008), we included SL as a covariate in the models investigating brain mass. We log-transformed SL and brain mass measures before running each model and mean-centred SL. Male and female guppies differ considerably in body size (mean SL \pm SE: wild males = 12.77 ± 0.13 mm, wild females = 15.66 ± 0.30 mm; Welch's $t_{96,9} = 8.76$, $p < 0.001$; laboratory males = 14.11 ± 0.15 mm, laboratory females = 28.26 ± 0.45 mm; Welch's $t_{29,19} = 29.7$, $p < 0.001$);

therefore, we ran separate analyses for males and females in each of our two studies. For the field-collected data, we included river (Aripo or Marianne), as well as the interaction between river and predation regime as factors. In all four models investigating brain mass, we tested for an interaction between SL and predation exposure on brain mass to test for the possibility of different allometric relationships across populations. This interaction was not significant for any of the models (all $p > 0.22$) and was subsequently dropped from the final analyses. We examined model residuals using QQ plots to look for violations of the homogeneity of variance or normality assumptions. All statistical analyses were performed in R version 3.2.2 (R Core Development Team, 2016), and graphs were produced in ggplot2 (Wickham, 2009).

2.4 | Ethics

Methods were approved by the Animal Care Committee of McGill University (protocols 2012-7133 and 2015-7708) and were conducted in accordance with the ethical guidelines from the Canadian Council on Animal Care and ABS/ASAB. Field sampling was approved by the Ministry of Agriculture, Land and Marine Resources of the Republic of Trinidad and Tobago. Guppies are neither endangered nor threatened and were abundant at all collection sites.

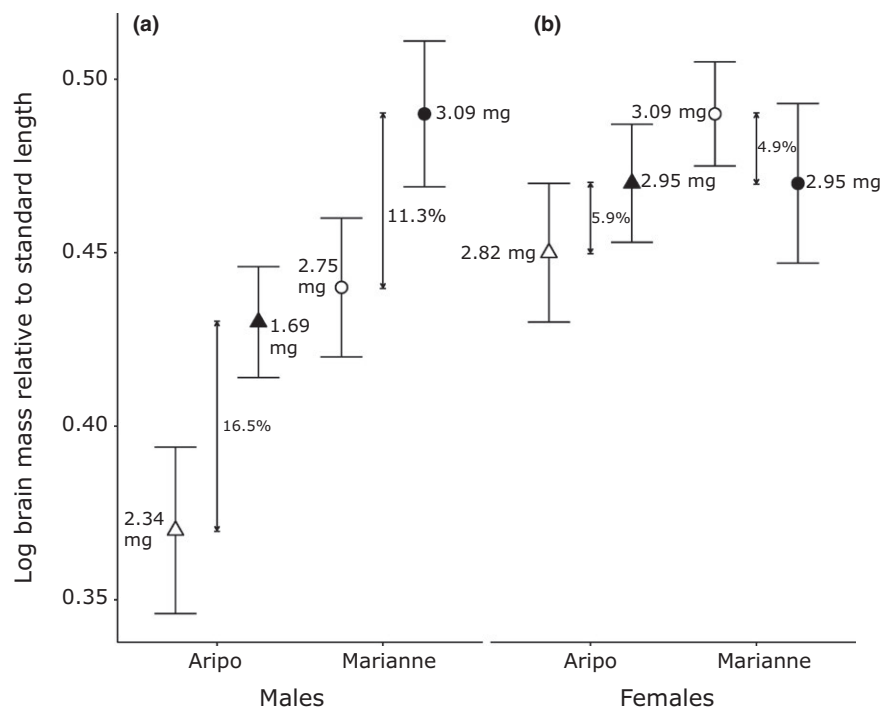


FIGURE 1 Expected (log-transformed) brain mass (\pm SE) of guppies captured in the wild. Expected values are for the mean standard length for each sex. Mean brain masses for each group are alongside the points, and percentage differences between groups are indicated with arrows. Males (a) from high-predation populations have larger brain masses for their body size than males from low-predation populations ($p = 0.05$). Males from the Marianne River had significantly heavier brains for their body size than males from the Aripo River ($p = 0.02$). Females (b) did not show a significant effect of predation regime or river. Filled symbols, environments with predators; open symbols, environments without predators; triangles, Aripo River; circles, Marianne River

TABLE 2 Estimates and standard error of fixed parameters and their interaction for the linear model with log-transformed brain mass for the guppy field population comparison. Estimates represent the difference in log-transformed brain mass between the level of a factor (identified in parenthesis) and the reference levels for categorical factors and are mean-centred for covariates. The reference levels were high predation and Aripo River. The standard length was log-transformed and mean-centred

Parameter	Estimate	SE	t-Value	p-Value
Males (<i>df</i> = 74)				
Intercept	0.43	0.016	26.37	<0.0001
Standard length	1.78	0.31	5.80	<0.0001
Predation (low)	-0.061	0.031	1.97	0.052
River (Marianne)	0.063	0.025	2.48	0.015
River × predation	0.0046	0.047	0.124	0.902
Females (<i>df</i> = 67)				
Intercept	0.47	0.017	27.93	<0.0001
Standard length	1.10	0.13	8.66	<0.0001
Predation (low)	-0.026	0.026	1.00	0.318
River (Marianne)	-0.0023	0.028	0.081	0.936
River × predation	0.050	0.037	1.35	0.181

Note. *df*, degrees of freedom.
p-Values ≤ 0.05 are shown in bold.

3 | RESULTS

3.1 | Field collections

We found that for an average body size, males collected from high-predation sites had brains 11.3% heavier in the Marianne River and 16.5% heavier in the Aripo River than males collected from low-predation sites in the same rivers ($p = 0.052$; Figure 1a; Table 2). Males from the Marianne River had 14.7% heavier brains than males from the Aripo River, but the interaction between predation regime and river was not significant (Figure 1a; Table 2). We found no evidence that predation regime or river was associated with relative brain mass in female guppies (Figure 1b; Table 2). Males from high-predation sites were significantly smaller bodied than low-predation males (mean $SL \pm SE$: high-predation males = 12.08 ± 0.14 ; low-predation males = 13.61 ± 0.14 ; $p < 0.001$; Table 3), but there was no similar significant difference in female body length (mean $SL \pm SE$: high-predation females = 16.31 ± 0.36 ; low-predation females = 14.80 ± 0.48 ; $p = 0.062$). Supporting Information Figure S1 illustrates the allometric relationships between brain mass and body length in the wild-caught fish.

3.2 | Developmental experiment

We found that for an average body size, males exposed to predation cues during development had brains 21.2% heavier than males exposed to control cues ($p = 0.011$; Figure 2a; Table 4). We found no evidence that exposure to predation cues during development

TABLE 3 Estimates and standard error of fixed parameters and their interaction for the linear model with log-transformed standard length for the guppy field population comparison. Estimates represent the difference in log-transformed standard length between the level of a factor (identified in parenthesis) and the reference levels. The reference levels were high predation and Aripo River

Parameter	Estimate	SE	t-Value	p-Value
Males (<i>df</i> = 74)				
Intercept	1.08	0.005	186.72	<0.0001
Predation (low)	0.056	0.0097	5.73	<0.0001
River (Marianne)	-0.0036	0.0095	0.38	0.705
River × predation	-0.0045	0.014	0.33	0.743
Females (<i>df</i> = 67)				
Intercept	1.17	0.016	73.15	<0.0001
Predation (low)	0.047	0.025	1.90	0.062
River (Marianne)	-0.031	0.027	1.15	0.253
River × predation	0.010	0.036	0.30	0.769

Note. *df*, degrees of freedom.
p-Values ≤ 0.05 are shown in bold.

influenced the relative brain mass of female guppies (Figure 2b; Table 4). Males exposed to predation cues were significantly larger bodied than males exposed to control cues (mean $SL \pm SE$: predator cue-exposed males = 14.51 ± 0.23 ; control cue males = 13.80 ± 0.17 ; $p = 0.014$; Table 5), but there was no significant difference in female body length (mean $SL \pm SE$: predator cue-exposed females = 28.13 ± 0.80 ; control cue females = 28.39 ± 0.49 ; $p = 0.65$). Supporting Information Figure S2 illustrates the allometric relationships between brain mass and body length in the laboratory-reared fish.

4 | DISCUSSION

Male guppies exposed to cues of predation risk in the laboratory, or actual predation risk in the wild, had larger brains for their body size than did males that did not have this experience. In contrast, we did not detect a consistent difference in relative brain mass between female guppies that were or were not exposed to real or simulated predation risk, suggesting that the effect of predation on relative brain mass is sex dependent in guppies and is weaker or absent in females. The population differences in males could be due to evolved differences; however, the parallel results from our laboratory experiment suggest that the difference in brain mass may be at least partially due to inducible plasticity in neural investment relative to investment in body size, triggered by exposure to predation cues during development. Notably, the effects of predator cues confined to early life were long-lasting, persisting throughout life.

Male guppies could hypothetically benefit from greater neural investment under predation threat if heavier brains relative to

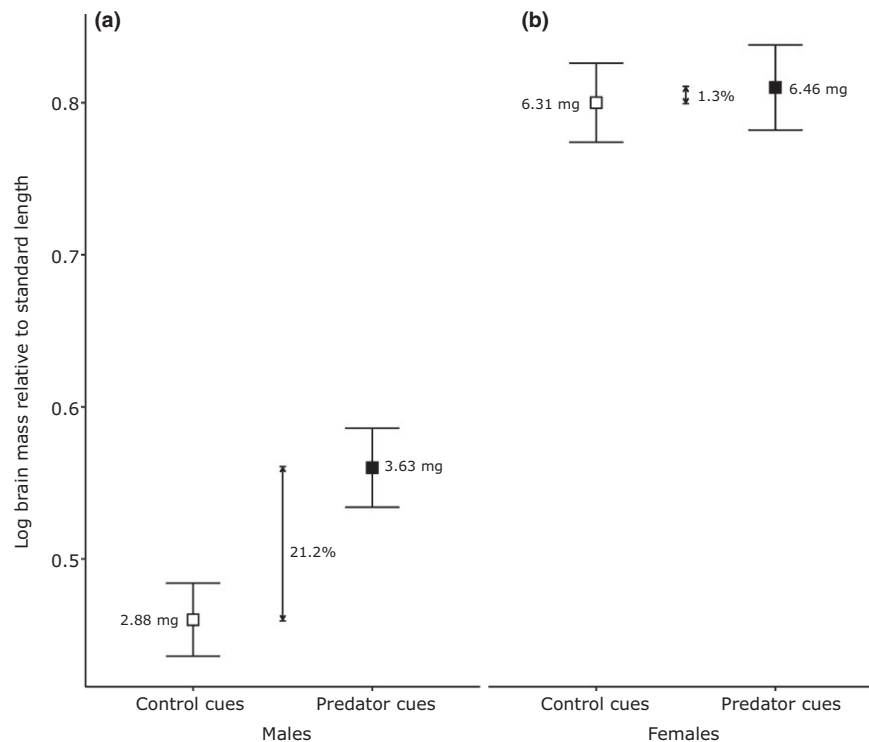


FIGURE 2 Expected (log-transformed) brain mass (\pm SE) of guppies experimentally exposed to predator or control cues during development in captivity. Expected values are for the mean standard length for each sex. Mean brain masses are alongside the points, and percentage differences between groups are indicated with arrows. Males (a) from the predator cue-exposed treatment had larger relative brain masses than males from the control treatment ($p = 0.01$). Females (b) did not show a significant effect of the predator cue treatment. Filled symbols, predator cue treatment; open symbols, control treatment

body size provide sensory, cognitive or motor benefits. It is possible that having a heavier brain may allow guppies to detect, assess, react to or learn about predation threats better and/or allow for simultaneous monitoring of predator threats while engaged in alternative activities such as courtship or foraging, similar to reports in

TABLE 4 Estimates and standard error of fixed parameters and their interaction for the linear model with log-transformed brain mass for the guppy laboratory developmental study. Estimates represent the difference in log-transformed brain mass between the level of a factor (identified in parenthesis) and the reference level for the categorical factor (the predator cue treatment) and are mean-centred for covariates. The standard length was log-transformed and mean-centred

Parameter	Estimate	SE	t-Value	p-Value
Males ($df = 46$)				
Intercept	0.56	0.026	21.16	<0.0001
Standard length	1.11	0.59	1.89	0.065
Predation (control)	-0.098	0.037	2.66	0.011
Females ($df = 21$)				
Intercept	0.81	0.028	28.86	<0.0001
Standard length	0.60	0.72	0.83	0.415
Predation (control)	-0.011	0.038	0.30	0.770

Note. df , degrees of freedom.
p-Values ≤ 0.05 are shown in bold.

birds (Møller & Erritzøe, 2014; Sol et al., 2007). A heavier brain may also allow individual guppies to better address social demands, such as group cohesion or coordination with conspecifics (Dunbar & Shultz, 2017). Guppies from high-predation populations do form more cohesive and coordinated groups (Ioannou, Ramnarine, & Torney, 2017), and group cohesion has antipredator benefits in prey fishes (Ioannou, Guttal, & Couzin, 2012; Krause & Ruxton,

TABLE 5 Estimates and standard error of fixed parameters and their interaction for the linear model with log-transformed standard length for the guppy laboratory developmental study. Estimates represent the difference in log-transformed standard length between the level of a factor (identified in parenthesis) and the reference level (the predator cue treatment)

Parameter	Estimate	SE	t-Value	p-Value
Males ($df = 46$)				
Intercept	1.16	0.0064	182.31	<0.00001
Predation (control)	-0.022	0.0086	-2.54	0.014
Females ($df = 21$)				
Intercept	1.46	0.0083	176.31	<0.0001
Predation (control)	-0.0052	0.011	-0.46	0.650

Note. df , degrees of freedom.
p-Values ≤ 0.05 are shown in bold.

2002). Interestingly, predation seems to select for a reduction in brain size in some other fish species, and the putative advantages of increased brain size in the face of predation risk thus certainly merit closer examination (Samuk et al., 2018; Walsh et al., 2016). Samuk et al. (2018) suggest that differences between studies could result from the type of antipredator responses employed and local ecological conditions, such as the availability of shelter. An experimental evolution study on guppies, tracking the effects of different predators on brain size, cognitive performance, social behaviour and antipredator defences across generations, with different antipredator responses available, would be a large undertaking but highly informative in this regard.

We found that males exposed to predators in the wild were smaller bodied than those from low-predation environments, replicating previous findings (Reznick & Endler, 1982). This raises the possibility that the change in relative brain mass we observed could reflect selection by predators for decreased overall body size. This could only explain our results if the brain was not reduced to the same degree as the rest of the body under predation threat, that is predation caused a differential effect on body vs. brain size, with the largest effect on body size. However, the results of our developmental study, in which predator-exposed males were larger than control males and yet relative brain mass was still greater, argue against a simple explanation in terms of body size. We are unsure why predator cues in the laboratory resulted in increased adult body size in male guppies, while exposure to genuine predation risk in the wild decreased male body size. Although guppies from high-predation populations forage less in standardized conditions (Botham et al., 2008), exposure to acute cues of high-predation risk induces short-term compensatory foraging (Elvidge, Ramnarine, & Brown, 2014). As fish in our developmental study were exposed to repeated acute predator cue exposures, this potentially explains the disparity between our two studies, although leaves open the question of why such an effect was not observed in females. The differential effects of predator cues vs. direct predator encounters and the effects of predation cues confined to early life compared to lifelong exposure are deserving of further investigation.

Why should predation affect brain mass in male guppies but not females? In guppies, males are more conspicuous, less social, bolder and poorer swimmers than females (Houde, 1997) and thus are more vulnerable to predation (Kotrschal, Buechel, et al., 2015). As a result, males may have more to gain from investment in neural tissues under predation threat. Alternatively, because females are slower to mature and longer lived (Magurran, 2005), they may show greater neural investment, regardless of predation risk. Relative brain size has been linked to life-history strategy in birds and mammals, with slower developing and longer-lived animals typically having larger brains for their body size (Bennett & Harvey, 2009; Iwaniuk & Nelson, 2003).

Some authors have been critical of studies of whole brain size (e.g., Chittka & Niven, 2009; Healy & Rowe, 2007; Logan et al., 2018). We agree that a more granular examination of specific brain regions, and other subtler aspects of neuroanatomy and neural organization, as well as the costs and benefits of brain enlargement, would add essential information to our understanding of neural

investment in guppies. Assessing whole brain mass does, however, have several advantages; for instance, measuring whole brains avoids problem of correctly determining the relevant homologous areas between taxa (van der Bijl & Kolm, 2016). Furthermore, while mosaic evolution of brain areas exists (Barton & Harvey, 2000), the size of different brain areas tends to correlate strongly with overall brain size (Finlay & Darlington, 1995; Kotrschal, Zeng, et al., 2017), so whole brain size can be a reasonable measure of neural investment, especially when the specific brain area of interest is uncertain (van der Bijl & Kolm, 2016). We argue that identifying effects on whole brain size can be a useful tool to identify relevant ecological factors affecting neural investment. Our current data show that whole brain mass varies across populations (see also Kotrschal, Deacon, et al., 2017) and responds to developmental conditions in guppies. Combined with the effects of artificial selection on brain mass in guppies (Kotrschal et al., 2013, 2014; Kotrschal, Buechel, et al., 2015; Kotrschal, Zeng, et al., 2017), these data suggest that relative brain mass is a relevant trait in guppies, encouraging more fine-scaled work in the future.

Our results contrast with some previous findings. For instance, Burns and Rodd (2008) did not find differences in size between the brains of female or male guppies collected from high- vs. low-predation wild populations. The reason for the discrepancy in the males is unclear, but it is worth noting that different methods for estimating brain size were used: Burns and Rodd measured the dorsal surface area of the telencephalon and optic tectum rather than brain mass. Kotrschal, Deacon, et al. (2017) found, as we did, that the density of fish predators across populations did not correlate with relative brain mass in female guppies; however, they did not examine males. Kotrschal et al. did find that the biomass of predatory prawns correlated positively with relative brain mass in females, suggesting that female brain mass may respond to threat from other types of predators.

Our field comparisons of high- and low-predation guppies came from only two replicate rivers. The parallel results in the two rivers, in the same sex, and the qualitative match with the developmental manipulation, again in the same sex, suggest, however, that predation is likely a key driver of the differences we observed in male brain mass. However, sampling of a greater number of rivers and a variety of other ecological conditions would clearly be a valuable follow-up. Additionally, the laboratory study was conducted on only a single-laboratory population, descended from a mixture of high-predation fish from two different rivers. It would be interesting to examine whether the developmental effects of predation cues differ between populations. We also note that the balance we used to weigh the brains was relatively coarse (0.1 mg listed repeatability, with more error likely at lower masses) given the small size of guppy brains (1.3–9.2 mg in our samples). However, any measurement error introduced by our instrument would not be systematic and therefore should reduce rather than increase our likelihood of detecting an effect. Indeed, it is possible that a subtle effect exists in the female brains which we failed to detect with our methodology.

It is not clear to what degree the interpopulation differences we observed in relative brain mass reflect local adaptation vs. phenotypic

plasticity. Environmental conditions can select for differences in neuroanatomy across populations; for example, black-capped chickadees, *Poecile atricapillus*, that live in harsher northern climates have larger hippocampal volumes than individuals from milder regions (Roth & Pravosudov, 2009), and these differences are retained in laboratory-reared offspring (Roth, LaDage, & Pravosudov, 2010). Guppies may have evolved increased brain mass under predation threat; however, our work suggests that plasticity can play an important role in determining brain mass in guppies, and therefore, the population differences that have been observed (Kotrschal, Deacon, et al., 2017; this study) may be partly or entirely due to a plastic response to cues of predation threat during development. Common garden experiments will be required to disentangle the contributions of selection and plasticity on relative brain mass in this species, ideally comparing plasticity across populations.

In conclusion, we found that male but not female guppies exposed to predators either naturally in the wild or experimentally in the laboratory have heavier brains for their body size than individuals that were not exposed to predators. Future work is required to determine the causes of this increased neural investment in male guppies and why this pattern is not observed in females. Our results highlight the potential importance for developmental plasticity in generating population differences in relative brain mass.

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CONFLICT OF INTERESTS

We have no competing interests to declare.

AUTHORS' CONTRIBUTIONS

A.R.R., I.L. and S.M.R. planned the study. A.R.R. conducted the field collections. I.L. conducted the developmental manipulations. A.R.R. dissected the fish and weighed the brains. A.R.R. and L.C.-T. analysed the data. L.C.-T. made the figures. A.R.R. wrote the first draft of the manuscript. All authors contributed to the final version of the manuscript.

DATA ACCESSIBILITY

Data are available from the Dryad Digital Repository <https://doi.org/10.5061/dryad.pq1js52> (Reddon, Chouinard-Thuly, Leris, & Reader, 2018).

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REFERENCES

- Barton, R. A. (1998). Visual specialization and brain evolution in primates. *Proceedings of the Royal Society B: Biological Sciences*, 265, 1933–1937. <https://doi.org/10.1098/rspb.1998.0523>
- Barton, R. A., & Harvey, P. H. (2000). Mosaic evolution of brain structure in mammals. *Nature*, 405, 1055–1058. <https://doi.org/10.1038/35016580>
- Bennett, P. M., & Harvey, P. H. (2009). Brain size, development and metabolism in birds and mammals. *Journal of Zoology*, 207, 491–509. <https://doi.org/10.1111/j.1469-7998.1985.tb04946.x>
- Benson-Amram, S., Dantzer, B., Stricker, G., Swanson, E. M., & Holekamp, K. E. (2016). Brain size predicts problem-solving ability in mammalian carnivores. *Proceedings of the National Academy of Sciences*, 113, 2532–2537. <https://doi.org/10.1073/pnas.1505913113>
- van der Bijl, W., & Kolm, N. (2016). Why direct effects of predation complicate the social brain hypothesis. *BioEssays*, 38, 568–577. <https://doi.org/10.1002/bies.201500166>
- van der Bijl, W., Thyselius, M., Kotrschal, A., & Kolm, N. (2015). Brain size affects the behavioural response to predators in female guppies (*Poecilia reticulata*). *Proceedings of the Royal Society B: Biological Sciences*, 282, 20151132. <https://doi.org/10.1098/rspb.2015.1132>
- Botham, M. S., Hayward, R. K., Morrell, L. J., Croft, D. P., Ward, J. R., Ramnarine, I., & Krause, J. (2008). Risk-sensitive antipredator behavior in the Trinidadian guppy, *Poecilia reticulata*. *Ecology*, 89, 3174–3185. <https://doi.org/10.1890/07-0490.1>
- Brandstätter, R., & Kotrschal, K. (2008). Brain growth patterns in four European cyprinid fish species (Cyprinidae, Teleostei): Roach (*Rutilus rutilus*), bream (*Abramis brama*), common carp (*Cyprinus carpio*) and sabre carp (*Pelecus cultratus*). *Brain Behavior and Evolution*, 35, 195–211. <https://doi.org/10.1159/000115867>
- Brown, G. E., & Godin, J.-G. J. (1999). Chemical alarm signals in wild Trinidadian guppies (*Poecilia reticulata*). *Canadian Journal of Zoology*, 77, 562–570. <https://doi.org/10.1139/z99-035>
- Brown, G. E., Paige, J. A., & Godin, J.-G. J. (2000). Chemically mediated predator inspection behaviour in the absence of predator visual cues by a characin fish. *Animal Behaviour*, 60, 315–321. <https://doi.org/10.1006/anbe.2000.1496>
- Burns, J. G., & Rodd, F. H. (2008). Hastiness, brain size and predation regime affect the performance of wild guppies in a spatial memory task. *Animal Behaviour*, 76, 911–922. <https://doi.org/10.1016/j.anbehav.2008.02.017>
- Burns, J. G., Saravanan, A., & Rodd, F. H. (2009). Rearing environment affects the brain size of guppies: Lab-reared guppies have smaller brains than wild-caught guppies. *Ethology*, 115, 122–133. <https://doi.org/10.1111/j.1439-0310.2008.01585.x>
- Chapman, L., Albert, J., & Galis, F. (2008). Developmental plasticity, genetic differentiation, and hypoxia-induced trade-offs in an African cichlid fish. *The Open Evolution Journal*, 2, 75–88. <https://doi.org/10.2174/1874404400802010075>
- Chittka, L., & Niven, J. (2009). Are bigger brains better? *Current Biology*, 19, R995–R1008. <https://doi.org/10.1016/j.cub.2009.08.023>
- Chouinard-Thuly, L., Reddon, A. R., Leris, I., Earley, R. L., & Reader, S. M. (2018). Developmental plasticity of the stress response in female but not in male guppies. *Royal Society Open Science*, 5, 172268. <https://doi.org/10.1098/rsos.172268>
- DePasquale, C., Neuberger, T., Hirrlinger, A. M., & Braithwaite, V. A. (2016). The influence of complex and threatening environments in

- early life on brain size and behaviour. *Proceedings of the Royal Society B: Biological Sciences*, 283, 20152564. <https://doi.org/10.1098/rspb.2015.2564>
- Diamond, M. C., Law, F., Rhodes, H., Lindner, B., Rosenzweig, M. R., Krech, D., & Bennett, E. L. (1966). Increases in cortical depth and glia numbers in rats subjected to enriched environment. *Journal of Comparative Neurology*, 128, 117–125. <https://doi.org/10.1002/cne.901280110>
- Dunbar, R. I. M., & Shultz, S. (2017). Why are there so many explanations for primate brain evolution? *Philosophical Transactions of the Royal Society B: Biological Sciences*, 372, 20160244. <https://doi.org/10.1098/rstb.2016.0244>
- Edmunds, M. (1974). *Defence in animals*. New York, NY: Longman Inc.
- Elfert, C., Farnworth, M., Schulz-Mirbach, T., Riesch, R., Bierbach, D., Klaus, S., ... Plath, M. (2015). Brain size variation in extremophile fish: Local adaptation versus phenotypic plasticity. *Journal of Zoology*, 295, 143–153. <https://doi.org/10.1111/jzo.12190>
- Elvidge, C. K., Ramnarine, I., & Brown, G. E. (2014). Compensatory foraging in Trinidadian guppies: Effects of acute and chronic predation threats. *Current Zoology*, 60, 323–332. <https://doi.org/10.1093/czoolo/60.3.323>
- Finlay, B., & Darlington, R. (1995). Linked regularities in the development and evolution of mammalian brains. *Science*, 268, 1578–1584. <https://doi.org/10.1126/science.7777856>
- Gonda, A., Herczeg, G., & Merilä, J. (2011). Population variation in brain size of nine-spined sticklebacks (*Pungitius pungitius*) – Local adaptation or environmentally induced variation? *BMC Evolutionary Biology*, 11, 75. <https://doi.org/10.1186/1471-2148-11-75>
- Gonda, A., Herczeg, G., & Merilä, J. (2013). Evolutionary ecology of intraspecific brain size variation: A review. *Ecology & Evolution*, 3, 2751–2764. <https://doi.org/10.1002/ece3.627>
- Gonda, A., Valimaki, K., Herczeg, G., & Merila, J. (2012). Brain development and predation: Plastic responses depend on evolutionary history. *Biology Letters*, 8, 249–252. <https://doi.org/10.1098/rsbl.2011.0837>
- Gotanda, K. M., Delaire, L. C., Raeymaekers, J. A. M., Pérez-Jvostov, F., Dargent, F., Bentzen, P., ... Hendry, A. P. (2013). Adding parasites to the guppy-predation story: Insights from field surveys. *Oecologia*, 172, 155–166. <https://doi.org/10.1007/s00442-012-2485-7>
- Harris, R. M., O'Connell, L. A., & Hofmann, H. A. (2016). Brain evolution, development, and plasticity. In S. V. Shepherd (Ed.), *The Wiley handbook of evolutionary neuroscience* (pp. 422–443). Chichester, UK: Wiley.
- Healy, S. D., & Rowe, C. (2007). A critique of comparative studies of brain size. *Proceedings of the Royal Society B: Biological Sciences*, 274, 453–464. <https://doi.org/10.1098/rspb.2006.3748>
- Heisenberg, M., Heusipp, M., & Wanke, C. (1995). Structural plasticity in the Drosophila brain. *Journal of Neuroscience*, 15, 1951–1960. <https://doi.org/10.1523/JNEUROSCI.15-03-01951.1995>
- Houde, A. E. (1997). *Sex, color, and mate choice in guppies*. Princeton, NJ: Princeton University Press.
- Ioannou, C. C., Guttal, V., & Couzin, I. D. (2012). Predatory fish select for coordinated collective motion in virtual prey. *Science*, 337, 1212–1215. <https://doi.org/10.1126/science.1218919>
- Ioannou, C. C., Ramnarine, I. W., & Torney, C. J. (2017). High-predation habitats affect the social dynamics of collective exploration in a shoaling fish. *Scientific Advances*, 3, 1–8. <https://doi.org/10.1126/sciadv.1602682>
- Iwaniuk, A. N., & Nelson, J. E. (2003). Developmental differences are correlated with relative brain size in birds: A comparative analysis. *Canadian Journal of Zoology*, 81, 1913–1928. <https://doi.org/10.1139/z03-190>
- Jerison, H. (1973). *Evolution of the brain and intelligence*. New York, NY: Academic Press.
- Kondoh, M. (2010). Linking learning adaptation to trophic interactions: A brain size-based approach. *Functional Ecology*, 24, 35–43. <https://doi.org/10.1111/j.1365-2435.2009.01631.x>
- Kotrschal, A., Buechel, S. D., Zala, S. M., Corral-Lopez, A., Penn, D. J., & Kolm, N. (2015). Brain size affects female but not male survival under predation threat. *Ecology Letters*, 18, 646–652. <https://doi.org/10.1111/ele.12441>
- Kotrschal, A., Corral-Lopez, A., Szidat, S., & Kolm, N. (2015). The effect of brain size evolution on feeding propensity, digestive efficiency, and juvenile growth. *Evolution*, 69, 3013–3020. <https://doi.org/10.1111/evo.12784>
- Kotrschal, A., Deacon, A. E., Magurran, A. E., & Kolm, N. (2017). Predation pressure shapes brain anatomy in the wild. *Evolutionary Ecology*, 31, 619–633. <https://doi.org/10.1007/s10682-017-9901-8>
- Kotrschal, A., Kolm, N., & Penn, D. J. (2016). Selection for brain size impairs innate, but not adaptive immune responses. *Proceedings of the Royal Society B: Biological Sciences*, 283, 20152857. <https://doi.org/10.1098/rspb.2015.2857>
- Kotrschal, A., Lievens, E. J., Dahlbom, J., Bundsen, A., Semenova, S., Sundvik, M., ... Kolm, N. (2014). Artificial selection on relative brain size reveals a positive genetic correlation between brain size and proactive personality in the guppy. *Evolution*, 68, 1139–1149. <https://doi.org/10.1111/evo.12341>
- Kotrschal, A., Rogell, B., Bundsen, A., Svensson, B., Zajitschek, S., Brännström, I., ... Kolm, N. (2013). Artificial selection on relative brain size in the guppy reveals costs and benefits of evolving a larger brain. *Current Biology*, 23, 168–171. <https://doi.org/10.1016/j.cub.2012.11.058>
- Kotrschal, A., Rogell, B., Maklakov, A. A., & Kolm, N. (2012). Sex-specific plasticity in brain morphology depends on social environment of the guppy, *Poecilia reticulata*. *Behavioral Ecology and Sociobiology*, 66, 1485–1492. <https://doi.org/10.1007/s00265-012-1403-7>
- Kotrschal, A., Zeng, H. L., van der Bijl, W., Öhman-Mägi, C., Kotrschal, K., Pelckmans, K., & Kolm, N. (2017). Evolution of brain region volumes during artificial selection for relative brain size. *Evolution*, 71, 2942–2951. <https://doi.org/10.1111/evo.13373>
- Krause, J., & Ruxton, G. D. (2002). *Living in groups*. Oxford, UK: Oxford University Press. <https://doi.org/10.1093/sysbio/sys022>
- Leris, I. (2016). *Early environment and the development of social behaviours in the Trinidadian guppy, Poecilia reticulata* (PhD thesis). Utrecht University, Utrecht, The Netherlands, pp. 1–169.
- Lima, S. L., & Dill, L. M. (1990). Behavioral decisions made under the risk of predation: A review and prospectus. *Canadian Journal of Zoology*, 68, 619–640. <https://doi.org/10.1139/z90-092>
- Logan, C. J., Avin, S., Boogert, N., Buskell, A., Cross, F. R., Currie, A., ... Montgomery, S. H. (2018). Beyond brain size: Uncovering the neural correlates of behavioral and cognitive specialization. *Comparative Cognition & Behavior Reviews*, 13, 55–89. <https://doi.org/10.3819/CCBR.2018.130008>
- MacLean, E. L., Hare, B., Nunn, C. L., Addessi, E., Amici, F., Anderson, R. C., ... Zhao, Y. (2014). The evolution of self-control. *Proceedings of the National Academy of Sciences*, 111, E2140–E2148. <https://doi.org/10.1073/pnas.1323533111>
- Magurran, A. E. (1998). Population differentiation without speciation. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 353, 275–286. <https://doi.org/10.1098/rstb.1998.0209>
- Magurran, A. E. (2005). *Evolutionary ecology: The Trinidadian guppy*. London: Oxford University Press. <https://doi.org/10.1093/acprof:oso/9780198527855.001.0001>
- Matthews, M., & Varga, Z. M. (2012). Anesthesia and euthanasia in zebrafish. *ILAR Journal*, 53, 192–204. <https://doi.org/10.1093/ilar.53.2.192>
- Møller, A. P., & Erritzøe, J. (2014). Predator-prey interactions, flight initiation distance and brain size. *Journal of Evolutionary Biology*, 27, 34–42. <https://doi.org/10.1111/jeb.12272>
- Öst, M., & Jaatinen, K. (2015). Smart and safe? Antipredator behavior and breeding success are related to head size in a wild bird. *Behavioural Ecology*, 26, 1371–1378. <https://doi.org/10.1093/beheco/arv093>

- R Core Development Team. (2016). *R: A language and environment for statistical computing*, 3.3.0. Vienna, Austria: R Foundation for Statistical Computing. Retrieved from <http://www.R-project.org/>.
- Reddon, A. R., Chouinard-Thuly, L., Leris, I., & Reader, S. M. (2018). Data from: Wild and laboratory exposure to cues of predation risk increase relative brain mass in male guppies. *Dryad Digital Repository*, <https://doi.org/10.5061/dryad.pq1js52>
- Reznick, D., Callahan, H., & Llauredo, R. (1996). Maternal effects on offspring quality in poeciliid fishes. *American Zoologist*, 36, 147. <https://doi.org/10.1093/icb/36.2.147>
- Reznick, D., & Endler, J. A. (1982). The impact of predation on life history evolution in Trinidadian guppies (*Poecilia reticulata*). *Evolution*, 36, 160–177. <https://doi.org/10.2307/2408156>
- Rosenzweig, M. R., & Bennett, E. L. (1969). Effects of differential environments on brain weights and enzyme activities in gerbils, rats, and mice. *Developmental Psychobiology*, 2, 87–95. <https://doi.org/10.1002/dev.420020208>
- Roth, T. C., LaDage, L. D., & Pravosudov, V. V. (2010). Learning capabilities enhanced in harsh environments: A common garden approach. *Proceedings of the Royal Society B: Biological Sciences*, 277, 3187–3193. <https://doi.org/10.1098/rspb.2010.0630>
- Roth, T. C., & Pravosudov, V. V. (2009). Hippocampal volumes and neuron numbers increase along a gradient of environmental harshness: A large-scale comparison. *Proceedings of the Royal Society B: Biological Sciences*, 276, 401–405. <https://doi.org/10.1098/rspb.2008.1184>
- Samuk, K., Xue, J., & Rennison, D. J. (2018). Exposure to predators does not lead to the evolution of larger brains in experimental populations of threespine stickleback. *Evolution*, 511, 307–314. <https://doi.org/10.1111/evo.13444>
- Schwartz, A. K., & Hendry, A. P. (2010). Testing the influence of local forest canopy clearing on phenotypic variation in Trinidadian guppies. *Functional Ecology*, 24, 354–364. <https://doi.org/10.1111/j.1365-2435.2009.01652.x>
- Sherry, D. F. (2006). Neuroecology. *Annual Reviews in Psychology*, 57, 167–197. <https://doi.org/10.1146/annurev.psych.56.091103.070324>
- Shultz, S., & Dunbar, R. I. (2006). Chimpanzee and felid diet composition is influenced by prey brain size. *Biology Letters*, 2, 505–508. <https://doi.org/10.1098/rsbl.2006.0519>
- Sol, D., Szekely, T., Liker, A., & Lefebvre, L. (2007). Big-brained birds survive better in nature. *Proceedings of the Royal Society B: Biological Sciences*, 274, 763–769. <https://doi.org/10.1098/rspb.2006.3765>
- Striedter, G. F. (2005). *Principles of brain evolution*. Sunderland: Sinauer Associates.
- Walsh, M. R., Broyles, W., Beston, S. M., & Munch, S. B. (2016). Predator-driven brain size evolution in natural populations of Trinidadian killifish (*Rivulus hartii*). *Proceedings of the Royal Society B: Biological Sciences*, 283, 20161075. <https://doi.org/10.1098/rspb.2016.1075>
- Wickham, H. (2009). *ggplot2: Elegant graphics for data analysis*. New York, NY: Springer.
- de Winter, W., & Oxnard, C. E. (2001). Evolutionary radiations and convergences in the structural organization of mammalian brains. *Nature*, 409, 710–714. <https://doi.org/10.1038/35055547>

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