Supplementary Text S5: On overclustering correction

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1 Overview

Here, we will verify whether the *dispersal* term of the orthogroup scores really penalizes overclustering. For that, we ran OrthoFinder (Emms and Kelly 2019) one more time using the previously described Brassicaceae data set, but now with an Markov inflation parameter (mcl) of 5. An mcl of 5 is usually considered too large, so we would expect orthogroup scores to be lower than, for instance, runs with mcl = 3. Our goal here is to verify if our hypothesis is true.

Loading required packages:

```
set.seed(123) # for reproducibility

library(here)
library(cogeqc)
library(ggpubr)
library(rstatix)
library(patchwork)
library(tidyverse)

source(here("code", "utils.R"))
```

2 Data acquisition

We ran OrthoFinder with the following code:

```
# Run OrthoFinder - default DIAMOND mode, mcl = 5
orthofinder -f data -S diamond -I 5 -o products/result_files/default_5 -og
```

Now, we will load our data to the R session as a list of \mathbf{cogeqc} -friendly orthogroup data frames.

```
# Extract tar.xz file
tarfile <- here("products", "result_files", "Orthogroups.tar.xz")
outdir <- tempdir()

system2("tar", args = c("-xf", tarfile, "--directory", outdir))

# Get path to OrthoFinder output
og_files <- list.files(
    path = outdir,
    pattern = "Orthogroups.*", full.names = TRUE
)

# Remove files for the ultrasensitive DIAMOND mode and add mcl=5
og_files <- c(
    og_files[c(2, 1, 3, 4)],
    here("products", "result_files", "Orthogroups_default_5.tsv.gz")
)</pre>
```

```
# Read and parse files
ogs <- lapply(og_files, function(x) {</pre>
    og <- read_orthogroups(x)</pre>
    og <- og %>%
        mutate(Species = stringr::str_replace_all(Species, "\\.", "")) %>%
        mutate(Gene = str_replace_all(
             Gene, c(
                 " \setminus \ [0-9] $" = "",
                 "\\.[0-9]\\.p$" = "",
                 "\t.t[0-9]$" = "",
                 "\\.q$" = ""
             )
        ))
    return(og)
})
names(ogs) <- c("1", "1.5", "2", "3", "5")
```

Next, we will load InterPro annotation from PLAZA 5.0 (Van Bel et al. 2022).

```
# Define function to read functional annotation from PLAZA 5.0
read_annotation <- function(url, cols = c(1, 3)) {</pre>
    annot <- readr::read_tsv(url, show_col_types = FALSE, skip = 8) %>%
        select(cols)
    names(annot)[1:2] <- c("Gene", "Annotation")</pre>
    return(annot)
}
# Get Interpro annotation
base <- "https://ftp.psb.ugent.be/pub/plaza/plaza_public_dicots_05/InterPro/"</pre>
interpro <- list(</pre>
    Athaliana = read_annotation(paste0(base, "interpro.ath.csv.gz")),
    Aarabicum = read_annotation(paste0(base, "interpro.aar.csv.gz")),
    Alyrata_cvMN47 = read_annotation(paste0(base, "interpro.aly.csv.gz")),
    Bcarinata_cvzd1 = read_annotation(paste0(base, "interpro.bca.csv.gz")),
    Crubella_cvMonteGargano = read_annotation(paste0(base, "interpro.cru.csv.gz")),
    Chirsuta = read_annotation(pasteO(base, "interpro.chi.csv.gz")),
    Sparvula = read_annotation(paste0(base, "interpro.spa.csv.gz"))
interpro <- lapply(interpro, as.data.frame)</pre>
```

3 Validating the overclustering correction

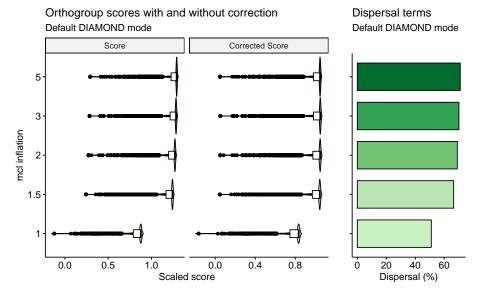
Now that we have all data we need (orthogroup data frames and domain annotations), let's calculate orthogroup scores. Here, we will use the function calculate_H_with_terms() from the file *utils.R*, which contains a slightly modified version of the function calculate_H() from **cogeqc**, but instead of updating the uncorrected scores with the corrected scores, it returns the dispersal terms and corrected scores and separate variables.

```
# Calculate orthogroup scores with and without correction for overclustering
og_homogeneity <- Reduce(rbind, lapply(seq_along(ogs), function(x) {</pre>
    mode <- names(ogs)[x]</pre>
    annotation <- Reduce(rbind, interpro) |> distinct()
    message("Working on ", mode)
    orthogroup_df <- merge(</pre>
        ogs[[x]],
        annotation,
        all.x = TRUE
    scores_df <- calculate_H_with_terms(</pre>
        orthogroup_df, correct_overclustering = TRUE, update_score = FALSE
    scores_df$Mode <- mode</pre>
    return(scores_df)
}))
og_homogeneity$Mode <- factor(
    og_homogeneity$Mode, levels = unique(og_homogeneity$Mode)
```

Next, let's visualize orthogroup scores with and without corrections, as well as look at the dispersal terms for each mode.

```
# Plot scores
p_scores <- og_homogeneity |>
   mutate(
       Score = (Score - min(Score) / (max(Score) - min(Score))),
       Score_c = (Score_c - min(Score_c) / (max(Score_c) - min(Score_c)))
   dplyr::select(Orthogroup, Score, `Corrected Score` = Score_c, Mode) |>
   pivot_longer(
        !c("Orthogroup", "Mode"),
       names_to = "Measure",
       values_to = "Score"
   ) |>
   mutate(
       Measure = factor(Measure, levels = c("Score", "Corrected Score"))
   ) |>
   ggpubr::ggviolin(
       x = "Mode", y = "Score",
       orientation = "horiz",
       fill = "Mode",
       palette = rev(c("#006D2C", "#31A354", "#74C476", "#BAE4B3", "#c7f2bf")),
       add = "boxplot", add.params = list(fill = "white")
   ) +
```

```
labs(
        x = "mcl inflation", y = "Scaled score",
        title = "Orthogroup scores with and without correction",
        subtitle = "Default DIAMOND mode"
    ) +
    facet_wrap(~Measure, scales = "free_x", nrow = 1) +
    theme(legend.position = "none")
# Plot dispersal terms
p_dispersal <- og_homogeneity |>
    dplyr::select(Mode, Dispersal) |>
    mutate(Dispersal = Dispersal * 100) |>
    dplyr::distinct() |>
    ggpubr::ggbarplot(
        x = "Mode", y = "Dispersal", stat = "identity",
        orientation = "horiz",
        fill = "Mode",
        palette = rev(c("#006D2C", "#31A354", "#74C476", "#BAE4B3", "#c7f2bf")),
    ) +
    labs(
        x = "", y = "Dispersal (%)",
        title = "Dispersal terms",
        subtitle = "Default DIAMOND mode"
    ) +
    theme(
        legend.position = "none",
        axis.text.y = element_blank()
# Combine plots
p_combined <- patchwork::wrap_plots(p_scores, p_dispersal, widths = c(2.5, 1))</pre>
p_combined
```



We can see that, without correction (homogeneity only), increasing the value for the *mcl* parameter leads to increasingly larger scores. However, as homogeneity increases, the dispersal also increases. After correcting for dispersal, larger values for the *mcl* parameter do not lead to higher orthogroup scores.

To verify that formally, let's perform a Mann-Whitney U test for differences in orthogroup scores for runs with mcl of 3 and 5 with and without correcting for dispersal.

As expected, without correcting for dispersal, using mcl = 5 leads to better orthogroup scores than using mcl = 3. However, after correction, orthogroup scores for mcl = 5 are worse than scores for mcl = 3, which is desired.

Session info

This document was created under the following conditions:

```
## - Session info -----
## setting value
## version R version 4.3.0 (2023-04-21)
## os Ubuntu 20.04.5 LTS
## system x86_64, linux-gnu
## ui X11
## language (EN)
## collate en_US.UTF-8
## ctype en_US.UTF-8
        Europe/Brussels
## tz
         2023-10-06
## date
## pandoc 3.1.1 @ /usr/lib/rstudio/resources/app/bin/quarto/bin/tools/ (via rmarkdown)
## package * version date (UTC) lib source
## abind
              1.4-5 2016-07-21 [1] CRAN (R 4.3.0)
## ape
                5.7-1 2023-03-13 [1] CRAN (R 4.3.0)
               0.1.10 2023-03-08 [1] CRAN (R 4.3.0)
1.4.1 2021-12-13 [1] CRAN (R 4.3.0)
## aplot
## backports
```

```
0.4.0
                                 2021-06-01 [1] CRAN (R 4.3.0)
    beeswarm
## BiocGenerics
                     0.46.0
                                 2023-04-25 [1] Bioconductor
                      1.30.21.1 2023-07-18 [1] CRAN (R 4.3.0)
## BiocManager
## BiocStyle
                   * 2.29.1 2023-08-04 [1] Github (Bioconductor/BiocStyle@7c0e093)
## Biostrings
                     2.68.0 2023-04-25 [1] Bioconductor
                     1.0-7 2021-04-24 [1] CRAN (R 4.3.0)
0.34 2023-05-09 [1] CRAN (R 4.3.0)
## bitops
##
    bookdown
## broom
                     1.0.4 2023-03-11 [1] CRAN (R 4.3.0)
## car
                     3.1-2 2023-03-30 [1] CRAN (R 4.3.0)
                     3.0-5 2022-01-06 [1] CRAN (R 4.3.0)
3.6.1 2023-03-23 [1] CRAN (R 4.3.0)
## carData
##
   cli
## codetools
                     0.2-19 2023-02-01 [4] CRAN (R 4.2.2)
## cogeqc
                    * 1.4.0 2023-04-25 [1] Bioconductor
                     1.4-2 2021-10-08 [1] CRAN (R 4.3.0)
2.1-0 2023-01-23 [1] CRAN (R 4.3.0)
##
    coin
##
    colorspace
                     1.5.2 2022-09-29 [1] CRAN (R 4.3.0)
## crayon
                     0.6.33 2023-07-07 [1] CRAN (R 4.3.0)
## digest
                   * 1.1.2 2023-04-20 [1] CRAN (R 4.3.0)
0.21 2023-05-05 [1] CRAN (R 4.3.0)
## dplyr
## evaluate
                     1.0.4 2023-01-22 [1] CRAN (R 4.3.0)
## fansi
                     2.1.1 2022-07-06 [1] CRAN (R 4.3.0)
1.1.1 2023-02-24 [1] CRAN (R 4.3.0)
## farver
## fastmap
                    * 1.0.0 2023-01-29 [1] CRAN (R 4.3.0)
## forcats
## generics
                     0.1.3 2022-07-05 [1] CRAN (R 4.3.0)
## GenomeInfoDb
                     1.36.0 2023-04-25 [1] Bioconductor
## GenomeInfoDbData 1.2.10 2023-04-28 [1] Bioconductor
## ggbeeswarm 0.7.2 2023-04-29 [1] CRAN (R 4.3.0)
## ggfun
                     0.0.9 2022-11-21 [1] CRAN (R 4.3.0)
                     * 3.4.1 2023-02-10 [1] CRAN (R 4.3.0)
0.1.0 2021-09-02 [1] CRAN (R 4.3.0)
## ggplot2
   ggplotify
## ggpubr
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                     0.6.4 2022-10-13 [1] CRAN (R 4.3.0)
## ggsignif
                     3.8.0 2023-04-25 [1] Bioconductor
1.6.2 2022-02-24 [1] CRAN (R 4.3.0)
##
   ggtree
## glue
## gridGraphics 0.5-1 2020-12-13 [1] CRAN (R 4.3.0)
                    0.3.3 2023-03-21 [1] CRAN (R 4.3.0)

* 1.0.1 2020-12-13 [1] CRAN (R 4.3.0)

1.1.3 2023-03-21 [1] CRAN (R 4.3.0)
## gtable
## here
## hms
## htmltools
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                     1.4.2 2023-04-07 [1] CRAN (R 4.3.0)
## igraph
                     2.34.0 2023-04-25 [1] Bioconductor
## IRanges
## jsonlite
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## knitr
                     1.43 2023-05-25 [1] CRAN (R 4.3.0)
                     0.4.2 2020-10-20 [1] CRAN (R 4.3.0)
## labeling
                     0.20-45 2021-09-22 [4] CRAN (R 4.2.0)
## lattice
## lazyeval
                     0.2.2
                                 2019-03-15 [1] CRAN (R 4.3.0)
## libcoin
                                 2021-09-27 [1] CRAN (R 4.3.0)
                      1.0-9
## lifecycle
                      1.0.3
                                 2022-10-07 [1] CRAN (R 4.3.0)
## lubridate
                  * 1.9.2 2023-02-10 [1] CRAN (R 4.3.0)
## magrittr
                      2.0.3 2022-03-30 [1] CRAN (R 4.3.0)
## MASS
                       7.3-58.2 2023-01-23 [4] CRAN (R 4.2.2)
```

```
## Matrix
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## matrixStats
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                      0.2-23
                                 2020-03-05 [1] CRAN (R 4.3.0)
## modeltools
## multcomp
                     1.4-25 2023-06-20 [1] CRAN (R 4.3.0)
## munsell
                     0.5.0 2018-06-12 [1] CRAN (R 4.3.0)
                     1.1-3 2021-10-08 [1] CRAN (R 4.3.0)
## mvtnorm
                     3.1-162 2023-01-31 [4] CRAN (R 4.2.2)
##
    nlme
##
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    patchwork
    pillar
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                     2.0.3 2019-09-22 [1] CRAN (R 4.3.0)
1.8.8 2022-11-11 [1] CRAN (R 4.3.0)
## pkgconfig
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##
    purrr
## R6
                     2.5.1 2021-08-19 [1] CRAN (R 4.3.0)
                     1.0.10
## Rcpp
                                 2023-01-22 [1] CRAN (R 4.3.0)
## RCurl
                     1.98-1.12 2023-03-27 [1] CRAN (R 4.3.0)
                   * 2.1.4 2023-02-10 [1] CRAN (R 4.3.0)
## readr
## reshape2
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                     1.1.1 2023-04-28 [1] CRAN (R 4.3.0)
2.23 2023-07-01 [1] CRAN (R 4.3.0)
2.0.3 2022-04-02 [1] CRAN (R 4.3.0)
## rlang
## rmarkdown
## rprojroot
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0.14 2022-08-22 [1] CRAN (R 4.3.0)
## rstatix
## rstudioapi
## S4Vectors
                     0.38.0 2023-04-25 [1] Bioconductor
## sandwich
                     3.0-2 2022-06-15 [1] CRAN (R 4.3.0)
                     1.2.1 2022-08-20 [1] CRAN (R 4.3.0)
1.2.2 2021-12-06 [1] CRAN (R 4.3.0)
## scales
## sessioninfo
## stringi
                     1.7.12 2023-01-11 [1] CRAN (R 4.3.0)
## stringr
                  * 1.5.0 2022-12-02 [1] CRAN (R 4.3.0)
                     3.5-3 2023-02-12 [4] CRAN (R 4.2.2)
1.1-2 2023-04-17 [1] CRAN (R 4.3.0)
## survival
   TH.data
## tibble
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                   * 1.3.0 2023-01-24 [1] CRAN (R 4.3.0)
## tidyr
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0.4.2 2022-12-18 [1] CRAN (R 4.3.0)
## tidyselect
## tidytree
## tidyverse
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## timechange
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## utf8
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                     0.6.3 2023-06-14 [1] CRAN (R 4.3.0)
0.4.5 2017-03-22 [1] CRAN (R 4.3.0)
## vctrs
## vipor
## withr
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## xfun
                     0.39 2023-04-20 [1] CRAN (R 4.3.0)
                   0.40.0 2023-04-25 [1] Bioconductor
## XVector
                      2.3.7
## yaml
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## yulab.utils
                    0.0.6 2022-12-20 [1] CRAN (R 4.3.0)
                     1.46.0 2023-04-25 [1] Bioconductor
## zlibbioc
                      1.8-12
## ZOO
                                2023-04-13 [1] CRAN (R 4.3.0)
##
## [1] /home/faalm/R/x86_64-pc-linux-gnu-library/4.3
## [2] /usr/local/lib/R/site-library
```

```
## [3] /usr/lib/R/site-library
## [4] /usr/lib/R/library
##
##
```

References

Emms, David M, and Steven Kelly. 2019. "OrthoFinder: Phylogenetic Orthology Inference for Comparative Genomics." *Genome Biology* 20 (1): 1–14.

Van Bel, Michiel, Francesca Silvestri, Eric M Weitz, Lukasz Kreft, Alexander Botzki, Frederik Coppens, and Klaas Vandepoele. 2022. "PLAZA 5.0: Extending the Scope and Power of Comparative and Functional Genomics in Plants." *Nucleic Acids Research* 50 (D1): D1468–74.